

New Phytologist Supporting Information

Article title: **C₃ plant carbon isotope discrimination does not respond to CO₂ concentration on decadal to centennial timescales**

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Article acceptance date: 13 October 2020

The following Supporting Information is available for this article:

Fig. S1 [CO₂] versus $\Delta^{13}\text{C}_{\text{plant}}$ values for historical species with medium-resolution records of Industrialization.

Fig. S2 $\Delta^{13}\text{C}_{\text{plant}}$ values of plants by genus.

Fig. S3 $\Delta^{13}\text{C}_{\text{plant}}$ values of plants by family.

Table S1 All data in .xlsx file. **(See separate file).**

Table S2 Range of climate variables and atmospheric parameters included in the historical portion of this study.

Table S3 T-test statistics (assuming unequal variances) comparing the means and ranges of $\Delta^{13}\text{C}_{\text{plant}}$ values for different taxa in this study.

Table S4 The relationship between $\delta^{13}\text{C}_{\text{CO}_2}$ and $\delta^{13}\text{C}_{\text{plant}}$ values for the historical specimens spanning Industrialization.

Table S5 Sensitivity (S given as $\% \text{ ppm}^{-1}$) for eight species with long historical record.

Table S6 The relationship between $\Delta^{13}\text{C}_{\text{plant}}$ values and non-barometric climate variables for historical and modern species sampled in this study.

Fig. S1 $[\text{CO}_2]$ values plotted against $\Delta^{13}\text{C}_{\text{plant}}$ for species with medium-resolution records of the period of Industrialization. Data are for: (a) *Pinus strobus* (Eastern white pine), (b) *Thuja sutchuenensis* (Sichuan arborvitae), (c) *Thuja koraiensis* (Korean arborvitae), (d) *Thuja standishii* (Japanese arborvitae), and (e) *Platycladus orientalis* (Chinese arborvitae). The outer panel shows change in $\Delta^{13}\text{C}_{\text{plant}}$ vs. $[\text{CO}_2]$ over Industrialization, while the inner panel shows the range and distribution of $\Delta^{13}\text{C}_{\text{plant}}$ values for this species. Each of the species occupies different geographic ranges and different ranges of climatic variability, but none shows a significant $\Delta^{13}\text{C}_{\text{plant}}$ response to rising $[\text{CO}_2]$ over the period of Industrialization.

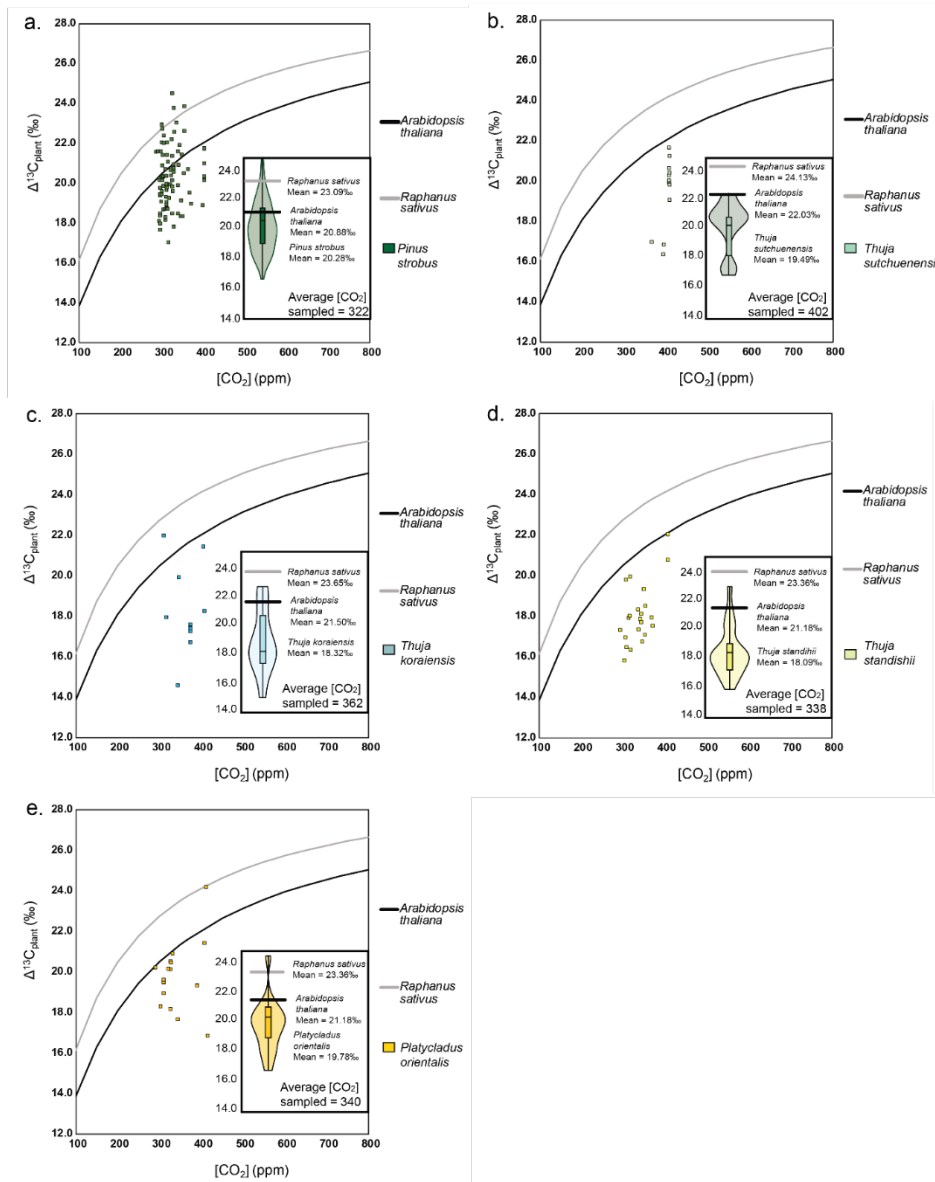


Fig. S2 Comparison of $\Delta^{13}\text{C}_{\text{plant}}$ values of plants as sampled in this study, the Schubert & Jahren (2012) model, and from the literature and sorted by genera. Angiosperms are shown to the left of the diagram, while gymnosperms are shown to the right. Genera for which $n < 10$ samples within the literature, were excluded from this figure. Xs denote mean values for each genus, and lines denote median values for each genus. Each box and whisker in this figure shows a different genus with $n \geq 15$ represented isotope values from this study, Sheldon et al. (2020) and literature. Boxes show the 75th percentile of data, while whiskers show the remaining 25th percentile of data. Individual dots show outliers.

