

1 **Supplementary Material**

2 **Analytical solution for the compartmental model**

3 When time  $(t) \leq$  time to maximal expansion ( $T_{max}$ ), the compartmental model in **Figure 2** is described by  
4 the equation  $dE/dt = \rho t$ , which has the analytical solution  $E(t) = R_0 \exp(\rho t)$ , where as described earlier,  $R_0$   
5  $= C_{max}/\text{fold}_x$ . This gives the exponential growth equation (Eq 1).

6 When  $t > T_{max}$ , the analysis that links the equations for the compartmental model to the analytical expression  
7 are provided below, where the rate at which effector cells decline is  $(\alpha-k)$ , the rate at which effector cells  
8 become memory cells is  $k$ , and the rate at which memory cells decline is  $\beta$ .

9 
$$dE/dt = -(\alpha-k)E - kE = -\alpha E$$

10 
$$dM/dt = \alpha E - \beta M$$

11 Rewritten in matrix notation and grouping terms gives:

12 
$$\frac{d}{dt} \begin{pmatrix} E \\ M \end{pmatrix} = \begin{pmatrix} -\alpha & 0 \\ k & -\beta \end{pmatrix} \begin{pmatrix} E \\ M \end{pmatrix}$$

13  
14 The eigenvalues of the above system are clearly  $\lambda_1 = -\alpha$  and  $\lambda_2 = -\beta$ , and the associated eigenvectors are  
15

16 
$$v_1 = \begin{pmatrix} 1 \\ -k/(\alpha - \beta) \end{pmatrix}, \quad v_2 = \begin{pmatrix} 0 \\ 1 \end{pmatrix}$$

17  
18 Eigenvector  $v_2$  is easy to check, and a check of  $v_1$  is shown below.

19  
20 
$$\begin{pmatrix} -\alpha & 0 \\ k & -\beta \end{pmatrix} \begin{pmatrix} 1 \\ -k/(\alpha - \beta) \end{pmatrix} = \begin{pmatrix} -\alpha \\ k + \beta k/(\alpha - \beta) \end{pmatrix} = \begin{pmatrix} -\alpha \\ \alpha k/(\alpha - \beta) \end{pmatrix} = -\alpha \begin{pmatrix} 1 \\ -k/(\alpha - \beta) \end{pmatrix}$$

21  
22 This results in the solution below, where  $C_1$  and  $C_2$  are 2 constants to be defined further below.  
23

24 
$$\begin{pmatrix} E(t) \\ M(t) \end{pmatrix} = C_1 \begin{pmatrix} 1 \\ -k/(\alpha - \beta) \end{pmatrix} e^{-\alpha(t-T_{max})} + C_2 \begin{pmatrix} 0 \\ 1 \end{pmatrix} e^{-\beta(t-T_{max})}$$

25  
26 The measurement variable is:

27

$$f(t - T_{\max}) = E(t - T_{\max}) + M(t - T_{\max}) = C_1 \left(1 - \frac{k}{\alpha - \beta}\right) e^{-\alpha t} + C_2 e^{-\beta t}$$

28

29

30 Also, at time  $T_{\max}$ , it is assumed that all cells are effector cells; thus,  $E(T_{\max}) =$  maximal concentration ( $C_{\max}$ )  
 31 (giving  $C_1 = C_{\max}$ ) and  $M(T_{\max}) = 0$  [giving  $C_2 = C_1 k / (\alpha - \beta) = C_{\max} k / (\alpha - \beta)$ ]. Note that in the analytical  
 32 formula,  $C_2 = C_{\max} \cdot F_B$ . Thus, the differential equation model is mapped to the analytical model:

