Gene name	Forward	Reverse
Atg7	CAGAAGAAGTTGAACGAGTA	CAGAGTCACCATTGTAGTAAT
Atg2b	CCTCCACTCTCAGAATCA	GTCCATCACACGAACATAA
Ulk1	ACCATTGTCTACCAGTGT	AGTGTCTTGTTCTTCTCATAA
LC3b	GCTCTTTGTTGGTGTGTA	TCTTCTGTTGCTGTTGTC
TFEB	CAGAAGCGAGAGCTAACAGAT	TGTGATTGTCTTTCTTCTGCCG
CTSA	GCTACCTCAGAGCATCGGAC	GTTAAGCCAAAGCACCACGG
GBA	GCCTCCCAGAAGAAGACACC	ATATCCCCTGGCTGACCCTT
GLA	TTGGGGTCAGAGCATTGGAC	AGTCATAACCTGCATCCCGC
PSAP	CAGCAGATGGTCTGGAGCAA	CAAGTTCCCAGCTTCGGTGA
CLCN7	CGAGATGCCTATCCACGCTT	CCCGGAAGAGCTTGAACACT
MCOLN1	TGGGCCAATGGATCAGCTTT	GTTCTTGTAACTGGCGCTGC
ATP6V1H	AAGACCAGCAGGTTCGCTAC	TGCAGAAAGGCTGAACAGGT
β-actin	GATCTGGCACCACACCTTCT	GGGGTGTTGAAGGTCTCAAA

S-Table 1. Primer information for qPCR used in this study

S-Table 2. Primer information for ChIP assay and EMSA used in this study

Chromatin Immunoprecipitation assay			
Gene name	Forward	Reverse	
Atg7	ACTGGGGCAAAGCAAAGGTA	CCATGGCTGTTCTGGACCTG	
Atg2b-Low	AGTCCGGGCGTCCGCCGGCG	GCGAGACCCAGACTCCCGGC	
Ulk1	CCAATTCCAGGACCCGACAC	GCAGACTGGGCATGTCAATG	
LC3b	AGTTAACAGATGCTCGCCCA	TGTGTGTGTCTCAGTCCGC	
TFEB-PPRE	TATCCTAGTGCCGGGAGCTT	TTTGGTTTCATCCGGCAGGT	
TFEB-CRE	ATCTAGTTGTAGAACAGCCA	ATCCCACCCCATAATTGCC	
Electrophoretic mobility shift assay			
LC3b-Foward	Biotin-ACCCTCGGTGACGCGCCGCGGGTCACCTGACTCGGCTGCG		
LC3b-Reverse	Biotin-CGCAGCCGAGTCAGGTGACTCGCGGCGCGTCACCGAGGGT		



S-Figure 1. Co-immunofluorescence staining of LC3b and HNF4 $\alpha$  in the liver tissue sections of WT and CREBH-KO mice. Co-immunofluorescence staining of LC3b and the hepatocyte-specific marker HNF4 $\alpha$  proteins in the frozen liver tissue sections from WT and CREBH-KO mice fed normal chow or fasted for 24 h. Liver tissue sections were stained with DAPI for nuclei (Blue). LC3b protein was visualized by green fluorescence, and HNF4 $\alpha$  protein was visualized by red fluorescence. Magnification: 40 ×.



