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10	Feedbacks between nitrogen fixation and soil organic matter increase ecosystem functions in
11	diversified agroecosystems
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- 32 Abstract

33 Nitrogen (N) losses from intensified agriculture are a major cause of global change, due to nitrate (NO_3) export and the eutrophication of aquatic systems as well as emissions of nitrous oxide (N_2O) 34 into the atmosphere. Diversified agroecosystems with legume cover crops couple N and carbon (C) 35 inputs to soil and reduce N pollution, but there is a need to identify controls on legume N₂ fixation 36 37 across ecosystems with variable soil conditions. Here, I tested the hypothesis that N mineralization from turnover of soil organic matter (SOM) regulates legume N₂ fixation across 10 farms that 38 spanned a gradient of SOM levels. I separated soil samples into two SOM fractions, based on size 39 and density, which are indicators of soil nutrient cycling and N availability (free particulate organic 40 matter and intra-aggregate particulate organic matter (POM)). This study indicates downregulation 41 of legume N₂ fixation in diversified agroecosystems with increasing N availability in intra-42 aggregate POM and increasing N mineralization. Intercropping the legume with a grass weakened 43 the relationship between N in POM and N₂ fixation due to N assimilation by the grass. Further, 44 45 mean rates of N and C mineralization across sites increased with two seasons of a legume-grass 46 cover crop mixture, which could enhance this stabilizing feedback between soil N availability and 47 N₂ fixation over time. These results suggest a potential mechanism for the diversity-ecosystem function relationships measured in long-term studies of agroecosystems, in which regular use of 48 legume cover crops increases total soil organic C and N and reduces negative environmental 49 impacts of crop production. 50

51 Key words: agroecosystem, biological nitrogen fixation, cover crop, legume, mineralization,
52 particulate organic matter, soil carbon, soil organic matter

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56 Introduction

Nitrogen (N) fertilizer use in agricultural landscapes is a major driver of global change.
Increases in reactive N production, dominated by agricultural intensification (Galloway et al. 2008),
and N applications that exceed plant need, contribute to eutrophication of surface waters and to

greenhouse gas emissions (Diaz and Rosenberg 2008, IPCC 2013). Despite decades of research and 60 61 technological development to improve N fertilizer use efficiency, N pollution is a persistent environmental problem (Diaz and Rosenberg 2008). Applying ecological principles to manage plant 62 63 diversity in agroecosystems for functions such as N retention may offer greater potential to mitigate pollution. For example, agroecosystems with legume N₂ fixation as a primary N source can reduce 64 65 N losses (Drinkwater et al. 1998, Syswerda et al. 2012) and sequester carbon (C) in soil (McDaniel et al. 2014, King and Blesh 2018). Given the potential for legume N sources to alleviate 66 environmental impacts, there is a critical need to understand patterns of, and ecological controls on, 67 68 legume N₂ fixation across variable agroecosystem conditions.

An ecological perspective offers insights into the mechanisms driving N losses from 69 intensified agriculture. Inorganic N fertilizers are vulnerable to leaching and gaseous losses, even 70 71 using best management practices, because they are applied in soluble forms that can be rapidly lost 72 if they exceed crop demand. Intensively managed agroecosystems typically have low stocks of soil organic matter (SOM), which exacerbates N losses because microbial processes that can cycle and 73 store N in soil are C limited (Drinkwater and Snapp 2007). In contrast, use of organic N sources, 74 75 particularly legume cover crops (i.e., non-harvested crops), offers greater promise for reducing N 76 losses (Syswerda et al. 2012, Blesh and Drinkwater 2013), because opportunities for microbial N 77 assimilation are increased by providing a high quality C input to soil (Fisk et al. 2015). Legume 78 cover crops also improve N retention by increasing the temporal diversity of crop rotations and 79 taking up N during periods when soil would otherwise be bare. For example, in a meta-analysis, Tonitto et al. (2006) showed that legume cover crops reduced NO_3^- leaching by 40%, compared to 80 81 the absence of a cover crop. A long-term (11 yrs) experiment in Michigan revealed that NO_3^{-1} 82 leaching was reduced by more than 60% in two management systems with greater biodiversity and legume N sources (Syswerda et al. 2012). In addition, use of legume cover crops as N sources can 83 reduce N₂O emissions compared to bare fallows (Han et al. 2017). Legume cover crops also 84 increase the spatial diversity of agroecosystems when planted in multi-species mixtures. When 85 legumes are combined with grass species, which are even stronger N scavengers, they can further 86 87 reduce NO₃ leaching (Kaye et al. 2019) and N₂O emissions (Basche et al. 2014). Overall, cover crops are considered central to an ecological approach to agroecosystem management. 88 Legume cover crops can sequester atmospheric CO₂ by increasing C inputs to 89 90 agroecosystems and building stocks of SOM (Poeplau and Don 2015, King and Blesh 2018). In some long-term experiments, management systems with legume cover crops do not have greater net 91

C inputs to soil compared to lower diversity rotations such as continuous grains, yet the diversified 92 93 systems still accumulate more SOM over time (Drinkwater et al. 1998, Grandy and Robertson 2007). Emerging understandings of soil C stabilization suggest a potential mechanism for these 94 findings (Cotrufo et al. 2013). Legume residues have low C:N ratios and are easily decomposed by 95 96 microbial communities. As a result, the fraction of legume cover crop residue that becomes SOM 97 may be higher if it increases the relative production of microbial byproducts and necromass – key components of stable SOM pools - for a given C input to soil (Schmidt et al. 2011, Cotrufo et al. 98 2013). Cover crops also increase the proportion of the year with plant root-microbe interactions in 99 100 the rhizosphere, and associated release of root exudates, extracellular enzyme production, and 101 presence of aggregate binding agents, which have previously been linked to accumulation of SOM (Tiemann et al. 2015, Lehmann et al. 2017) and to soil nutrient cycling capacity (Clarholm 1985, 102 103 Drinkwater and Snapp 2007, Kuzyakov and Blagodatskaya 2015). In particular, legume cover crops can increase fractions of SOM that turnover more quickly than the total SOM pool, especially 104 particulate organic matter (POM). POM pools respond to changes in management on year to 105 106 decadal scales, correlate with soil N mineralization rates, and are early indicators of longer-term 107 accumulation of SOM (Cambardella and Elliott 1992, Wander 2004, Luce et al. 2016). Legume cover crops can therefore enhance multiple ecosystem functions. 108

109 Despite these benefits, farmers continue to rely on inorganic fertilizers partly because they 110 have more predictable outcomes for crop production. Nitrogen inputs from legume cover crops 111 depend on ecological processes and are inherently more variable. There is a need, then, to increase predictive understanding of legume N₂ fixation inputs from cover crops. In natural ecosystems, key 112 ecological controls on N₂ fixation include high levels of soil inorganic N, low availability of non-N 113 nutrients that may limit legume growth or fixation (e.g., P, Fe, K, Mo), preferential grazing of 114 legumes by herbivores, and shade intolerance of legumes compared to other plant functional groups 115 (Vitousek et al. 2013). Biotic interactions can also influence these mechanisms. For example, the 116 percentage of legume N from the atmosphere is mediated by interactions with grasses. Grasses are 117 strong competitors for soil N, reducing soil inorganic N pools and causing the legume to fix more of 118 its N (Li et al. 2009). However, grasses may also compete with legumes for soil nutrients such as P, 119 120 or resources like light, reducing legume biomass production (Cech et al. 2010, Poffenbarger et al. 2015). In agroecosystems, which typically have lower diversity than natural ecosystems, many of 121 these constraints are alleviated through management. However, nutrients such as P or Fe can limit 122 legume biomass production, and it is well known that inorganic N fertilizers inhibit legume N₂ 123

