STANDARDIZING CYP2D6 GENOTYPE TO PHENOTYPE TRANSLATION:

CONSENSUS RECOMMENDATIONS FROM THE CLINICAL

PHARMACOGENETICS IMPLEMENTATION CONSORTIUM (CPIC) AND DUTCH PHARMACOGENETICS WORKING GROUP (DPWG)

Short Running Title: THE *CYP2D6* GENOTYPE TO PHENOTYPE STANDARDIZATION PROJECT

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Conflicts of Interest

The authors declared no competing interests for this work.

ABSTRACT

Translating *CYP2D6* genotype to metabolizer phenotype is not standardized across clinical laboratories offering pharmacogenetic testing and pharmacogenetic clinical practice guidelines such as the Clinical Pharmacogenetics Implementation Consortium (CPIC) and the Dutch Pharmacogenetics Working group (DPWG). The genotype to phenotype translation discordance between laboratories and guidelines can cause discordant CYP2D6 phenotype assignments and thus lead to inconsistent therapeutic recommendations and confusion among patients and

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clinicians. A modified-Delphi method was used to obtain consensus for a uniform system for translating *CYP2D6* genotype to phenotype among a panel of international CYP2D6 experts. Experts with diverse involvement in CYP2D6 interpretation (clinicians, researchers, genetic testing laboratorians, and pharmacogenetic implementers; n = 37) participated in conference calls and surveys. After completion of seven surveys, a consensus (>70%) was reached with 82% of the CYP2D6 experts agreeing to the final *CYP2D6* genotype to phenotype translation method. Broad adoption of the proposed *CYP2D6* genotype to phenotype translation method by guideline developers such as CPIC and DPWG and clinical laboratories as well as researchers will result in more consistent interpretation of CYP2D6 genotype.

INTRODUCTION

Cytochrome P450 2D6 (CYP2D6) is directly involved in the metabolism of ~20% of currently approved medications ¹, and genetic variation in the *CYP2D6* gene has been implicated in the efficacy and/or toxicity of many drugs. Consequently, the highly polymorphic *CYP2D6* gene is the focus of several Clinical Pharmacogenetics Implementation Consortium (CPIC) and/or Dutch Pharmacogenetics Working Group (DPWG) clinical practice guidelines on 15 widely used medications, including selective serotonin reuptake inhibitors, tricyclic antidepressants, atomoxetine, codeine, tramadol, tamoxifen, ondansetron and tropisetron ²⁻⁸. Recently, CPIC and DPWG reported some discrepancies in their guidelines, primarily related to how certain *CYP2D6* genotypes or diplotypes (from here on referred to as 'genotype') were translated into metabolizer phenotype or metabolizer status (from here on referred to as 'phenotype'). **Figure 1** describes the process used to translate identified *CYP2D6* genetic variants into phenotypes. Given that the clinical recommendations for CYP2D6 in CPIC and DPWG guidelines are based on phenotype, the assignment of CYP2D6 phenotype based on genotype is a critical aspect for consistent clinical implementation.

Translating *CYP2D6* genotype to phenotype is also not standardized across clinical laboratories offering pharmacogenetic (PGx) testing. Current systems used to translate genotype to phenotype rely on the star (*) allele nomenclature (defining which variant(s) are present in an allele), and the assignment of function to the star alleles (i.e., increased, normal, decreased, or no function) with inferring phenotype based on the identified genotype. Some systems utilize the activity score (AS) system for assignment of phenotype where each allele is assigned an "activity value" ranging from 0 to 1 (e.g., 0 for no function, 0.5 for decreased function and 1.0 for normal function) ¹⁰. In addition, given that the *CYP2D6* allele can also have variable copy number, the activity value of an allele is multiplied by the number of gene copies (i.e. x2, x3, etc. if copy number is known, or x2 as a default value if copy number is unknown and reported as xN). As such, the *CYP2D6* AS is the sum of the activity values assigned to each allele ¹⁰ (**Figure 1**). Within the CPIC guidelines, the *CYP2D6* AS is then translated into a phenotype using the following classification system: individuals with an AS of 0 are poor metabolizers (PMs), those

with a score of 0.5 are intermediate metabolizers (IMs), those with a score of 1.0, 1.5 and 2.0 are normal metabolizers (NMs) and those with a score >2 are ultrarapid metabolizers (UMs) (later referred to as the "CPIC method") (**Table 1**).

While the CPIC guidelines and some clinical laboratories categorize an AS of 1.0 as CYP2D6 NMs, other clinical laboratories and the DPWG guidelines consider an AS of 1.0 as CYP2D6 IMs ^{6-8, 11} (**Table 1**). Differences also exist for the AS value that separates NMs from UMs. Consequently, the different ways of inferring CYP2D6 phenotype between laboratories and guidelines can cause discordant CYP2D6 phenotype assignments and thus lead to inconsistent therapeutic recommendations. To minimize confusion, it is important to maintain standardized CYP2D6 phenotype translation from genotype data. As such, the purpose of this project was to harmonize the systems used by the CPIC and DPWG and reach consensus among an international panel of *CYP2D6* experts regarding the standardization of how to translate *CYP2D6* genotype to phenotype.

MATERIALS AND METHODS

The Delphi survey technique is an established approach for seeking expert consensus on a given topic ¹²⁻¹⁴. The method uses a series of repeated structured questionnaires or "rounds". Each round provides written, systematic refinement of expert opinion, where feedback of group opinion is provided after each round ¹⁵. Delphi survey technique guidelines proposed by Hasson et al. were consulted in the design of the project ¹⁶. The method used is this study is often referred to as a "modified-Delphi" as a major modification to the Delphi technique consists of beginning the process with a set of carefully selected options versus an open-ended questionnaire.

