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# 45 List of Abbreviations:

- 46 APRI, AST to platelet ratio index. BMI, body mass index. CT, computed tomography. DXA,
- 47 dual-energy X-ray absorption. FHS, Framingham Heart Study. LPR, liver-phantom ratio. LPRS,
- 48 lipodystrophy polygenic risk score. NAFLD, nonalcoholic fatty liver disease. MGI, Michigan
- 49 Genomics Initiative.
- 50
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59

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- Andrew Wright: study design, data analysis and interpretation, and drafting of the manuscript
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#### 77 Abstract:

78 Background: Up to 25% of patients with nonalcoholic fatty liver disease (NAFLD) are not obese but may have a fat or muscle composition that predisposes them to NAFLD. Our aim was to 79 80 determine whether body composition parameters associate with NAFLD and to identify genetic 81 contributors to this association. Methods: This study included two cohorts. The first included 82 2,249 participants from the Framingham Heart Study who underwent a computed tomography 83 scan to evaluate hepatic steatosis, dual-energy X-ray absorptiometry testing to assess body composition, and clinical examination. Body composition parameters were normalized to total 84 body weight. A subset of participants underwent genotyping with an Affymetrix 550K SNP 85 array. The second cohort, Michigan Genomics Initiative, included 19,239 individuals with 86 87 genotyping on the Illumina HumanCoreExome v.12.1 array and full electronic health record data. Results: Using sex-stratified multivariable linear regression, greater central body fat 88 associated with increased hepatic steatosis, while greater lower extremity body fat associated 89

with decreased hepatic steatosis. Greater appendicular lean mass was associated with decreased 90 hepatic steatosis in men but not in women. A polygenic risk score for lipodystrophy (regional or 91 global loss of adipose tissue) was associated with increased hepatic steatosis, increased liver 92 fibrosis, and decreased lower extremity fat mass. Conclusions: Greater central body fat 93 associated with increased hepatic steatosis while greater lower extremity body fat and, in men, 94 greater appendicular lean mass were associated with decreased hepatic steatosis. A genetic risk 95 score for lipodystrophy was associated with NAFLD and liver fibrosis. Our results suggest that 96 buffering of excess energy by peripheral fat and muscle may protect against NAFLD and liver 97 fibrosis in the general population. 98

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#### 263/275 words 100

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#### Introduction 101

Nonalcoholic fatty liver disease (NAFLD) is characterized by excessive triglyceride 102 accumulation in the liver in the absence of significant alcohol use or other underlying cause (1). 103 NAFLD is the most common chronic liver disease worldwide, affecting 20-40% of the general 104 population, and is associated with metabolic conditions such as obesity, diabetes, and 105 dyslipidemia (2-4). Unfortunately, treatment options remain limited and a better understanding 106 107 of the pathophysiology underlying NAFLD will be critical in developing more effective treatments. 108

109

While NAFLD is associated with obesity, approximately 25% of patients with NAFLD are not 110 obese (5). This finding suggests that not all fat contributes equally to NAFLD risk: it may be 111 regional adiposity rather than overall adiposity that contribute to liver steatosis. Visceral fat is 112 113 associated with increased risk of NAFLD and progression to hepatic fibrosis (6-8). In contrast, 114 gluteofemoral and lower extremity fat correlate with decreased transaminases and may be protective against hepatic steatosis (9-11). Despite the importance of different fat depots in 115 NAFLD, the literature on lower extremity fat in NAFLD is limited to studies of a few hundred 116 subjects, mostly in Asian populations. 117

118

Skeletal muscle mass may also protect against NAFLD. Sarcopenia, a condition of low skeletal 119 muscle mass, has been linked to increased risk of NAFLD and advanced fibrosis (12-14). 120

- 121 Skeletal muscle function too may influence NAFLD: greater hand grip strength has been linked
- to decreased NAFLD prevalence (15). Further, NAFLD is associated with substitution of adipose
- tissue in skeletal muscle (16) and increased insulin resistance of skeletal muscle (17). However,
- again, the literature on muscle fat and NAFLD risk is limited to only small studies of a few
- hundred subjects and has not been studied in a Western population.
- 126