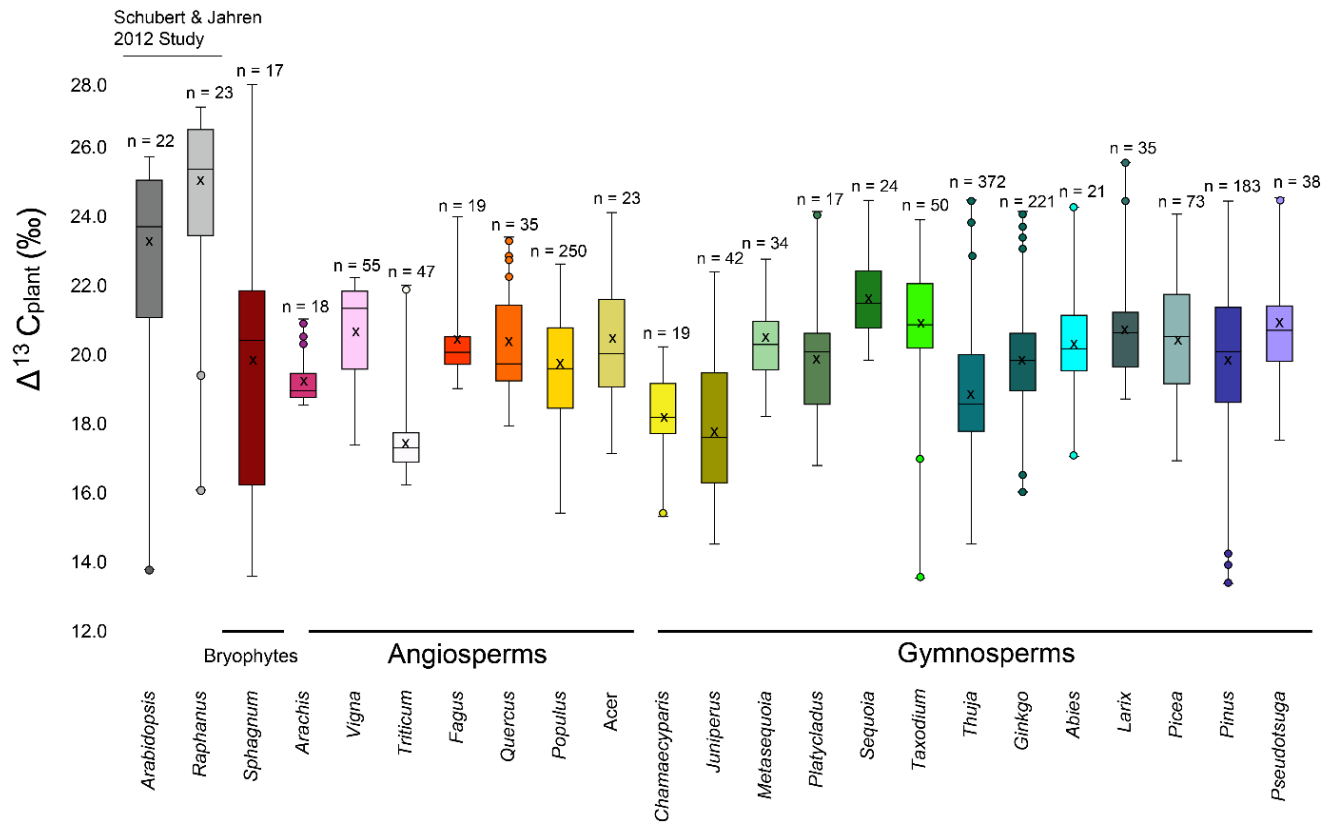


Fig. S3 Comparison of $\Delta^{13}\text{C}_{\text{plant}}$ values of plants as sampled in this study, the Schubert & Jahren (2012) model, and from the literature and sorted by family. Angiosperms are shown to the left of the diagram, while gymnosperms are shown to the right. Families for which $n < 10$ samples within the literature were excluded from this figure, except for “other Brassicaceae” – members of the same family as *Arabidopsis* and *Raphanus*, for comparison. Xs denote mean values for each genus. Each box and whisker in this figure shows a different family with $n \geq 15$ represented isotope values from this study, Sheldon et al. (2020) and literature (except for Brassicaceae, which is included to show the discrepancy between model species and other Brassicaceae). Boxes show the 75th percentile of data, while whiskers show the remaining 25th percentile of data.