$$C_2 = C_{\max} \cdot F_B = C_{\max} k / (\alpha - \beta)$$

$$k = F_B \cdot (\alpha - \beta)$$

33

34

35 **Semi-mechanistic Model Selection**

36 The semi-mechanistic model presented here is not the only model that gives rise to exponential  
 37 growth followed by biexponential decay. If memory cells were produced during the expansion phase as  
 38 well or if cells could transition back and forth from the effector to memory state, then the analytic solution  
 39 to such a model would have the same functional form. It is also possible that terminal phase is not due to  
 40 formation of memory cells, but rather the low level of constant proliferation in response to the continual  
 41 production of CD19 antigen; the kinetics of this process would look similar. Furthermore, DeBoer and  
 42 Perelson<sup>4</sup> also showed in equations 8-10 of their paper that rather than have expansion stop at  $T_{\max}$ ,  
 43 one could use a “cascade” model of cell division, modeling each generation of expansion complex model  
 44 and realistic model with successive generations of expansion. The cascade model could be approximated  
 45 by the simpler model used here; the advantage of the model used here is it is simpler to implement and to  
 46 present.

47

48 **Derivation of AUC to approximate  $C_{\max}$**

49 In the scenario where no comedications are given, the area under the curve (AUC) from time 0 to time  
 50 “ $T_{\text{end}}$ ” is given by the equation below, where  $R = C_{\max} / \text{fold}_x$ ,  $A = (1 - F_B) \cdot C_{\max}$ , and  $B = F_B \cdot C_{\max}$ . The final  
 51 equation below shows that in this model formulation, AUC is proportional to  $C_{\max}$ ; thus, covariates that affect  
 52  $C_{\max}$  also impact the AUC. In the derivation below, we switch from an equality to an approximation because

53 with the parameters fit here,  $\exp(\rho \cdot T_{\max}) = \text{foldx} = 3900 \gg 1$ , such that  $[\exp(\rho \cdot T_{\max}) - 1] / \exp(\rho \cdot T_{\max}) \approx$

54 1

55

56

$$\begin{aligned}
 AUC_{0-T_{\text{end}}} &= \int_0^{T_{\text{max}}} R \exp(\rho t) dt + \int_{T_{\text{max}}}^{T_{\text{end}}} A \exp(-\alpha(t - T_{\text{max}})) + B \exp(-\beta(t - T_{\text{end}})) dt \\
 &= \left[ \frac{R}{\rho} \exp(\rho t) \right]_0^{T_{\text{max}}} + \left[ -\frac{A}{\alpha} \exp(-\alpha(t - T_{\text{max}})) - \frac{B}{\beta} \exp(-\beta(t - T_{\text{end}})) \right]_{T_{\text{max}}}^{T_{\text{end}}} \\
 &= \left[ \frac{R}{\rho} (\exp(\rho \cdot T_{\text{max}}) - 1) \right] + \left[ \frac{A}{\alpha} (1 - \exp(-\alpha(T_{\text{end}} - T_{\text{max}}))) + \frac{B}{\beta} (1 - \exp(-\beta(T_{\text{end}} - T_{\text{max}}))) \right]
 \end{aligned}$$

substituting in the equations for  $R$ ,  $A$ , and  $B$  gives

$$\begin{aligned}
 &= \left[ \frac{C_{\text{max}} (\exp(\rho \cdot T_{\text{max}}) - 1)}{\rho \exp(\rho \cdot T_{\text{max}})} \right] \\
 &\quad + \left[ \frac{(1 - F_B) \cdot C_{\text{max}}}{\alpha} (1 - \exp(-\alpha(T_{\text{end}} - T_{\text{max}}))) + \frac{F_B \cdot C_{\text{max}}}{\beta} (1 - \exp(-\beta(T_{\text{end}} - T_{\text{max}}))) \right] \\
 &= C_{\text{max}} \cdot \left[ \frac{1 (\exp(\rho \cdot T_{\text{max}}) - 1)}{\rho \exp(\rho \cdot T_{\text{max}})} \right. \\
 &\quad \left. + \frac{(1 - F_B)}{\alpha} (1 - \exp(-\alpha(T_{\text{end}} - T_{\text{max}}))) + \frac{F_B}{\beta} (1 - \exp(-\beta(T_{\text{end}} - T_{\text{max}}))) \right] \\
 &\approx C_{\text{max}} \left[ \frac{1}{\rho} + \frac{(1 - F_B)}{\alpha} (1 - \exp(-\alpha(T_{\text{end}} - T_{\text{max}}))) + \frac{F_B}{\beta} (1 - \exp(-\beta(T_{\text{end}} - T_{\text{max}}))) \right] \\
 &\text{as } T_{\text{end}} \rightarrow \infty
 \end{aligned}$$

