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Phytochemical changes in milkweed induced by elevated CO₂ alter wing morphology but not toxin sequestration in monarch butterflies

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42 **Abstract:**

- 43 1. Environmental change has the potential to influence trophic interactions by altering the
44 defensive phenotype of prey.
- 45 2. Here, we examine the effects of a pervasive environmental change driver, elevated
46 atmospheric concentrations of CO₂ (eCO₂), on toxin sequestration and flight morphology
47 of a specialist herbivore.
- 48 3. We fed monarch butterfly larvae, *Danaus plexippus*, foliage from four milkweed,
49 *Asclepias*, species of varying chemical defense profiles grown under either ambient or
50 eCO₂. We also infected a subset of these herbivores with a protozoan parasite,
51 *Ophryocystis elektroscirrha*, to understand how infection and environmental change
52 combine to alter herbivore defenses. We measured changes in phytochemistry induced by
53 eCO₂ and assessed cardenolide, toxic steroid, sequestration and wing morphology of
54 butterflies.
- 55 4. Monarchs compensated for lower plant cardenolide concentrations under eCO₂ by
56 increasing cardenolide sequestration rate, maintaining similar cardenolide composition
57 and concentrations in their wings under both CO₂ treatments. We suggest that these

- 58 increases in sequestration rate are a byproduct of compensatory feeding aimed at
59 maintaining a nutritional target in response to declining dietary quality.
- 60 5. Monarch wings were more suitable for sustained flight (more elongated) when reared on
61 plants grown under eCO₂ or when reared on *A. syriaca* or *A. incarnata* rather than on *A.*
62 *curassavica* or *A. speciosa*. Parasite infection engendered wings less suitable for
63 sustained flight (wings became rounder) on three of four milkweed species. Wing loading
64 (associated with powered flight) was higher on *A. syriaca* than on other milkweeds,
65 whereas wing density was lower on *A. curassavica*. Monarchs that fed on high
66 cardenolide milkweed developed rounder, thinner wings, which are less efficient at
67 gliding flight.
- 68 6. Ingesting foliage from milkweed high in cardenolides may provide protection from
69 enemies through sequestration yet come at a cost to monarchs manifested as lower
70 quality flight phenotypes: rounder, thinner wings with lower wing loading values.
- 71 7. Small changes in morphology may have important consequences for enemy evasion and
72 migration success in many animals. Energetic costs associated with alterations in defense
73 and morphology may, therefore, have important consequences for trophic interactions in
74 a changing world.

75

76 **Key-words:** *Asclepias*, cardenolides, environmental change, elevated CO₂, *Danaus plexippus*,
77 monarch butterfly, predator-prey, plant secondary metabolites.

78 **Introduction**

79 Environmental change influences trophic interactions through multiple mechanisms (Gilman,
80 Urban, Tewksbury, Gilchrist, & Holt, 2010; Tylianakis, Didham, Bascompte, & Wardle, 2008).
81 For example, elevated atmospheric carbon dioxide (eCO₂) has both direct and indirect effects on
82 organismal physiology, and behavior. Plants grown under eCO₂ produce tissues with lower
83 nitrogen concentrations (Drake, Gonzalez-Meler, & Long, 1997; Robinson, Ryan, & Newman,
84 2012), causing herbivores to increase the amount of foliage they consume (Docherty et al., 1996;
85 Johnson, Lopaticki, & Hartley, 2014). Moreover, eCO₂ also changes the composition and
86 concentration of plant secondary metabolites (PSMs) (Klaiber, Dorn, & Najar-Rodriguez, 2013;
87 Ryan, Rasmussen, & Newman, 2010). Because catabolizing PSMs is energetically costly,
88 changes in these compounds affect the ecology of herbivores (Hunter, 2016). In general,

89 herbivore growth, fecundity, and survival decline under eCO₂ (Jamieson et al., 2017; Robinson
90 et al., 2012). Higher trophic levels are also affected by CO₂-induced shifts in plant quality
91 (Facey, Ellsworth, Staley, Wright, & Johnson, 2014; Ode & Crompton, 2013; Ode, Johnson, &
92 Moore, 2014), often mediated by shifts in prey nutrition and growth (Chen, Ge, & Parajulee,
93 2005; Klaiber, Najar-Rodriguez, Dialer, & Dorn, 2013). For example, eCO₂ reduces alfalfa
94 nutritional quality, which increases armyworm larval development times, resulting in asynchrony
95 between larvae and parasitoid wasps, ultimately reducing parasitoid fitness (Dyer *et al.* 2013).
96 Elevated CO₂ may also benefit higher trophic levels by inhibiting herbivore defense and escape
97 (Hentley, Vanbergen, Hails, Jones, & Johnson, 2014).