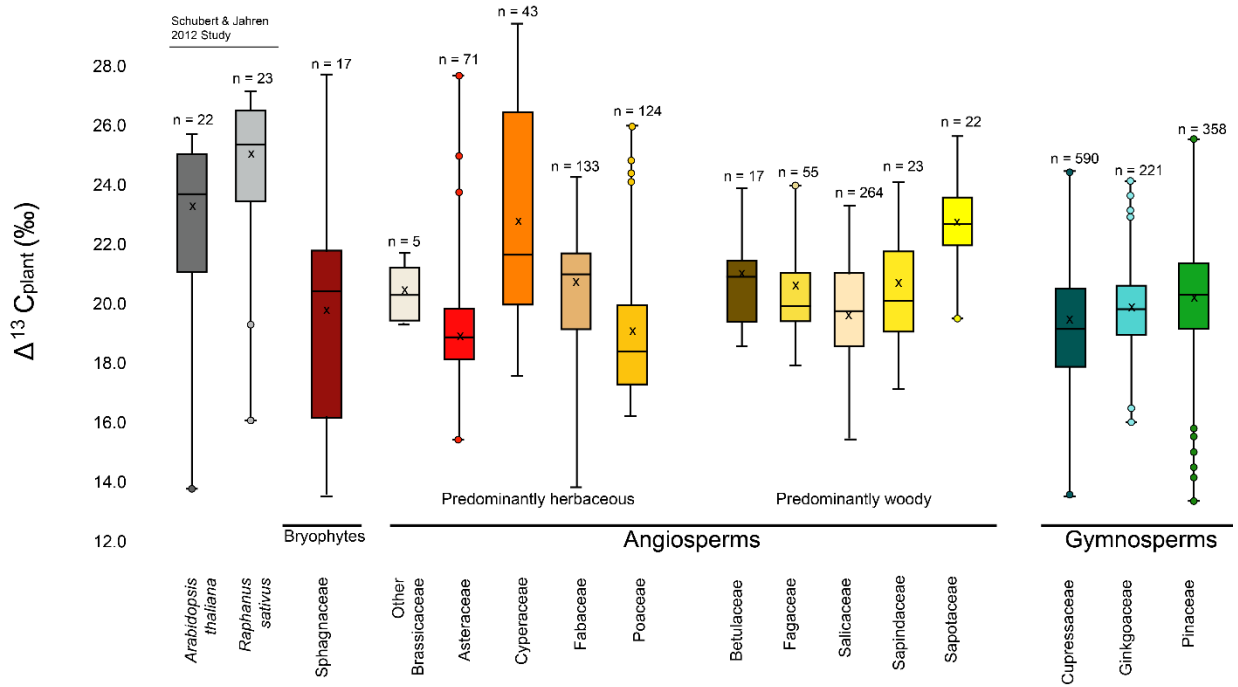


Table S1 Isotope data measurements for specimens collected in the Earth Systems Science Lab (tab labeled *ESS Lab*) and from literature (tab labeled *Literature*). References for literature and ESS data shown in tab labeled *References*. $\delta^{13}\text{C}_{\text{plant}}$ values and $\Delta^{13}\text{C}_{\text{plant}}$ values (calculated using Eq. 2) are listed for ESS Lab and Literature values when known. Climate (PRISM Climate Group 2004) and barometric data (Etheridge et al. 1998; Keeling et al. 2001; White et al. 2015; Eggleston et al. 2016), year, collector, associated publications are listed when known and applicable. N/a denotes information not available. Eq. 2 (see details in main text):

$$\Delta^{13}\text{C}_{\text{plant}} = \frac{\delta^{13}\text{C}_{\text{CO}_2} - \delta^{13}\text{C}_{\text{plant}}}{1 + \frac{\delta^{13}\text{C}_{\text{plant}}}{1000}}$$

(for full table, see separate file).

Table S2 Range of climate variables included in the historical portion of this study, as collected from PRISM Climate Group (2004), White et al. (2015), and Eggleston (2016).

Climate, Atmospheric or Temporal Variable	Minimum	Maximum	Mean
Time (Year)	1804	2019	1971
[CO ₂] (ppm)	283	411	351
δ ¹³ C _{CO₂} (‰)	-8.7	-6.4	-7.6
Latitude (°N)	9.9	68.2	42.5
Longitude (°E)	-152.3	139.9	-86.7
Mean Annual Precipitation (mm yr ⁻¹)	193	4368	999
Mean Annual Temperature (°C)	-5.6	25.0	8.0
Maximum Summer Temperature (°C)	13.3	35.9	26.2
Elevation (m)	4	2766	967
Annual Minimum Vapor Pressure Deficit (hPa)	0.18	6.32	1.23
Annual Maximum Vapor Pressure Deficit (hPa)	0.63	21.87	10.58

Table S3 Results from t-tests (assuming unequal variances) comparing the means and ranges of *Arabidopsis* (a) and *Raphanus* (b) and families highly sampled from the literature dataset. N/a denotes that the comparison to the model species is not applicable (in the cases that it would involve comparing the model species to itself).

Taxon	Mean	Standard Deviation	Sample Size	t-Stat	p(T<=t) two-tail value
<i>Arabidopsis</i>	22.66	10.65	22	n/a	n/a
Asteraceae	18.76	1.78	68	5.44	4.14 x 10 ⁻⁵
Other Brassicaceae	20.37	0.93	5	2.80	0.02
Cupressaceae	19.28	3.49	584	4.83	7.72 x 10 ⁻⁵
Cyperaceae	20.78	11.01	212	2.58	3.39 x 10 ⁻³
Fabaceae	20.61	2.80	134	2.89	7.76 x 10 ⁻³
Fagaceae	20.34	1.97	54	3.22	4.25 x 10 ⁻³
Ginkgoaceae	19.93	2.21	221	3.89	7.88 x 10 ⁻⁴
Pinaceae	20.24	3.45	356	3.45	2.28 x 10 ⁻³
Poaceae	18.88	4.20	119	5.25	3.79 x 10 ⁻⁵
Salicaceae	19.77	2.60	261	4.12	4.48 x 10 ⁻⁴
Taxon	Mean	Standard Deviation	Sample Size	t-Stat	p(T<=t) two-tail value
<i>Raphanus</i>	24.47	8.16	23	n/a	n/a
Asteraceae	18.76	1.78	68	9.23	2.37 x 10 ⁻⁹
Other Brassicaceae	20.37	0.93	5	5.33	1.37 x 10 ⁻⁴
Cupressaceae	19.28	3.49	584	8.64	1.07 x 10 ⁻⁸
Cyperaceae	20.78	11.01	212	5.80	1.23 x 10 ⁻⁵
Fabaceae	20.61	2.80	134	6.30	1.25 x 10 ⁻⁶
Fagaceae	20.34	1.97	54	6.61	5.34 x 10 ⁻⁷
Ginkgoaceae	19.93	2.21	221	7.52	1.21 x 10 ⁻⁷
Pinaceae	20.24	3.45	356	7.01	3.79 x 10 ⁻⁷
Poaceae	18.88	4.20	119	8.95	2.60 x 10 ⁻⁹
Salicaceae	19.77	2.60	261	7.79	6.69 x 10 ⁻⁸

Table S4 Results showing the relationship between $\delta^{13}\text{C}_{\text{CO}_2}$ and $\delta^{13}\text{C}_{\text{plant}}$ for each species studied over Industrialization. The first row shows the universal relationship proposed by Arens et al. (2000), and the second shows a generalized relationship that incorporates all 8 of the species we tested. Specimens sampled post-2017 were excluded to focus on the historical portion of this study.