57

$$AUC_{0-\infty} \approx C_{\text{max}} \left( \frac{1}{\rho} + \frac{1 - F_B}{\alpha} + \frac{F_B}{\beta} \right)$$

58

59 **Covariate Analysis – Further Details**

60 Sex, race, and weight were included because they were demographic parameters that are typically  
61 included in a model-based analysis of exposure. Weight-adjusted dose, too, is a factor that for standard  
62 drugs correlates with exposure. Down’s Syndrome was included because ALL is more prevalent in  
63 Down’s syndrome patients. Prior lymphodepleting therapy and prior stem cell transplant were included  
64 because it’s possible that both could have affected the CAR-T cell ability to expand. Per protocol patients  
65 received fixed, prespecified weight or surface area-adjusted lymphodepleting (LD) fludarabine and  
66 cyclophosphamide chemotherapy dosing. A small percent (listed in ELIANA publication) of patients  
67 received non-FC LD chemotherapy or non LD chemotherapy if blood counts were very low. In this  
68 subset, 85 patients received fludarabine and cyclophosphamide, 2 received only cyclophosphamide, and  
69 3 received neither.

70 For practical reasons T cell numbers were not included as a covariate in this analysis. While the inclusion  
71 of T cell numbers as a covariate could alter  $C_{max}$ , it is unlikely that it would impact the primary  
72 conclusions of the effect of tocilizumab and corticosteroids on the rate of expansion. However, this  
73 question warrants study in future analyses. If it is possible to collect data on tumor burden or CD19  
74 antigen density, this data should be included in assessing the impact on  $C_{max}$  and if the follow-up is long  
75 enough, then  $\beta$  as well. The impact of missing this information in this study is that we may have larger  
76 portion of unexplained variability in the exposure.

77 The relationship between  $C_{max_i}$  ( $C_{max}$  for patient  $i$ ), the random effect  $\eta_i$  with variance  $\omega^2$ , and the covariate  
78  $j$  is given below. Here,  $x_{ij}$  denotes the covariate  $j$  value for patient  $i$ ,  $z_{ij}$  is a transformation of  $x_{ij}$ , and  $\theta_j$   
79 denotes the size of the covariate effect.

80

81 (4) 
$$C_{max_i} = C_{max} \cdot \exp(\eta_i) \cdot \prod_j z_{ij}$$

82

83 For continuous covariates (eg, dose),

84

85 (5) 
$$z_{ij} = (x_{ij}/x_{median,j})^{\theta_j}$$

86

87 For dichotomous covariates,  $x_i$  is assigned either 0 or 1 (eg,  $x_{ij} = 0$  for no tocilizumab;  $x_{ij} = 1$  for  
88 tocilizumab). Then  $z_{ij}$  is given by

89

90 (6) 
$$z_{ij} = (\theta_j)^{x_{ij}}$$

91

92 For categorical covariates with  $n > 2$  categories (eg, race)  $n - 1$  dichotomous variables ( $y_{ijk}$ ) are created  
93 for each patient  $i$ , with  $k$  going from 1 to  $n - 1$ . Then  $z_{ij}$  is given by