- fixation because of the greater C cost for acquiring N from symbiotic N₂ fixation compared to
 assimilating soil inorganic N (Schipanski et al. 2010, Gelfand and Robertson 2015).
- In diversified agroecosystems, SOM pools including POM are central to soil nutrient cycling 126 dynamics, yet most research on N₂ fixation in agriculture has focused on inorganic N supplied by 127 128 synthetic fertilizer, while the role of N mineralized from decomposition of SOM remains poorly 129 understood (Schipanski and Drinkwater 2011). Diversified agroecosystems that rely on legumes and other organic N sources have different N cycling dynamics from fields with synthetic fertilizers, 130 which experience episodic inputs of inorganic N that saturate the root zone. Organic and diversified 131 farms have larger N inputs than natural ecosystems, to balance the N removed in harvested crops, 132 but inorganic N is released more gradually in the rhizosphere through microbial turnover of SOM. 133 Further, N cycling rates differ among farms with SOM of differential quantity and quality. For 134 instance, agroecosystems with a long organic management history may have low standing pools of 135 inorganic N because of tight coupling between plant and microbial productivity (Paterson 2003), 136 but high mineralization rates due to accumulation of SOM and enhanced nutrient cycling capacity 137 (Berthrong et al. 2013). Given the potential for legumes to build POM pools, and the critical role 138 that SOM pools such as POM play in nutrient cycling, relationships between POM, N 139 mineralization, and legume N₂ fixation are important to understand. 140

The goal of this study was to identify ecosystem-scale controls on legume N₂ fixation in the 141 context of diversified agroecosystems. Determining patterns in N2 fixation across variable soil 142 143 conditions could inform management of legume N sources, and increase understanding of feedback mechanisms that underpin the efficiency of legumes demonstrated by long-term studies. In a field 144 145 experiment on 10 farms spanning a gradient of soil fertility levels due to distinct management legacies, I separated soil samples into two SOM fractions, based on size (> 53 µm) and density, 146 which are indicators of internal nutrient cycling and N availability (light or "free" POM; and POM 147 that is physically-protected inside microaggregates) (Marriott and Wander 2006b). Here, I used 148 these two POM fractions to assess relationships between N availability from turnover of SOM and 149 legume N₂ fixation (measured using stable isotope methods) across a gradient of SOM levels, 150 holding management constant. I hypothesized that legume N₂ fixation would decrease with 151 increasing levels of N in POM, due to enhanced microbial processing of SOM and increased N 152 153 mineralization with larger POM pools (Fig. 1). Furthermore, I hypothesized that intra-aggregate POM should account for more variation in this relationship relative to light fraction POM because it 154 turns over more slowly and is considered a better indicator of longer-term changes in soil N cycling 155

- 156 capacity due to management (Wander et al. 1994, Schipanski et al. 2010). Finally, I expected
- 157 interspecific competition in a legume-grass mixture to reduce the effect of POM N on N_2 fixation,
- due to soil N assimilation by the fibrous root system of the intercropped grass (Fig. 1).
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160 Materials and methods

161 Experimental design

I partnered with 10 vegetable farmers in southeastern Michigan to establish an on-farm 162 experiment identifying patterns in N₂ fixation by a legume cover crop, hairy vetch (Vicia villosa L.), 163 grown alone and in mixture with a grass cover crop, cereal rye (Secale cereale L.), across a soil 164 gradient. The farms had been in organic vegetable production from three to 14 years at the start of 165 the experiment, and analysis of baseline soil properties from each experimental field reflected a soil 166 fertility gradient, which resulted primarily from distinct management histories (Table 1, Table 2). 167 Nine of the 10 fields were Alfisols, and one was a Mollisol. Six of the 10 fields had previous 168 compost inputs from poultry litter or other sources of composted manure. Across farms, total 169 organic C varied from 33.1 to 64.1 Mg ha⁻¹, plant available phosphorus (P) was 8 times greater on 170 the highest P site (112 mg kg⁻¹) than on the lowest (14.3 mg kg⁻¹). The size and N concentration of 171 POM pools also ranged widely, particularly for the protected POM pool, indicating that there were 172 differences in the quality of SOM on the farms. Soil inorganic N pools $(NO_3 - N + NH_4 + N)$ ranged 173 from 2.6-14.4 mg kg⁻¹. 174

175 Management details for the experiment, planted in one field per farm, were determined collaboratively with farmers. In summer 2015, after analysis of baseline soil samples, farmers 176 177 planted the field to a crop in the *Amaryllidaceae* family—either onion or garlic. The *Allium* crop 178 was fertilized with compost inputs on four of the farms, and the remaining farms applied no fertility inputs for this crop. No additional fertility amendments were added to fields for the remainder of the 179 experiment, except for the N inputs from legume N2 fixation. Following crop harvest, fields were 180 divided into four replicate blocks to establish the experimental treatments for the cover crop along 181 with a no-cover crop control. The fields ranged in size from 0.05 to 1.4 ha, but the measurements 182 were constrained to a 130 m² area to minimize variability in field area across farms. Farmers 183 planted the fields in a mixture of cereal rye (seeding rate of 56 kg ha⁻¹) and hairy vetch (seeding rate 184 185 of 25 kg ha⁻¹) between August 21- August 28, 2015. Vetch seeds were inoculated with N-Dure (Verdesian Life Sciences, Cary, NC) at approximately 4 g kg⁻¹ seed. Seeds were surface broadcast 186 and lightly incorporated. The cover crop overwintered, and by mid-May, 2016, farmers mowed and 187

then rototilled the field to incorporate the cover crop, and planted a primary crop in the

189 *Cucurbitaceae* family between May 27 - June 4, 2016. Following crop harvest, the cover crop

190 mixture was planted for a second season between August 30 - September 14, 2016 using the same

191 method as in year one. The cover crop was terminated by mowing and rototillage in May, 2017,

192 following final sampling.

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194 Baseline soil sampling

A composite, baseline soil sample from each experimental field was analyzed to determine 195 initial soil conditions and characterize soil fertility status in spring of 2014 or fall 2015 on all 196 197 participating farms (Table 1, Table 2). Approximately 15-20 soil cores (2 cm diameter by 20 cm depth) were composited per site. I estimated bulk density from the fresh weight of 8 cores per field, 198 which was later adjusted for soil moisture. Soil was processed immediately for soil moisture, 199 determined gravimetrically by drying at 105 °C for 48 hours, and triplicate soil subsamples were 200 sieved to 2 mm for analysis of extractable inorganic N (NO₃⁻ and NH₄⁺) with 2 M KCl. The amount 201 of NH_4^+ and NO_3^- in each sample was analyzed colorimetrically on a discrete analyzer (AQ2, Seal 202 203 Analytical, Mecuon, WI). Triplicate soil subsamples were also sieved to 2 mm for a 7-day anaerobic N mineralization incubation, an indicator of potential N availability from decomposition 204 of organic matter, followed by extraction with 2 M KCl and colorimetric analysis of NH₄⁺. 205 206 Potentially mineralizable N (N-min) was calculated by subtracting the initial amount of NH₄⁺ in the 207 soil from the NH₄⁺ released during the 7-day incubation (Drinkwater et al. 1996).