A number of genes have been implicated in NAFLD and some of these genes also influence 127 body composition (18). For example, individuals with NAFLD in the presence of the PNPLA3 128 I148M variant are less frequently obese than those with the ancestral allele at PNPLA3 (19, 20). 129 More recently, several genetic contributors to lipodystrophy have been identified. Lipodystrophy 130 is characterized by global or selective deficiency of adipose tissue in the absence of malnutrition 131 132 or a catabolic state (21). While most patients with NAFLD are not overtly "lipodystrophic", NAFLD is itself a form of ectopic fat accumulation and is highly prevalent in patients with 133 familial lipodystrophy (21, 22). Lipodystrophy was previously viewed primarily through the lens 134 of rare familial diseases, but relative lipodystrophy may also exist in the general population as a 135 136 continuous trait (23). Non-familial lipodystrophy is heritable and a previously-reported lipodystrophy polygenic risk score (LPRS) predicts insulin resistance and decreased lower 137 138 extremity adiposity, a feature of lipodystrophy (24). Whether people with an increased polygenic lipodystrophy score store more fat in the liver is not known. 139

140

We hypothesize that NAFLD may be a marker of partial lipodystrophy in the population. We test
whether body composition—specifically, fat distribution and muscle bulk, strength, and fat
content—associate with NAFLD in a large, well-characterized European-ancestry cohort, the
Framingham Heart Study (FHS). Further, we test whether individuals with higher lipodystrophy
polygenic scores have higher prevalence of NAFLD and liver fibrosis using FHS and another
cohort, Michigan Genomics Initiative (MGI).

- 147
- 148 Methods
- 149
- 150 *Ethics statement*

151 All FHS participants provided written informed consent approved by the Boston University

152 Institutional Review Board and Hebrew SeniorLife Institutional Review Board. All MGI

153 participants provided written informed consent approved by the institutional review board of the

154 University of Michigan (Ann Arbor, MI). All research performed in this paper was approved by

the institutional review board of the University of Michigan (Ann Arbor, MI).

156

157 Cohorts

This study included two cohorts. The first was FHS, a multigenerational prospective cohort study 158 of residents in and around Framingham, Massachusetts characterizing a broad array of 159 160 phenotypes related to cardiovascular health (25). We included the FHS Offspring and Generation 3 sub-cohorts. Between 1995 and 1998, 3492 participants from the Offspring cohort completed 161 the seventh clinical examination (exam 7). Between 2008 and 2011, 3399 participants from 162 Generation 3 completed the second clinical examination (exam 2). These examinations included 163 164 a detailed medical history, physical examination, collection of blood specimens, and measurement of anthropometric data including hand grip strength assessment (for the Offspring 165 166 cohort, hand grip strength measurements were collected separately) (25). Selected subjects participated on sub-studies that involved additional testing including multidetector CT scan, 167 168 whole-body DXA scan, and quadriceps strength testing. We excluded participants who reported excess alcohol use (>21 alcoholic drinks per week for men and >14 alcoholic drinks per week for 169 170 women). The physical activity index is a composite score calculated based on participant responses to questions regarding different levels of physical activity and sleep patterns over a 24 171 172 hour period (26). Grip and quadriceps strength were measured as described previously (27, 28). 173

In FHS, a subset of participants underwent genotyping with a 550K SNP array (Affymetrix 500K
Dual GeneChip and 50K gene-centered MIP set) (29). Imputation was performed using the 1000
Genomes cosmopolitan panel March 2012(v3) on the Michigan Imputation Server
(<u>https://imputationserver.sph.umich.edu/index.html</u>) (30).

178

179 MGI is a prospective cohort with ongoing enrollment; all patients undergoing elective surgery at

180 Michigan Medicine (Ann Arbor, Michigan) are potentially eligible for enrollment in this cohort.

181 Enrollment involves genotyping of peripheral blood on the Illumina HumanCoreExome v.12.1

array, a GWAS and exome array consisting of >500,000 SNPs (31). In addition, full laboratory
information and billing codes are available.

184

#### 185 Hepatic steatosis and muscle attenuation assessment

186 Between 2008 and 2011, multidetector abdominal CT scans (64-slice, General Electric Health

187 Care) were performed as described previously (32). The mean attenuation (Hounsfield units)

188 from three regions in the liver as well as from a calibration control (phantom) was calculated.

189 The liver-phantom ratio (LPR) was calculated by dividing the mean hepatic attenuation by the

attenuation of the calibration control ("phantom"). LPR  $\leq 0.33$  was used to define NAFLD as

reported previously (33). Muscle attenuation was measured at the left and right paraspinous

muscles at the mid-abdominal level as previously described (34).

