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| 9 | Potamotrygoninae) reveals effects of continent-scale paleogeographic change and drainage |
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29

30 ABSTRACT

Aim: Paleogeographic changes have had profound effects on the evolution and diversity of the
 Neotropical biota. However, the influence of marine incursions on the origin, diversification and
 distribution of fishes is still incompletely understood. We investigate the biogeographical and
 chronological patterns of diversification for the marine-derived Neotropical freshwater stingrays
 (subfamily Potamotrygoninae) at a continental scale.

6 *Location:* Neotropics, South America.

7 Taxa: Neotropical freshwater stingrays. Subfamily Potamotrygoninae (Myliobatiformes:

8 Chondrichthyes).

9 Methods: We generated a time-calibrated phylogeny for 35 of the 38 valid species of Neotropical

10 freshwater stingrays, from most of the major river basins of South America, using four genes.

11 We used BEAST2 to determine the chronology of population and species divergence events, and

12 'BioGeoBEARS' to infer historical biogeographic patterns.

13 *Results:* The Potamotrygoninae originated during the early/middle Miocene in the upper Amazon

14 region. We recover clades associated with particular geographic areas and detect a recurrent

15 pattern of upper Amazon clades sister to clades in adjacent basins. The timing of dispersals from

16 the upper Amazon to adjacent areas corresponds with the end of the Pebas wetlands. Lower

17 Amazon and Shield associated taxa are relatively young.

Main conclusions: We propose that the origin of the Neotropical freshwater stingrays is related to marine incursions that occurred during the Oligocene/Miocene. Subsequent diversification of Potamotrygoninae occurred in the Pebas wetland system in the upper Amazon with colonization of adjacent basins. These movements were generally unidirectional, with few lineages returning to the upper Amazon, and we speculate that ecological factors drove this pattern. We observed a burst of potamotrygonine diversification 5 Ma that appears to be related to the modern channelization of the Amazon Basin.

25 Keywords: Amazon, Guiana Shield, Marine incursions, Orinoco, Paleogeography, Parana-

26 Paraguay, Pebas Wetlands, South America.

27

1 1. INTRODUCTION

2 The Neotropics are home to the greatest diversity of freshwater fishes in the world (Reis et 3 al. 2016). While dominated by fishes that have an ancient association with freshwater, such as the catfishes, characiforms, and weakly electric fishes, the Neotropical ichthyofauna also 4 includes an intriguing component derived from marine-derived lineages. These are lineages that 5 are endemic to continental freshwater habitats but are most closely related to clades that are 6 7 predominantly distributed in marine environments, including taxa such as anchovies, flatfishes, pufferfishes, drum, needlefishes, and stingrays (Lovejoy et al. 2006; Bloom and Lovejoy, 2017). 8 This biogeographic pattern is mirrored by South American non-fish groups, such as dolphins, 9 10 manatees, sponges, crustaceans, and mollusks (Nuttall, 1990, Cassens et al. 2000; Hamilton et al. 11 2001; Vermeij and Wesselingh, 2002; Cavalcanti et al., 2019). Many of the South American 12 marine-derived lineages have colonized and diversified in freshwater environments, and are widely distributed across the continent, suggesting considerable age. 13

14 The timing and mechanisms involved in the origin of marine-derived lineages into South America are topics of ongoing investigation. Earlier explanations for the origin of marine 15 lineages evoked the invasion of the South American fluvial system via estuaries and river mouths 16 (Roberts, 1972). Our present understanding is that freshwater invasions might have been 17 18 associated with paleogeographical events such as changes in sea level and marine incursions 19 (Lovejoy et al. 1998; Wesselingh and Hoorn, 2011; Bloom and Lovejoy, 2011, 2017). Replication of phylogenetic patterns and congruence of age estimates for cladogenetic events 20 21 leading to the origin of marine-derived lineages provide important clues regarding these 22 hypotheses. Invasions via river mouths could occur at any time, involve any large river system, 23 and, as dispersal events depend on intrinsic properties of lineages – we should not expect replication of phylogenetic patterns and age across lineages. South America has experienced 24 intermittent marine incursion events dating as far as the upper Cretaceous, with peaks of marine 25 influence during the Paleocene, Eocene, Miocene and Pleistocene (Shephard et al. 2010; 26 Wesselingh and Hoorn, 2011; Louterbach et al., 2014). Biogeographic studies propose incursions 27 during the Eocene (the Pozo incursions) and most importantly incursions related to the Pebas 28 29 Mega-Wetland System of the Miocene as events that facilitated invasions of marine-lineages into