The POM > 53 µm was isolated from triplicate 40 g subsamples of unsieved, air-dried soil 208 209 using a combined size and density fractionation method. To isolate the light fraction POM (also called free POM), the subsamples were first gently shaken for 1 h in sodium polytungstate (1.7 g 210 cm⁻³), allowed to settle for 16 h, and free POM floating on top of the solution was removed by 211 aspiration. To isolate intra-aggregate POM (i.e., physically-protected POM), the remaining soil 212 213 sample was shaken with 10% sodium hexametaphosphate to disperse soil aggregates and then rinsed through a 53 µm filter (Marriott and Wander 2006b). The material larger than 53 µm was 214 retained, and the protected POM was separated from sand by decanting. The C and N of both POM 215 fractions were measured on an ECS 4010 CHNSO Analyzer (Costech Analytical Technologies, 216 217 Inc.; Valencia, CA). Total soil C and N (to 20 cm) were measured by dry combustion on a Leco TruMac CN Analyzer (Leco Corporation, St. Joseph, MI). Subsamples of ~100g of sieved dried soil 218 219 were analyzed for particle size, pH, Bray-1 P, and K at the A & L Great Lakes Laboratories, Inc.

(Fort Wayne, IN). As an indicator of microbial activity and SOM quality, C mineralization (C-min)
was determined by a short-term aerobic incubation of rewetted soil (i.e., the flush of CO₂ during a
1-day incubation), using a method modified from Franzluebbers et al. (2000) (Appendix S1).

223

224 Legume N_2 fixation

During the cover crop seasons, I established three small plots of 1 m² within each replicate 225 block in each field (i.e., 12 small plots per farm) to estimate biological N₂ fixation by the vetch 226 grown in mixture and monoculture based on ¹⁵N natural abundance (Shearer and Kohl 1986). Soon 227 after establishment of the cover crop mixture (e.g., in late September), all vetch plants were hand-228 229 weeded out of one set of small plots to establish a sole rye treatment, and all rye plants were handweeded out of another set of plots for sole vetch stands. A third small plot was left in the mixture, 230 and the background field was also planted to the vetch-rye mixture, except for the no cover crop 231 control. 232

Aboveground biomass in vetch and vetch-rye mixture treatments was sampled from the small plots between 25 April - 6 May 2016, and between 9 - 18 May 2017, from a 0.25 m² section of each small plot, avoiding plot edges. Plant biomass was cut at the soil surface, separated by species, and dried at 60 °C for 48 h. Dried shoot biomass was first ground in a Wiley Mill to 2mm, and was then pulverized using a cyclone mill and analyzed on a continuous flow Isotope Ratio Mass Spectrometer (Stable Isotope Facility, UC Davis) for total C and N content, and for δ^{15} N. The %N derived from fixation was calculated using the following model:

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%N from fixation =
$$100*((\delta^{15}N_{ref} - \delta^{15}N_{legume}) / (\delta^{15}N_{ref} - B))$$

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where $\delta^{15}N_{ref}$ is the $\delta^{15}N$ signature of the reference plant (cereal rye), $\delta^{15}N_{legume}$ is the $\delta^{15}N$ signature 243 of the legume, and B is defined as the $\delta^{15}N$ signature of a legume when dependent solely on 244 atmospheric N₂ B values were determined by growing vetch in the greenhouse in a N-free medium 245 (Appendix S1). The *B* value for vetch was -0.44. Total N input from vetch N₂ fixation (kg N ha⁻¹) 246 was calculated for mixture and monoculture treatments using aboveground biomass, % N, and %N 247 from fixation. The ¹⁵N natural abundance method requires sufficient enrichment of the plant-248 249 available soil N pool to accurately estimate the % of legume N derived from fixation (i.e., from the atmosphere). There was significant separation in δ^{15} N values between the legume (vetch) and the 250

reference plant (sole cereal rye) in mixture and monoculture treatments (Appendix S1: Fig. S1). For
example, in 2016 the separation in mixture ranged from about 2 - 6 ‰ across sites.

I estimated the quantity of fixed N transferred from hairy vetch to cereal rye in the vetch-rye mixture by comparing the δ^{15} N of rye in mixture with the δ^{15} N of rye in monoculture plots:

% rye N from transfer = $100[(\delta^{15}N_{rye monoculture} - \delta^{15}N_{rye mix})/(\delta^{15}N_{rye monoculture} - C)]$ 255 where C is the δ^{15} N value of the integrated fractionation of δ^{15} N during N₂ fixation in legume roots 256 and between the roots and sampled shoots of the rye. C is typically estimated as the lowest $\delta^{15}N$ 257 value of grass grown in mixture, or 0, whichever is lower. Here, C was set to 0 because all $\delta^{15}N$ 258 values were positive for cereal rye. Transfer of fixed N from vetch to rye can occur through 259 260 multiple mechanisms: (i) turnover of vetch root tissue and assimilation of mineralized N by the neighboring rye; (ii) exudation of soluble N compounds by the vetch and uptake by the rye; and (iii) 261 transfer of N through mycorrhizal associations. The natural abundance method has limitations for 262 quantifying N transfer between legumes and non-legumes because of the assumption that the 263 transferred N has a similar δ^{15} N value to that measured in the shoots of the legume, which may not 264 be valid (Peoples et al. 2015). However, this approach provides a relative estimate of fixed N 265 assimilated by rye in the mixture treatment across sites. 266

267

268 Final soil analysis

269 Between 9 May - 18 May 2017, immediately before terminating the cover crop and the 270 experiment, a final composite soil sample (8-10 cores) was collected from each replicate block on each farm in the cover crop mixture and no cover crop control. The soil samples were analyzed for 271 272 extractable inorganic N pools, N-min and C-min, total C and N, Bray-1 P, and free and protected 273 POM fractions as described for the baseline soil sampling. To determine changes in soil fertility and SOM pools with the cover crop mixture across farms, an effect size was calculated by subtracting 274 the final value for each soil parameter measured in the mixture treatment from the final value 275 measured in the no cover crop control at the May 2017 sampling. 276

277

278 Statistical analysis

Statistical analyses were computed in R (The R Foundation for Statistical Consulting,
Vienna, Austria). To test for differences in vetch N₂ fixation between mixture and monoculture,
one-way ANOVA analyses were performed for each experimental year using the *lme4* package for
linear, mixed-effect models with treatment as a fixed effect, and block nested in farm as a random

- effect. To test the hypothesis that increasing POM N concentrations would decrease vetch N₂
- fixation, I used linear regression to model vetch N_2 fixation in mixture and monoculture in 2016 as
- a function of SOM pools and other properties expected to control legume N_2 fixation (e.g., soil P,
- textural properties), excluding two extreme outliers for vetch N₂ fixation. Model comparisons were
- made by assessing goodness of fit with the Akaike Information Criterion (AIC). After 2016, one of
- the participating farmers decided to leave farming, so there were 10 farms in year one (2016) and
- nine farms in year two (2017). Results are reported as statistically significant at $\alpha = 0.05$.
- 290