For the Delphi method used (**Figure 2**), *CYP2D6* expert members of the CPIC and DPWG were solicited by email invitation, as well as other international investigators with published expertise in *CYP2D6* pharmacogenetics. In addition, experts were solicited by posting a description of the project on the PharmGKB and CPIC websites.

Experts were invited to participate in a series of surveys using an internet-based survey tool (SurveyMonkey Inc, Palo Alto, CA; http://www.surveymonkey.com), supplemented with multiple live webinars that were used to explain the survey and solicit feedback. The webinars were designed to facilitate understanding of the survey to encourage completion; towards the end of the process an additional webinar was used to assist in developing consensus. Each survey also included questions regarding the expert's workplace setting and degree of *CYP2D6* expertise (i.e., role in clinical pharmacogenetics, time devoted to *CYP2D6*). Responses were included in the analysis if the respondent provided their name and contact information, which were necessary to enable follow up with the respondent for the subsequent round (but not disclosed). Responses were tabulated as numeric counts and frequencies for each phase to determine whether consensus was reached. Consensus was defined as 70% of experts agreeing; this level of agreement has been considered appropriate in previous Delphi studies ¹⁷⁻¹⁹.

Phase 0: Assessment

The objective of the assessment phase was to define areas of discordance between assignment of CYP2D6 phenotype based on genotype. The Genetic Testing Registry (GTR) ²⁰ was queried for laboratories performing clinical *CYP2D6* genetic testing, and emails requesting participation in a survey were sent to each laboratory. The survey consisted of 16 questions regarding *CYP2D6* genotype interpretation including questions regarding current methods used to translate *CYP2D6* genotype to phenotype (see ²¹ for the laboratory survey questions). Clinical practice guidelines (CPIC and DPWG) were evaluated for systems used to translate genotype to phenotype. References used in the evidence tables of CPIC guidelines and additional literature were evaluated for differences in AS between 0.5 versus 1 and 1 versus 2 and consequences of *CYP2D6*10*-containing genotypes on AS assignment. Results were presented to the CYP2D6 experts on the first conference call.

Phase 1: Development

The objective of the development phase was to determine *CYP2D6* genotype to phenotype translation options for evaluation and assess initial expert opinions on current systems being used by clinical laboratories and available PGx guidelines. Given that the discordance between genotype to phenotype translation is mostly related to the AS of 0.5, 1, and 2 and disagreements

regarding the activity value assigned to *CYP2D6*10*, the first expert conference call provided examples of pharmacokinetic studies with AS data and additional studies comparing *CYP2D6*10* activity (see ²¹ for this spreadsheet). Experts were required to either attend the live conference call or to listen to the recorded version and asked to provide feedback and additional references if warranted.

Phase 2. Prioritization

The expert opinions discussed in the development phase were used to inform the prioritization phase with the final objective to select a genotype to phenotype translation system to which at least 70% of the experts agreed upon. Survey 1 asked specifically if experts thought there was a clinically significant difference between AS of 1 versus 2 and 0.5 versus 1, and if there was a rationale to use a lower activity value for AS calculation (i.e. 'downgrade' the activity value from 0.5 to 0.25) for some *CYP2D6* alleles (e.g. *CYP2D6*9*, *10, *17, *29, *41) to more accurately reflect activity relative to other *CYP2D6* alleles. Survey 1 also presented five different systems for translating *CYP2D6* genotype to phenotype to assess expert opinion of each system. All questions required expert explanation, references and examples to support the opinion. The results from Survey 1 were presented on a conference call and discussed, and a subsequent call presented two methods for assigning AS (i.e., AS ordinal groups versus continuous percentage activity values). The results from Surveys 2 and 3 were used to prioritize one method to move into the refinement phase. Results including expert comments from previous surveys were provided with each survey.

Phase 3 and 4. Refinement and Consensus

Based on the results from Survey 3, Surveys 4 to 6 were used to refine the details of the selected approach. Experts were asked a series of questions related to AS definitions for each CYP2D6 phenotype. A summary of comments from previous surveys was provided and experts were asked to review the comments prior to responding to subsequent questions.

Phase 5. Validation

Once consensus was reached, results were presented on a member-wide CPIC call and posted to the CPIC website for two months to allow for public comment. PharmGKB also blogged about the project and solicited feedback. Feedback was presented to the experts on a subsequent conference call and discussed. Survey 7 measured acceptance of incorporation of the feedback into the previous consensus system. The final survey (Survey 8) measured the level of acceptance of the final *CYP2D6* genotype to phenotype translation system.

RESULTS

Expert Panel Composition

A total of 37 CYP2D6 experts participated in the project with 27 completing Survey 1, 28 completing Survey 2, 24 completing Survey 3, 25 completing Survey 4, 27 completing Survey 5, 31 completing Survey 6, 23 completing Survey 7, and 27 completing Survey 8. Not all experts participated in each round and some experts participated in the initial or early rounds, but not in the later rounds or vice versa. The participants represented a diverse group of self-identified experts with varying levels of CYP2D6 expertise (**Table 2**), and an international representation: 59% were from the United States, 27% from Europe, and 11% from other countries (i.e., South Korea, Japan, Canada, and Australia). The study was facilitated by representatives from both CPIC (n=5) and the DPWG (n=3) leadership.