193

#### 194 Body composition assessment

Whole-body and regional measures of lean mass and fat mass were obtained by DXA scan (GE 195 Lunar Prodigy fan beam densitometer) as described previously (28, 35). For the Offspring 196 cohort, these DXA scans were obtained from 1996-2001. For Generation 3, they were obtained 197 in 2010 and 2011. The DXA protocol was the same between the two cohorts. Lower extremity 198 fat mass was a reported measure that combined the fat mass in both legs. Total fat mass was also 199 reported. Appendicular lean mass was calculated by combining bilateral upper and lower 200 extremity lean mass. Central fat mass was calculated by subtracting the bilateral upper and lower 201 extremity fat mass from the whole body total fat mass. These measures were scaled to body 202 weight by dividing the respective values by each participant's weight in kilograms and reported 203 204 as a percentage.

205

### 206 *Clinical and laboratory measurements*

The age of the participant documented at the time of the clinical examination was used for the analysis. Body mass index (BMI) was defined as weight (kg)/height (m<sup>2</sup>). Diabetes was defined by the presence of a fasting glucose  $\geq 126$  mg/dL, hemoglobin A1c  $\geq 6.5\%$ , medical history of

210 physician-diagnosed diabetes, or receiving medication for the treatment of diabetes.

211 Hypertension was defined as a systolic blood pressure  $\geq$  140 mmHg, diastolic blood pressure  $\geq$ 

212 90 mmHg, physician-diagnosed hypertension, or receiving antihypertensive therapy. Metabolic

syndrome was diagnosed based on the National Cholesterol Education Program's Adult
Treatment Panel III guidelines (36).

215

In both FHS and MGI, the AST to platelet ratio index (APRI) score was used for noninvasive 216 assessment of liver fibrosis. In MGI, we defined cirrhosis based on presence of an ICD-9 code 217 218 (571.5, 571.2, and 571.6), ICD-10 code (K74.X, K70.2-4, and K71.7), or a text search for cirrhosis. Text search of radiology and pathology reports was performed for the character 219 "cirrho," and participants with that character were flagged as having cirrhosis with the following 220 exceptions: (1) if the word "without" or "no" appeared in the same sentence as "cirrho," subjects 221 were considered to not have cirrhosis; (2) if the words "primary biliary cirrhosis" appeared in a 222 sentence, that sentence was ignored for text search purposes; and (3) if the words "evaluate," 223 "assess," or "rule out" appeared in a sentence with "cirrho," that sentence was ignored for text 224 search purposes. A gastroenterologist (V.L.C.) manually reviewed 200 randomly-selected text 225 strings and identified no false positive cirrhosis diagnoses. 226

227

#### 228 Statistical analysis: non-genetic

229 Differences in characteristics between participants with and without NAFLD were determined

using a t-test for continuous variables and chi-square test for proportions.

231

For the graphs of % NAFLD vs. central fat mass, we computed sex-specific percentiles of central body fat mass and identified the percentage of participants within each percentile with NAFLD, as defined by LPR < 0.33. These percentages were then stratified separately by high vs. low appendicular lean mass or lower extremity fat mass (i.e. above vs. below sex-stratified median). Univariable linear regression was performed on % NAFLD vs. central fat mass and graphed as a smoothed linear model (Fig. 2). Addition of quadratic terms did not improve the regression (p >0.05 for all comparisons).

239

240 Multivariable linear regression analysis was performed to determine the relationship between

liver steatosis (as measured by negative LPR) and measures of body composition and strength.

- 242 For these analyses negative LPR (increased liver steatosis) was treated as the dependent
- 243 (outcome) variable. Note that a higher LPR is associated with decreased liver steatosis, so that

- *positive* beta values for covariates actually imply *decreased* liver steatosis. Since this is counterintuitive, to increase readability, we used negative LPR as the dependent variable, so that 245
- positive beta values imply *increased* liver steatosis. The primary independent (exposure) 246
- variables were (1) appendicular lean mass, (2) lower extremity fat mass, (3) central fat mass, (4) 247
- grip strength, (5) quadriceps strength, and (6) muscle steatosis (negative muscle attenuation, for 248
- reasons similar to those for LPR, as above). These were inverse normally transformed in order to 249
- improve interpretability and treated as continuous independent variables (18). B-values for body 250
- composition parameters were reported as the effect of one rank unit (1/6<sup>th</sup> of the total variation of 251
- that trait) on LPR. In sensitivity analyses, we ran these regressions with non-transformed 252
- covariates and the results were qualitatively the same (data not shown). Regression analyses 253
- were stratified by sex. Proportion of variation explained by variables was estimated by 254
- 255 comparing sums of squares for individual variables in the model with the total sum of squares.
- 256