291 **Results**

292 Biological N₂ fixation in mixture and monoculture

Using stable isotope methods, the mean percentage of vetch N from fixation across all farms 293 was high in both cover crop seasons, ranging from 76 - 92%, and was significantly higher in 294 mixture than in monoculture (Fig. 2). In 2017, the mean % of vetch N from fixation was higher than 295 in 2016, and in 2017 vetch produced approximately half as much biomass compared to the 2016 296 vetch cover crop. The difference in vetch biomass production between years was reflected in mean 297 total aboveground N accumulation in the treatments (Fig. 2), which was 103 and 109 kg ha⁻¹ in 298 vetch mixture and monoculture in 2016, respectively, but 69 and 56 kg ha⁻¹ in mixture and 299 monoculture in 2017. In 2016, total mean N assimilation by rye in mixture was 51.5 kg N ha⁻¹ 300 (range of 16.6 - 111.7 kg N ha⁻¹ across sites), with an estimated 4.6 kg ha⁻¹ of that N derived from 301 302 vetch N₂ fixation, on average. In 2017, the rye grown in mixture assimilated 39.9 kg N ha⁻¹, on average, with an estimated 7.6 kg ha⁻¹ of the N transferred from vetch. Overall, total N 303 304 accumulation between the mixture and monoculture treatments was not significantly different in 305 either year; however, total N supply from N₂ fixation was significantly greater in the sole legume stand due to greater legume biomass than in the mixed stand. 306

- Across farms, aboveground vetch N₂ fixation in 2016 ranged from 1.4 to 177 kg N ha⁻¹ in mixture and from 16 to 232 kg N ha⁻¹ in monoculture, and was highly correlated with vetch aboveground biomass ($R^2 = 0.94$ in mixture, and 0.74 in monoculture). In 2017, vetch N₂ fixation in mixture varied from 1.5 to 78 kg N ha⁻¹, and from 3 to 126 kg N ha⁻¹ in monoculture, across individual farms. The relationships between legume shoot biomass and total fixed N in shoots were similarly strong in 2017 ($R^2 = 0.92$ in mixture and 0.86 in monoculture).
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314 Controls on N_2 fixation across the soil gradient

- Linear regression models using the baseline soil properties as predictors, and vetch N₂ 315 316 fixation in 2016 as the dependent variable, showed that two of the soil properties were significant predictors of the amount of shoot N fixed (in kg N ha⁻¹) by vetch (Appendix S1: Table S1). N 317 concentration in the free POM pool was not a significant predictor of N_2 fixation (P = 0.294 for sole 318 vetch; P = 0.421 for vetch in mixture), however, I found a significant and negative correlation 319 between the N concentration in the protected POM pool and vetch N₂ fixation in mixture with a 320 regression coefficient of -44.16 (P = 0.002), and also in monoculture (-41.39, P = 0.026). The 321 amount of shoot N fixed by vetch was positively correlated with plant available P concentration 322 across the farm sites (P = 0.006 in mixture, and P < 0.001 in monoculture). Together, these soil 323 324 properties explained 42% (sole vetch) and 47% (vetch mixture) of the variation in vetch N₂ fixation. For vetch in mixture, the best fit model had soil P, and the C:N ratio of the protected POM pool 325 (9.32, P < 0.001) rather than POM N concentration. This model explained 54% of the variation in 326 vetch N₂ fixation in mixture; however, the C:N of the protected POM was only a marginally 327 significant predictor of vetch N_2 fixation in monoculture (P = 0.05). 328 There was also a significant, negative relationship between the N concentration of the 329
- protected POM pool and the % of vetch shoot N derived from fixation (Fig. 3). The slope of this 330 relationship was steeper for sole vetch than for vetch grown in mixture (Fig. 3). For sole vetch, the 331 % N from fixation also exhibited a significant and negative relationship with N-min, although it was 332 weaker than the relationship with POM (Appendix S1: Fig. S2; $R^2 = 0.11$; P = 0.04), and there was 333 no relationship between N-min and % N from fixation for vetch in mixture (P = 0.617). In contrast 334 to the positive relationships between soil P and the amount of fixed N in vetch shoots in mixture 335 336 and monoculture (i.e., driven by biomass production), there was no relationship between soil P and 337 the % of vetch above ground N derived from fixation (Appendix S1: Fig. S3) for either mixture (P =(0.793) or monoculture treatments (P = 0.612) 338
- 339

340 Changes in labile organic matter pools with a legume-grass mixture

Finally, I asked how two seasons of the overwintering vetch-rye cover crop mixture
influenced the indicators of soil fertility measured at baseline (Fig. 4). There was a significant
increase in mean C and N mineralization rates compared to the no cover crop control across farms.
The mean size of both POM pools (free and protected POM) also increased, as did the N
concentration of the protected POM pool (shown as a significant decrease in C:N ratio). At the final
soil sampling across farm sites, there was a strong positive relationship between protected POM N

and N-min (Fig. 5), where POM N accounted for half of the variation in potentially mineralizable N. There were also weaker, but significantly positive relationships between both POM pools (in g kg dry soil⁻¹) and N-min (free POM, R²=0.33; P<0.0001; and protected POM, R² = 0.16, P = 0.007). Mean soil NH₄⁺ and NO₃⁻ concentrations, and plant available P concentration, did not change with the cover crop.

352

353 Discussion

Strategic management of plant diversity in agroecosystems for increased ecosystem 354 functioning (Storkey et al. 2015, Finney and Kaye 2017, Blesh 2018) could help alleviate the 355 widespread environmental costs of intensified agriculture. Use of legume cover crops can build soil 356 organic C and N (Drinkwater et al. 1998, King and Blesh 2018), thereby supplying N to crops while 357 also sequestering atmospheric CO₂, and reducing NO₃⁻ leaching to aquatic systems and N₂O 358 emissions. Given that legume N₂ fixation varies with soil properties and interspecific interactions, 359 predictive understanding of this variation is needed to inform managing legumes as a N source for 360 crop production. Physiological controls on N₂ fixation have been fairly well characterized, but 361 362 ecological controls (i.e., controls influenced by interactions between legumes and other organisms, or between legumes and the environment) are less well understood (Vitousek et al. 2013), 363 364 particularly in diversified agroecosystems. For example, numerous greenhouse and field 365 experiments have shown that increasing inorganic N reduced nodulation and legume N₂ fixation 366 (e.g., Van Kessel and Hartley 2000, Goss et al. 2002, Salvagiotti et al. 2008). However, there remains a lack of studies across gradients of SOM levels (cf. Schipanski et al. 2010, Blesh 2018)-a 367 368 critical gap because diversified agroecosystems do not have pulse additions of inorganic N but instead have more gradual release of N through decomposition of SOM (Schipanski and Drinkwater 369 2011). This study shows a strong stabilizing feedback between N in intra-aggregate SOM (protected 370 POM) and N₂ fixation, due to increased N mineralization from enhanced microbial activity fostered 371 by greater organic matter inputs to soil. These results increase understanding of ecosystem-scale 372 rates and regulation of N₂ fixation across working agroecosystems. 373