98
99 Many specialist herbivores have evolved mechanisms to co-opt PSMs for their own defense
100 (Dyer & Deane Bowers, 1996; Opitz & Müller, 2009; Petschenka & Agrawal, 2016).
101 Sequestration by insect herbivores involves the modification, transfer, and storage of toxic
102 compounds at high metabolic costs, potentially reducing immune responses to other enemies
103 (Greeney, Dyer, & Smilanich, 2012; Smilanich, Dyer, Chambers, & Bowers, 2009). Monarch
104 butterflies, *Danaus plexippus*, store toxic steroids (cardenolides) derived from the foliage of their
105 milkweed, *Asclepias*, host plants (Reichstein, Euw, Parsons, & Rothschild, 1968). Cardenolides
106 disrupt the function of Na⁺/K⁺-channels in animal cells (Agrawal, Petschenka, Bingham, Weber,
107 & Rasmann, 2012) and render monarchs well-defended prey (Malcolm & Brower, 1989). The
108 concentration and composition of cardenolides sequestered by monarchs depends on milkweed
109 species, the amount of tissue consumed, and sequestration efficiency (Agrawal, Ali, Rasmann, &
110 Fishbein, 2015; Bowers & Collinge, 1992; Camara, 1997; Malcolm, 1990, 1994). Environmental
111 factors that alter phytochemistry and consumption rates could also influence sequestration and
112 affect vulnerability to enemies. Despite a growing body of work illustrating the effects of
113 environmental change on milkweed chemistry (Andrews, 2015; Tao, Berns, & Hunter, 2014;
114 Vannette & Hunter, 2011), little is known about how monarch sequestration will respond to
115 future environmental conditions (*but see* Tao & Hunter, 2015).

116
117 In addition to sequestration, the seasonal migration of monarchs may also reduce mortality
118 imposed by natural enemies. In eastern North America, monarchs migrate up to 4,500 km from
119 their summer breeding grounds to overwintering sites in Mexico every fall (Urquhart & Urquhart

120 1978; Brower & Malcolm 1991; Flockhart *et al.* 2017). For many flying animals, foraging,
121 escape, and migration are strongly influenced by wing size, shape, and wing loading; the ratio
122 between body mass and wing area (Berwaerts, Van Dyck, & Aerts, 2002; Robert Dudley, 2002).
123 Subtle changes in wing size and shape can affect drag, lift, and flight behavior (Srygley &
124 Thomas, 2002). To maximize energy use efficiency, flying animals both glide and actively
125 propel (Kovac, Vogt, Ithier, Smith, & Wood, 2012; Park, Bae, Lee, Jeon, & Choi, 2010). Larger,
126 more elongated wing shapes, with high wing length to width ratio (aspect ratio), allow for
127 optimal gliding flight (Kerlinger, 1989). Migratory eastern N. American monarchs have larger
128 and more elongated wings (higher aspect ratios) than non-migratory conspecifics (Altizer &
129 Davis, 2010; Li, Pierce, & de Roode, 2016). Migratory monarchs also have higher wing loading
130 values, correlated with larger energy reserves for stronger powered flight (Dudley & Srygley,
131 2008). However, despite an extensive body of literature detailing the importance of dietary
132 chemistry for insect fitness (Awmack & Leather, 2002), few studies have explored the effects of
133 diet on wing morphology and flight ability (Boggs & Freeman, 2005; Johnson, Solensky,
134 Satterfield, & Davis, 2014; Pellegrons, Van Dongen, Van Dyck, & Lens, 2009). Food restriction
135 reduces monarch wing size (Johnson et al., 2014), but no study to date has examined the effects
136 of changing phytochemistry on monarch wing morphology.

137
138 Natural enemies may also influence toxin sequestration and morphology of herbivores. While
139 sequestration can reduce insect immunity to parasites (*reviewed in* Greeney *et al.* 2012), it is
140 unclear how parasite infection alters sequestration. Throughout their range, monarchs suffer
141 infection by a debilitating, protozoan parasite, *Ophryocystis elektroscirrha*, that reduces monarch
142 lifespan, fecundity, and flight ability (Bradley & Altizer, 2005; de Roode, Chi, Rarick, & Altizer,
143 2009). Heavily infected butterflies are more likely to die during migration, termed “migratory
144 culling” (Altizer, Hobson, Davis, De Roode, & Wassenaar, 2015). Thus, monarchs that do arrive
145 at overwintering grounds support lower pathogen loads (Altizer, Bartel, & Han, 2011). For
146 moderately infected monarchs, there is no clear effect of infection on wing morphology (Bradley
147 & Altizer, 2005). *Ophryocystis elektroscirrha* likely depletes the energy reserves necessary for
148 flight (Altizer et al., 2015). However, additional stressors, such as reductions in diet quality
149 induced by eCO₂, may influence the impact of infection on flight ability.

150

151 Here, we investigate the effects of eCO₂ on the defensive phenotype of monarch butterflies. We
152 fed larvae foliage from four milkweed species with varying phytochemistry grown under either
153 ambient CO₂ (aCO₂) or eCO₂. We also infected a subset of monarchs with *O. elektroscirra*, to
154 understand how infection and environmental change combine to alter monarch defense,
155 including wing traits associated with migration. We measured changes in phytochemistry
156 induced by eCO₂ and assessed sequestration and morphology of butterflies. We expected
157 sequestration profiles to mirror changes in plant chemistry induced by eCO₂ because factors that
158 alter phytochemistry and consumption rates should also influence the types and amounts of
159 PSMs monarchs sequester. We also predicted that changes in cardenolides and reductions in the
160 nutritional quality of larval host-plants grown under eCO₂ (Decker, de Roode, & Hunter, 2018;
161 Robinson et al., 2012) would cause declines in the quality of the insect flight phenotype: smaller,
162 thinner and rounder wings with lower wing loading values. Feeding on lower quality food with
163 different types and amounts of cardenolides may engender a metabolic cost inflicting stress upon
164 the insect and inducing morphological changes. Finally, we hypothesized that the metabolic costs
165 of infection would exacerbate any deleterious effects of eCO₂ on cardenolide sequestration or
166 wing morphology.