Phases 0 and 1: Assessment and Development

Email invitations were sent to 43 clinical testing laboratories who reported performing *CYP2D6* genotype testing to the GTR. A total of 15 laboratories completed a survey regarding how their laboratory translated *CYP2D6* genotype to phenotype. Of those, 47% (n=7) reported using the CPIC method for translating *CYP2D6* genotype to phenotype (i.e., AS of 1.0 is classified as NM). Of the eight laboratories (53%) not using the CPIC method (i.e., AS of 1.0 is classified as IM), six disclosed their *CYP2D6* genotype to phenotype translation methods (**Table 1**). Full results can be found at reference ²¹. Experts participated in an initial conference call during which results from the laboratory survey were reported, evidence supporting differences in AS of 0.5, 1, and 2 was presented and available information regarding *CYP2D6*10* activity was shared. Finally, options for a system for translating genotype to phenotype were discussed.

Phase 2: Prioritization

In Survey 1, 93% (n=25) of the experts agreed that there is a clinically significant difference between a CYP2D6 AS of 1 and 2, and 78% (n=21) agreed that there also is a significant difference between an AS of 0.5 and 1. Among the experts agreeing to the need to downgrade some alleles to an activity value of 0.25 (53% (n=14)), 85% selected CYP2D6*10 and 50% selected CYP2D6*41. Based on the first conference call discussion, Survey 1 included five potential options to move forward (Supplemental Figure S1). However, no method reached consensus (>70%). Comments and Survey 1 results were made available to all participants and discussed on the second conference call. Based on feedback provided after the second call, a third call was held to discuss using a 'percentage activity system' versus the AS system (see Discussion). After receiving feedback from several of the experts, CPIC and the DPWG recommended to the experts to proceed with the use of the AS system to which 94% (n=29) agreed in Survey 2. Also in Survey 2, 42% (n=12) preferred a system that classifies AS of 0.5 and 1 as IMs, 38% (n=11) preferred to create a new phenotype group for AS of 1, 7% (n=2) thought both methods would be acceptable, and 14% (n=4) recommended another method. Experts were asked to provide their rationale for their responses and Survey 2 results were discussed on a conference call. Survey 2 results can be found at reference (22). Based on the conference call discussion and Survey 2 results, CPIC and DPWG representatives recommended to proceed with the use of AS and to downgrade the activity value of some alleles (currently limited to CYP2D6*10). Using an activity value of 0.25 for AS calculation to more accurately reflect the considerably decreased activity of CYP2D6*10 results in the introduction of additional AS groups. In Survey 3, the majority of experts (96%) agreed to create an activity value of 0.25 category, and 88% agreed to the assignment of AS 0.5 to 1 as an IM.

Phase 3 and 4: Refinement and Consensus

Surveys 4 and 5 were used to refine the new system discussed above. Specifically, the experts discussed how to integrate the new AS groups (i.e. 0.25, 0.75, 1.25, 1.75, 2.25) that are introduced by the addition of an activity value of 0.25 for AS calculations into the four phenotype categories (i.e., PM, IM, NM and UM). As shown in **Supplemental Table S1**, Survey 5 included two options. Fourteen (52%) of the experts chose option 1, 11 (41%) chose option 2,

and two (7%) disagreed with both options. Because the experts favored option 1 (52% vs 41%), the CPIC and DPWG representatives recommended option 1 on Survey 6; this decision was supported by expert comments regarding the small contribution of an activity score of 0.25 to clinically appreciable activity to the overall function. Experts agreed with the option and consensus was reached (27 (87%) agreed while four experts (13%) disagreed) (**Table 3**).

Phase 5. Validation

After two months of accepting public comments, two issues were revisited for consideration: 1) inclusion of a rapid metabolizer (RM) phenotype group between NM and UM and 2) use of contiguous AS values to define each phenotype (i.e. no gaps between AS categories). After discussion on a conference call and Survey 6, 87% (n=20) of the experts rejected the introduction of a RM phenotype group while 70% agreed to use contiguous AS ranges to define CYP2D6 phenotype based on genotype. Experts were also asked regarding the range for PMs: the majority (61%; n=14) favored to define PMs as having two no function alleles (AS=0), 30% (n=7) favored defining AS<0.25 as PMs, and 9% indicated "I do not know". Results were discussed on a subsequent conference call on which the experts also discussed and agreed on the contiguous ranges for the other phenotype groups (Survey 7; 82% of the participants (n=22) agreeing to the final assignments shown in **Table 3**).

DISCUSSION

We engaged a diverse group of international CYP2D6 experts to establish a standardized method for translating CYP2D6 genotype to metabolizer phenotype. The major focus of this working group was to harmonize how to translate CYP2D6 genotype into phenotype; a secondary aim was to explore how currently used systems could be improved. This international group of experts consisted of representatives of academia and industry including clinical genetic testing laboratories. Also, individuals with experience in implementing CYP2D6 pharmacogenetics into clinical practice and electronic health records at large hospitals were included to assess the impact of the project on past or ongoing CYP2D6 implementation efforts. Importantly, the final CYP2D6 translation method presented in Table 3 will be incorporated into the CYP2D6 tables on www.cpicpgx.org and used in all new and updated CPIC and DPWG guidelines. We also recommend that this system be considered as standard practice across all areas of clinical

pharmacogenetics, including clinical genetic testing laboratories. We also strongly encourage PGx researchers to use this standardized method to report their findings as this will greatly facilitate future data collection from the literature and comparison of data.