As predicted, with increasing N concentrations in protected POM across the gradient, I found a significant and negative relationship with both the % of vetch N from fixation, and total fixed N in shoots. There was also a significantly positive relationship between POM N concentration and N-min, and a stabilizing feedback between N-min and % N from fixation for vetch in monoculture, supporting the hypothesis that POM increases N mineralization from 379 microbial processing of SOM. These results highlight a potential mechanism by which legume N 380 sources in diversified agroecosystems better reduce surplus N and N losses (Drinkwater et al. 1998, 381 Blesh and Drinkwater 2013), by responding to changes in SOM following transitions from conventional to diversified management (Fig. 1). Protected POM N moderated vetch N₂ fixation in 382 383 sole and mixed stands, while light fraction POM was not a significant predictor of BNF. This was 384 somewhat surprising because both POM pools are considered indicators of soil fertility. A related experiment on a subset of the same farms showed that the free POM pool was negatively related to 385 legume biomass within different cover crop mixtures, in addition to finding a negative relationship 386 between protected POM N and both legume biomass and shoot N fixed (Blesh 2018). However, 387 results from this study supported the hypothesis that the feedback with protected POM N would be 388 stronger. Free POM turns over relatively quickly and reflects recent C inputs to soil (Wander 2004). 389 Consequently, intra-aggregate POM may be a more robust indicator of soil nutrient cycling capacity 390 because it turns over more slowly and can better reflect longer-term changes in SOM due to 391 management (Marriott and Wander 2006a, von Lützow et al. 2006). Schipanski et al. (2010) 392 measured soybean N2 fixation across a soil gradient, and found weak evidence that the % of 393 394 soybean N from fixation was regulated by soil N availability from POM because soil N availability was secondary to the effect of soil texture on N₂ fixation. In their study, differences in soil texture 395 396 were confounded with management history and the resulting fertility gradient. In this study, 397 although textural properties varied across farms (e.g., from 11-32% clay), there were no significant 398 relationships between % clay and % N from fixation. Furthermore, across farms in this study, there were no significant relationships between sand or clay content and the measured POM pools, 399 400 emphasizing the role of management in driving the amount of POM in soils (Marriott and Wander 401 2006b).

402 The hypothesis that the relationship between POM N and N₂ fixation would be influenced by biotic interactions was also supported. The slope of the relationship between POM N and the % 403 404 of vetch N from fixation was steeper for the sole vetch treatment. Furthermore, the regression relationships between N-min and % N from fixation were significant only for sole vetch and not for 405 vetch in mixture (Appendix S1: Fig. S2). This suggests that the effect of N availability from 406 407 turnover of POM was moderated by growing vetch in mixture with rye (Fig. 3). A previous study 408 testing different cover crop mixture treatments on eight of these farms also found negative correlations between legume biomass production in mixtures (for winter pea and crimson clover) 409 410 and the protected POM N pool; however, in that experiment soil properties did not predict the % of

411 N from fixation (i.e., the N₂ fixation rate) for any species (Blesh 2018). Another study focused on 412 intercropped red clover also found no relationship between labile soil N pools and the % N from fixation (Schipanski and Drinkwater 2011). The lack of relationships with % N from fixation in 413 these other studies could be due in part to intercropping with non-legume species. Here, the 414 intercropped rye competed with the vetch for soil inorganic N, taking up an average of 51.5 kg N 415 ha⁻¹ in 2016 (with an estimated 46.9 kg ha⁻¹ of that N derived from soil), which also caused vetch to 416 fix N at a higher rate in mixture (Fig. 2). In contrast to % N from fixation, the effect of POM N on 417 total fixed N in aboveground vetch biomass (in kg N ha⁻¹) was stronger in mixture than monoculture 418 (Appendix S1: Table S1). This likely occurred because rye was the stronger competitor in mixture 419 at high soil N levels. For instance, across farms, rye aboveground biomass (both total, and as a % of 420 mixture biomass) increased with the protected POM N concentration in soil (F = 10.6, P = 0.002421 and F = 16.8, P = 0.0002, respectively), which reduced vetch biomass—the primary driver of total 422 fixed N inputs (Herridge et al. 2008). 423

Beyond the regulatory effect of N from turnover of POM, legume biomass production across 424 farms, and thus total N₂ fixed, was limited by P and increased with greater soil P concentrations 425 (Appendix S1: Fig. S2). Together, POM N and soil P concentrations explained 42-54% of the 426 variation in vetch N₂ fixation (Appendix S1: Table S1), similar to earlier findings with different 427 428 legume cover crop mixtures on these farms (Blesh 2018). However, there was no evidence that the N₂ fixation process itself is regulated by P availability (i.e., there was no relationship between soil P 429 430 concentration and the % of plant N from fixation), which supports other studies (Vitousek et al. 2002). Although the model R² values were relatively high, especially for data from working 431 432 agroecosystems, there remains substantial unexplained variation that could result from several 433 factors. First, additional variation in N₂ fixation could result from physiological controls at smaller 434 scales, such as soil O₂ availability (Layzell and Hunt 1990). Second, rhizobial strains differ in their effectiveness, and may have differed in vetch nodules across farms, because background soil 435 populations often occupy nodules despite inoculation (Thies et al. 1991). Consequently, applying 436 one B-value across sites, to account for isotopic fractionation, may not fully meet the assumptions 437 of the natural abundance method. This method also assumes that the reference plant (rye) was not 438 affected by free living N₂ fixation in the rhizosphere, a topic of renewed attention in agroecosystem 439 440 research on perennial grasses (Roley et al. 2018, Smercina et al. 2019). However, given the experimental design, in which the reference plants were in small plots nested within a larger field 441

442 inoculated with rhizobia, and given that the farm management histories often included legumes in443 crop rotation, this assumption is likely valid.

Analysis of SOM pools and fluxes following two years of the cover crop mixture showed 444 that, on average, the cover crop increased the size of both POM pools, and rates of microbial 445 446 activity (C-min and N-min; Fig. 4). Further, POM N and N-min had a strong relationship across 447 sites. These dynamics would potentially enhance the stabilizing feedback identified here and increase N cycling efficiency over time. The modeling of vetch N₂ fixation as a function of baseline 448 soil properties – to determine if the POM gradient resulting from distinct management histories 449 regulated N₂ fixation – focused on vetch N₂ fixation in 2016, because soil properties started 450 451 changing quickly in response to the cover crop mixture treatment (Fig. 4). This increase in soil N cycling may also help explain the large reduction in vetch biomass in the second year. These 452 453 changes in soil properties with the cover crop support the hypothesis that the mechanism driving the stabilizing feedback was increased microbial activity and turnover of N with larger pools of intra-454 aggregate POM. 455

456 Results from this experiment suggest that protected POM represents an ecologically meaningful SOM fraction that regulates N₂ fixation in diversified agroecosystems because it is a 457 key source of inorganic N through N mineralization. The POM that is occluded in aggregates often 458 459 has a low C:N ratio, suggesting capacity for both mineralization and stabilization of SOM (Marriott 460 and Wander 2006a, Hatton et al. 2012, Cotrufo et al. 2013). Occlusion within micro-aggregates can 461 stabilize SOM by making it physically inaccessible to decomposers (von Lützow et al. 2006). Although accumulation of SOM with legumes is frequently observed (Fornara and Tilman 2008, 462 463 McDaniel et al. 2014), POM that is protected in soil aggregates may exhibit faster turnover rates in diversified agroecosystems than typically assumed based on experiments finding mean residence 464 times of 10-80 years for occluded POM (Collins et al. 2000, Puget et al. 2000, Dorodnikov et al. 465 2011). Legume cover crops and cover crop mixtures increase the presence of living roots in 466 agroecosystems. Root C inputs can contribute to aggregation and stabilization of SOM, but roots 467 may also cause priming effects and other rhizosphere interactions that increase turnover of stable 468 469 SOM pools, notably by root growth through aggregates or in hotspots with large rhizodeposits 470 (Clarholm 1985, Paterson 2003, Kuzyakov and Blagodatskaya 2015). For instance, plants can 471 increase N mineralization in the rhizosphere (Cheng 2009) by adjusting levels of soluble C compounds released through root exudates (Hamilton and Frank 2001). In this study, the increased 472 473 N and C mineralization rates with the cover crop mixture relative to the no cover crop control; the