Measurement	Slope	Intercept
Arens et al. (2000)	1.05	-18.72
Total Historical Dataset (pre-2017)	1.22	-17.10
<i>Pinus strobus</i>	1.58	-15.54
<i>Platyclusus orientalis</i>	1.47	-15.72
<i>Populus tremuloides</i>	1.08	-18.77
<i>Thuja koraiensis</i>	0.36	-22.84
<i>Thuja occidentalis</i>	1.31	-15.66
<i>Thuja plicata</i>	0.95	-18.53
<i>Thuja standishii</i>	2.46	-6.77
<i>Thuja sutchuenensis</i>	6.42	27.19

Table S5 Sensitivity (S given as $\% \text{ ppm}^{-1}$) for eight species with long historical record. S is compiled for the entire range of $[\text{CO}_2]$ values spanning the collection, and as pre-1960 (acceleration; Keeling et al. 2001) and post-1960 ranges. Expected sensitivity is calculated using the best-fit function, $S (\% \text{ ppm}^{-1}) = 0.21(28.26)^2 / [28.26+0.21(\text{CO}_2 + 25)]^2$ from Schubert & Jahren's (2012) study using data from both *Arabidopsis thaliana* and *Raphanus sativus* growth chamber experiments. S-column cells highlighted in blue indicate actual sensitivities far more negative than expected. S-column cells in red indicate sensitivities far more positive than expected, and S-column cells in white indicate sensitivities matching what is expected. N/a is used when there are no samples collected of the species during the specified time period.

Genus	Species	Age (Range in $[\text{CO}_2]$)	S ($\%/\text{ppm}$)	Standard Deviation ($\%/\text{ppm}$)
<i>Pinus</i>	<i>strobis</i>	Total (286-395)	-0.0117	3.019
		<1960 (286-320)	-0.0421	n=51
		>1960 (320-395)	0.0014	
<i>Platycladus</i>	<i>orientalis</i>	Total (290-408)	0.0077	1.642
		<1960 (290-320)	-0.0045	n=12
		>1960 (320-408)	0.0346	
<i>Populus</i>	<i>tremuloides</i>	Total (288-411)	0.0078	9.078
		<1960 (288-320)	-0.0441	n=79
		>1960 (320-411)	-0.0011	
<i>Thuja</i>	<i>koraensis</i>	Total (311-408)	-0.0411	0.536
		<1960 (311-320)	-0.4977	n=6
		>1960 (320-408)	0.0495	
<i>Thuja</i>	<i>occidentalis</i>	Total (283-408)	0.0128	4.497
		<1960 (283-320)	0.0417	n=85
		>1960 (320-408)	-0.0012	
<i>Thuja</i>	<i>plicata</i>	Total (289-411)	0.0161	4.164
		<1960 (289-320)	-0.0085	n=163
		>1960 (320-411)	0.0338	
<i>Thuja</i>	<i>standishii</i>	Total (292-408)	0.0333	1.583
		<1960 (292-320)	0.0592	n=15
		>1960 (320-408)	0.0474	
<i>Thuja</i>	<i>sutchuenensis</i>	Total (366-408)	0.0851	0.203
		<1960 (n/a)	n/a	n=4
		>1960 (366-408)	0.0851	
Expected Sensitivity		<1960 (280-320)	0.0181	
		>1960 (320-408)	0.0139	

Table S6 Results showing the statistical relationship between carbon isotope discrimination and non-[CO₂] climate variables for each species studied over Industrialization for each species studied over Industrialization. Specimens collected in modern times were included. N/a denotes when climate variables were not accessible for the specified species.

a						
Species	R ² value					
	Mean Annual Precipitation (mm yr ⁻¹)	Mean Annual Temperature (°C)	Maximum Summer Temperature (°C)	Elevation (m)	Latitude (°N)	
<i>Pinus strobus</i>	4.00 x 10 ⁻³	0.01	0.01	0.06	0.06	
<i>Platycladus orientalis</i>	7.00 x 10 ⁻⁴	0.01	0.74	0.10	3.47 x 10 ⁻⁶	
<i>Populus tremuloides</i>	0.05	0.01	3.00 x 10 ⁻³	0.08	0.15	
<i>Thuja occidentalis</i>	1.00 x 10 ⁻³	0.01	4.00 x 10 ⁻³	1.00 x 10 ⁻³	0.01	
<i>Thuja koraiensis</i>	0.15	0.76	0.32	0.10	0.37	
<i>Thuja plicata</i>	0.06	0.02	0.07	0.04	0.05	
<i>Thuja standishii</i>	0.55	0.34	n/a	0.03	0.25	
<i>Thuja sutchuenensis</i>	0.03	0.06	n/a	0.44	0.01	
Overall historical study	0.02	0.01	0.01	0.00	4.00 x 10 ⁻³	
b						
Species	p-value					
	Mean Annual Precipitation (mm yr ⁻¹)	Mean Annual Temperature (°C)	Maximum Summer Temperature (°C)	Elevation (m)	Latitude (°N)	
<i>Pinus strobus</i>	0.62	0.33	0.43	0.04	0.03	
<i>Platycladus orientalis</i>	0.93	0.77	0.00	0.27	0.99	
<i>Populus tremuloides</i>	1.00 x 10 ⁻³	0.20	0.50	7.88 x 10 ⁻⁶	8.87 x 10 ⁻¹⁰	
<i>Thuja occidentalis</i>	0.72	0.30	0.47	0.66	0.23	
<i>Thuja koraiensis</i>	0.52	0.05	0.43	0.53	0.08	
<i>Thuja plicata</i>	2.00 x 10 ⁻³	0.09	3.00 x 10 ⁻³	8.00 x 10 ⁻³	0.01	
<i>Thuja standishii</i>	0.09	0.22	n/a	0.58	0.06	
<i>Thuja sutchuenensis</i>	0.56	0.46	n/a	0.02	0.75	
Overall historical study	4.00 x 10 ⁻³	0.65	0.24	0.88	0.01	