Throughout the project several issues and challenges were identified and discussed in detail (**Table 4**) as follows. 1) Lengthy discussions entailed the possibility of generating a new phenotype group for AS=0.5 as patients with genotypes consisting of one decreased and one nonfunctional allele appear to have lower activity compared to those with genotypes giving rise to an AS of 1. Concerns were raised that combining AS of 0.5 and 1.0 in research studies may mask potentially significant differences among these AS groups since there are considerably fewer subjects with an AS of 0.5. The introduction of a new phenotype group describing patients between PM and IM, was however, rejected by the CPIC and DPWG representatives based on the CPIC term standardization project which determined that five phenotype groups are sufficient ¹⁸; the majority of experts also rejected the introduction of an additional phenotype group. 2) A number of factors weighed into the decision to reclassify AS of 1 from NM to IM. Since published studies vary on how subjects with an AS of 1 are grouped (NM vs IM) it is difficult to compare AS of 0.5-1 vs 2 or AS of 1 versus 2 with confidence to support differences in outcomes between these groups. In addition, more laboratories also currently classify an AS of 1 as IMs and not NMs (**Table 1**), indicating that classifying an AS of 1 as IM may be minimally disruptive to most research and clinical laboratories. 3) While the goal is to have a translation system that is agnostic to the drug used, the experts realized that certain genotypes may need recommendations that differ from their 'drug-agnostic' phenotype group assignment. To address this challenge, recommendations from CPIC and the DPWG can be different for certain drugs (see CYP2D6*10-containing genotypes in CPIC tamoxifen guideline for example 3), or for a particular AS group if warranted. In other words, a recommendation can be based on the AS versus the phenotype group. Therefore, it is extremely important that clinical laboratories not only report phenotype, but also detail the patient's genotype and sequence variations tested (see Bousman et al. for guidance of how to select a PGx test ²²).

A secondary goal of the project was to re-evaluate the activity values assigned to alleles with decreased function. Currently, a value of 0.5 bins decreased function alleles together regardless

of the percentage activity they retain compared to the CYP2D6.1 (wild-type) protein product. The majority of values used today for AS calculation are based on the original report by Gaedigk et al that was published 11 years ago ¹⁰. The prospect of lowering the value assigned to *CYP2D6*10*, an allele that is anecdotally known to have 'little' activity was reviewed earlier, but the authors did not find sufficient evidence to downgrade this allele based on evidence available six years ago ²³. Since then, there was mounting evidence suggesting that *CYP2D6*10* not only consistently conveys decreased function across substrates, but also appears to be, on average, considerably lower compared to other decreased function alleles. Thus, using an activity value of 0.5 for AS calculation for *CYP2D6*10*-containing genotypes may overestimate the metabolic capacity of patients with *CYP2D6*10/*10* or *10/no function genotypes. Assigning an activity value of 0.25 to the *CYP2D6*10* allele for AS calculation will group *CYP2D6*10/*10* as AS=0.5 and *10/no function as AS=0.25 (opposed to AS=1 and AS=0.5 respectively), which more precisely aligns with the level of reduction of enzyme activity.

Notably, there are other star (*) alleles that harbor the *CYP2D6*10* defining variant (100C>T; rs1065852) in combination with other variants that, to the best of current knowledge, do not impact function or have decreased function on their own (e.g. 1023C>T), which are currently classified by CPIC as "decreased" function and thus, receive a value of 0.5 for AS calculation (e.g., *CYP2D6*49*, *54, *65, *72). Note that positions are provided according to the genomic *CYP2D6* RefSeq NG_008376.3, the numbering system recommended by PharmVar ²⁴. There are also a number of alleles currently with g.100C>T labeled as "uncertain" function (e.g., *37, *52, *64, *87, *94, *95). Most experts recommended that these alleles should also receive an activity value of 0.25; however, concerns were raised by some of the experts regarding the lack of evidence (i.e. in vitro or in vivo studies) for most of these alleles (e.g. 100C>T in combination with other SNP(s) may obliterate function, or compensate for the decreased function caused by 100C>T)^{10,25}. Thus, other *CYP2D6* alleles containing the 100C>T variant besides *10 will be assessed as part of future CPIC guideline development. At that time functional status and values for activity score calculations will be assigned for these alleles; other alleles will also be reviewed and re-assessed during this process.

It was also discussed whether genotype to phenotype translation should be standardized across all CYP450 enzymes. Currently, the AS is applied to *CYP2D6* for which it was originally devised to accommodate a large (and growing) number of alleles with varying activity and was widely adopted after being published ¹⁰; hence, it was a natural decision for CPIC to adopt this system. The AS was eventually also adopted for *DPYD* to accommodate the vast number of sequence variants that emerged for this gene. As shown in **Supplemental Table S2**, other *CYP* genes have their distinct systems to translate genotype to phenotype. There was no consensus among the group whether this would be a desirable goal because a major revision towards a *CYP*-wide system may pose a major challenge for clinical reporting and implementation with unclear benefits.

Feedback included the suggestion to add a RM phenotype group. One argument for having a RM phenotype group was that certain genotypes may have increased activity compared to NMs (e.g., *1x2/*41), but less than UMs (e.g., *1x2/*2); it was also argued that the introduction of an RM group would be in alignment with *CYP2C19*. However, the experts felt that there was not enough evidence to differentiate between two "increased function" phenotypes (rapid and ultrarapid) for CYP2D6 and thus, these groups would not be clinically useful.