negative relationships observed between protected POM N and vetch N₂ fixation, and between N-474 475 min and N₂ fixation; and the positive relationship between POM N and N-min, all suggest that cover crops access N from SOM pools that are considered stable (i.e., protected for decades or 476 longer; Wander 2004; von Lützow et al. 2006) through interactions with microorganisms in the 477 rhizosphere. This finding supports other studies reporting that intra-aggregate POM is related to N 478 479 mineralization rates on organic and diversified farms (e.g., Willson et al. 2001, Marriott and Wander 2006a), which tend to have greater root presence in space and/or time depending on 480 functional diversity of crop rotations. 481

482 There is an urgent need to develop more sustainable agroecosystems to avoid crossing critical biophysical thresholds, including degradation of ecosystem processes on which agriculture 483 depends (IPCC 2013). Central to realizing this goal is increasing plant diversity in agroecosystems, 484 particularly diversity of plant functional traits including N₂ fixation (Isbell et al. 2017, Blesh 2018). 485 More on-farm studies, across even wider gradients of soil and management conditions, are needed 486 to further develop and refine predictive models of N inputs from legume N₂ fixation. Legume cover 487 crops represent a feasible opportunity for increasing agroecosystem biodiversity and ecosystem 488 functions because they can be grown between cultivation of primary crops. Results from this on-489 farm experiment reveal stabilizing feedbacks that help explain the smaller N losses measured in 490 491 agroecosystems with legume N sources. Over time, agroecosystems with regular use of legume 492 cover crops will likely reach a new steady state for SOM, at which point legume N₂ fixation rates 493 may be lower, or legumes may be less competitive in cover crop mixtures, suggesting that farmers could adapt their management towards other functional groups of cover crops. Cover crops could be 494 495 adopted more broadly with appropriate policy incentives to increase agroecosystem diversity and meet environmental sustainability and food security goals. 496

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505 Literature cited

- Basche, A. D., F. E. Miguez, T. C. Kaspar, and M. J. Castellano. 2014. Do cover crops increase or
 decrease nitrous oxide emissions? A meta-analysis. Journal of Soil and Water Conservation
 69:471-482.
- Berthrong, S. T., D. H. Buckley, and L. E. Drinkwater. 2013. Agricultural management and labile
 carbon additions affect soil microbial community structure and interact with carbon and
 nitrogen cycling. Microbial Ecology 66:158-170.
- 512 Blesh, J. 2018. Functional traits in cover crop mixtures: biological nitrogen fixation and
 513 multifunctionality. Journal of Applied Ecology 55:38-48.
- Blesh, J., and L. Drinkwater. 2013. The impact of nitrogen source and crop rotation on nitrogen
 mass balances in the Mississippi River Basin. Ecological Applications 23:1017-1035.
- Cambardella, C., and E. Elliott. 1992. Particulate soil organic-matter changes across a grassland
 cultivation sequence. Soil Science Society of America Journal 56:777-783.
- 518 Cech, P. G., P. J. Edwards, and H. Olde Venterink. 2010. Why is abundance of herbaceous legumes
 519 low in African savanna? A test with two model species. Biotropica 42:580-589.
- Cheng, W. 2009. Rhizosphere priming effect: Its functional relationships with microbial turnover,
 evapotranspiration, and C–N budgets. Soil Biology and Biochemistry 41:1795-1801.
- 522 Clarholm, M. 1985. Interactions of bacteria, protozoa and plants leading to mineralization of soil
 523 nitrogen. Soil Biology and Biochemistry 17:181-187.
- Collins, H., E. Elliott, K. Paustian, L. Bundy, W. Dick, D. Huggins, A. Smucker, and E. Paul. 2000.
 Soil carbon pools and fluxes in long-term corn belt agroecosystems. Soil Biology and
 Biochemistry 32:157-168.
- 527 Cotrufo, M. F., M. D. Wallenstein, C. M. Boot, K. Denef, and E. Paul. 2013. The Microbial
- 528 Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition
- with soil organic matter stabilization: do labile plant inputs form stable soil organic matter?Global Change Biology 19:988-995.
- Diaz, R. J., and R. Rosenberg. 2008. Spreading dead zones and consequences for marine
 ecosystems. Science 321:926-929.
- Dorodnikov, M., Y. Kuzyakov, A. Fangmeier, and G. L. Wiesenberg. 2011. C and N in soil organic
 matter density fractions under elevated atmospheric CO2: turnover vs. stabilization. Soil
 Biology and Biochemistry 43:579-589.

- Drinkwater, L. E., C. A. Cambardella, J. D. Reeder, and C. W. Rice. 1996. Potentially mineralizable
 nitrogen as an indicator of biologically active soil nitrogen. Pages 217-229 Methods for
 assessing soil quality. SSSA Special Publication 49.
- Drinkwater, L. E., and S. S. Snapp. 2007. Nutrients in agroecosystems: rethinking the management
 paradigm. Advances in Agronomy 92:163-186.
- Drinkwater, L. E., P. Wagoner, and M. Sarrantonio. 1998. Legume-based cropping systems have
 reduced carbon and nitrogen losses. Nature 396:262-265.
- Finney, D. M., and J. P. Kaye. 2017. Functional diversity in cover crop polycultures increases
 multifunctionality of an agricultural system. Journal of Applied Ecology 54:509-517.
- Fisk, L., L. Barton, D. Jones, H. Glanville, and D. Murphy. 2015. Root exudate carbon mitigates
 nitrogen loss in a semi-arid soil. Soil Biology and Biochemistry 88:380-389.
- Fornara, D. A., and D. Tilman. 2008. Plant functional composition influences rates of soil carbon
 and nitrogen accumulation. Journal of Ecology 96:314-322.
- Franzluebbers, A., R. Haney, C. Honeycutt, H. Schomberg, and F. Hons. 2000. Flush of carbon
 dioxide following rewetting of dried soil relates to active organic pools. Soil Science Society
 of America Journal 64:613-623.
- 552 Galloway, J. N., A. R. Townsend, J. W. Erisman, M. Bekunda, Z. Cai, J. R. Freney, L. A.
- Martinelli, S. P. Seitzinger, and M. A. Sutton. 2008. Transformation of the nitrogen cycle:
 recent trends, questions, and potential solutions. Science 320:889-897.
- Gelfand, I., and G. P. Robertson. 2015. A reassessment of the contribution of soybean biological
 nitrogen fixation to reactive N in the environment. Biogeochemistry 123:175-184.
- Goss, M., A. de Varennes, P. Smith, and J. Ferguson. 2002. N2 fixation by soybeans grown with
 different levels of mineral nitrogen, and the fertilizer replacement value for a following crop.
 Canadian Journal of Soil Science 82:139-145.
- Grandy, A. S., and G. P. Robertson. 2007. Land-use intensity effects on soil organic carbon
 accumulation rates and mechanisms. Ecosystems 10:59-74.
- Hamilton, E. W., and D. A. Frank. 2001. Can plants stimulate soil microbes and their own nutrient
 supply? Evidence from a grazing tolerant grass. Ecology 82:2397-2402.
- Han, Z., M. T. Walter, and L. E. Drinkwater. 2017. N2O emissions from grain cropping systems: a
 meta-analysis of the impacts of fertilizer-based and ecologically-based nutrient management
 strategies. Nutrient cycling in agroecosystems 107:335-355.