Neotropical freshwaters (Lovejoy et al. 2010; Wesselingh and Hoorn, 2011; Bloom and Lovejoy,
 2011, 2017).

The Pebas Mega-Wetland System (hereafter Pebas wetlands or system) was an 3 epicontinental marine/freshwater system covering more than one million km², spanning from the 4 Caribbean to southern South America (Fig. 1) (Hoorn, Wesselingh, Ter Steege, et al. 2010; 5 Hoorn, Wesselingh, Hovikoski, et al. 2010; Shephard et al. 2010; Wesselingh and Hoorn, 2011; 6 7 Bernal et al. 2019). This system covered the western Amazon region during most of the Miocene (23-10 Ma) (Hoorn, Wesselingh, Hovikoski, et al. 2010; Bernal et al. 2019), but its southernmost 8 boundary is uncertain. Some authors have suggested it extended as far as southern Argentina, 9 connecting to the Paranaense Sea (Aleman and Ramos, 2000; Brea and Zucol, 2011), while 10 11 others argue that it extended only as far as present-day Bolivia (Wesselingh and Hoorn, 2011; 12 Bernal et al. 2019). The giant wetlands of the Pebas experienced oscillating levels of marine 13 influence, as evidenced by sediments characteristic of both freshwater and tidal conditions (Hovikoski et al. 2010) and a rich fossil record of freshwater to marine-associated lineages, 14 ranging from invertebrates (Wesselingh et al. 2006; Wesselingh and Ramos, 2010) and plants 15 16 (Hoorn, 2006; Bernal et al. 2019), to crocodylians (Salas-Gismondi et al. 2015). Complex 17 salinity gradients would have offered an ideal setting for the evolution of marine to freshwater 18 transitions, and the connection of the Pebas system to the Caribbean region would have offered 19 proto marine-derived lineages a portal to the upper Amazon (Nuttal, 1990; Lovejoy et al. 2006; 20 Bloom and Lovejoy, 2017; Bernal et al. 2019).

21 Aside from the effects of the Pebas system on the origin of marine-derived lineages in South America, this vast wetland also likely affected biogeographic patterns of any aquatic lineages 22 23 that occurred there. The Pebas System was fed by rivers flowing from the Andes and older shield drainages of the western Amazon, with periodic connections to the Caribbean Sea, until the 24 establishment of the transcontinental Amazon River in the late Miocene (Figueiredo et al. 2009; 25 Shephard et al. 2010; Wesselingh and Hoorn, 2011; Hoorn et al. 2017). The Pebas was thus a 26 27 likely dispersal corridor for aquatic organisms throughout the proto-Amazon region, allowing 28 movements between continental drainages that were previously and subsequently isolated 29 (Bernal et al. 2019). However, while studies have hypothesized the role of the Pebas system in

isolating faunas (e.g., Hubert and Renno, 2006; Solomon et al. 2008); its influence on the
 distribution and potential dispersal of freshwater fish species has been less explored.