In 2016, CPIC published a consensus project aimed to standardize terms describing allele function and phenotype ¹⁸. Prior to this project various terms were used for allele function and phenotype, which impeded reporting and sharing of test results across clinical laboratories and electronic health records. Based on the results of this project, Systematized Nomenclature of Medicine-Clinical Terms (SNOMED-CT) and Logical Observation Identifiers Names and Codes (LOINC) terms were created for use in the electronic health record to facilitate efficient reporting of pharmacogenetic results. Although the 2016 project did not address standardization of the translation of genotype to phenotype, pharmacogenetic experts were asked whether they favor a four or five major category phenotype system. The majority of participants (91%, n=48) agreed to four categories (see all survey results at ²¹). The CYP2D6 experts and CPIC and DPWG representatives considered this result suggesting that adding an additional phenotype category may not be widely accepted by the pharmacogenetic community.

The use of a contiguous AS scale for defining metabolizer phenotype was addressed at two stages during the Delphi process. Early in the process (call #3, Survey 2) a group of experts advocated for an alternative system referred to as the 'percentage activity' system. Similar to the AS, in the percentage activity system each allele is assigned a value on the scale of 0 (no activity) to 1 (normal activity); however, in the percentage activity system values are assigned in increments of 0.1 instead of 0.5 (now 0.25) (Figure 3). Also, instead of calculating the sum of the two activity values to calculate the AS, the values would be averaged and multiplied by 100 for the percentage activity system so that each patient's CYP2D6 metabolic capacity is described on a percentage activity scale of 0% (analogous to AS=0) to 100% (AS=2.0) or higher. It was argued that the percentage activity system may be more intuitive to clinicians. While such a system may ultimately be more precise, there are a number of hurdles. For example, the determination of activity for an allele is difficult as is and to discriminate activity on a scale of 10% increments seems impossible as there are no data for the vast majority of alleles at this point in time. Second, there is a broad range of inter-individual variability among subjects within the same genotype group ^{10, 25} and third, even if activity could be determined on a 10% scale, percent activities may still need to be translated into a limited number of phenotyping categories for feasibility of clinical implementation.

Given these challenges, the experts came to consensus on Survey 2 to move forward with the AS system mainly due to limited data for estimating percentage activity of individual alleles, with a general interest in future work that moves the field in the direction of more precise activity estimates as well as the prospect of developing more sophisticated dosing algorithms that are based on population pharmacokinetic and dynamic models taking genotype along other pertinent factors into account.

The second discussion of a 'contiguous scale' system was held after the public comment period (Survey 7), after thresholds for each phenotype had already been agreed upon. Given the possibility of future allelic re-estimates or percentage activities, the experts defined the consensus scale contiguously, such that all potential values of AS have a consensus phenotype translation. For example, given that an AS=0 is PM and an AS=0.25 is IM, would an AS=0.2 be

a PM or IM? Thus, the contiguous consensus scale (**Table 3**) can accommodate any future scores regardless of the number of groups or system used.

A central aim of this project was to continue the previously reported and ongoing efforts dedicated to standardizing inconsistent components related to clinical pharmacogenetics, including genetic testing, interpretation, recommendations, and implementation ^{18, 26-29}. Importantly, we strongly encourage all pharmacogenetics stakeholders to adopt the consent *CYP2D6* translation system that has emerged from this project. Broad adoption of the proposed *CYP2D6* translation system by clinical laboratories as well as researchers will ultimately lead to reduced interlaboratory discrepancies, increased consistency in *CYP2D6* reporting thus more consistent test interpretation. The performance of this system will also be measurable over time based on the metrics from the College of American Pathologists (CAP) Pharmacogenetic Proficiency Survey, as *CYP2D6* genotyping/phenotyping has historically had the greatest interlaboratory variability among the commonly tested pharmacogenetic genes ³⁰. However, we acknowledge that adopting this process, if distinct from a previous reporting protocol, may also result in laboratory cost and effort to modify workflows and reconcile previously reported *CYP2D6* results based on prior translation systems.

Healthcare institutions that have already implemented *CYP2D6* genotyping using the CPIC method will be affected by this new system as follows: 1) Patients with a *CYP2D6* AS of 1.0 who were previously assigned an NM phenotype will now have to be reassigned an IM phenotype and patients with a *CYP2D6* AS of 2.25 who were previously assigned an UM phenotype assigned as NM; 2) *CYP2D6* interpretive reports as well as all applicable educational materials pertaining to an AS of 1.0 (or 2.25) will need to be updated. Because the former change will necessitate substantial efforts in order to back-track patients and inform them of their new phenotype assignment, some institutions may elect not to inform previously tested patients of their new re-assigned CYP2D6 phenotype.

The Delphi method is a powerful tool that was developed to build consensus among and to develop standards across different disciplines ^{12, 13, 15}. Key risks to the validity of a Delphi study include overestimating the expertise of participants and attrition across the consensus rounds.

Given that each participant had some CYP2D6 pharmacogenetic expertise and 51% of survey respondents indicated that they spend >26% of their time devoted to work related to CYP2D6, we believe to have had adequate CYP2D6 expertise among our survey participants. Although attrition rates were not defined a priori, 76% of the experts participated in Survey 7 (participation averaged 74% for Surveys 1-6) and relative to other Delphi panels and the recommended minimum panel size, our final consensus panel was relatively large (suggested minimum for expert panels is ten participants), which reinforces the validity of our results ³¹. To reduce bias, especially the authority or reputation of specific individuals, Delphi panel participants are often kept anonymous throughout the process. Although survey creators and analysts were not blinded to participants, identifying information was not shared among survey participants. The only occasions of participant identification were in between surveys when nonblinded email invitations were sent to participants in conference calls and webinars during which interim results were discussed. Because many survey results were close the CPIC and DPWG representatives discussed options for the next survey based on previous results and comments from the experts, which resulted in recommendations of limited choices to move forward. However, experts still had to agree to the option.