S-Figure 2. Autophagosome size analysis as well as levels of CREBH and CREB in the livers of WT and CREBH-KO mice. (A) The sizes of autophagosomes in the liver tissues of WT and CREBH-KO mice under feeding or after fasting. Transmission electron micrograph (TEM) analysis was performed with liver tissue sections from CREBH-KO and WT mice under feeding or after 14h-fasting. The size of autophagosome in the liver of WT or CREBH-KO mice was quantified with the TEM images using ImageJ software. The autophagosome diameter, which reflect the size of autophagosome, was calculated based on the scale bar of the TEM image. Data represent mean  $\pm$  SD (n= 12 for WT-fed; 15 for KO-fed, WT-fast, and KO-fast). The change in hepatic autophagosome size between WT and CREBH KO mice was not statistically significant. (B) Expression and cleavage of CREBH protein were increased in mouse livers in response to fasting. WT and CREBH-KO mice were subjected to feeding or fasting for 6, 12, or 24 h before they were euthanized for liver tissue collection. Western blot analysis was performed with liver protein lysates pooled from three mice per group per genotype to determine the levels of the CREBH precursor [CREBH (P)] and its cleaved form [CREBH (A)]. Quantification of the CREBH precursor and cleaved proteins in the livers of WT mice in the fed state or after fasting for 6, 12, and 24 h. The CREBH protein signals, determined by Western blotting densitometry, were normalized to the  $\beta$ -actin signal. The fold changes in the levels of the precursor or activated form of CREBH were determined by comparing the protein signals to those obtained under the fed condition (defined as 1). (C) Western blot analysis of CREB with WT and CREBH-KO mouse liver protein lysates pooled from three mice per group per genotype. Levels of GAPDH were determined as a loading control.



S-Figure 3. CREBH regulate expression of the key autophagy genes in the liver in response to fasting. (A) Quantification of LC3b, ATG7, ATG2b, P62, and Ulk1 proteins in the liver tissues of WT mice in the fed state or after fasting for 6, 12, and 24 h. Protein signals, determined by Western blotting densitometry in Figure 3, were normalized to the  $\beta$ -actin signal. The fold changes in protein levels or ratios were determined by comparing the protein signals to those obtained under the fed condition (defined as 1). CREBH (P), CREBH precursor; CREBH (A), activated/cleaved form of CREBH; LC3-I, a cytosolic form of LC3b; LC3-II, lipid-modified LC3b form. (B) Quantification of LC3b, ATG7, ATG2b, P62, and Ulk1 proteins in the livers of CREBH-KO and WT mice infected with Ad overexpressing GFP or activated CREBH. Protein signals, determined by Western blotting densitometry in Figure 3, were normalized to the  $\beta$ -actin signal. The fold changes in protein signals to the section signal and Ulk1 proteins in the livers of CREBH-KO and WT mice infected with Ad overexpressing GFP or activated CREBH. Protein signals, determined by Western blotting densitometry in Figure 3, were normalized to the  $\beta$ -actin signal. The fold changes in protein levels or ratios were determined by comparing the protein signals to those obtained under the fed condition (defined as 1).



S-Figure 4. CREBH regulate expression of the genes involved in lysosomal biogenesis in the liver upon fasting. (A-C) Expression levels of the genes encoding functions in lysosomal hydrolase and accessory (A), lysosomal membrane (B), and lysosomal acidification (C) in the livers of CREBH-KO and WT mice under the feeding condition or after fasting for 6, 12, or 24 h. mRNA expression levels were determined by qPCR. Fold changes in mRNA levels were determined by comparison to the mRNA levels in one of the WT control mice under the feeding condition. Data represent mean  $\pm$  SEM (n=3). \* P $\leq$ 0.05. FD, feeding condition.

S-Figure 5. CRE- or PPRE- binding sites in the promoter regions of human and mouse autophagy genes

## Human LC3b gene promoter

### Mouse *Lc3b* gene promoter

Red highlighted: CRE-binding sequences; Transcription start site. Blue highlighted: PPRE-binding sequences