References

- Araus, J. L., Reynolds, M. P., & Acevedo, E. (1993). Leaf posture, grain yield, growth, leaf structure, and carbon isotope discrimination in wheat. *Crop Science*, 33(6), 1273-1279.
- Arens N. C., Jahren A. H. (2000). Carbon isotope excursion in atmospheric CO₂ at the Cretaceous-Tertiary boundary: evidence from terrestrial sediments. *Palaios*, 15(4), 314-322.
- Bai E., Boutton T., Liu F., Wu X., Archer S. (2008). Variation in woody plant $\delta^{13}\text{C}$ along a topographic gradient in a subtropical savanna parkland. *Oecologia*, 156:479-489.
- Berling D. J., Woodward F. I. (1995). Stomatal responses of variegated leaves to CO₂ enrichment. *Annals of Botany*, 75(5), 507-511.
- Berling D. J. (1996). ^{13}C discrimination by fossil leaves during the late-glacial climate oscillation 12-10 ka BP: measurements and physiological controls. *Oecologia*, 108(1), 29-37.
- Bonal D., Sabatier D., Montpied P., Tremeaux D., & Guehl J. M. (2000). Interspecific variability of $\delta^{13}\text{C}$ among trees in rainforests of French Guiana: functional groups and canopy integration. *Oecologia*, 124, 454-468.
- Brooks J. R., Flanagan L. B., Buchmann N., & Ehleringer J. R. (1997). Carbon isotope composition of boreal plants: functional grouping of life forms. *Oecologia*, 110:301-311.
- Broadmeadow M. S. J., Griffiths H., Maxwell C., & Borland A. M. (1992). The carbon isotope ratio of plant organic material reflects temporal and spatial variations in CO₂ within tropical forest formations in Trinidad. *Oecologia*, 89(3), 435-441.
- Bruhl J. J., & Wilson K. L. (2007). Towards a comprehensive survey of C₃ and C₄ photosynthetic pathways in Cyperaceae. *Aliso: A Journal of Systematic and Evolutionary Botany*, 23(1), 99-148.
- Buchmann N., Guehl J. M., Barigah T. S., & Ehleringer J. R. (1997). Interseasonal comparison of CO₂ concentrations, isotopic composition, and carbon dynamics in an Amazonian rainforest (French Guiana). *Oecologia*, 110, 120-131.
- Chevillat V. S., Siegwolf R. T. W., Pepin S., & Körner C. (2005). Tissue-specific variation of $\delta^{13}\text{C}$ in mature canopy trees in a temperate forest in central Europe. *Basic and applied ecology*, 6, 519-534.
- Collister J. W., Rieley G., Stern B., Eglinton G., & Fry B. (1994). Compound-specific $\delta^{13}\text{C}$ analyses of leaf lipids from plants with differing carbon dioxide metabolisms. *Organic Geochemistry*, 21, 619-627.

- Condon A. G., Richards R. A., & Farquhar G. D. (1993). Relationships between carbon isotope discrimination, water use efficiency and transpiration efficiency for dryland wheat. *Australian Journal of Agricultural Research*, 44(8), 1693-1711.
- Dawson T. E., Ehleringer J. R. (1993). Isotopic enrichment of water in the “woody” tissues of plants: implications for plant water source, water uptake, and other studies which use the stable isotopic composition of cellulose. *Geochimica et Cosmochimica Acta*, 57(14), 3487-3492.
- De Lillis M., Matteucci G., & Valentini R. (2004). Carbon assimilation, nitrogen, and photochemical efficiency of different Himalayan tree species along an altitudinal gradient. *Photosynthetica*, 42, 597-605.
- DeLucia E. H., & Schlesinger W. H. (1991). Resource-use efficiency and drought tolerance in adjacent Great Basin and Sierran plants. *Ecology*, 72, 51-58.
- Diefendorf, A. F., Mueller, K. E., Wing, S. L., Koch, P. L., & Freeman, K. H. (2010). Global patterns in leaf ^{13}C discrimination and implications for studies of past and future climate. *Proceedings of the National Academy of Sciences*, 107(13), 5738-5743.
- Dodd M. B., Lauenroth W. K., & Welker J. M. (1998). Differential water resource use by herbaceous and woody plant life-forms in a shortgrass steppe community. *Oecologia*, 117, 504-512.
- Donovan L. A., Ehleringer J. R., (1991). Ecophysiological differences among juvenile and reproductive plants of several woody species. *Oecologia*, 86, 594-597.
- Dungait J. A. J., Docherty G., Straker V., & Evershed R. P. (2008). Interspecific variation in bulk tissue, fatty acid and monosaccharide $\delta^{13}\text{C}$ values of leaves from a mesotrophic grassland plant community. *Phytochemistry*, 69, 2041-2051.
- Dupouey J. L., Leavitt S., Choisnel E., & Jourdain S., (1993). Modelling carbon isotope fractionation in tree rings based on effective evapotranspiration and soil water status. *Plant, Cell & Environment*, 16(8), 939-947.
- Eggleston, S., Schmitt, J., Bereiter, B., Schneider, R., & Fischer, H. (2016). Evolution of the stable carbon isotope composition of atmospheric CO_2 over the last glacial cycle. *Paleoceanography*, 31(3), 434-452.
- Ehleringer J. R., & Cerling T. E. (1995). Atmospheric CO_2 and the ratio of intercellular to ambient CO_2 concentrations in plants. *Tree physiology*, 15(2), 105-111.
- Ehleringer J. R., Lin Z. F., Field C. B., Sun G. C., Kuo C. Y. (1987). Leaf carbon isotope ratios of plants from a subtropical monsoon forest. *Oecologia*, 72, 109-114.