- Hatton, P.-J., M. Kleber, B. Zeller, C. Moni, A. F. Plante, K. Townsend, L. Gelhaye, K. Lajtha, and
 D. Derrien. 2012. Transfer of litter-derived N to soil mineral–organic associations: evidence
 from decadal 15N tracer experiments. Organic Geochemistry 42:1489-1501.
- Herridge, D. F., M. B. Peoples, and R. M. Boddey. 2008. Global inputs of biological nitrogen
 fixation in agricultural systems. Plant and Soil 311:1-18.
- 572 IPCC. 2013. Climate Change 2013. The Physical Science Basis. Summary for Policymakers. United
 573 Nations, <u>http://www.ipcc.ch/</u>.
- Isbell, F., P. R. Adler, N. Eisenhauer, D. Fornara, K. Kimmel, C. Kremen, D. K. Letourneau, M.
 Liebman, H. W. Polley, and S. Quijas. 2017. Benefits of increasing plant diversity in
 sustainable agroecosystems. Journal of Ecology 105:871-879.
- Kaye, J., D. Finney, C. White, B. Bradley, M. Schipanski, M. Alonso-Ayuso, M. Hunter, M.
 Burgess, and C. Mejia. 2019. Managing nitrogen through cover crop species selection in the
 US mid-Atlantic. PloS one 14:e0215448.
- King, A. E., and J. Blesh. 2018. Crop rotations for increased soil carbon: perenniality as a guiding
 principle. Ecological Applications 28:249-261.
- Kuzyakov, Y., and E. Blagodatskaya. 2015. Microbial hotspots and hot moments in soil: concept &
 review. Soil Biology and Biochemistry 83:184-199.
- Layzell, D. B., and S. Hunt. 1990. Oxygen and the regulation of nitrogen fixation in legume
 nodules. Physiologia Plantarum 80:322-327.
- Lehmann, A., W. Zheng, and M. C. Rillig. 2017. Soil biota contributions to soil aggregation. Nature
 ecology & evolution 1:1828.
- Li, Y.-Y., C.-B. Yu, X. Cheng, C.-J. Li, J.-H. Sun, F.-S. Zhang, H. Lambers, and L. Li. 2009.
 Intercropping alleviates the inhibitory effect of N fertilization on nodulation and symbiotic
 N 2 fixation of faba bean. Plant and Soil 323:295-308.
- Luce, M. S., J. K. Whalen, N. Ziadi, and B. J. Zebarth. 2016. Net nitrogen mineralization enhanced
 with the addition of nitrogen-rich particulate organic matter. Geoderma 262:112-118.
- Marriott, E. E., and M. Wander. 2006a. Qualitative and quantitative differences in particulate
 organic matter fractions in organic and conventional farming systems. Soil Biology and
 Biochemistry 38:1527-1536.
- Marriott, E. E., and M. M. Wander. 2006b. Total and labile soil organic matter in organic and
 conventional farming systems. Soil Science Society of America Journal 70:950-959.

- McDaniel, M., L. Tiemann, and A. Grandy. 2014. Does agricultural crop diversity enhance soil
 microbial biomass and organic matter dynamics? A meta-analysis. Ecological Applications
 24:560-570.
- Paterson, E. 2003. Importance of rhizodeposition in the coupling of plant and microbial
 productivity. European Journal of Soil Science 54:741-750.
- Peoples, M. B., P. M. Chalk, M. J. Unkovich, and R. M. Boddey. 2015. Can differences in 15N
 natural abundance be used to quantify the transfer of nitrogen from legumes to neighbouring
 non-legume plant species? Soil Biology and Biochemistry 87:97-109.
- Poeplau, C., and A. Don. 2015. Carbon sequestration in agricultural soils via cultivation of cover
 crops–A meta-analysis. Agriculture, Ecosystems & Environment 200:33-41.
- Poffenbarger, H. J., S. B. Mirsky, R. R. Weil, J. E. Maul, M. Kramer, J. T. Spargo, and M. A.
- Cavigelli. 2015. Biomass and nitrogen content of hairy vetch-cereal rye cover crop mixtures
 as influenced by species proportions. Agronomy Journal 107:2069-2082.
- Puget, P., C. Chenu, and J. Balesdent. 2000. Dynamics of soil organic matter associated with
 particle-size fractions of water-stable aggregates. European Journal of Soil Science 51:595 605.
- Roley, S. S., D. S. Duncan, D. Liang, A. Garoutte, R. D. Jackson, J. M. Tiedje, and G. P. Robertson.
 2018. Associative nitrogen fixation (ANF) in switchgrass (Panicum virgatum) across a
 nitrogen input gradient. PloS one 13:e0197320.
- Salvagiotti, F., K. G. Cassman, J. E. Specht, D. T. Walters, A. Weiss, and A. Dobermann. 2008.
 Nitrogen uptake, fixation and response to fertilizer N in soybeans: A review. Field Crops
 Research 108:1-13.
- Schipanski, M. E., and L. E. Drinkwater. 2011. Nitrogen fixation of red clover interseeded with
 winter cereals across a management-induced fertility gradient. Nutrient cycling in
 agroecosystems 90:105-119.
- Schipanski, M. E., L. E. Drinkwater, and M. P. Russelle. 2010. Understanding the variability in
 soybean nitrogen fixation across agroecosystems. Plant and Soil 329:379-397.
- Schmidt, M. W., M. S. Torn, S. Abiven, T. Dittmar, G. Guggenberger, I. A. Janssens, M. Kleber, I.
 Kogel-Knabner, J. Lehmann, D. A. Manning, P. Nannipieri, D. P. Rasse, S. Weiner, and S.
- E. Trumbore. 2011. Persistence of soil organic matter as an ecosystem property. Nature
 478:49-56.

- Shearer, G., and D. H. Kohl. 1986. N2-fixation in field settings: estimations based on natural 15N
 abundance. Australian Journal of Plant Physiology 13:699-756.
- Smercina, D. N., S. E. Evans, M. L. Friesen, and L. K. Tiemann. 2019. To fix or not to fix: controls
 on free-living nitrogen fixation in the rhizosphere. Applied and Environmental Microbiology
 85:e02546-02518.
- Storkey, J., T. Döring, J. Baddeley, R. Collins, S. Roderick, H. Jones, and C. Watson. 2015.
 Engineering a plant community to deliver multiple ecosystem services. Ecological
 Applications 25:1034-1043.
- Syswerda, S. P., B. Basso, S. K. Hamilton, J. B. Tausig, and G. P. Robertson. 2012. Long-term
 nitrate loss along an agricultural intensity gradient in the Upper Midwest USA. Agriculture,
 Ecosystems & Environment 149:10-19.
- Thies, J. E., P. W. Singleton, and B. B. Bohlool. 1991. Influence of the size of indigenous rhizobial
 populations on establishment and symbiotic performance of introduced rhizobia on fieldgrown legumes. Applied and Environmental Microbiology 57:19-28.
- Tiemann, L., A. Grandy, E. Atkinson, E. Marin-Spiotta, and M. McDaniel. 2015. Crop rotational
 diversity enhances belowground communities and functions in an agroecosystem. Ecology
 Letters 18:761-771.
- Tonitto, C., M. B. David, and L. E. Drinkwater. 2006. Replacing bare fallows with cover crops in
 fertilizer-intensive cropping systems: a meta-analysis of crop yield and N dynamics.
 Agriculture, Ecosystems and Environment 112:58-72.
- Van Kessel, C., and C. Hartley. 2000. Agricultural management of grain legumes: has it led to an
 increase in nitrogen fixation? Field Crops Research 65:165-181.