244

Analyses were performed using R version 3.4.4 (R Foundation for Statistical Computing, 257

- Vienna, Austria; www.r-project.org) with the tidyverse package (www.tidyverse.org). A two-258
- sided p value of 0.05 was used to determine statistical significance. 259
- 260

#### Genetic analysis 261

Only participants of European ancestry were included in genetic analyses. First, principal 262 components were calculated based on LASER/TRACE (https://laser.sph.umich.edu), using the 263 World imputed reference panel (37). To exclude individuals who did not cluster with the 264 European group, individuals with |Z| score |>3 for any of the first three principal components 265 266 were removed. Then, the principal components were recalculated on remaining individuals using the European panel. 267

268

LPRS was calculated as previously reported (24). In brief, LPRS was the total number of 269 270 disease-causing alleles at each of 53 previously-reported single nucleotide polymorphisms (SNPs) each individual carried. These SNPs were selected based on being associated with 271 272 increased serum insulin, decreased high-density lipoprotein cholesterol, and increased triglycerides; they were tested and shown to associate with decreased lower extremity fat 273 274 indicative of lipodystrophy (24). In cases when the genotype at that SNP was imputed rather than

directly genotyped, we used dose, i.e. probability of having that given genotype at the SNP. First, 275 we calculated in FHS the percentage of participants with NAFLD (defined as LPR  $\leq 0.33$  (38)) 276 as a function of LPRS, and performed logistic regression using proportion NAFLD as the 277 278 dependent variable and number of risk alleles as the independent variable. This was graphed as a smoothed linear model (Fig. 3A). There was no improvement in the model after addition to 279 280 quadratic terms for number of risk alleles. Next, LPRS was used as an independent variable for phenotypes including the continuous traits of hepatic steatosis (negative LPR) and APRI, as well 281 282 as the binary traits of cirrhosis or NAFLD. These models were adjusted for age, age<sup>2</sup>, and the first ten principal components (to account for ethnic differences) and either stratified by or 283 adjusted for sex as well. 284

285

### 286 **Results**

287 *Study population* 

Fig. 1 illustrates FHS participant selection for this study. Data on CT-measured hepatic steatosis, whole body DXA scan, quadriceps and hand grip strength, physical activity index, and clinical

examination were available from 1,389 individuals from Generation 3 and 1,032 from Offspring.

After excluding individuals with excess alcohol intake (Methods), 1,300 individuals from

Generation 3 and 949 individuals from Offspring remained for a total of 2,249 individuals.

293

Overall, the cohort was 49% male with mean age  $58.5 \pm 11.8$  years (Supp. Table 1). Prevalence

of NAFLD was 28.3%. Table 1 shows clinical parameters stratified by presence vs. absence of

296 NAFLD. Participants with NAFLD were older and more frequently male, and had higher

297 prevalence of diabetes, hypertension, and the metabolic syndrome, as well as expected

differences in biochemical profiles (p < 0.05 for all comparisons).

299

300 Body composition and muscle strength

Table 1 depicts body composition and muscle strength in FHS participants with or without

302 NAFLD in univariate analyses. In the overall cohort, participants with NAFLD had greater

amounts of total fat and central fat, and smaller amounts of total lean mass and appendicular (i.e.

arms and legs) lean mass (Table 1; p < 0.0001 for all). There was no difference in lower

extremity fat mass (p = 0.26). Grip strength was greater in participants with NAFLD (p = 0.007)

while there was no difference in quadriceps strength based on NAFLD status (*p* = 0.92, Table 1).
Sex-stratified analysis was fairly similar overall. However, among women, grip strength no
longer differed based on NAFLD status (Supp. Table 2). Among men, lower extremity fat was
higher and grip strength lower in those with NAFLD(Supp. Table 3).

310

#### 311 *Effect of body composition on NAFLD and fibrosis*

Next, we sought to identify whether differences in body composition associated with increased 312 risk of NAFLD in FHS. Because central fat, lower extremity fat, and appendicular lean mass are 313 correlated with one another, we investigated whether they independently affected hepatic 314 steatosis after adjustment for one another. Fig. 2 shows percentage of participants with NAFLD 315 as a function of percentile of central fat mass, stratified by sex and either lower extremity fat 316 mass or appendicular lean mass status. In all analyses, higher central body fat associated with 317 greater NAFLD prevalence (p < 0.0001). High lower extremity body fat associated with lower 318 NAFLD prevalence in both men and women (p < 0.05). High appendicular lean mass associated 319 with lower NAFLD prevalence in men (p < 0.05), but not in women (p = 0.16). 320

