ORIGINAL ARTICLE

Fat necrosis with an associated lymphocytic infiltrate represents a histopathologic clue that distinguishes cellular dermatofibroma from dermatofibrosarcoma protuberans

Shula A. Schechter MD 💿

Department of Pathology, University of Michigan, Ann Arbor, Michigan

Correspondence

Shula A. Schechter, MD, Department of Pathology, University of Michigan, 2800 Plymouth road, Building 35-1411, Ann Arbor, MI 48109-2800. Email: sschecht@med.umich.edu

Abstract

Background: Cellular dermatofibromas (CDFs) and dermatofibrosarcoma protuberans (DFSP) can be challenging to differentiate from one another. Morphologically, both entities commonly extend into the subcutis, exhibit high cellularity with limited cytologic atypia and have a mixed fascicular-to-storiform growth pattern. We sought to evaluate the significance of fat necrosis with an associated lymphocytic infiltrate as a histopathologic clue for distinguishing CDFs from DFSP.

Scott C. Bresler MD, PhD 💿 | Rajiv M. Patel MD 💿

Methods: We identified cases in our pathology database with a primary diagnosis of CDF or DFSP. Punch or excisional biopsy specimens with extension into the subcutis were selected. Previously biopsied lesions and specimens that did not interact with the subcutis were excluded. Histopathologic features were evaluated in hematoxylin and eosin stained sections.

Results: Fat necrosis with lymphocytic infiltrate was present in 20/20 cases of CDF. None of the 20 DFSP cases had fat necrosis with lymphocytic infiltrate although 4/20 had fat necrosis alone.

Conclusions: Fat necrosis with associated lymphocytic response can aid in the distinction between CDF and DFSP.

KEYWORDS

cellular dermatofibroma, dermatofibrosarcoma protuberans, fat necrosis, fibrous histiocytoma, histopathology

1 | INTRODUCTION

Dermatofibromas (DFs) are common benign dermal proliferations of fibroblasts and histiocytes. Characteristic histopathologic features include sparing of the papillary dermis, epidermal hyperplasia with basilar hyperpigmentation, entrapment of peripheral collagen bundles, and an admixed chronic inflammatory infiltrate. Of the many variants of DFs, cellular dermatofibromas (CDFs) are relatively uncommon, accounting for approximately 5% to 10% of dermatofibromas.^{1,2} In comparison to conventional DFs, CDFs tend to be larger in size and have a higher recurrence rate that ranges from 10% to 26%.^{1,3} Histopathologic features differentiating CDFs from conventional DFs

include: increased cellularity, frequent extension into the subcutis, increased mitotic rate, and a fascicular-to-storiform growth pattern.¹ These features overlap with characteristics of dermatofibroma sarcoma protuberans (DFSP) and can create a diagnostic dilemma.

DFSPs are low-grade, locally aggressive cutaneous sarcomas comprised of monotonous spindle cells. The cells are characteristically arranged in a storiform pattern with neoplastic cells infiltrating subcutaneous tissue and encasing individual adipocytes resulting in a honeycomb appearance. DFSPs are considered neoplasms of intermediate potential with a high rate of local recurrence of up to 50%.⁴ However, metastasis is an uncommon event, which usually occurs in the setting of fibrosarcomatous transformation.⁵ 914 WILEY JOURDOF P

TABLE 1	Comparison of clinical features of CDF and DFSP
---------	---

CDF	DFSP	
34	49	
10-61	27-86	
8:12	9:11	
Location (%)		
8/20 (40)	7/20 (35)	
5/20 (25)	1/20 (5)	
5/20 (25)	10/20 (50)	
0/20 (0)	2/20 (10)	
	34 10-61 8:12 8/20 (40) 5/20 (25) 5/20 (25)	

Immunohistochemical stains and molecular studies can aid in distinguishing CDFs from DFSPs. CD34 positivity occurs in approximately 90% of DFSPs but is also present in up to 25% of CDFs.^{3,6} However, CD34 positivity in CDFs is limited to the periphery as compared to the diffuse pattern of staining in DFSPs. Also, in contrast to CDFs, immunohistochemistry of DFSP shows an absence of CD163 and factor XIIIa,^{3,7} although factor XIIIa positivity in dermal dendrocytes can confound interpretation. In diagnostically challenging cases, ancillary molecular tests can be exploited and the presence of COL1A1-PDGFB fusions in DFSPs by can aid in distinguishing DFSPs from CDFs.