3 The Neotropical freshwater stingrays (subfamily Potamotrygoninae) represent the only extant, obligate freshwater clade of elasmobranchs (Carvalho et al. 2016). Composed of 38 4 species in four genera (Silva and Loboda, 2019, Loboda et al. In press) potamotrygonines are 5 6 found in almost every major river basin in South America (Carvalho et al. 2003), with maximum 7 species diversity in the Amazon (28 species). The Potamotrygoninae exhibit considerable 8 ecological diversity, ranging from piscivores to mollusk and insect feeders (Kolmann et al. 2016, Fontenelle et al. 2017; Rutledge et al., 2019), and ranging in size from the >1m Potamotrygon 9 brachvura to the <25cm Wallace's stingray, or arraia-cururu (Potamotrygon wallacei) (Castello 10 et al. 2016; Rosa et al. 2016) (Fig. 2). The beautiful dorsal color patterns of many species make 11 12 them desirable aquarium specimens in the international ornamental fish trade (Fontenelle and 13 Carvalho, 2016). The sister taxon of the potamotrygonines is composed of two marine species of Styracura, from the eastern Pacific and western Atlantic (Lovejoy, 1996; Lovejoy et al., 1998; 14 Carvalho et al. 2016, Fontenelle et al. 2017). The Potamotrygoninae are hypothesized to have 15 16 originated because of marine incursions into the upper Amazon and Pebas system associated 17 regions during the Miocene (Carvalho et al. 2004, Lovejoy et al. 1998, 2006), and subsequently 18 spread to other river systems in South America. However, other authors argue for an older origin, 19 based on fossil evidence from the Eocene that is attributed to potamotrygonines (Frailey, 1986; 20 Adnet et al. 2014), molecular estimates that show a mid-late Eocene origin for potamotrygonines 21 (Bloom and Lovejoy, 2017), and a morphology-based phylogeny that includes fossil stingrays 22 and places the age of potamotrygonines in the Eocene (Carvalho et al. 2004).

23 The broad geographic distribution and diversity of the Potamotrygoninae, combined with 24 their potential to reveal insights regarding the origins of marine-derived lineages and patterns of 25 faunal connectivity throughout South America, make them an excellent candidate for 26 biogeographic investigation. We used a comprehensive dated molecular phylogeny for the 27 subfamily Potamotrygoninae, including 35 of the 38 species spanning their entire known 28 distribution, to investigate the historical biogeography of the group. We investigated the 29 hypothesis that Neotropical freshwater stingrays originated and diversified in conjunction with 30 marine incursions into the Pebas wetland system in the upper Amazon, and then dispersed to

adjacent basins. We tested whether potamotrygonines originated during the Miocene and
 whether the ancestral distribution of the group overlapped with the area of the Pebas wetlands
 (upper Amazon). Additionally, we investigated patterns of biogeographic connectivity in relation
 to the distribution of potamotrygonines in other river systems, including the rivers of the Guiana
 and Brazilian shields, the lower Amazon, and the Parana-Paraguay basins.

6

7 2. MATERIAL AND METHODS

8 2.1 Taxon sampling, DNA extraction, polymerase chain reaction (PCR) amplification, and 9 sequencing

10 We analyzed a total of 350 specimens representing 35 out of 38 currently recognized species of the Potamotrygoninae with extensive geographic coverage of the subfamily distribution. We 11 12 also included the two marine species of the genus Styracura (Styracurinae), the marine sister 13 group of Potamotrygoninae, thus sampling 37 out of 40 species in Potamotrygonidae. As 14 outgroups, we included Urobatis halleri from the eastern Pacific, U. jamaicensis, from the 15 western Atlantic, and Taeniura lymma, from the Indo-Pacific, based on previous phylogenetic hypotheses (Lovejoy et al. 1998; Margues, 2001; Aschliman et al. 2012). A complete list of 16 17 species and localities is provided in Table S1 in Supporting Information.

We extracted genomic DNA and PCR amplified fragments of three mitochondrially encoded
(mtDNA) genes: ATP synthase membrane subunit 6 and 8 (*atpase*), cytochrome b (*cytb*) and
cytochrome c oxidase I (*col*); and one nuclear ribosomal internal transcribed spacer 1 (*its-1*).
Details regarding molecular protocols are provided in Table S2.