321

We then performed multivariable linear regressions to determine whether body composition 322 323 parameters independently associated with hepatic steatosis as a continuous variable in FHS. We used as minimal covariates in all models age, physical activity, alcoholic drinks per week, and 324 325 cohort. On multivariable analysis, in both men and women, greater central fat mass associated with more liver steatosis, while greater lower extremity fat mass associated with less liver 326 327 steatosis (Table 2 and Supp. Table 4). In men, but not in women, greater appendicular lean mass associated with less hepatic steatosis (Table 2 and Supp. Table 4). In both sexes, greater 328 329 paraspinal muscle fat associated with increased hepatic fat (Table 2 and Supp. Table 4). These 330 findings persisted in models adjusting for minimal covariates, central fat mass, lower extremity fat mass, appendicular lean mass, and either hand grip strength, quadriceps strength, or muscle 331 attenuation (Supp. Table 4). In no model did quadriceps strength or hand grip strength associate 332 333 with hepatic steatosis (Supp. Table 4).

334

For reference, among women, one standard deviation of liver steatosis corresponds to an LPR of0.06. Thus, the fact that in women each rank unit of central fat mass was associated with a

change in liver steatosis of 0.02 implies significant explanatory power. Among men, the liver

steatosis standard deviation was 0.07, and the  $\beta$  values associated in men with each inverse-

normalized unit of central fat mass (0.03), lower extremity fat (-0.01), and appendicular lean

mass (-0.01) were relatively large. The combination of central fat, lower extremity fat,

appendicular lean mass, and muscle attenuation accounted for 14.4% of variation in liver

342 steatosis in women and 18.1% in men.

343

344 We also tested whether these fat depots associated with APRI, a noninvasive marker of fibrosis,

in FHS. In men, greater central body fat associated with increased APRI: each rank unit increase

was associated with 0.031 increase in APRI (95% CI 0.003-0.061). There were no other

347 associations between body composition and APRI in men. In women, there was no association

348 between body composition and APRI.

349

350 Genetic lipodystrophy risk score

351 We further explored whether genetic predisposition to partial lipodystrophy influences liver-

related phenotypes in FHS and MGI, using a lipodystrophy polygenic risk score (LPRS)

353 (Methods). In FHS, mean (SD) LPRS was 52.8 (4.3) and in MGI, 55.1 (4.6). Participants with

NAFLD had higher LPRS than those without NAFLD (53.2 vs. 52.7, p = 0.006). We validated that, consistent with conferring a partial lipodystrophy phenotype, higher LPRS associated with dyslipidemia and insulin resistance and decreased lower extremity fat (p < 0.001 for all), but did not affect central fat or overall body mass index (Fig. 3A). These findings held when men and

- 358 women were analyzed separately (Supp. Figs. 1-2).
- 359

360 Unadjusted NAFLD prevalence increased significantly with increasing LPRS in the overall

cohort (Fig. 3B; p < 0.005). This association remained in women (p < 0.005) but not in men (p =

0.16; Supp. Figs. 1 and 2). On multivariable linear regression, higher LPRS was associated with

increased liver steatosis in the overall FHS cohort (Fig. 3A) and among women (Supp. Fig. 1A).

In men, LPRS did not associate with liver steatosis but the trend was in the same direction as in

women (Supp. Fig. 2A). Adjusted odds ratio for NAFLD per allele of LPRS in the overall cohort

was 1.04 (95% CI 1.01-1.06); overall, individuals in the 90<sup>th</sup> percentile for LPRS were 36% more

367 likely to have NAFLD than those in the 10<sup>th</sup> percentile.

368

Finally, we examined the effect of the LPRS on liver fibrosis. We first performed linear 369 370 regression with APRI as the dependent and LPRS as the independent variable in FHS, but the association was not significant. Therefore, we tested this hypothesis in a hospital based cohort, 371 the Michigan Genomics Initiative (MGI) (n = 19,239). In the overall MGI cohort, each allele of 372 LPRS was associated with OR 1.02 for cirrhosis diagnosis (95% CI 1.00-1.03, p = 0.03; Fig. 373 3C). Individuals in the 90th percentile of LPRS were 22% more likely to have cirrhosis than those 374 in the 10<sup>th</sup> percentile. Adjusted OR among men was 1.02 (95% CI 1.00-1.04, p = 0.03; Supp. Fig. 375 2C) and among women there was no significant association though consistent direction of effect 376 (OR 1.01, 95% CI 0.99-1.03; Supp. Fig. 1C). After adjustment, higher LPRS was associated with 377 greater APRI (Fig. 3A), indicating increased fibrosis. While the association was not significant 378 379 when men and women were analyzed separately, the directions of effect trended in the same direction (p = 0.06 and 0.10 in women and men, respectively; Supp. Figs. 1-2). 380 381