167 **Materials and Methods**

168 ***Milkweed and Monarch Source Materials***

169 We analyzed the wings of monarchs reared on milkweeds grown under ambient (400 ppm) or
170 elevated (760 ppm) CO₂ at the University of Michigan Biological Station (UMBS). We provide
171 details of the UMBS CO₂ array in Decker *et al.* (2018). In 2015, we grew four milkweed species
172 in a 40 chamber array (Drake *et al.* 1989), with 20 chambers maintained at aCO₂ and 20 at eCO₂.
173 Within those chambers, we grew milkweed that varied in cardenolide concentrations, ranging
174 from high to low: *A. curassavica*, *A. syriaca*, *A. speciosa*, and *A. incarnata*. Seeds were planted
175 in the UMBS greenhouse and, 18 days later, seedlings were transferred outside into the chamber
176 array for the remainder of the experiment. Each chamber contained 24 milkweeds (4 species x 6
177 plants per species).

178
179 The monarchs used in this study were the F₁ offspring of seven full-sib crosses between monarch
180 lineages from eastern N. America (St Marks, FL). Monarch larvae were assigned to one of 16
181 treatments (2 parasite treatments x 4 host plant species x 2 levels of CO₂ x 20 replicate

182 chambers) making for 320 monarchs reared in total. Not all monarchs survived to adulthood and
183 some inoculated monarchs resisted infection, inflating the sample size of uninfected monarchs.
184 Final sample sizes varied for each species by CO₂ treatment between N=19-27 for uninfected
185 and N=5-11 for infected monarchs (Table S1 in the Supporting Information). Each individual
186 monarch larva was assigned three plants of the same treatment grown in the same chamber on
187 which to complete development.

188
189 Monarch larvae were inoculated with *O. elektroscirra* on cuttings from one of their three
190 assigned plants. After 42 days of growth in the array, we placed plant cuttings in individual
191 containers kept under aCO₂. A darkened monarch egg (darkening indicates eggs ready to hatch)
192 was attached to a leaf on each cutting to ensure that neonates consumed their assigned plant
193 before parasite inoculation. Three days after hatching, larvae were inoculated with *O.*
194 *elektroscirra* following the methods of de Roode *et al.* (2008). Parasites originated from one
195 lineage collected from an eastern N. American, wild-caught butterfly. Control larvae were fed
196 leaf disks of the same size with no spores. Foliar chemistry samples were taken from each plant
197 at the same time as inoculations (see below) and assumed to adequately reflect the chemistry of
198 the other two assigned plants the larvae consumed in later instars.

199
200 Larvae were fed cuttings from their assigned host-plants *ad libitum* until pupation. Adult
201 butterflies were sexed, weighed, and kept in 5.75 x 9.5 cm glassine envelopes at 15°C for the
202 remainder of their adult lives (de Roode, Gold, & Altizer, 2007). Three weeks after death, we
203 removed and stored the monarch wings at -20°C, and estimated parasite spore load from bodies
204 using established methods following de Roode *et al.* (2008). We scanned the left forewing of
205 each monarch on an HP scanJet 6300C (Hewlett-Packard, Palo Alto, USA), weighed the wing,
206 and stored it in 1mL centrifuge tubes for cardenolide analyses.

207 208 ***Cardenolide Chemical Analysis***

209 We quantified foliar cardenolide concentrations (Vannette & Hunter, 2011; Zehnder & Hunter,
210 2009) from the first milkweed that monarchs consumed, and in left forewings, because wing
211 cardenolides correlate tightly with body cardenolide concentrations and wings are typically the

212 first tissues tasted by bird predators upon attack (Fink & Brower 1981). We detail our chemical
213 analysis in Appendix S1 in the Supporting Information.

214

215 *Wing Morphometrics*

216 We measured forewing morphometrics because monarchs position forewings to cover their
217 hindwings during soaring flight (Altizer & Davis, 2010), allowing forewing size and shape to
218 have the largest influence on flight ability. Additionally, preliminary work established that
219 milkweed chemistry only affects forewing morphology (Berns, Zelditch, & Hunter, 2014).

220

221 We scanned the left forewing of each specimen with a ruler for scale and calibrated Adobe
222 Photoshop (Adobe, San Jose, USA) to calculate distance measures based on a pixel-to-millimeter
223 ratio. We took four basic measures of forewing morphology: first, length of the butterfly wing
224 from wing apex to thorax insertion (mm); second, width of the forewing at the longest axis
225 perpendicular to the length measurement (mm); third, total forewing area (mm²); and fourth,
226 wing perimeter (mm, Fig. S1 in the Supporting Information). For slightly damaged wings, we
227 estimated wing edges to create a complete outline. Butterflies with substantial wing damage were
228 discarded from all analyses.

229

230 We calculated two metrics of forewing shape: wing aspect ratio (length divided by width), and
231 roundness (area to perimeter ratio: $4\pi \text{ area}/\text{perimeter}^2$) (Altizer & Davis 2010). We also
232 calculated wing loading (body mass/wing area), a common aeronautical measure indicative of
233 maneuverability and flight performance. Finally, we examined butterfly wing density which we
234 termed specific wing area (wing area/wing mass).