22

23 2.2 Phylogenetic analysis and dating inference

We used BEAST2 (v2.4.7, Bouckeart et al. 2014) to estimate phylogeny and divergence times under an uncorrelated log-normal relaxed clock (Drummond et al. 2006), allowing rates to vary among branches. Genes were unlinked and set to a GTR model with a gamma distributed rate of heterogeneity for each gene individually, as determined by jModelTest 2 (Darriba et al. 1 2012). We used a Yule Model prior for rates of cladogenesis and ran four independent analyses

2 for 100 million generations, sampling every 10000 generations. We used Tracer (v1.6,

3 Drummond and Rambaut, 2007) to evaluate convergence and to verify if effective sample size

4 (ESS) was at least 200 for all parameters. We discarded 25% of the initial sample as burn-in. The

5 runs were combined using LogCombiner (v2.4.7, Bouckeart et al. 2014) and the maximum

6 credibility tree was generated in TreeAnnotator (v2.4.7, Bouckeart et al. 2014).

7 We used four different dating priors in the BEAST2 analysis. We used the earliest-known proposed fossil for the potamotrygonine subfamily, Potamotrygon ucayalensis (Adnet et al. 8 2014) from the middle Eocene (~41 Ma), and the fossils of P. contamanensis, P. canaanorum 9 and P. rajachloeae (Chabain et al. 2017) from the late Oligocene - early Miocene (~30-20 Ma). 10 11 We assigned a log-normal distributed prior for the origin of the subfamily Potamotrygoninae using these fossils, and to accommodate date uncertainty, set this prior to have an offset of 20.0, 12 13 standard deviation (S) of 0.75 and mean (M) of 12.0. We also used the closure of the 14 Panamanian Isthmus as a splitting point between Urobatis halleri and U. jamaicensis and 15 between Styracura schmardae and S. pacifica. We used the start (~15 Ma) and end (~3 Ma) of 16 the isolation process between the Pacific and the Atlantic (Lundberg et al. 1998; Leigh and 17 Vermeij, 2014), as a log-normal prior, with offset of 2.5, S of 0.7 and M of 2.0. We used the uplift of the eastern Cordillera in Colombia to date the split between Potamotrygon magdalenae 18 19 (from the Magdalena River basin) and P. yepezi (from the Maracaibo Lake basin). We set this log-normal prior based on the start (~12 Ma) and end (~5-3 Ma) of the uplift of this mountain 20 21 range (Hoorn et al. 1995; Gregory-Wodzicki, 2000), with an offset of 2.5, S of 0.75 and M of 22 3.0. Finally, we used the uplift of the Merida Andes as a splitting point between P. magdalenae + 23 P. yepezi and the rest of the subfamily. The Merida Andes uplift interrupted the connectivity 24 between the rivers draining from Colombia and Venezuela to the rest of the Amazon Basin around 10-8 Ma (Lundberg et al. 1998; Hoorn, Wesselingh, Hovikoski, et al. 2010). We set this 25 prior as a log-normal distribution, with offset of 7.0, S of 0.5 and M of 3.5. We used log-normal 26 27 distributions for the geological priors because, even without hard bounds, we wanted to shape a higher distribution of probability around the "end point" of each isolation event but not ignore 28 29 the possibility of earlier divergence (Ho and Phillips, 2009).

30

1 2.3 Biogeography and diversification

2 To infer biogeographical history, a probabilistic model of geographical range evolution was 3 performed using the R package 'BioGeoBEARS' (Matzke, 2013). This package uses both Maximum Likelihood and Bayesian approaches to estimate ancestral range probabilities for the 4 nodes of a dated phylogeny based on distributions of extant species. We used 13 freshwater 5 biogeographical areas, as well as a "Marine" area for the outgroups (Fig. 3), which were 6 7 determined based on areas of endemism proposed by Lundberg et al. (1998), Albert et al. (2011) and Dagosta and de Pinna (2017). We pruned our time-calibrated Bayesian phylogeny by 8 selecting a single individual to represent monophyletic groups within each area, resulting in a 9 tree with 150 terminals that was used for the 'BioGeoBEARS' analyses. 10