### 382 Discussion

In summary, we show that greater central fat mass associated with increased hepatic steatosis, while greater lower extremity fat mass and appendicular lean mass associate with less hepatic steatosis. In addition, greater paraspinal muscle fat was associated with increased hepatic steatosis. Overall, these four body composition parameters accounted for a substantial proportion of variation in hepatic steatosis: 14% in women and 18% in men. Finally, higher LPRS led to increased hepatic steatosis and fibrosis in the population, with a 36% and 22% increased risk, respectively, in individuals with high vs. low LPRS.

390

Our findings suggest that NAFLD may be a marker of partial lipodystrophy in the population. Lipodystrophy is classically thought of as a rare monogenic disease, but partial lipodystrophy (or even differences in fat depot distribution) may exist as a continuous trait in the population (21, 24). We found that LPRS associates with increased hepatic steatosis and fibrosis. Further, among participants with NAFLD, non-obese participants had a lower appendicular fat than did obese subjects. Together, these findings imply that inadequate appendicular adipose tissue may contribute to NAFLD and fibrosis.

398

Interestingly, we found LPRS associates with increased hepatic steatosis in women but not men. 399 This may be because men have lesser lower extremity fat (7.7% vs. 13.2%) and consequently 400 401 greater total lean mass (67% vs. 56%) and appendicular lean mass (31% vs. 24%) than women. 402 We note that while there was no statistical association between appendicular lean mass and hepatic steatosis in women, the direction of effect was the same in both men and women 403 404 suggesting that in both genders muscle mass is protective against NAFLD. Thus, it may be that in men muscle may be able to buffer excess calories more than in women who have less mass 405 and thus use fat to buffer excess calories. A genetic decrease in lower extremity fat may 406 therefore confer proportionally greater risk for NAFLD in women than in men who already have 407 a small amount of lower extremity fat depot. 408

409

We also found that muscle steatosis associates with NAFLD, likely because when excess energy cannot be buffered by adipose tissue it may be stored in ectopic fat depots such as muscle and liver. Unlike muscle steatosis however, muscle strength as measured by quadriceps and hand grip strength did not in this study correlate with NAFLD. Thus, while it appears that muscle fat is associated with increased hepatic steatosis and muscle mass with decreased steatosis, muscle strength does not appear to associate with NAFLD.

416

Consistent with earlier findings, in our study greater central (visceral) adiposity associates with 417 418 increased prevalence of NAFLD while greater lower extremity adiposity associates with decreased NAFLD prevalence (10, 39). The mechanisms underlying these differences in disease 419 420 risk based on fat location remain incompletely-characterized but may relate to differences in macrophage and cytokine profiles in visceral fat and direct blood flow from visceral fat to the 421 422 liver via the portal circulation (40, 41). In both men and women, body composition metrics explained a substantial proportion of variation in hepatic steatosis (18% and 14%, respectively). 423 Additional studies will be required to better understand the biology underlying these 424 relationships. 425

426

427 Our study is limited by including only participants of European ancestry. The association

428 between hepatic steatosis and LPRS may only reflect an association with one particular form of

429 lipodystrophy and may not be generalizable to all lipodystrophy. Finally, DXA cannot

distinguish between subcutaneous lower extremity fat and deeper lower extremity fat layers, or
between muscle and other lean tissues such as skin and connective tissue, though there is no

432 clear pathophysiologic reason non-muscle lean tissue would be related to NAFLD.

433

434 Strengths of the study include that it is a large, population-based study, which increases the 435 generalizability of our findings. In addition, CT and DXA are excellent quantitative noninvasive 436 measurements of hepatic steatosis and body composition, respectively, allowing rigorous testing 437 of how body composition relates to NAFLD. Full genotypic information was available for 438 genetic analysis. We also were able to assess for effects of the LPRS on fibrosis using two 439 independent methods.

440

In conclusion, we demonstrated a novel association between partial lipodystrophy and liver steatosis and fibrosis in the population. We also report a connection between CT-measured hepatic steatosis and muscle steatosis, and to our knowledge this is the first such report in a Caucasian population. These results suggest that interventions directed at increasing muscle quantity, decreasing overall fat burden, or shifting fat distribution toward appendicular fat may be beneficial in reducing NAFLD and preventing its complications.