235

236 *Analyses*

237 We used linear mixed models (LMMs; R version 3.3.2.; package: lme4) to assess effects of our
238 treatments on phytochemistry, toxin sequestration, and wing morphology. Chamber identity was
239 included as a random effect in all models (Littell, Stroup, & Freund, 2002; Vannette & Hunter,
240 2011). For models with monarch traits, we also included monarch lineage as a random effect
241 because it was not specifically manipulated and had uninformative factor levels. We transformed
242 all (dependent and independent) variables when necessary to achieve homogeneity of variance,

243 simplified models when possible, and visually inspected model residuals to confirm best fit
244 (Crawley, 2012). We present model structures, results, and random effects in Table S3 in the
245 Supporting Information.

246
247 To determine the effects of CO₂ and milkweed species on foliar cardenolide concentration, we
248 ran LMMs with foliar cardenolide concentration (square root transformed) as a response variable
249 and CO₂ treatment and milkweed species as fixed effects. Because there was a significant
250 interaction between milkweed species and CO₂, we analyzed each plant species separately to
251 determine which species drove the interaction. We used similar LMMs to test for effects of CO₂,
252 host-plant species and infection on monarch wing cardenolides (square-root-transformed).
253 Additionally, we tested whether CO₂ treatment or infection altered the relationship between
254 foliar and wing cardenolides by including these factors (CO₂ treatment and infection) as fixed
255 effects in an LMM with foliar cardenolide concentration (square root transformed) as an
256 independent variable and wing cardenolide concentration (square root transformed) as the
257 dependent variable. A significant interaction between foliar cardenolide concentration and either
258 of the factors (CO₂ treatment or infection) indicates a change in the slope of the relationship
259 between plant and butterfly cardenolides dependent on that factor.

260
261 Some monarchs exposed to *O. elektroscirra* resist infection and are spore-free as adults. We
262 compared the wing cardenolides of control monarchs (never exposed to the parasite) with those
263 of monarchs that were exposed to the parasite but had no spores. We ran an LMM with monarch
264 cardenolide concentration (square root transformed) as the dependent variable and parasite
265 exposure (control and exposed but uninfected) as a fixed effect. There were no significant
266 differences between resistant monarchs and control monarchs in the sequestration of total
267 cardenolides ($F_{1,190} = 0.90$, $p = 0.345$). Therefore, in all analyses of sequestration, we grouped
268 these two monarch treatments (control and exposed but uninfected) into one “uninfected” status.
269 We followed a similar procedure to determine whether or not monarch sex influenced
270 sequestration chemistry and found no effect of monarch sex on sequestered cardenolide
271 concentrations ($F_{1,250} = 0.24$, $p = 0.624$). Therefore, monarch sex was not included in models
272 that explored treatment effects on toxin sequestration.

273

274 We used permutational multivariate analysis of variance (PerMANOVA; Anderson, 2001) to
275 compare the effects of CO₂ treatment, milkweed species and, for butterfly cardenolides,
276 infection status on the assemblage of cardenolide compounds produced in milkweed and
277 sequestered by monarchs. For plant cardenolide composition, we performed a PerMANOVA
278 with CO₂ treatment, milkweed species and their interaction as independent variables, and Bray-
279 Curtis distance of percentage weight of each foliar cardenolide peak as dependent variables. We
280 chose Bray-Curtis as our ordination because it includes relative abundance information and
281 accounts for peak identity. To determine which factors altered wing cardenolide composition, we
282 performed a PerMANOVA with CO₂ treatment, milkweed species, infection status and their
283 interactions as independent variables, and the Bray-Curtis distance of percentage weight of each
284 sequestered wing cardenolide peak as dependent variables.

285
286 We followed Altizer & Davis (2010) and used Principal Component Analysis (PCA) to reduce
287 butterfly morphology measures into one PCA axis explaining forewing size (PCA-size) and
288 another PCA axis explaining forewing shape (PCA-shape). Forewing length, width, area and
289 perimeter were used to create the PCA-size axis that explained 99.6% of the total variance, while
290 forewing area and roundness were used to create the PCA-shape axis that explained 95.2% of the
291 total variance. Higher values of PCA-size represent larger wings, and higher values of PCA-
292 shape represent more elongated wings. We first ran LMMs similar to those used above for
293 sequestration results, to test whether there was a difference between control monarchs and those
294 that did not become infected in the size and shape of their wings. We ran two LMMs with either
295 PCA-shape or PCA-size as the dependent variables and parasite exposure (control or cleared) as
296 a fixed effect to examine the possibility that resisting infection could result in morphological
297 differences. There were no significant differences between resistant monarchs and control
298 monarchs in the size ($F_{1,166} = 0.35$, $p = 0.552$) and shape of wings ($F_{1,180} = 1.90$, $p = 0.170$). We
299 then ran models with these PCA axes as response variables, and CO₂ treatment, milkweed
300 species, infection and monarch sex as fixed effects. Due to sample size limitations (see Table
301 S1), we could not include the four-way interaction in any of our full models, but we did initially
302 include all other interaction terms between CO₂ treatment, milkweed species, infection, and
303 monarch sex.

304

305 To examine effects of the treatments on monarch wing loading and specific wing area, we ran
306 LMMs with wing loading (log-transformed) and specific wing area as response variables, with
307 identical fixed effects as above. We ran two LMMs with either wing loading or specific wing
308 area as response variables and CO₂ treatment, milkweed species, infection, and monarch sex as
309 fixed effects.