Vitousek, P. M., K. Cassman, C. Cleveland, T. Crews, C. B. Field, N. B. Grimm, R. W. Howarth,
R. Marino, L. Martinelli, and E. B. Rastetter. 2002. Towards an ecological understanding of
biological nitrogen fixation. Pages 1-45 *in* E. W. Boyer and R. W. Howarth, editors. The

- Nitrogen Cycle at Regional to Global Scales: Report of the International SCOPE Project.
 Springer.
- Vitousek, P. M., D. N. Menge, S. C. Reed, and C. C. Cleveland. 2013. Biological nitrogen fixation:
 rates, patterns and ecological controls in terrestrial ecosystems. Philosophical Transactions
 of the Royal Society of London B: Biological Sciences 368:20130119.
- von Lützow, M., I. Kögel-Knabner, K. Ekschmitt, E. Matzner, G. Guggenberger, B. Marschner, and
 H. Flessa. 2006. Stabilization of organic matter in temperate soils: mechanisms and their

- relevance under different soil conditions–a review. European Journal of Soil Science
 57:426-445.
- Wander, M. 2004. Soil organic matter fractions and their relevance to soil function. Soil organic
 matter in sustainable agriculture. CRC Press, Boca Raton, FL:67-102.
- 665 Wander, M., S. Traina, B. Stinner, and S. Peters. 1994. Organic and conventional management
- effects on biologically active soil organic matter pools. Soil Science Society of AmericaJournal 58:1130-1139.
- Willson, T., E. Paul, and R. Harwood. 2001. Biologically active soil organic matter fractions in
 sustainable cropping systems. applied soil ecology 16:63-76.
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671 Data Availability

Data are available from the Dryad Digital Repository: <u>https://doi.org/10.5061/dryad.70q4744</u>

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Ivailable fr

Table 1. Baseline soil properties of each field, prior to planting the experiment on the 10 farms in the study, measured from composite soil cores sampled to 20-cm depth. Abbreviations: C-min = short-term C mineralization; C:N = C-to-N ratio; N-min = potentially mineralizable N; POM = particulate organic matter; TOC = total organic carbon; TN = total soil nitrogen.

Farm	тос	TN	N-min	C-min	Free POM	C:N Free POM	Protected POM	C:N Protected POM	Protected POM N
	Mg ha ⁻¹	Mg ha ⁻¹	mg kg ⁻¹ wk ⁻¹	μg CO ₂ -C g ⁻¹ d ⁻¹	g kg-1		g kg-1		mg kg-1
1	38.8	4.8	9.9	56.5	3.0	20.4	4.4	12.8	72.5
2	52.8	5.0	11.7	111.6	2.6	24.5	4.3	14.5	55.6
3	64.1	6.2	23.5	84.5	13.9	16.6	11.4	14.1	139.7
4	56.8	5.0	10.1	59.0	5.2	37.3	9.1	13.5	61.6
5	47.0	4.9	14.3	65.5	13.8	16.6	3.7	13.8	31.5
6	47.3	5.1	10.8	45.5	4.3	25.1	3.1	18.9	45.0
7	58.9	5.8	16.9	70.1	14.4	18.6	3.7	18.6	28.7
8	39.8	4.1	14.4	54.8	9.0	19.0	2.2	15.5	33.5
9	50.3	5.1	14.9	53.8	11.7	25.5	3.7	19.7	28.4
10	33.1	3.7	7.0	31.5	5.6	23.2	2.1	17.8	117.0

Table 2. Additional baseline soil properties of each field, prior to planting the experiment on the 10 farms in the study, measured from composite soil cores sampled to 20-cm depth.

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Farm	$NO_3-N + NH_4-N$	Р	K	рН	sand	silt	clay	Soil order
	mg kg-1	mg kg-1	mg kg-1		°⁄0	%	%	
0	3.0	45.0	100.8	7.0	74.0	15.0	11.0	Alfisol
2	2.6	18.3	78.5	6.3	48.0	28.0	24.0	Mollisol
3	13.9	43.0	162.0	7.5	60.5	23.5	16.0	Alfisol
(()	5.2	18.0	31.8	7.2	61.5	22.0	16.5	Alfisol
5	13.8	84.0	82.0	7.1	60.0	26.0	14.0	Alfisol
6	4.3	14.3	94.5	7.1	36.5	34.5	29.0	Alfisol
\mathbf{Q}	14.4	65.0	173.3	7.4	49.5	27.5	23.0	Alfisol
8	9.0	25.0	95.8	6.4	65.5	22.0	12.5	Alfisol
9	11.7	58.0	110.3	6.4	38.0	30.5	31.5	Alfisol
10	5.6	111.5	130.3	7.0	74.0	15.0	11.0	Alfisol

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Figure legends.

Figure 1. Conceptual diagram of the hypothesized stabilizing feedback between particulate organic matter (POM) pools and legume N_2 fixation. In soils with a legacy of low plant diversity, POM pools are small and the supply of plant-available N from microbial turnover of SOM is small, which leads to high rates of legume N_2 fixation. Regular use of legume cover crops in diversified agroecosystems over time is expected to increase the size of POM pools, which are an energy source for microorganisms. The size of the microbial community (MC) and rates of microbial turnover of POM are therefore also expected to increase. Greater mineralization of N from POM increases the plant-available N pool, and ultimately decreases legume reliance on N_2 fixation, because of the C-cost of acquiring N through the symbiotic relationship with rhizobia. The overall effect is greater N availability for crop assimilation from internal nutrient cycling processes. The right side of the figure also shows the hypothesized moderating effect of intercropping the legume with a grass cover crop (dashed lines). The grass assimilates soil N, drawing down the plant-available N pool and maintaining higher rates of N_2 fixation (i.e., the brown dashed line around the N_2 fixation box).

Figure 2. Top panel: Percentage of vetch N derived from fixation. Bottom panel: Total N assimilation in vetch-rye mixture biomass (mixture) and in vetch biomass (monoculture), separated into N derived from vetch N_2 fixation and N from the soil. Values are means +/- standard error.

Figure 3. Regression relationships for N concentration in the protected POM pool and the % of vetch shoot N derived from fixation, for sole vetch (left) and vetch grown in mixture with rye (right).

Figure 4. Mean change across nine farms in properties that reflect soil nutrient cycling dynamics, calculated as the difference between the vetch-rye cover crop mixture treatment and the no cover crop control, shown in their measured units with 95% confidence intervals.

Figure 5. Regression relationship between the N concentration in the protected POM pool and the rate of potentially mineralizable N across farm sites at the final sampling time.



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-C U. Manu Author



Mean change with cover crop mixture