447

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### 572 Figure Legends

- 573
- **Figure 1**: Study design flowchart. CT, computed tomography. DXA, dual-energy X-ray
- absorptiometry. PAI, physical activity index.
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**Figure 2**: Effect of central body fat, lower extremity body fat, and appendicular lean mass on

- 578 nonalcoholic fatty liver disease prevalence. Percentage of participants with nonalcoholic fatty
- 579 liver disease (NAFLD) based on sex-specific percentile of central fat mass. (A-B) Stratified
- based on lower extremity fat (LEF) mass above or at the median ("high") vs. below the median

("low"), for women (A) and men (B). (C-D) Stratified based on appendicular lean mass (ALM) 581 above or at the median ("high") vs. below the median ("low"), for women (C) and men (D). 582 583 Shaded areas represent 95% confidence intervals.

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Figure 3: Effect of LPRS on multiple traits. (A) Percentage of FHS participants with NAFLD 585 based on number of LPRS alleles. (B) Percentage of MGI participants with cirrhosis based on 586 number of LPRS alleles. The shaded area represents the 95% confidence interval. (C) Forest plot 587 of associations between LPRS and multiple traits. The scale on the x axis is the allele effect size 588 (B) of one LPRS allele, divided by the standard deviation of the specific parameter; i.e. what 589 proportion of standard deviation is accounted for by each additional allele of LPRS. Error bars 590 depict 95% confidence interval. Liver steatosis represents negative liver-phantom ratio. Muscle 591 steatosis represents negative muscle attenuation in Hounsfeld units. All traits except APRI were 592 593 measured in FHS; APRI was measured in MGI. HDL, high-density lipoprotein. TRIG, triglycerides. LDL, low-density lipoprotein. HOMA-IR, homeostatic model of insulin resistance. 594

BMI, body mass index. 595

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# Table 1: Clinical and laboratory characteristics

Characteristic	Overall	No NAFLD	NAFLD	P value for no NAFLD	
	(n = 2249)	(n = 1613)	(n = 636)	vs. NAFLD	
Age (years)	58.5 (11.8)	58.1 (11.8)	59.3 (11.6)	0.028	
Male (%)	48.6%	44.8%	58.2%	< 0.0001	
Hypertension (%)	31.8%	25.6%	47.3%	< 0.0001	
Diabetes (%)	14.9%	9.8%	27.5%	< 0.0001	
Metabolic syndrome (%)	26.1%	15.2%	50.9%	< 0.0001	
Body mass index (kg/m <sup>2</sup> )	28.1 (5.0)	26.9 (4.4)	31.1 (5.1)	< 0.0001	
Systolic blood pressure	120.2 (15.0)	110 5 (15 0)	124 4 (15 2)	- 0.0001	
(mmHg)	120.2 (15.9)	118.5 (15.8)	124.4 (15.5)	< 0.0001	
Diastolic blood pressure	74.2 (0.2)	72.2(0.0)	76.0 (0.6)	< 0.0001	
(mmHg)	74.3 (9.2)	/3.3 (8.9)	76.9 (9.0)		
Hemoglobin (g/dL)	13.9 (1.3)	13.8 (1.3)	14.0 (1.3)	0.001	
Platelets $(10^9/L)$	239.4 (61.2)	239.0 (62.1)	240.3 (59.2)	0.67	
Hemoglobin A1c (%)	5.5 (0.5)	5.5 (0.4)	5.8 (0.8)	< 0.0001	
Creatinine (mg/dL)	0.91 (0.23)	0.90 (0.23)	0.93 (0.23)	0.007	
Fasting glucose (mg/dL)	99.8 (19.1)	97.0 (15.9)	106.8 (24.0)	< 0.0001	
Total cholesterol (mg/dL)	187.1 (36.0)	188.4 (35.0)	183.9 (38.0)	0.01	
Triglycerides (mg/dL)	117.0 (75.3)	103.6 (54.7)	150.9 (104.2)	< 0.0001	
High-density lipoprotein	$c_0 \circ (10 A)$	(2,7,(10,5))	52.0.(1.00)	. 0. 0001	
(mg/dL)	60.0 (18.4)	62.7 (18.5)	53.0 (16.2)	< 0.0001	
Alanine aminotransferase	24.2(14.7)	(12.0)	20.0.(1.7)	. 0. 0001	
(U/L)	24.2 (14.7)	21.9 (13.2)	29.9 (10.7)	< 0.0001	