94

95 (7) 
$$z_{ij} = \prod_k (\theta_{jk})^{y_{ijk}}$$

96



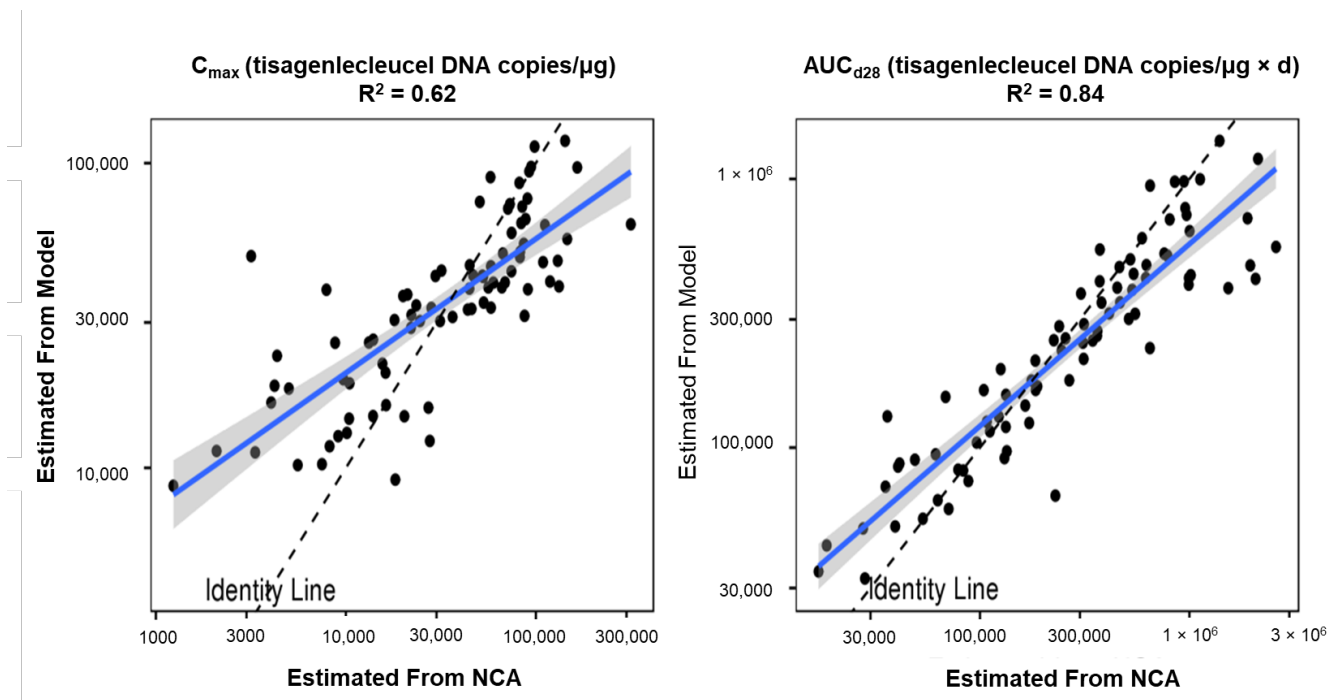
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98
99
100 Model MLXTRAN file
101
102 INPUT:
103 parameter = {foldx, Tmax, Cmax, Ftoci, Fster, alpha, FB, beta}
104 regressor = {TOCI1T, STERSTT}
105
106 EQUATION:
107
108 ; Figure out whether tocilizumab or steroids come first
109 ; assign first comedication to t1,F1 and second to t2,F2
110 ;
111 ; If patient does not receive TOCILIZUMAB, then TOCI1T=99999 in dataset
112 ; If patient does not receive STEROIDS, then STERSTT=99999 in dataset
113     if TOCI1T <= STERSTT
114         t1 = min(TOCI1T,Tmax)
115         F1 = Ftoci
116         t2 = min(STERSTT,Tmax)
117         F2 = Fster
118     else
119         t1 = min(STERSTT,Tmax)
120         F1 = Fster
121         t2 = min(TOCI1T,Tmax)
122         F2 = Ftoci
123     end
124
125 ; Compute constants for solving system
126 ; Note that: foldx = exp(rho*tau)
127     rho = log(foldx)/Tmax
128     R0 = Cmax/exp(rho*Tmax)
129     R1 = R0*exp( rho* t1 )
130     R2 = R1*exp(F1 *rho*(t2 -t1))
131     CmaxAdjust = R2*exp(F1*F2*rho*(Tmax -t2))
132
133     AA = (1-FB)*CmaxAdjust