We have frequently observed fat necrosis with an associated lymphocytic infiltrate in CDFs and sought to evaluate this as histopathologic clue for distinguishing CDFs from DFSPs, in conjunction with other histopathologic features.

2 MATERIALS AND METHODS

With approval from University of Michigan Institutional Review Board, cases of CDF and DFSP were identified through a retrospective search of a University of Michigan Department of Pathology database for the period between 2008 and 2017. Punch or excisional biopsy specimens with extension into the subcutis were selected. Previously biopsied lesions and specimens that did not interact with the subcutis were excluded. The diagnosis was confirmed using hematoxylin and eosin-stained sections of paraffin-embedded formalin-fixed tissue of each case and previously performed immunohistochemical stains. Histopathologic features of each case were evaluated, including the presence of peripheral collagen trapping, honeycombing, chronic inflammation (including lymphocytes or plasma cells within the lesion), floret-like giant cells, epidermal hyperplasia, Grenz zone, and fat necrosis with associated lymphocytic infiltrate.

RESULTS 3

We identified 40 cases, of which 20 were CDFs and 20 were DFSPs. Clinical characteristics of the cohort are summarized in Table 1. CDF patients were younger than DFSP patients, with mean ages of 34 and

TABLE 2 Comparison of pathologic features of CDF and DFSP

	CDF (%)	DFSP (%)
Fat necrosis with lymphocytic infiltrate	20/20 (100)	0/20 (0)
Peripheral collagen trapping	20/20 (100)	14/20 (70)
Chronic inflammation	17/20 (85)	6/20 (30)
Epidermal hyperplasia	15/20 (75)	9/20 (45)
Grenz zone	10/20 (50)	9/20 (45)
Hemosiderin	5/20 (25)	2/20 (20)
Floret-like giant cells	4/20 (20)	1/20 (5)
Necrosis	2/20 (10)	0/20 (0)
Honeycombing	2/20 (10)	17/20 (85)

49, respectively (Table 1). CDFs and DFSPs were commonly located on the trunk or the lower limb/limb girdle; each affected equal pronortions of men and women

CDFs and DFSPs differed in a number of histopathologic features (Table 2; Figure 1). Fat necrosis with associated lymphocytic infiltrate was seen in all cases of CDF (20/20) and none of the cases of DFSP (0/20). The lymphocytic infiltrate was consistently observed in areas of fat necrosis and was present at the junction of the dermis and subcutis, often with perivascular accentuation. Although fat necrosis was seen in 4/20 cases of DFSP, these cases showed a pseudomembranous type of fat necrosis without any associated lymphocytic infiltrate. However, pseudomembranous fat necrosis does not seem to be specific for DFSP, as this pattern was also observed in many cases of CDF. Honeycombing was more common in DFSP (17/20 vs 2/20 with CDF). Peripheral collagen trapping occurred in 14/20 cases of DFSP and all cases of CDF (Table 2: Figure 2). The presence of floret-like giant cells, Grenz zone, epidermal hyperplasia, or hemosiderin-laden macrophages also did not distinguish between CDF and DFSP.

Immunohistochemistry showed a diffuse pattern of CD34 staining in all cases of DFSP. Among CDF cases, 6/20 demonstrated patchy peripheral CD34 staining, on the other hand, 1/20 had weak CD34 positivity throughout with a more typical accentuation at the periphery.

DISCUSSION 4

CDFs and DSFPs share a number of clinical and histopathological features. Nevertheless, it is imperative to differentiate CDF from DFSP, which is more likely to be locally aggressive and requires wide local excision.