310

311 Finally, we tested whether cardenolide sequestration affected wing shape, loading, and density.
312 We ran three LMMs with either PCA-shape, wing loading or wing density (log-transformed) as
313 dependent variables, and monarch cardenolide concentration (square root transformed),
314 milkweed species and their interaction as fixed effects.

315 **Results**

316 *Milkweed Host-Plant Chemistry*

317 Of the 252 milkweeds that supported surviving monarchs (see Table S1), 114 produced
318 measurable cardenolides. To address the large occurrence of zeros in our chemistry dataset, we
319 followed methods detailed in Appendix S1. There was no effect of CO₂ treatment on the
320 proportion of plants that produced detectable foliar cardenolides (CO₂: $\chi^2 = 0.82$, $p = 0.366$).
321 *Asclepias incarnata* and *A. syriaca* were significantly more likely to express zero cardenolides
322 (species: $\chi^2 = 19.52$, $p = 0.0002$). The interaction term between CO₂ and species was not
323 significant (CO₂ * species: $\chi^2 = 1.49$, $p = 0.684$). We therefore chose to exclude the plants that
324 did not produce cardenolides from further analyses of phytochemistry.

325

326 The 114 plants with detectable cardenolides included 65 *A. curassavica*, 19 *A. syriaca*, 18 *A.*
327 *speciosa*, and 12 *A. incarnata*. Elevated CO₂ changed the concentration of foliar cardenolides in
328 a species-specific manner (species*CO₂: $F_{3,106} = 3.05$, $p = 0.032$, Fig. 1a). Under eCO₂ there was
329 a 52% decline in the foliar cardenolide concentrations of *A. curassavica* ($F_{1,36} = 13.43$, $p =$
330 0.0008 , Fig. 1a). Cardenolide concentrations in *A. syriaca* ($F_{1,13} = 1.0847$, $p = 0.32$), *A. speciosa*
331 ($F_{1,13} = 0.76$, $p = 0.399$) and *A. incarnata* ($F_{1,11} = 0.01$, $p = 0.910$) remained unaffected by
332 eCO₂. Across CO₂ treatments, *A. curassavica* produced the highest cardenolide concentrations,
333 while *A. incarnata* produced the lowest (species: $F_{3,106} = 71.72$, $p < 0.0001$, Fig. 1a). Milkweed
334 species varied in the assemblage of cardenolides that they produced (PERMANOVA, species:
335 $F_{3,110} = 24.16$, $R^2 = 0.39$, $p = 0.001$). In addition, the effect of CO₂ treatment on cardenolide

336 composition varied among milkweed species (PERMANOVA, CO₂*species: F_{3, 110} = 2.26, R² =
337 0.037, p = 0.004).

338

339 ***Monarch Wing Chemistry***

340 Monarchs maintained constant wing cardenolide concentrations between CO₂ treatments
341 (CO₂*species: F_{3,214}=1.60, p=0.191, Fig. 1b) despite the decline in foliar cardenolide
342 concentration in *A. curassavica* induced by eCO₂ (Fig. 1a). Critically, monarchs feeding on
343 milkweed foliage grown under eCO₂ sequestered more cardenolides per unit cardenolide
344 available in host-plants (plant cardenolides* CO₂: F_{1,109}=5.54, p=0.020, Fig. 1c). When feeding
345 on *A. syriaca*, monarchs infected with parasites sequestered 20% less cardenolide in their wings
346 than did uninfected monarchs (infection*species: F_{3,228}=2.71, p=0.0462, Fig. S2 in the
347 Supporting Information). CO₂ treatment, milkweed species and infection did not interact to
348 influence the concentration of cardenolides sequestered by monarchs (CO₂*species*infection: F
349 _{3,225} = 0.83, p = 0.4803).

350

351 Despite the interactive effects of CO₂ treatment and milkweed species on the composition of
352 foliar cardenolides, the cardenolide profiles sequestered by monarchs were influenced only by
353 milkweed species (PERMANOVA, species: F_{3, 247} = 157.00, R² = 0.65, p = 0.001). Neither
354 eCO₂ alone (PERMANOVA, CO₂: F_{1, 250} = 2.37, R² = 0.003, p = 0.073), nor its interaction with
355 plant species (PERMANOVA, CO₂*species: F_{3, 247} = 1.41, R² = 0.006, p = 0.149), influenced the
356 cardenolide profiles sequestered by monarchs.

357

358 ***Monarch Wing Morphology***

359 Monarch wings were more elongated (higher values of PCA-Shape) when larvae fed on
360 milkweed grown under eCO₂ (CO₂: F_{1, 214} = 15.82, p < 0.0001, Fig. 2a) or when larvae
361 consumed *A. syriaca* or *A. incarnata* (species: F_{3, 212} = 3.78, p = 0.011, Fig. 2c). Additionally,
362 the wings of female butterflies were more elongated than those of males (sex: F_{1, 213} = 15.50, p =
363 0.0001, Fig. 2b).