In conclusion, consensus among an international panel of CYP2D6 experts regarding the standardization of translating *CYP2D6* genotype to phenotype was achieved. Moving forward, CPIC and DPWG will use this system in their practice guidelines. As most pharmacogenetic clinical recommendations are based on phenotype, we anticipate that broad adoption of the proposed *CYP2D6* genotype to phenotype translation framework will minimize discrepant *CYP2D6* test results and inconsistent therapeutic recommendations.

STUDY HIGHLIGHTS

•What is the current knowledge on the topic? Translating *CYP2D6* genotype to metabolizer phenotype is not standardized across clinical laboratories offering pharmacogenetic testing and pharmacogenetic clinical practice guidelines.

- •What question did this study address? The purpose of this project was to harmonize the systems used by the CPIC and DPWG and reach consensus among an international panel of *CYP2D6* experts regarding the standardization of how to translate *CYP2D6* genotype to phenotype.
- •What does this study add to our knowledge? We engaged a diverse group of international CYP2D6 experts to establish a standardized method for translating *CYP2D6* genotype to metabolizer phenotype.
- •How might this change clinical pharmacology or translational science? Broad adoption of the proposed *CYP2D6* genotype to phenotype translation method by guideline developers such as CPIC and DPWG and clinical laboratories as well as researchers will result in more consistent interpretation of CYP2D6 genotype.

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AUTHOR CONTRIBUTIONS

K.E.C., K.S., M.W-C., J.J.S., C.E.H., T.E.K., R.S.G., M.V.R., S.A.S., D.L.H., H-J.G., and A.G. wrote the manuscript; K.E.C., K.S., M.W-C., J.J.S., R.S.G., M.V.R., H-J.G., and A.G. designed the research; K.E.C., K.S., M.W-C., and A.G. performed the research; K.E.C., K.S., M.W-C., and A.G. analyzed the data.

REFERENCES

Saravanakumar, A., Sadighi, A., Ryu, R. & Akhlaghi, F. Physicochemical Properties,
 Biotransformation, and Transport Pathways of Established and Newly Approved Medications: A

- Systematic Review of the Top 200 Most Prescribed Drugs vs. the FDA-Approved Drugs Between 2005 and 2016. *Clin Pharmacokinet*. (2019). doi: 10.1007/s40262-019-00750-8
- 2. Brown, J.T., *et al.* Clinical Pharmacogenetics Implementation Consortium (CPIC) Guideline for CYP2D6 Genotype and Atomoxetine Therapy. *Clin Pharmacol Ther.* **106**, 94-102 (2019).
- 3. Goetz, M.P., et al. Clinical Pharmacogenetics Implementation Consortium (CPIC) Guideline for CYP2D6 and Tamoxifen Therapy. *Clin Pharmacol Ther.* **103**, 770-777 (2018).
- 4. Bell, G.C., et al. Clinical Pharmacogenetics Implementation Consortium (CPIC) guideline for CYP2D6 genotype and use of ondansetron and tropisetron. Clin Pharmacol Ther. 102, 213-218 (2017).
- 5. Hicks, J.K., et al. Clinical pharmacogenetics implementation consortium guideline (CPIC) for CYP2D6 and CYP2C19 genotypes and dosing of tricyclic antidepressants: 2016 update. *Clin Pharmacol Ther.* **102**, 37-44 (2017).
- 6. Crews, K.R., et al. Clinical Pharmacogenetics Implementation Consortium guidelines for cytochrome P450 2D6 genotype and codeine therapy: 2014 update. *Clin Pharmacol Ther.* **95**, 376-382 (2014).
- 7. Hicks, J.K., et al. Clinical Pharmacogenetics Implementation Consortium (CPIC) Guideline for CYP2D6 and CYP2C19 Genotypes and Dosing of Selective Serotonin Reuptake Inhibitors. *Clin Pharmacol Ther.* **98**, 127-134 (2015).
- 8. Swen, J.J., et al. Pharmacogenetics: from bench to byte--an update of guidelines. *Clin Pharmacol Ther.* **89**, 662-673 (2011).
- 9. Bank, P.C.D., et al. Comparison of the Guidelines of the Clinical Pharmacogenetics

 Implementation Consortium and the Dutch Pharmacogenetics Working Group. Clin Pharmacol

 Ther. 103, 599-618 (2018).
- 10. Gaedigk, A., et al. The CYP2D6 activity score: translating genotype information into a qualitative measure of phenotype. *Clin Pharmacol Ther.* **83**, 234-242 (2008).
- 11. AmpliChip™ CYP450 Package Insert. 2005.
- Dalkey, N. & Helmer, O. An experimental application of the Delphi method to the use of experts. *Management Science.* **9**, 458-467 (1963).
- 13. Beretta, R. A critical review of the Delphi technique. *Nurse Researcher*. **3**, 79-89 (1996).
- 14. Green, B., Jones, M., Hughes, D. & Williams, A. Applying the Delphi technique in a study of GPs' information requirements. *Health & social care in the community*. **7**, 198-205 (1999).