Clinical features such as age, sex, and lesion location have been of limited value in discriminating between CDF and DFSP.^{1,3,8-11} DFSP sometimes presents as a raised plaque, but it may also present as a dermal nodule similar to CDF. Although patients who present with DFSP are older on average than those who present with CDF, both CDF and DFSP can occur in individuals of any age. Furthermore, they appear to occur equally in males and in females. DFSP may be

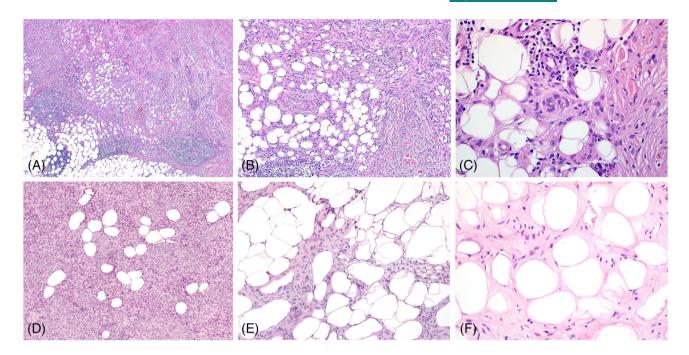


FIGURE 1 A and B, CDF with fat necrosis and associated lymphocytic infiltrate at the junction of the dermis and subcutis with perivascular accentuation (C) (H&E, \times 40 and \times 100, \times 400). D, DFSP with "honeycomb" infiltration of fat without necrosis or inflammation (H&E, \times 100). E and F, DFSP with fat necrosis without lymphocytic infiltrate (H&E, \times 200)

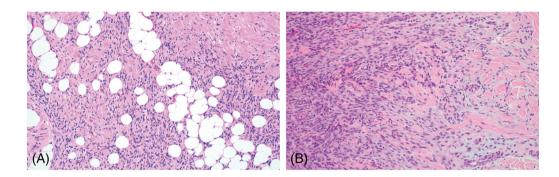


FIGURE 2 A, DFSP with peripheral collagen trapping (H&E, ×200). B, CDF with peripheral collagen trapping (H&E, ×200)

somewhat more frequent on the trunk than CDF, but with both CDF and DFSP occurrence is commonly on an extremity and less often on the head and neck.^{1,3,8-11}The clinical presentations of CDF and DFSP in our cohort are similar to those reported in previous studies and do not help to distinguish between CDFs and DSFPs.

Traditionally, CDFs and DSFPs have also been difficult to distinguish based on histopathology. CDFs are cellular proliferations of spindled cells with a fascicular-to-storiform growth pattern. They commonly extend into the subcutis, mimicking DFSP, which also has a storiform appearance. However, in the present study, we found that histopathologic features did allow CDFs to be differentiated from DFSPs (Table 2; Figure 1). In all 20 cases of CDF in our sample, we identified a lymphocytic infiltrate in association with fat necrosis, on the other hand, this finding was not observed in any of the 20 cases of DFSP. Other helpful histopathologic features in differentiating CDF from DFSP were honeycombing and chronic inflammation. Consistent with previous descriptions of CDF, floret-like giant cells, grenz zone, epidermal hyperplasia, and hemosiderin-laden macrophages did not reliably distinguish between CDF and DFSP.^{1,11} Peripheral collagen trapping was present in all CDF cases in our sample but, unexpectedly, it was also present in the majority of DFSP cases (Table 2; Figure 2).

The etiology of CDF is unclear; however, contributors to DF development have been studied and debated. Based on the presence of clonal aberrations and recurrent translocations, at least a subset of DFs is probably neoplastic.¹²⁻¹⁴ It has also been suggested that DFs are a reactive process arising in response to inflammation or trauma.^{15,16}

Fat necrosis with associated lymphocytic response has not been described previously but occurs consistently in CDF in our cohort. The absence of this finding in DFSP has important clinical implications as it may serve as an additional histopathologic clue to distinguish between CDF and DFSP.