364

365 While *O. elektroscirra* infection had no independent effect on forewing shape (infection: F_{1, 212}
366 = 0.90, p=0.3550), infected monarchs from aCO₂ plants had rounder wings than butterflies from

367 eCO₂ infection treatments (infection*CO₂: F_{1,212} = 9.46, p = 0.002, Fig. 3a). Moreover, infected
368 monarchs had rounder wings than uninfected monarchs when feeding on *A. curassavica*, *A*
369 *syriaca*, and *A. incarnata*, but had more elongated wings than uninfected monarchs fed *A.*
370 *speciosa* (infection*species: F_{3,212} = 4.54, p = 0.004, Fig. 3b). Finally, there were minor
371 differences between male and female butterflies in wing shapes dependent on plant species and
372 infection (sex*infection*species: F_{3,213} = 2.96, p = 0.033, Fig. S3 in the Supporting
373 Information). However, the three-way interaction term explained only a small portion of variance
374 in the model when compared to the main effects reported above. Wing aspect ratio correlated
375 most strongly with PCA-shape ($r = 0.999$, N = 237, p < 0.0001), thus, we summarize the model
376 results for wing aspect ratios in Table 1 and Table S2 in the Supporting Information.

377
378 In contrast to wing shape, none of our treatments affected the size of monarch wings. Male
379 wings were only slightly significantly larger than those of females (sex: F_{1,231} = 3.47, p =
380 0.064). Likewise, wing sizes were unaffected by CO₂ treatment (CO₂: F_{1,31} = 0.31, p = 0.579),
381 milkweed host plant species (species: F_{3,202} = 2.09, p = 0.102), infection status (infection: F_{1,226}
382 = 2.42, p = 0.121) or the interaction between these treatments (CO₂ *species* infection: F_{3,204} =
383 0.98, p = 0.403).

384
385 The wing loading values of male monarchs were 5% higher than those of female monarchs (sex:
386 F_{1,15} = 17.13, p = 0.0008). Notably, monarchs reared on *A. syriaca* had a 5% higher wing
387 loading than did those reared on other milkweed species (species: F_{3,15} = 4.77, p = 0.0153, Fig.
388 4a). Wing loading was unaffected by CO₂ treatment (CO₂: F_{1,11} = 3.01, p = 0.112), or parasite
389 infection (infection: F_{1,15} = 3.80, p = 0.07).

390
391 Female monarch wings were 6% denser than male monarch wings (sex: F_{1,228} = 15.74, p <
392 0.0001). Monarchs fed *A. curassavica* had the thinnest wings while those fed *A. syriaca* had the
393 densest wings (species: F_{3,199} = 2.66, p = 0.0492, Fig. 4b). Interestingly, the wings of infected
394 monarchs were 7% less dense than the wings of uninfected monarchs (infection: F_{1,221} = 20.65,
395 p < 0.0001, Fig. 4c). Specific wing area was unaffected by CO₂ treatment (CO₂: F_{1,31} = 0.02, p
396 = 0.897).

397

398 In the simplified model exploring effects of cardenolide sequestration on wing shape, we found
399 no effects of monarch cardenolide sequestration (monarch cardenolides: $F_{1,230} = 0.05$, $p = 0.816$),
400 plant species (species: $F_{3,228} = 2.19$, $p = 0.090$) or their interaction (monarch
401 cardenolides*species: $F_{3,230} = 1.62$, $p = 0.184$, Table S4 in the Supporting Information) on wing
402 shape. Interestingly, monarchs that sequestered higher concentrations of cardenolides had lower
403 wing loading values (monarch cardenolides: $F_{1,15} = 8.41$, $p = 0.011$, Fig. 5). Also, those
404 monarchs feeding on higher cardenolide milkweed had lower wing loading values (species: $F_{3,15}$
405 $= 4.11$, $p = 0.025$), but there were no species-dependent effects of monarch cardenolide
406 sequestration on wing loading (monarch cardenolides*species: $F_{3,15} = 2.82$, $p = 0.073$). Finally,
407 there was a marginally significant trend of less dense wings produced in monarchs sequestering
408 higher concentrations of cardenolides ($F_{1,229} = 3.59$, $p = 0.059$). There were no effects of either
409 milkweed species (species: $F_{3,227} = 1.95$, $p = 0.122$) or the interaction on wing density (monarch
410 cardenolides*species: $F_{3,230} = 1.15$, $p = 0.330$).

411

412 **Discussion**

413 Rapid environmental change may influence trophic interactions by altering the defensive
414 phenotype of prey. Here, we demonstrate that: 1) monarchs maintain the concentration and
415 composition of cardenolides that they sequester despite changes in the phytochemistry of one
416 milkweed species induced by eCO₂. 2) Aspects of monarch morphology important to flight
417 ability such as wing shape, loading, and density vary in response to eCO₂, milkweed host plant
418 species, infection, and sex. 3) Feeding on high cardenolide milkweed is associated with the
419 formation of rounder, thinner wings, which are less efficient at gliding flight. We suggest that
420 changes in sequestration rates under eCO₂ are a byproduct of compensatory feeding aimed at
421 maintaining a nutritional target in response to declining diet quality. Additionally, monarchs
422 exhibit the cost of sequestering higher concentrations of cardenolides through declining wing
423 loading values. Small changes in wing morphology can have important consequences for
424 migration success (Bradley & Altizer, 2005), including migratory escape from parasites.
425 Therefore, changes in sequestration and morphology may have consequences for monarch
426 defense and migration in a changing world.