- Ehleringer J. R., Phillips S. L., Schuster W. S., & Sandquist D. R. (1991). Differential utilization of summer rains by desert plants. *Oecologia*, 88(3), 430-434.
- Ehleringer J. R., Phillips S. L., & Comstock J. P. (1992). Seasonal variation in the carbon isotopic composition of desert plants. *Functional Ecology*, 6, 396-404.
- Ehleringer J. R., Hall A. E., Farquhar G. D. (Eds.). (1993). Stable isotopes and plant carbon-water relations (Vol. 109129). San Diego: Academic Press.
- Escudero A., Mediavilla S., Heilmeyer H. (2008). Leaf longevity and drought: avoidance of the costs and risks of early leaf abscission as inferred from the leaf carbon isotopic composition. *Functional Plant Biology*, 35, 705-713.
- Etheridge, D. M., Steele, L. P., Langenfelds, R. L., Francey, R. J., Barnola, J. M., & Morgan, V. I. (1998). Historical CO₂ records from the Law Dome DE08, DE08-2, and DSS ice cores. *Trends: a compendium of data on global change*, 351-364.
- Franco A. C., Duarte H. M., Geßler A., de Mattos E. A., Nahm M., Rennenberg H., Lüttge U. (2005). In situ measurements of carbon and nitrogen distribution and composition, photochemical efficiency and stable isotope ratios in *Araucaria angustifolia*. *Trees*, 19(4), 422-430.
- Friend A. D., Woodward F. I., & Switsur V. R. (1989). Field measurements of photosynthesis, stomatal conductance, leaf nitrogen and $\delta^{13}\text{C}$ along altitudinal gradients in Scotland. *Functional Ecology*, 117-122.
- Garten C. T., & Taylor G. E. (1992). Foliar $\delta^{13}\text{C}$ within a temperate deciduous forest: spatial, temporal, and species sources of variation. *Oecologia*, 90, 1-7.
- Gerdol R., Iacumin P., Marchesini R., & Bragazza L. (2000). Water- and nutrient-use efficiency of a deciduous species, *Vaccinium myrtillus*, and an evergreen species, *V. vitis-idaea*, in a subalpine dwarf shrub heath in the southern Alps, Italy. *Oikos* 88(1), 19-32.
- Gutierrez M. V., & Meinzer F. C. (1994). Carbon isotope discrimination and photosynthetic gas exchange in coffee hedgerows during canopy development. *Functional Plant Biology*, 21(2), 207-219.
- Guy R. D., & Reid D. M. (1986). Photosynthesis and the influence of CO₂-enrichment on $\delta^{13}\text{C}$ values in a C₃ halophyte. *Plant, Cell & Environment*, 9(1), 65-72.
- Hall A. E., Richards R. A., Condon A. G., Wright G. C., & Farquhar G. D. (1994). Carbon isotope discrimination and plant breeding. *Plant breeding reviews*, 12(81), 113.
- Hansen D., & Steig E. (1993). Comparison of water-use efficiency and internal leaf carbon dioxide concentration in juvenile leaves and phyllodes of *Acacia koa* (leguminosae) from Hawaii, estimated by two methods. *American Journal of Botany*, 80(10), 1121-1125.

- He C.-X., Li J.-Y., Zhou P., Guo M., & Zheng Q.-S. (2008). Changes of leaf morphological, anatomical structure and carbon isotope ratio with the height of the Wangtian Tree (*Parashorea chinensis*) in Xishuangbanna, China. *Journal of Integrative Plant Biology* 50(2), 168-173.
- Hemming D., Yakir D., Ambus P., Aurela M., Besson C., Black K., Gross, P. (2005). Pan-European $\delta^{13}\text{C}$ values of air and organic matter from forest ecosystems. *Global Change Biology*, 11(7), 1065-1093.
- Holtum J. A. M., & Winter K. (2005). Carbon isotope composition of canopy leaves in a tropical forest in Panama throughout a seasonal cycle. *Trees*, 19(5), 545-551.
- Huc R., Ferhi A., & Guehl J. M. (1994). Pioneer and late stage tropical rainforest tree species (French Guiana) growing under common conditions differ in leaf gas exchange regulation, carbon isotope discrimination and leaf water potential. *Oecologia*, 99(3-4), 297-305.
- Hultine K. R., & Marshall J. D. (2000). Altitude trends in conifer leaf morphology and stable carbon isotope composition. *Oecologia*, 123(1), 32-40.
- Inagaki Y., Miura S., & Kohzu A. (2004). Effects of forest type and stand age on litterfall quality and soil N dynamics in Shikoku district, southern Japan. *Forest Ecology and Management*, 202(1-3), 107-117.
- Ismail A. M., & Hall, A. E. (1993). Inheritance of carbon isotope discrimination and water-use efficiency in cowpea. *Crop Science*, 33(3), 498-503.
- Ismail A. M., Hall A. E., & Bray E. A. (1994). Drought and pot size effects on transpiration efficiency and carbon isotope discrimination of cowpea accessions and hybrids. *Functional Plant Biology*, 21(1), 23-35.
- Johnson R. C., & Tieszen L. L. (1993). Carbon isotope discrimination, water relations, and gas exchange in temperate grass species and accessions. In *Stable isotopes and plant carbon-water relations* (pp. 281-296). Academic Press.
- Keeling, C. D., Piper, S. C., Bacastow, R. B., Wahlen, M., Whorf, T. P., Heimann, M., & Meijer, H. A. (2005). Atmospheric CO_2 and $^{13}\text{CO}_2$ exchange with the terrestrial biosphere and oceans from 1978 to 2000: Observations and carbon cycle implications. In *A history of atmospheric CO_2 and its effects on plants, animals, and ecosystems* (pp. 83-113). Springer, New York, NY.
- Keeling, C. D., Piper, S. C., Bacastow, R. B., Wahlen, M., Whorf, T. P., Heimann, M., & Meijer, H. A. (2005). Terrestrial biosphere and oceans from 1978 to 2000: observations and carbon cycle implications. *A History of Atmospheric CO_2 and Its Effects on Plants, Animals and Ecosystems*, 2, 83-113.