11 We performed biogeographical inference using six different models: DEC, DEC+J; 12 DIVAlike; DIVAlike+J; BAYAREAlike; and BAYAREAlike+J. The likelihood-based 13 Dispersal-Extinction Cladogenesis (DEC – Ree and Smith, 2008; Matzke, 2013) model assumes 14 that derived lineages after cladogenesis will inherit a single-range area, which can be a subset of 15 the ancestor's range; DIVAlike, a likelihood version of the DIVA model (Ronquist and Sanmartin, 2011), allows for derived lineages to inherit more than one area as their range, but 16 does not allow for the area of these lineages to be a subset of the ancestor's range; 17 18 BAYAREAlike is a likelihood version of the BAYAREA (Landis et al. 2013) and assumed that 19 no range evolution occurs at cladogenesis, and the derived lineages inherit the same range of the ancestral state. The parameter "J" adds founder-event speciation to each of the presented models 20 21 (see Table 1 and Matzke, 2013, 2014 for additional details). We set the parameter 22 max range size to five, and we included the null range parameter which allows ranges to consist 23 of zero areas, a state of transition in the matrix of DEC models. We used 'BioGeoBEARS' to calculate the log likelihood (lnL) and the corrected Akaike Information Criterion (AICc) to 24 choose the best fitting biogeographical model. 25

26

27 **3. RESULTS**

28 3.1 Origin and biogeography of the Neotropical freshwater stingrays

1 The phylogenetic analysis (Figs. 3, S1, S2) recovered Neotropical freshwater stingrays 2 (Potamotrygoninae) as a monophyletic group that is most closely related to the marine subfamily 3 Styracurinae (composed of two species that occur in coastal waters on either side of the Isthmus 4 of Panama) (Fig. 3). The date of divergence between the freshwater Potamotrygoninae and the marine Styracurinae was estimated at 26.4 Ma (ranging 32.1-20.6 Ma), dating the origin of the 5 6 Neotropical freshwater stingrays to the late Oligocene to early Miocene, coincident with the start 7 of the Pebas wetlands period (Figs. 3, S2). The best fitting model for was determined to be 8 DEC+J (Table 1), and we recover the most probable ancestral range for the family 9 Potamotrygonidae as marine (Fig. S3), while the most probable ancestral range for the 10 Potamotrygoninae is the upper Amazon (Figs. 3, S3).

11

12 **3.2 Diversification and biogeographic patterns within the Neotropical freshwater stingrays**

13 Within Potamotrygoninae, the first divergence is between the clades

14 Heliotrygon+Paratrygon (5 species) and Plesiotrygon+Potamotrygon (33 species), with a date

15 of 22.45 (20.6/25.1) Ma (early Miocene) (Fig. 3). Both clades are reconstructed to have most

16 likely ancestrally occupied the upper Amazon (Fig. 3), and both contain lineages that occur in

17 most of the major South American river drainages.

In the *Heliotrygon+Paratrygon* clade, *Heliotrygon* occurs only in the upper Amazon region (Peru). Its sister lineage *Paratrygon* is much more widespread, occurring across multiple basins of the Orinoco and Amazon, and exhibits distinct, well-defined clades corresponding to major river basins (Figs. 3, S3). In *Plesiotrygon+Potamotrygon*, the genus *Plesiotrygon* is restricted to the Amazon Basin, occurring from the upper Amazon (Peru) to the lower portions of the main channel of the Amazon River. *Potamotrygon* is the most diverse and widely distributed genus, with a range that encompasses the whole distribution of the subfamily (Figs. 3, S3).

According to our biogeographic analysis, the two major lineages of Potamotrygoninae originated in the upper Amazon region and then expanded their distributions to adjacent drainages during the Miocene Pebas wetlands period. Our analysis suggests that freshwater stingray lineages first occupied the Negro/Branco, the trans-Andean region, the Parana-Paraguay, and the Brazilian Shield (Tocantins) prior to 10 Ma (Figs. 3, S3).

1 The late Miocene, after the end of the Pebas period (~10 Ma), roughly corresponds to the 2 time of origin of the modern trans-continental route of the Amazon River. The biogeographical 3 reconstruction provides evidence of the first colonization of the Orinoco Basin and several 4 Brazilian Shield drainages during this period (Figs. 3, S3). We also find evidence that 5 colonization of areas that had already been occupied by freshwater stingray lineages took place 6 at this time.