### Human ATG7 gene promoter

CTTACAGGCCAGACAGAGAAACTTGTCATAATCACTGCTGTATCTTCAACACCTGTCCAG TGCTTTGCACGCGCAGAGTATCTAACCAAGTATTGAGAGAGGGAATAACTTTATCTCACTGA CAGTGAGGAGCTGTGAGAATGATGGCTGGTTTAGAAAAGGCAGGTTGGTCACTGTCGACG TTCACTGGCCTTTTCCTACTAAAATTCTCATCTCCTGGCTCTCCACACCTGCCACCCTGA TGGCCCCTGTGCTGCGTTTGATGCCGCCTCTCCTGGAGAATGACCATGGTGATCATTCCT GTCATCCTCTGAAATCAAAAGAGAGAACGTGGGCACTTTCTTAAAAGCCTGAAGGGAATG TAGACATTCCGACATCTGGTAAGGGAGACGCTCTCCATCGCTTCCCCGGGGGGCGTCACC GCCCCTGATGCCCCGCCTCTCACGACTCAAGTTCCTCCACTGGTGCTGCGTGCTGCG GGCGCATTTCCCCGGCATGCCTCTGCTCCCTCCCACGCGCCGCCTCTCCAGTGGCAA GGCGCATTTCCCCGGCATGCCTCTGCTCCCTTGCCACGCGCCGCCTCCCAGTGGCAA

Red highlighted: CRE-binding sequences; Transcription start site. Blue highlighted: PPRE-binding sequence

## Mouse Atg7 gene promoter

 GTTATTGCGGTTGTGGGGGACCCTTTTTGTCGTTGTTGTTGTAGTGCGCATGCGCGCCGCGCGCTTCCGCGTT $TGTGTGGGGCTGGGGTTA \underline{TTACGTCA} TTGGGCCGCGCGCGCGCGCGCGCGCGCGCGCGCTGGGTTGCC$ 

Red highlighted: CRE-binding sequences; Blue highlighted: PPRE-binding sequence

## Human ATG2b gene promoter

GGTTTTGGATGATTAGGCTGTGTGTGCGTGTGAGAATGATCACATG<mark>TGGCGT</mark>CATGCTTTGTACAGAGC CTCAGACCACTGGGCCTCGTCCAGTGAGAGAGTCCTCTCTGGCGACATCACACGCGGAGCAGCCAGGGGC CACCTTAGATCTCAGATCTCTCAGAGCAATACTTTTCTGAACTGCCACTGTGCCTGGGTGGTTGG<u>GTTG</u> <u>GTGTC</u>ATGCTTCTGACTAGAGTAGATCGCGCATGTCCACCAGTGATACGTTGAGTCCTTACAGTTCCCC CCATGGAGTCCCATAAGCAGCTCCATCGAGATCTGTCAGCAAGTTGCAGGACCCCACAATGTTCTGAC ATGTTAAGACCCCCTTACATGACGAGTAGAGAGGCAGCTGAGGCCACAACCGTGTCTTCCTCTTGAAT GGAGCTAACTCGGAACCCCCGTTTTCTCTTCCTTTCTGCCCACCACTGAACATTGCCTTT<u>TAGATAACT</u> CAGTGTTTCTTCTAGATGTCATAGCAATAGACTTTCACTTTCATGAAGTTTGGGTACGATTTGGATTCT

### Mouse Atg2b gene promoter

Red highlighted: CRE-binding sequences; Transcription start site.

# Human ULK1 gene promoter

### Mouse *Ulk1* gene promoter

Red highlighted: CRE-binding sequences; 📥 Transcription start site.

## Human TFEB gene promoter

Blue highlighted: PPRE-binding sequences;  $rac{ranscription start site}$ .