427

428 *Monarchs increase sequestration rate under eCO₂*

429 We demonstrate that monarchs can increase their rate of cardenolide sequestration under eCO₂
430 (Fig. 1). Specifically, monarchs sequester a constant concentration and composition of
431 cardenolides from *A. curassavica* despite a 52% reduction in foliar cardenolides and changes in
432 foliar cardenolide composition induced by eCO₂. Monarchs are known to maintain constant
433 concentrations of the cardenolides that they sequester from *A. curassavica* in response to nutrient
434 deposition, another environmental change driver (Tao & Hunter, 2015). Herbivores regulate
435 sequestration by altering both the total amount of foliage consumed and sequestration efficiency
436 (Camara, 1997). Notably, herbivorous arthropods maintain target ratios of carbohydrates to
437 protein in their diet through behavioral shifts in consumption (Simpson et al., 2015). Therefore, it
438 is possible that the monarchs in our study increased the amount of foliage consumed to
439 compensate for reductions in foliar nutrient content under eCO₂ (Hunter, 2001; Johnson et al.,
440 2014; Lincoln, Sionit, & Strain, 1984; Zavala, Nability, & DeLucia, 2013).

441

442 ***Environment influences monarch wing morphology***

443 Our treatments altered indices of monarch wing morphology (wing shape, loading, and density)
444 that are important to both aerial maneuverability and long-distance flight (Berwaerts et al.,
445 2002). Notably, both CO₂ treatment and milkweed species influenced wing shape contingent
446 upon parasite infection status (Figs 2 & 3). Under aCO₂, infection induced rounder wings
447 lowering the mean aspect ratio from 1.95 ± 0.002 to 1.93 ± 0.004 . However, under eCO₂ both
448 infected and uninfected monarchs developed more angular wings (1.97 ± 0.005 , 1.96 ± 0.002 ,
449 Fig. 3a). No study to date has specifically tested the effects of wing shape on the probability of
450 successful migration from start to finish in monarchs. However, field sampling indicates that
451 earlier arriving migrants tend to have larger, more elongated wing shapes suggesting that this
452 flight phenotype increases migration success (Satterfield & Davis, 2014). Therefore, future
453 environmental conditions may induce the formation of wing shapes that improve monarch flight
454 efficiency despite infection.

455

456 We also report effects of milkweed species on monarch wing shape, loading, and density, similar
457 to findings in other flying insects (Benítez, Vargas, & Püschel, 2015; Davis & de Roode, *in*
458 *Revision*; Soto, Carreira, Soto, & Hasson, 2008). Importantly, monarch wings are rounder and
459 less dense when larvae are reared on *A. curassavica*, a plant exotic to N. America and increasing

460 in prevalence in the southern US (Satterfield, Maerz, & Altizer, 2015). This species of milkweed
461 does not senesce in autumn, and contributes to a loss of monarch migratory behavior as
462 butterflies encounter viable foliage during their late-season stopovers (Satterfield et al., 2018,
463 2015). Our data suggest that the offspring of those sedentary monarchs fed *A. curassavica* will
464 develop lower quality flight phenotypes, perhaps furthering the loss of migratory behavior.

465
466 Small differences in wing morphology that affect the efficiency of flight could have large
467 consequences for monarch migration success (Bradley & Altizer, 2005). Eastern N. American
468 monarchs migrate up to 4,500 km through soaring and active flight (Brower & Malcolm, 1991;
469 Gibo, 1986). Monarchs must take shelter during adverse weather conditions, utilize wind
470 patterns, and cross large expanses of unsuitable habitat (Garland & Davis, 2002; Gibo & Pallett,
471 1979). Therefore, any factor that causes monarchs to remain grounded during beneficial flying
472 conditions or reduces the amount of time monarchs may stay aloft over unsuitable habitat will
473 significantly reduce migration success. In our study, eCO₂ eliminates the shape difference
474 between infected and uninfected individuals, inducing more elongated wings in both groups. If
475 infected individuals become more efficient gliders under environmental change, this might
476 decrease migratory culling which reduces pathogen prevalence seasonally in the N. American
477 monarch population (Altizer et al., 2011; Bartel, Oberhauser, de Roode, & Altizer, 2011).

478
479 Although our treatments altered monarch wing shape, we detected no effect of diet or infection
480 on wing size. All of the butterflies used in this study originated from the migratory eastern N.
481 American population. Therefore our data substantiate previous studies demonstrating strong
482 selection for larger wings imposed by migration distance within this population (Altizer & Davis,
483 2010; Li et al., 2016). Forewing size is likely more important to migration success than wing
484 shape because it is conserved among all our treatments and is selected for in the migratory
485 populations of monarchs (Altizer & Davis, 2010; Flockhart et al., 2017; Li et al., 2016).
486 However, aerodynamic theory suggests that wing shape can be important for maneuverability
487 and energy conservation (Pennycuick, 2008). Because our monarchs were constrained within
488 these constant wing sizes, perhaps the amount of differentiation in the angular nature of wing
489 shape was limited. Further studies exploring the plasticity of wing shape and size in response to

490 host-plant and environmental conditions among different populations of monarchs across the
491 globe are needed to better address this idea.

492

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498

499 **Authors' Contributions:**

500 LED, MDH & JCdR designed the experiment. JCdR provided butterflies and parasites. LED,
501 AJS & MDH collected and analyzed the data. LED wrote the manuscript; all authors contributed
502 to drafts.

503

504 **Data Accessibility:** Data are available in the Dryad Digital Repository.