ORCID

 Shula A. Schechter
 https://orcid.org/0000-0001-9202-5405

 Scott C. Bresler
 https://orcid.org/0000-0003-2504-466X

 Rajiv M. Patel
 https://orcid.org/0000-0002-1521-4947

REFERENCES

- Calonje E, Mentzel T, Fletcher CD. Cellular benign fibrous histiocytoma. Clinicopathologic analysis of 74 cases of a distinctive variant of cutaneous fibrous histiocytoma with frequent recurrence. *Am J Surg Pathol.* 1994;18(7):668-676.
- Han TY, Chang HS, Lee JH, Lee WM, Son SJ. A clinical and histopathological study of 122 cases of dermatofibroma (benign fibrous histiocytoma). *Ann Dermatol*. 2011;23(2):185-192.
- Gaufin M, Michaelis T, Duffy K. Cellular dermatofibroma: clinicopathologic review of 218 cases of cellular dermatofibroma to determine the clinical recurrence rate. *Dermatol Surg.* 2019;45(11):1359-1364.
- Fletcher CD, Hogendoorn P, Mertens F, Bridge J. WHO Classification of Tumours of Soft Tissue and Bone. 4th ed. Lyon, France: IARC Press; 2013.
- Hoesly PM, Lowe GC, Lohse CM, Brewer JD, Lehman JS. Prognostic impact of fibrosarcomatous transformation in dermatofibrosarcoma protuberans: a cohort study. J Am Acad Dermatol. 2015;72(3): 419-425.
- Goldblum JR, Tuthill RJ. CD34 and factor-XIIIa immunoreactivity in dermatofibrosarcoma protuberans and dermatofibroma. *Am J Dermatopathol*. 1997;19(2):147-153.
- Sachdev R, Sundram U. Expression of CD163 in dermatofibroma, cellular fibrous histiocytoma, and dermatofibrosarcoma protuberans: comparison with CD68, CD34, and factor XIIIa. J Cutan Pathol. 2006; 33(5):353-360.
- 8. Bowne WB, Antonescu CR, Leung DH, et al. Dermatofibrosarcoma protuberans: a clinicopathologic analysis of patients treated and followed at a single institution. *Cancer*. 2000;88(12):2711-2720.

- Kreicher KL, Kurlander DE, Gittleman HR, Barnholtz-Sloan JS, Bordeaux JS. Incidence and survival of primary dermatofibrosarcoma protuberans in the United States. *Dermatol Surg.* 2016;42(Suppl 1): S24-S31.
- Criscito MC, Martires KJ, Stein JA. Prognostic factors, treatment, and survival in dermatofibrosarcoma protuberans. JAMA Dermatol. 2016; 152(12):1365-1371.
- 11. Volpicelli ER, Fletcher CD. Desmin and CD34 positivity in cellular fibrous histiocytoma: an immunohistochemical analysis of 100 cases. *J Cutan Pathol.* 2012;39(8):747-752.
- Hui P, Glusac EJ, Sinard JH, Perkins AS. Clonal analysis of cutaneous fibrous histiocytoma (dermatofibroma). J Cutan Pathol. 2002;29(7): 385-389.
- Chen TC, Kuo T, Chan HL. Dermatofibroma is a clonal proliferative disease. J Cutan Pathol. 2000;27(1):36-39.
- Plaszczyca A, Nilsson J, Magnusson L, et al. Fusions involving protein kinase C and membrane-associated proteins in benign fibrous histiocytoma. *Int J Biochem Cell Biol*. 2014 Aug;53:475-481.
- Calonje E. Is cutaneous benign fibrous histiocytoma (dermatofibroma) a reactive inflammatory process or a neoplasm? *Histopathology*. 2000; 37(3):278-280.
- Zelger BG, Zelger B. Dermatofibroma (fibrous histiocytoma): an inflammatory or neoplastic disorder? *Histopathology*. 2001;38(4): 379-381.

How to cite this article: Schechter SA, Bresler SC, Patel RM. Fat necrosis with an associated lymphocytic infiltrate represents a histopathologic clue that distinguishes cellular dermatofibroma from dermatofibrosarcoma protuberans. *J Cutan Pathol.* 2020;47(10):913–916. https://doi.org/10.

1111/cup.13744