- 15. von der Gracht, H.A. Consensus measurement in Delphi studies Review and implications for future quality assurance. *Technological Forecasting & Social Change*. **79**, 125-1536 (2012).
- 16. Hasson, F., Keeney, S. & McKenna, H. Research guidelines for the Delphi survey technique. *Journal of advanced nursing*. **32**, 1008-1015 (2000).
- 17. Slade, S.C., Dionne, C.E., Underwood, M. & Buchbinder, R. Standardised method for reporting exercise programmes: protocol for a modified Delphi study. *BMJ Open.* **4**, e006682 (2014).
- 18. Caudle, K.E., *et al.* Standardizing terms for clinical pharmacogenetic test results: consensus terms from the Clinical Pharmacogenetics Implementation Consortium (CPIC). *Genet Med.* **19**, 215-223 (2017).
- 19. Henderson, E.J. & Rubin, G.P. Development of a community-based model for respiratory care services. *BMC Health Serv Res.* **12**, 193 (2012).
- 20. Rubinstein, W.S., *et al.* The NIH genetic testing registry: a new, centralized database of genetic tests to enable access to comprehensive information and improve transparency. *Nucleic Acids Res.* **41**, D925-935 (2013).
- 21. CPIC. CYP2D6 genotype to phenotype standardization project. 2019; June 24.
- 22. Bousman, C.A., Zierhut, H. & Muller, D.J. Navigating the Labyrinth of Pharmacogenetic Testing: A Guide to Test Selection. *Clin Pharmacol Ther.* (2019). doi: 10.1002/cpt.1432
- 23. Hicks, J.K., Swen, J.J. & Gaedigk, A. Challenges in CYP2D6 phenotype assignment from genotype data: a critical assessment and call for standardization. *Curr Drug Metab.* **15**, 218-232 (2014).
- 24. Gaedigk, A., et al. The Evolution of PharmVar. Clin Pharmacol Ther. 105, 29-32 (2019).
- 25. Montane Jaime, L.K., Lalla, A., Steimer, W. & Gaedigk, A. Characterization of the CYP2D6 gene locus and metabolic activity in Indo- and Afro-Trinidadians: discovery of novel allelic variants. *Pharmacogenomics.* **14**, 261-276 (2013).
- 26. Caudle, K.E., *et al.* Standardization can accelerate the adoption of pharmacogenomics: current status and the path forward. *Pharmacogenomics*. **19**, 847-860 (2018).
- 27. Kalman, L.V., et al. Pharmacogenetic allele nomenclature: International workgroup recommendations for test result reporting. *Clin Pharmacol Ther.* **99**, 172-185 (2016).
- 28. Pratt, V.M., et al. Recommendations for Clinical CYP2C19 Genotyping Allele Selection: A Report of the Association for Molecular Pathology. *J Mol Diagn*. **20**, 269-276 (2018).
- 29. Moriyama, B., et al. Clinical Pharmacogenetics Implementation Consortium (CPIC) Guidelines for CYP2C19 and Voriconazole Therapy. Clin Pharmacol Ther. **102**, 45-51 (2017).

- 30. Wu, A.H. Genotype and phenotype concordance for pharmacogenetic tests through proficiency survey testing. *Arch Pathol Lab Med.* **137**, 1232-1236 (2013).
- 31. Okoli, C. & Pawlowski, S. The Delphi method as a research tool: an example, design considerations and applications. *Information and management*. **42**, 15-29 (2004).
- 32. Wang, D., *et al.* Common CYP2D6 polymorphisms affecting alternative splicing and transcription: long-range haplotypes with two regulatory variants modulate CYP2D6 activity. *Hum Mol Genet*. **23**, 268-278 (2014).

FIGURE LEGENDS

FIGURE 1. PROCESS FOR TRANSLATION OF CYP2D6 GENOTYPE TO PHENOTYPE $_$

^aDiplotype describes the combination of two alleles (or haplotypes) which can involve multiple variants. Diplotype and genotype, a term that technically describes variation at a single nucleotide position, are often used interchangeably. Since genotype is the more commonly used term, it is used throughout this report.

FIGURE 2. MODIFIED DELPHI PROCESS

^aComments from each round were made available to all experts and discussed on conference calls.

FIGURE 3. COMPARISON OF THE CPIC METHOD AND PERCENTAGE ACTIVITY METHOD FOR TRANSLATING *CYP2D6* GENOTYPE TO PHENOTYPE.

Thin lines represent different ways to translate AS into phenotype and the bold lines represent the recommended CYP2D6 genotype to phenotype translation consensus system. PM, poor metabolizer; IM, intermediate metabolizer; NM, normal metabolizer; UM, ultrarapid metabolizer

SUPPLEMENTARY INFORMATION TITLES

(SUPPLEMENT TO STANDARDIZING CYP2D6 GENOTYPE TO PHENOTYPE TRANSLATION: CONSENSUS RECOMMENDATIONS FROM THE CLINICAL PHARMACOGENETICS IMPLEMENTATION CONSORTIUM (CPIC) AND DUTCH PHARMACOGENETICS WORKING G)

SUPPLEMENTARY MATERIALS. Figure S1, Tables S1-S2

TABLE 1. COMPARISON OF SYSTEMS USED FOR CYP2D6 GENOTYPE TO PHENOTYPE TRANSLATION

	CPIC	DPWG	System 1 ^a	System 2	System 3	System 4
+			(n=1)	(n=1)	(n=3)	(n=1)
	Activity Scores					
UM	>2	≥3	≥3	≥3	>2	Not tested
NM to			2.25 < x <	2.5		
UM			3			
NM	1 to 2	1.5 to 2.5	$1.75 \le x \le$	2	1.5 to 2	1 ^b to 2
			2.25			
IM to NM	_		1.25 < x <	1.5		
	\mathbf{H}		1.75			
IM	0.5	0.5 to 1	0.75≤x≤1.2	1	0.5 to 1	0.5 to 1 ^b
			5			
PM to IM			0 <x<0.75< th=""><th>0.5</th><th></th><th></th></x<0.75<>	0.5		
PM	0	0	0	0	0	0

UM: ultrarapid metabolizer; NM: normal metabolizer; IM: intermediate metabolizer; PM: poor metabolizer; Activity Score (AS) ranges shown in gray are not reported. n refers to the number of laboratories that reported using the system.