- Kloeppel B. D., Gower S. T., Treichel I. W., & Kharuk S. (1998). Foliar carbon isotope discrimination in *Larix* species and sympatric evergreen conifers: a global comparison. *Oecologia*, 114(2), 153-159.
- Körner C., Farquhar G. D., Roksandic Z. (1988). A global survey of carbon isotope discrimination in plants from high altitude. *Oecologia*, 74(4), 623-632.
- Kohorn L. U., Goldstein G., & Rundel P. W. (1994). Morphological and isotopic indicators of growth environment - variability in $\delta^{13}\text{C}$ in *Simmondsia chinensis*, a dioecious desert shrub. *Journal of Experimental Botany*, 45(12), 1817-1822.
- Leffler A. J., & Enquist B. J. (2002). Carbon isotope composition of tree leaves from Guanacaste, Costa Rica: comparison across tropical forests and tree life history. *Journal of Tropical Ecology*, 18(1), 151-159.
- Li Z.-H., Leavitt S. W., Mora C. I., & Liu R.-M., (2005). Influence of earlywood-latewood size and isotope differences on long-term tree-ring $\delta^{13}\text{C}$ trends. *Chemical Geology*, 216(3-4), 191-201.
- Lockheart M. J., Van Bergen P. F., & Evershed R. P. (1997). Variations in the stable carbon isotope compositions of individual lipids from the leaves of modern angiosperms: implications for the study of higher land plant-derived sedimentary organic matter. *Organic Geochemistry*, 26(1-2), 137-153.
- Marshall J. D., & Zhang J. (1994). Carbon isotope discrimination and water-use efficiency in native plants of the north-central Rockies. *Ecology*, 75(7), 1887-1895.
- Martin B., & Thorstenson Y. R. (1988). Stable carbon isotope composition ($\delta^{13}\text{C}$), water use efficiency, and biomass productivity of *Lycopersicon esculentum*, *Lycopersicon pennellii*, and the F1 hybrid. *Plant Physiology*, 88(1), 213-217.
- Martin B., Bytnerowicz A., & Thorstenson Y. R. (1988). Effects of air pollutants on the composition of stable carbon isotopes, $\delta^{13}\text{C}$, of leaves and wood, and on leaf injury. *Plant Physiology*, 88(1), 218-223.
- Mayland H. F., Johnson D. A., Asay K. H., & Read J. J. (1993). Ash, carbon isotope discrimination, and silicon as estimators of transpiration efficiency in crested wheatgrass. *Functional Plant Biology*, 20(3), 361-369.
- McArthur J. V., & Moorhead K. K. (1996). Characterization of riparian species and stream detritus using multiple stable isotopes. *Oecologia* 107(2), 32-238.
- Merven C. (2015). Isotope ecology of temperate conifers (Masters' Thesis, University of Michigan, Ann Arbor, MI, United States of America).

- Mole S., Joern A. (1994). Feeding behavior of graminivorous grasshoppers in response to host-plant extracts, alkaloids, and tannins. *Journal of chemical ecology*, 20(12), 3097-3109.
- Mooney H. A., Bullock S. H., & Ehleringer J. R. (1989). Carbon isotope ratios of plants of a tropical dry forest in Mexico. *Functional Ecology*, 3, 137-142.
- Morgan J. A., LeCain D. R., McCaig T. N., & Quick J. S. (1993). Gas exchange, carbon isotope discrimination, and productivity in winter wheat. *Crop Science*, 33(1), 178-186.
- PRISM Climate Group, Oregon State University. (2004). <http://prism.oregonstate.edu>, created 4 Feb 2004.
- Rubino, M., Etheridge, D. M., Trudinger, C. M., Allison, C. E., Battle, M. O., Langenfelds, R. L., Steele, L. P., Curran, M., Bender, M., White, J. W. C. & Jenk, T. M. (2013). A revised 1000 year atmospheric $\delta^{13}\text{C}$ -CO₂ record from Law Dome and South Pole, Antarctica. *Journal of Geophysical Research: Atmospheres*, 118(15), 8482-8499.
- Rao R. N., Williams J. H., Wadia K. D. R., Hubick K. T., & Farquhar G. D. (1993). Crop growth, water-use efficiency and carbon isotope discrimination in groundnut (*Arachis hypogaea* L.) genotypes under end-of season drought conditions. *Annals of applied Biology*, 122(2), 357-367.
- Nagy L., & Proctor J. (2000). Leaf $\delta^{13}\text{C}$ signatures in heath and lowland evergreen rain forest species from Borneo. *Journal of Tropical Ecology*, 16, 757-761.
- Osorio J., & Pereira J. S. (1994). Genotypic differences in water use efficiency and ^{13}C discrimination in *Eucalyptus globulus*. *Tree Physiology*, 14(7-8-9), 871-882.
- Panek J. A. (1996). Correlations between stable carbon-isotope abundance and hydraulic conductivity in Douglas-fir across a climate gradient in Oregon, USA. *Tree Physiology*, 16(9), 747-755.
- Peñuelas J., & Azcón-Bieto J. (1992). Changes in leaf $\Delta^{13}\text{C}$ of herbarium plant species during the last 3 centuries of CO₂ increase. *Plant, Cell & Environment*, 15(4), 485-489.
- Polley H. W., Johnson H. B., & Mayeux H. S. (1995). Nitrogen and water requirements of C₃ plants grown at glacial to present carbon dioxide concentrations. *Functional Ecology*, 86-96.
- Royles J., Horwath A. B., & Griffiths H. (2014). Interpreting bryophyte stable carbon isotope composition: Plants as temporal and spatial climate recorders. *Geochemistry, Geophysics, Geosystems*, 15(4), 1462-1475.
- Rundel P. W., Stichler W., Zander R. H., & Ziegler H. (1979). Carbon and hydrogen isotope ratios of bryophytes from arid and humid regions. *Oecologia*, 44(1), 91-94.