134     BB = FB*CmaxAdjust
135
136 ; Analytical solution for system
137     if t<0
138         ylin = R0
139     elseif t < t1
140         ylin = R0*exp(rho*t)
141     elseif t < t2
142         ylin = R1*exp(F1*rho*(t-t1))
143     elseif t < tmax
144         ylin = R2*exp(F1*F2*rho*(t-t2))
145     else
146         ylin = AA*exp(-alpha*(t-Tmax)) + BB*exp(-beta*(t-Tmax))
147     end
148
149     logy = log(ylin)
150
151 OUTPUT:
152 output = {logy}
153

```

154 **Figure S1** Correlation between direct and model-based estimates of maximal concentration ( $C_{max}$ ) and  
155 area under the curve at day 28 ( $AUC_{d28}$ ). The data-driven estimates of  $C_{max}$  and  $AUC_{d28}$  were calculated  
156 in patients who had  $\geq 21$  days of follow-up data.  $C_{max}$  was the maximum quantitative polymerase chain  
157 reaction value in the first 28 days, and  $AUC_{d28}$  was calculated using the trapezoidal rule. The model-  
158 based estimate for  $C_{max}$  was derived directly from the *post hoc* estimate of the  $C_{max}$  parameter. The  
159 model-based  $AUC_{d28}$  was calculated using the *post hoc* estimates of all the cellular kinetics and the  