^bNM AS=1 is a combination of a fully functional allele plus a no function allele; IM AS=1 is a combination of two decreased function alleles.

^aThis laboratory utilizes a propriety system of values to determine AS.

Author M

76%-100%

	No. (%) respondents
	(n=37)
Workplace setting	
Laboratory test interpretation	3 (8)
Nonprofit or academic hospital	14 (38)
Reference/clinical laboratory	7 (19)
Research or clinical institute	3 (8)
University	10 (27)
% time related to work involving CY	/P2D6
0%-5%	2 (5)
6%-25%	16 (43)
26%-50%	11 (30)
51%-75%	6 (16)

2 (5)

TABLE 3. FINAL CONSENSUS CYP2D6 GENOTYPE TO PHENOTYPE TRANSLATION COMPARED TO PREVIOUSLY REPORTED CPIC AND DPWG METHODS

Inferred CYP2D6	Previous CPIC	Previous	Consensus	Consensus	Examples of CYP2D6 diplotypes for
Phenotype	Definition	DPWG	Definition	Contiguous	consensus translation method
-	(AS)	Definition	(AS)	Definition	
\overline{O}		(AS)		(AS)	
Ultrarapid	>2	>2.5	>2.25	>2.25	*1/*1xN, *1/*2xN ^b , *2 ^a /*2xN ^b , *1x2/*9
Metabolizer					
Normal Metabolizer	1-2	1.5-2.5	1.25	$1.25 \le x \le 2.25$	*1/*10
			1.5		*1/*41, *1/*9
M			2.0		*1/*1, *1/*2
			2.25		*2x2/*10
Intermediate	0.5	0.5-1	0.25	0 < x <1.25	*4/*10
Metabolizer			0.5		*4/*41, *10/*10,
			0.75		*10/*41
0			1		*41/*41, *1/*5,
Poor Metabolizer	0	0	0	0	*3/*4, *4/*4, *5/*5, *5/*6

^aCYP2D6*2 is currently considered to be a normal function allele by CPIC and DPWG; however, this function assignment has been challenged ³² and some laboratories report CYP2D6*2 function differently. Function of this allele will be reassessed as additional data become available.

^bN is categorical and indicates the number of copy variants, e.g. *1x2, *1x3, etc.

TABLE 4. DISCUSSION POINTS RAISED BY CYP2D6 EXPERTS DURING DELPHI PROCESS

Discussion Points	Pros	Cons
Addition of CYP2D6 rapid metabolizer phenotype Addition of new phenotype group between IM and PM	 Certain genotypes may have increased activity compared to NMs but less than UMs Addition of RM group would be in alignment with CYP2C19 Combining AS of 0.5 and 1.0 in research studies may mask potentially significant differences among these AS groups 	 Not enough evidence to differentiate between two increased function phenotypes (RM vs UM) and not clinically useful Many studies combine AS of 0.5 and 1.0 into one phenotype group Majority of experts rejected the addition of an additional phenotype group May not be clinically useful and would be outside the terms developed in the term standardization project 18
Changing AS of 1 from NM to IM	 More likely accepted by experts More clinical laboratories are already reporting an AS of 1.0 as an IM Providing recommendations for a "normal metabolizer" is confusing and 	• Institutions and laboratories following the CPIC guidelines will need update this phenotype translation and potentially re-contact previously tested patients to inform them of the

currently two CPIC guidelines provide	phenotype change with any associated
separate recommendations for AS of	clinical recommendations
1.0	
CYP2D6*10 has been characterized as	Institutions and laboratories following
	the CPIC guidelines will need update
decreased function across substrates	this phenotype translation and
More flexibility for assigning AS and	potentially re-contact previously tested
therefore, recommendations	patients to inform them of the
	phenotype change with any associated
	clinical recommendations
Can accommodate any future values	No currently used activity scores fall
of AS	in between the already listed ranges
	(e.g., there is no AS of 0.1)
May be more intuitive to clinicians	Little to no data exist for the vast
May be more accurate	majority of alleles to discriminate
	activity on a scale of 0.1 (10%
	increments)
	Broad range of interindividual
	variability among subjects within the
	same genotype group
	Percent activities may still need to be
	 CYP2D6*10 has been characterized as an allele conveying significantly decreased function across substrates More flexibility for assigning AS and therefore, recommendations Can accommodate any future values of AS May be more intuitive to clinicians

	translated into a limited number of
	phenotyping categories in order to
7	follow CPIC and/or DPWG guidelines

UM: ultrarapid metabolizer; RM: rapid metabolizer; NM: normal metabolizer; IM: intermediate metabolizer; PM: poor metabolizer;

CPIC: Clinical Pharmacogenetics Implementation Consortium; DPWG: Dutch Pharmacogenetics Working Group; AS: activity score

ACTIVITY VALUE STAR ALLELE **ALLELE GROUP** 18475-4+ CYP206*4 no function 10000>/ DIPLOTYPE" **ACTIVITY VALUES ACTIVITY SCORE** PHENOTYPE CYF206*1/*4 CYP206*1 - 1 1+0-1 CYP2D6 CYF2D6*6 - D intermediate metabolizer $cts_12692_f1.tif$