- Sandquist D. R., & Cordell S. (2007). Functional diversity of carbon-gain, water-use, and leaf-allocation traits in trees of a threatened lowland dry forest in Hawaii. *American Journal of Botany*, 94(9), 1459-1469.
- Schubert B. A., & Jahren A. H. (2012). The effect of atmospheric CO₂ concentration on carbon isotope fractionation in C₃ land plants. *Geochimica et Cosmochimica Acta*, 96, 29-43.
- Sheldon N. D., Smith SY, Stein R. A., & Ng M. (2020). Carbon isotope ecology of gymnosperms and implications for paleoclimatic and paleoecological studies. *Global and Planetary Change*, 184, 103060, 1-17.
- Simpson D. A., Muasya A. M., Chayamarit K., Parnell J. A., Suddee S., Wilde B. D., & Pooma R. (2005). *Khaosokia caricoides*, a new genus and species of Cyperaceae from Thailand. *Botanical Journal of the Linnean Society*, 149(3), 357-364.
- Smedley M. P., Dawson T. E., Comstock J. P., Donovan L. A., Sherrill E., Cook C. S., & Ehleringer J. R. (1991). Seasonal carbon isotope discrimination in a grassland community. *Oecologia*, 85(3), 314-320.
- Sternberg L. O., Deniro M. J., & Johnson H. B. (1984). Isotope ratios of cellulose from plants having different photosynthetic pathways. *Plant Physiology*, 74(3), 557-561.
- Terwilliger V. J. (1997). Changes in the $\delta^{13}\text{C}$ values of trees during a tropical rainy season: some effects in addition to diffusion and carboxylation by Rubisco? *American Journal of Botany*, 84(12), 1693-1700.
- Toft N. L., Anderson J. E., & Nowak R. S. (1989). Water use efficiency and carbon isotope composition of plants in a cold desert environment. *Oecologia*, 80(1), 11-18.
- Tu T. T. N., Kürschner W. M., Schouten S., & Van Bergen P. F. (2004). Leaf carbon isotope composition of fossil and extant oaks grown under differing atmospheric CO₂ levels. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 212(3-4), 199-213.
- Uemura A., Harayama H., Koike N., & Ishida A. (2006). Coordination of crown structure, leaf plasticity and carbon gain within the crowns of three winter-deciduous mature trees. *Tree Physiology*, 26(5), 633-641.
- Ueno O., Samejima M., Muto S., & Miyachi S. (1988). Photosynthetic characteristics of an amphibious plant, *Eleocharis vivipara*: expression of C₄ and C₃ modes in contrasting environments. *Proceedings of the National Academy of Sciences*, 85(18), 6733-6737.
- Valentini R., Anfodillo T., & Ehleringer J. R. (1994). Water sources and carbon-isotope composition ($\delta^{13}\text{C}$) of selected tree species of the Italian Alps. *Canadian Journal of Forest Research*, 24(8), 1575-1579.

- Valentini R., Mugnozza G. E. S., & Ehleringer J. R. (1992). Hydrogen and carbon isotope ratios of selected species of a Mediterranean Macchia ecosystem. *Functional Ecology*, 627-631.
- Van de Water P., Leavitt S., & Betancourt J. (2002). Leaf $\delta^{13}\text{C}$ variability with elevation, slope aspect, and precipitation in the southwest United States. *Oecologia*, 132(3), 332-343.
- Van de Water P. K., Leavitt S. W., & Betancourt J. L. (1994). Trends in stomatal density and $^{13}\text{C}/^{12}\text{C}$ ratios of *Pinus flexilis* needles during last glacial-interglacial cycle. *Science*, 264(5156), 239-243.
- Welker J. M., Wookey P. A., Parsons A. N., Press M. C., Callaghan T. V., & Lee J. A. (1993). Leaf carbon isotope discrimination and vegetative responses of *Dryas octopetala* to temperature and water manipulations in a High Arctic polar semi-desert, Svalbard. *Oecologia*, 95(4), 463-469.
- White J. W., Castillo J. A., Ehleringer J. R., Garcia J. A. C., & Singh S. P. (1994). Relations of carbon isotope discrimination and other physiological traits to yield in common bean (*Phaseolus vulgaris*) under rainfed conditions. *The Journal of Agricultural Science*, 122(2), 275-284.
- White, J.W.C., Vaughn, B.H., Michel, S.E. (2015). University of Colorado, Institute of Arctic and 720 Alpine Research (INSTAAR), Stable Isotopic Composition of Atmospheric Carbon Dioxide (^{13}C and ^{18}O) from the NOAA ESRL Carbon Cycle Cooperative Global Air Sampling Network, 722 1990-2014, Version: 2015-10-26.
- Williams D. G., & Ehleringer J. R. (1996). Carbon isotope discrimination in three semi-arid woodland species along a monsoon gradient. *Oecologia*, 106(4), 455-460.
- Williams D. G., & Ehleringer J. R. (2000). Carbon isotope discrimination and water relations of oak hybrid populations in southwestern Utah. *Western North American Naturalist*, 121-129.
- Winter K., Troughton J. H. (1978). Carbon assimilation pathways in *Mesembryanthemum nodiflorum* L. under natural conditions. *Zeitschrift für Pflanzenphysiologie*, 88(2), 153-162.
- Wright G. C., Rao R. C., & Farquhar G. D. (1994). Water-use efficiency and carbon isotope discrimination in peanut under water deficit conditions. *Crop Science*, 34(1), 92-97 (1994).
- Zhang J., Marshall J. D., & Jaquish B. D. (1993). Genetic differentiation in carbon isotope discrimination and gas exchange in *Pseudotsuga menziesii*. *Oecologia*, 93(1), 80-87.
- Zhang J., Fins L., & Marshall J. D. (1994). Stable carbon isotope discrimination, photosynthetic gas exchange, and growth differences among western larch families. *Tree Physiology*, 14(5), 531-539.

Zibulski, R., Wesener F., Wilkes H., Plessen B., Pestryakova L. A., & Herzschuh, U. (2019). C/N ratio, stable isotope ($\delta^{13}\text{C}$, $\delta^{15}\text{N}$), and n-alkane patterns of brown mosses along hydrological gradients of low-centred polygons of the Siberian Arctic. *Biogeosciences*, *14*(6), 1617-1630.