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730 **SUPPORTING INFORMATION**

731 Additional supporting information may be found in the online version of this article.

732

733 Appendix S1. Details of cardenolide extraction and the generalized linear mixed model used to
734 determine the distribution of plants that produced undetectable cardenolides.

735

736 Figure S1. An illustration of the four basic morphometric measures taken in Adobe Photoshop.

737

738 Figure S2. Effects of infection by *Ophryocystis elektroscirrha* on the total concentration of
739 cardenolides sequestered by monarchs fed four species of milkweed.

740

741 Figure S3. The interaction between milkweed host-plant species, infection and sex on a
742 composite measure of monarch forewing shape.

743

744 Table S1. Sample sizes of surviving monarchs that were used to explore the effects of milkweed
745 species, elevated CO₂, and infection, on toxin sequestration and wing morphology

746

747 Table S2. The three-way interaction between treatments on mean monarch wing Aspect Ratios

748

749 Table S3. Table of all models used in this study with accompanying ANOVA tables.

750

751 Table S4. Table of model results testing the relationship between cardenolide sequestration in
752 wings and milkweed species on monarch wing shape, loading and density.

753 **Captions**

754 **Table 1.** The a) main and b) interactive effects of CO₂ treatment, butterfly sex, and host plant on
755 monarch wing aspect ratios (wing length/wing width), a component of monarch wing shape.
756 Data are the means ±1 SE. Aspect ratios range between 1.6 to 2.1 but average around 1.93 in
757 eastern N. American monarch populations. Simplified linear mixed effects model structure is as
758 follows: Aspect Ratio ~ CO₂ + butterfly sex + infection + milkweed species + CO₂ * infection +
759 milkweed species*infection + sex*infection*milkweed species + random effects = chamber,
760 monarch lineage. We do not present nonsignificant main effects of infection. Table S2 presents
761 the three-way interaction.

762
763 **Fig. 1.** Effects of eCO₂ on (a) foliar cardenolide concentrations (mg/g dry mass), (b) monarch
764 wing cardenolide concentrations (mg/g dry mass), and (c) the relationship between foliar and
765 wing cardenolide concentrations. Bars represent mean values ±1 SE. Traits were transformed
766 before analyses but are presented as untransformed values for ease of interpretation. Grey bars
767 represent plants grown under aCO₂ and orange bars are those from eCO₂ or the monarchs that
768 fed on those plants. Milkweed species codes: CUR = *A. curassavica*, SYR = *A. syriaca*, SPE =
769 *A. speciosa*, INC = *A. incarnata*.

770
771 **Fig. 2.** The main effects of (a) CO₂ treatment, (b) sex and (c) milkweed species on a composite
772 measure of monarch forewing shape. Points represent mean PCA-shape values ±1 SE. With
773 increasing PCA-shape values wings become more elongated and angular. Milkweed species
774 codes are the same as above.

775
776 **Fig. 3.** The interactions between (a) CO₂ treatment and infection by OE, and (b) milkweed
777 species and infection on a composite measure of monarch forewing shape. Points represent mean
778 PCA-shape values ±1 SE. Red points indicate mean shape values of infected monarchs while
779 blue points represent uninfected monarchs. Milkweed species codes are the same as above.

780
781 **Fig. 4.** The effects of milkweed species on monarch (a) wing loading (body mass/wing area) and
782 (b) specific wing area (wing area/wing mass), a measure of wing density. (c) The effects of OE
783 infection on specific wing area. Bars represent mean values ±1 SE. Higher specific wing area
784 values indicate wings that are less dense. Milkweed species codes are the same as above.

785

786 **Fig. 5.** The relationship between the concentration of cardenolides sequestered in monarch wings
787 and wing loading (body mass/wing area). Points represent individual monarchs and colors
788 correspond to the species of milkweed host-plants fed to each monarch. Milkweed species codes
789 are the same as above.

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Tables:

Table 1.

Main Effects			Two-way Interactions		
CO₂ treatment	$F_{1,214} = 15.82$	$p < 0.0001$	Infection*CO₂	$F_{1,212} = 9.46$	$p = 0.0024$
	Aspect Ratio		Infection Status	CO₂ treatment	Aspect Ratio
Ambient	1.95 ± 0.002		Infected	Ambient	1.93 ± 0.004
Elevated	1.96 ± 0.002			Elevated	1.97 ± 0.005
Butterfly Sex	$F_{1,213} = 15.50$	$p < 0.0001$	Uninfected	Ambient	1.95 ± 0.002
	Aspect Ratio			Elevated	1.96 ± 0.002
Female	1.97 ± 0.002		Infection*Species	$F_{3,212} = 4.54$	$p = 0.004$
Male	1.93 ± 0.002		Infection Status	Milkweed Species	Aspect Ratio
Milkweed Species	$F_{3,212} = 3.78$	$p = 0.0113$			
	Aspect Ratio		Infected	A. curassavica	1.93 ± 0.008
A. curassavica	1.94 ± 0.003			A. incarnata	1.95 ± 0.005
A. incarnata	1.97 ± 0.003			A. speciosa	1.97 ± 0.006
A. speciosa	1.95 ± 0.004			A. syriaca	1.94 ± 0.0102
A. syriaca	1.96 ± 0.003		Uninfected	A. curassavica	1.95 ± 0.003
				A. incarnata	1.97 ± 0.003
				A. speciosa	1.94 ± 0.004
				A. syriaca	1.96 ± 0.003