160 formula above for  $AUC_{0-T_{end}}$  (where  $T_{end} = 28$ ). Significant correlation between the noncompartmental  
161 analysis (NCA) and model-based estimates was observed.  $T_{end}$ , time of final measurement of transgene  
162 copy number. The blue line and shaded area correspond to the linear regression with 90% confidence  
163 intervals. The dotted line is the identity line.

164



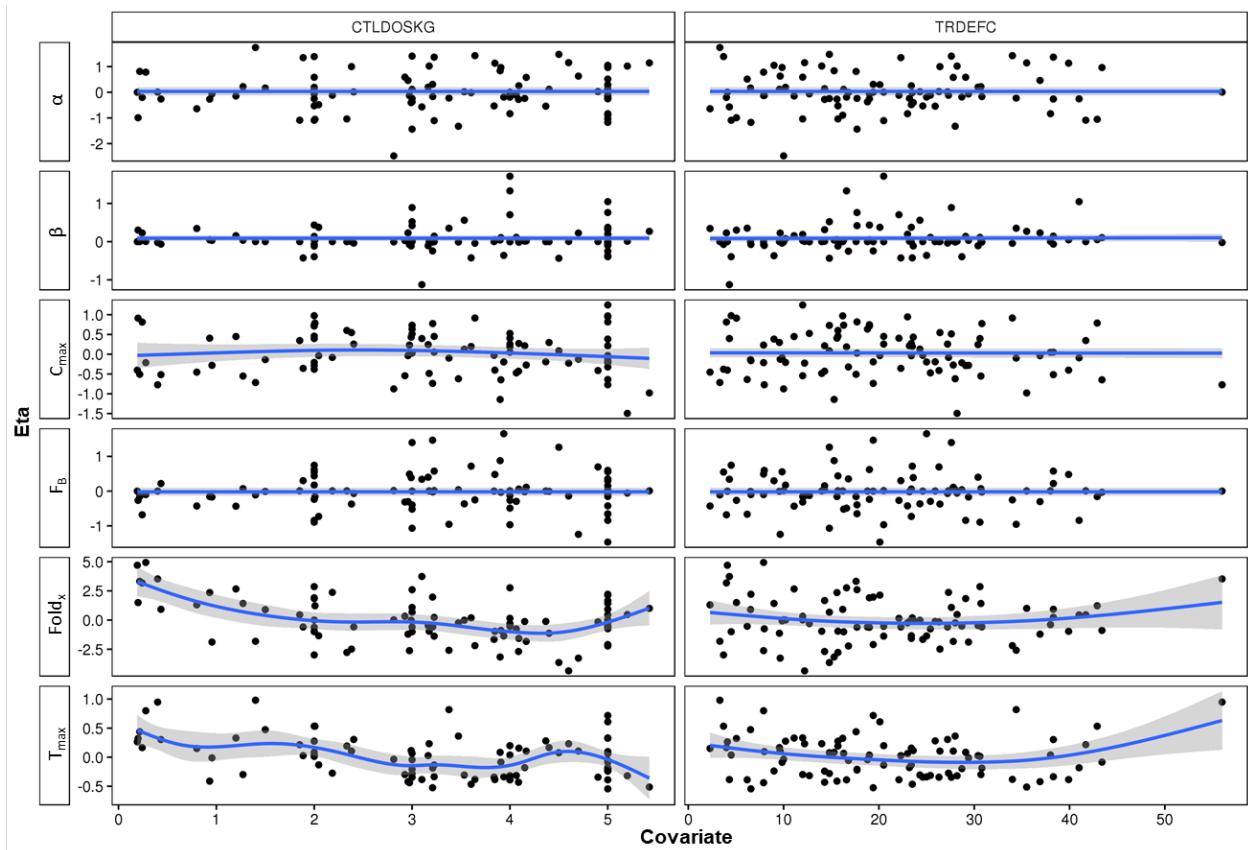
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166

167 **Figure S2** Exploratory plots for covariate analysis. **(a)** Continuous covariates plotted against *post hoc*  
 168 random-effects estimates (Eta). CTLDOSKG is defined as the dose of tisagenlecleucel per kg. TRDEFC  
 169 is defined as the transduction efficiency. **(b)** Categorical covariates plotted against *post hoc* random-  
 170 effects estimates. Downs denotes Down syndrome, LDFLUD denotes that the patient received  
 171 fludarabine lymphodepleting therapy, Ster denotes that the patient received corticosteroids, Stud denotes  
 172 the study (ELIANA, B2202; ENSIGN, B2205J), and Toci denotes that the patient received tocilizumab.  