#### Mouse *Tfeb* gene promoter

CAAAAGGGCAAGATCCAGGTCATGTACCACATTCCAAACGGAGACAGGATATATCGTGAAAACGGTG GGCTCTTGAGCAAGGGGTTTTACATCTCACACCTGCCGGATGAAACCAAAAATCATGCTCCGAGCACA GCTCCGTCAGATTGAGGGCACTATAATCAACTAATGAATTCTAGGCCACCTAGGGTTATTGGGTGAGT CTGTCTCCACGAAACAAAGCAATGGTAGAAATAACAATAAACCAGGTTTTAAGGCCAAGCACGTGAA CCCTGGAGTGTGGCTTCAGTGCTCTCGCAGACCTTGCCTGGAAATCTCACAAAGAGAACTGTTTTGATT ACAGTTAAGATATCCTGCCTTCCTTTATAACCTTTTACCTAATCAATTACAACAGTATCTGCTACACCC CAGGAAACGTCCCGTGATCTTATTTAAATTTCTCAGCAGTCCTTTGAAGATAACTGCGGGACTCACCAA AACAAAAGTATGGCTATGGTAAAAGACAAGGAGAAGCCCTTGCCCAGTTTCTCTGTCAACTTCCTCTT AGTGGTATACACTTTTAATCCCAGCACTTGAAAGACAGGTGGATCTCTGTGAATTTAAGGCCAGACTG **ATACTTACA**TAAATATATATTTAATAAACTAAAGCTCTGCTTCATACTGGCTTGTCCTGAAATTATAAT AATAATTAATAGAGCAACAACAATAATATT**GTTATTATTATTTA**ACAAAGAAATCAAAACTCTCCCC AAGTGGAAGTTGCTAAGGGATAGGGTAACTTCTCAGGTTGCTGCTGGCAATTATGGGGGGTGGGATCCT ATATTTTTCTCACACTTTCTGGAAAAGCAGCTTGCTATTGGTCCTTTTTCAAGATGTAATAATATAGAA AGTCATATCACGGGCCATCATTGACAGAAAAAAGAGAGAAAAATCTGAAAAGGGAATAAAGACAAA CGTTTAAAGTTAAAAATTGACTCCTAGGGAACAGCGGCCAGTAGAATGGAACTGCAAATCCCGGCGA GCCCTTCGCGGCTGCGACGGGGACTACATTTCCCAGCGGGCACAGCGGTAGGCCTATGGGGGGGCGGTC 

Red highlighted: CRE-binding sequences; Blue highlighted: PPRE-binding sequence



**S-Figure 6. CREBH regulates rhythmic expression of the genes involved in lysosomal biogenesis in the liver of mice under the circadian clock.** (A-F) Rhythmic expression levels of *CTSA*, *GBA*, *GLA*, *PSAP*, *CLCN7*, *MCOLN1*, and *ATP6V1H* genes in the livers of CREBH-KO and WT control mice across a 24-h circadian cycle. mRNA expression levels were determined by qPCR. Fold changes in mRNA levels were determined by comparison to levels in one of the WT control mice at the starting circadian time point. Each expression point represents a pooled liver sample from 3 to 5 mice per group per time point. Black and white bars represent circadian dark-light phases.



S-Figure 7. Analyses of cAMP levels in CREBH-KO or Bmal1-LKO mouse livers and CREBH mRNA levels in human NASH patient livers. (A) Levels of cAMP in liver tissues of CREBH-KO, Baml1-LKO, and WT control mice determined by Cyclic AMP Competitive ELISA Kit (Invitrogen). Each bar donates mean  $\pm$  SEM (n=4). Total protein concentrations (mg/mL) of homogenized liver tissues were determined for normalizing the sample loading. The mean value was expressed as pmol cAMP per mg of total liver proteins. (B) Expression levels of *CREBH* mRNA in livers of human patients with non-alcoholic steatohepatitis (NASH) and control individuals without NASH. The analysis presented in this graph was conducted on the dataset obtained from the National Center for Biotechnology Information (NCBI) Gene Expression Omnibus (GEO) Database (www.ncbi.nlm.nih.gov/geo/). The dataset we analyzed was deposited by Ahrens et al (*Cell Metab* 2013, 18: 296-302). Liver samples were obtained from patients undergoing liver biopsy for suspected NAFLD or intraoperatively for assessment of liver histology. Control samples were obtained from individuals excluded for liver malignancy and with liver histology that demonstrated absence of both cirrhosis and malignancy {Ahrens, 2013 #73}. Expression levels of mRNAs were based on the mRNA microarray data obtained from the GSE48452 dataset from the GEO database. The mean relative intensity for CREBH mRNA is shown. Each bar donates mean  $\pm$  SEM (n=12 for Control and 14 for NASH). \*  $P \leq 0.05$ .