Aspartate aminotransferase	225(10.2)	21.9(10.4)	24.1(0.8)	< 0.0001
(U/L)	22.5 (10.3)	21.8 (10.4)	24.1 (9.8)	< 0.0001
Total bilirubin (mg/dL) 0.49 (0.27)		0.49 (0.26)	0.50 (0.30)	0.26
Gamma-glutamyltransferas	se	25 5 (22 5)	40.2(40.6)	< 0.0001
(U/L)	29.7 (33.7)	25.5 (25.5)	40.3 (49.0)	< 0.0001
Albumin (g/dL)	4.5 (0.3)	4.5 (0.3)	4.5 (0.3)	0.63
Liver-phantom ratio	0.34 (0.06)	0.37 (0.03)	0.27 (0.07)	< 0.0001
NAFLD, nonalcoholic fatty	y liver disease			
Table 2: Body composition	and strength metrics			
Characteristic	Overall	No NAFLD	NAFLD	P value for no NAFLD vs.
	(n = 2249)	(n = 1613)	(n = 636)	NAFLD
Total body fat (kg)	27.5 (10.2)	25.4 (9.1)	33.0 (10.7)	< 0.0001
Central body fat (kg)	15.8 (5.7)	14.4 (5.1)	19.6 (5.8)	< 0.0001
Lower extremity fat (kg)	8.4 (3.6)	8.0 (3.4)	9.4 (4.1)	< 0.0001
Total lean mass (kg)	48.2 (11.5)	46.7 (11.1)	52.0 (11.7)	< 0.0001
Appendicular lean mass	21.4(5.0)	20.8(5.8)	22.2(6.1)	< 0.0001
(kg)	21.4 (3.9)	20.8 (3.8)	23.2 (0.1)	
Total body fat/weight	24.40(0.2)	22.5% (0.4)	26.00/ (8.5)	< 0.0001
(kg/kg*100%)	34.4% (9.3)	55.5% (9.4)	30.970 (8.3)	
Central body fat/weight	10.70/ (4.8)	18 00/ (4 0)	21.80/(4.0)	< 0.0001
(kg/kg*100%)	19.7% (4.8)	18.9% (4.9)	21.8% (4.0)	
Lower extremity fat/	10.50/ (2.8)	10,60/ (2,9)	10 40/ (2 7)	0.26
weight (kg/kg*100%)	10.3% (3.6)	10.0% (3.8)	10.4% (3.7)	
Total lean mass/weight	61.1% (9.5)	62.1% (9.6)	58.6% (8.8)	< 0.0001

(kg/kg*100%)							
Appendicular lean mass/	27.00((4.8))		27 40/ (4 8)		2610/(4.5)		< 0.0001
weight (kg/kg*100%)	27.0% (4.8)		27.4% (4.8)		20.1% (4.3)		
Hand grip strength (kg)	35.6 (12.3)		35.1 (12.2)		36.7 (12.4)		0.007
Quadriceps muscle	25.5 (8.6)		25 5 (9 5)	25.5(0,0)	0.92		
strength (kg)			23.3 (8.3)		23.3 (9.0)		
Muscle attenuation	49.2 (7.3)		<b>70</b> 0 ( <i>C</i> 0)				< 0.0001
(Hounsfield units)			50.0 (6.8)		47.4 (7.9)		
NAFLD, nonalcoholic fatty liver disease							
Table 3: linear regression on	hepatic steatosi	S					
Daramatar	Beta coefficien	t					
	Women	Men					
Central fat index	0.0193 *	0.0272 *					
Lowor oxtromity fat							
Lower extremity fat	-0.0072 *	-0.012 *					
Annondicular loon mass							
Appendicular lean mass	-0.0038 <sup>NS</sup>	-0.0106	*				
index							
Muscle steatosis	0.0013 *	0.0009 *					

Linear regression on hepatic steatosis, as defined as liver-phantom ratio (see Methods for details). Muscle steatosis was defined as negative muscle attenuation (Hounsfeld units). All four traits were inverse normalized. Beta coefficients correspond to the effect of one rank unit (approximately 1/6 This article is protected by copyright. All rights reserved

of the total variation). Analysis was stratified by sex. Covariates were age,  $age^2$ , physical activity index, drinks per week, and cohort (i.e. Offspring vs. Generation 3), and the four parameters in the above table. NS, not significant; \* p < 0.05

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