 173  $C_{max}$ , maximal concentration;  $F_B$ , fraction of transgene copies present during the decline at the gradual  
 174 rate  $\beta$ , starting from  $T_{max}$ ;  $fold_x$ , fold expansion; HSCT, hematopoietic stem cell transplant;  $T_{max}$ , time to  
 175 maximal expansion. The blue line and shaded area shows the loess smoothed line; no obvious trends are  
 176 observed.

177

178 (a)

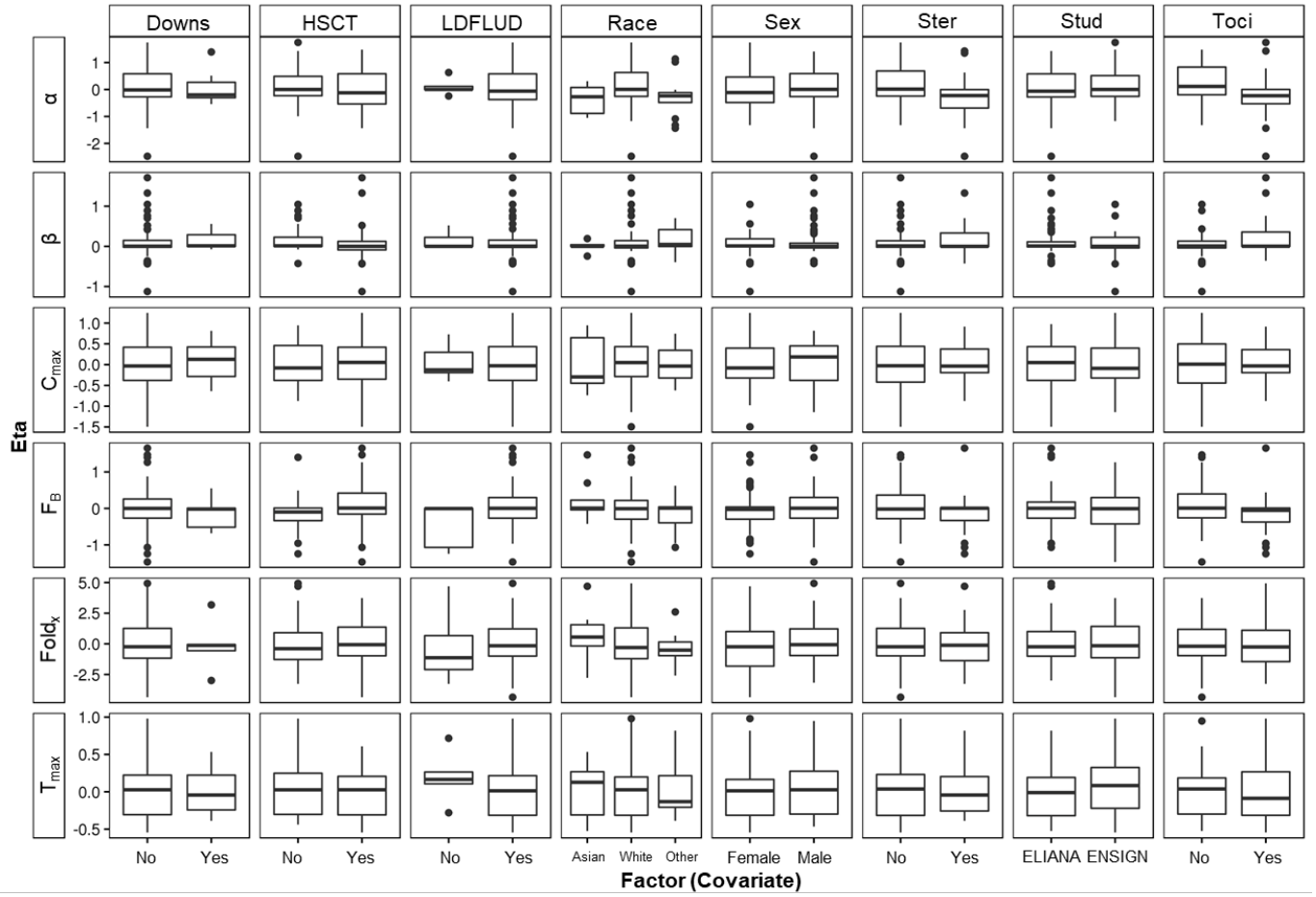
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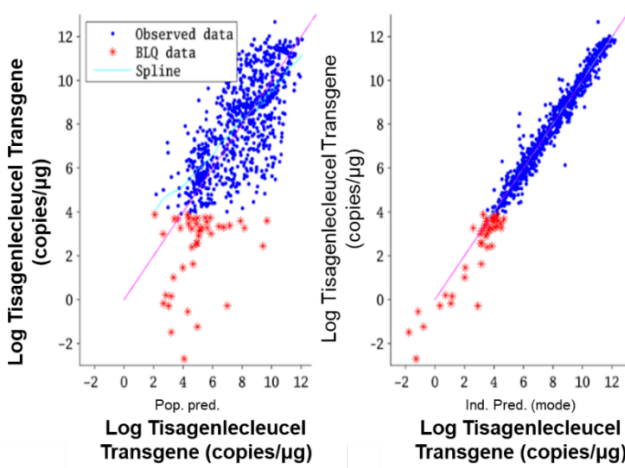
181 (b)



182

183 **Figure S3** Predictions vs observations. **(a)** Blue dots are quantifiable values, and red asterisks signify  
 184 simulated residuals below the limit of quantification (BLQ). **(b)** Blue dots are quantifiable values, and red  
 185 asterisks signify simulated residuals BLQ. The green solid lines in the pdf plot demonstrate the true  
 186 residual distribution, and the black lines that nearly overlie the green lines denote the theoretical  
 187 distribution. IWRES, individual weighted residual; NPDE, normalized distribution prediction errors; pdf,  
 188 probability distribution function.

189 **(a)**



191 **(b)**