7 These reconstructions show that during and after the Pliocene, potamotrygonine lineages 8 colonized the lower portion of the main channel of the Amazon River (lower Amazon), drainages associated with the Essequibo and Eastern Guyana, and the Xingu, Tapajós and 9 Parnaíba basins for the first time, and there was ongoing exchange of lineages to and from 10 11 drainages that had been colonized earlier. Our results show evidence that the lineages in the 12 Essequibo and Eastern Guyana originated from the upper Amazon and Orinoco (Figs. 3, S3d). 13 Lineages in the lower portion of the main channel of the Amazon River and Brazilian Shield 14 drainages originated from the upper Amazon and Negro/Branco ranges. These recent lineages 15 form groups based on locality and/or biogeographical areas, comprising specimens belonging to 16 different species but that are from the same area (Fig. S3). We find evidence for Pliocene 17 colonization of the Parana-Paraguay lineages by lineages associated with upper Amazon (Peru) 18 drainages and Brazilian Shield drainages.

19

20 4. DISCUSSION

21 4.1 Origins of the Potamotrygoninae coincides with the Pebas wetlands.

22 The origins and diversification of marine-derived lineages in South American freshwaters 23 have been attributed to both invasions of river mouths (Roberts, 1972) and marine incursion 24 events that occurred intermittently since the Cretaceous (Frailey, 1986; Carvalho et al. 2004; 25 Lovejoy et al. 2006; Bloom and Lovejoy, 2011). Several authors have linked marine incursions 26 during the Miocene to origins of marine derived taxa in South American freshwaters, including 27 fishes, mollusks, crustaceans, and mammals (Nuttall, 1990; Cassens et al. 2000; Hamilton et al. 28 2001; Vermeij and Wesselingh, 2001; Bloom and Lovejoy, 2017; Cavalcanti et al., 2019). Bloom 29 and Lovejoy (2017) estimated the ages of 10 Neotropical marine-derived fish lineages and found

1 that while most transitioned to freshwater during the Oligocene, a few origins were estimated as 2 Miocene. Our analysis dates the age of the split of the Neotropical freshwater stingrays from 3 their marine sister group to 26 Ma, with error estimates between 32 and 21 Ma, roughly the 4 boundary between the Oligocene and Miocene. At that time, the proto-Amazon was draining into the Caribbean region, before the closure of the Panama Isthmus, and intense marine intrusions 5 6 related to the Pebas system, which dominated the western Amazon region, allowed for biotic 7 interchange of faunas between marine and freshwater environments (Hoorn, Wesselingh, Hovikoski, et al. 2010; Leigh et al. 2014). The results place the origin of Potamotrygoninae to a 8 time when the western Amazon was dominated by the Pebas wetlands, corroborating our initial 9 10 hypothesis that this lineage originated in conjunction with marine incursions into the Pebas 11 wetlands.

Our results also indicate that initial diversification of potamotrygonines took place in the
Pebas wetlands. The origin of two major lineages of Potamotrygoninae

14 (*Heliotrygon+Paratrygon* and *Plesiotrygon+Potamotrygon*), as well as the origin of the

15 individual potamotrygonine genera, date to the Miocene. The estimated ancestral ranges for

16 Potamotrygoninae, *Heliotrygon+Paratrygon* and *Plesiotrygon+Potamotrygon* are all recovered

17 as upper Amazon (Figs. 3, S3). Indeed, until approximately 10 Ma, the proposed end of the

18 dominance of the Pebas wetlands, all ancestral ranges are estimated to be either upper Amazon

19 or Negro/Branco (Fig. S3), matching the range of paleo-floodplains in the Sub-Andean foreland

20 (Figueiredo et al. 2009; Wesselingh and Hoorn, 2011; Lima and Ribeiro, 2011). Thus, our results

21 suggest that initial diversification and distribution of potamotrygonines is associated with Pebas

22 wetland habitats. We suggest that the initial burst of potamotrygonine diversification may have

23 been associated with the dynamics of the Pebas. Salinity fluctuations and their deleterious effects

on incumbent Neotropical freshwater fish lineages, referred to as a competition trough (Vermeij

and Wesselingh, 2002; Lovejoy et al., 2006; Bloom and Lovejoy, 2017), may have opened new

26 niche space for salinity-tolerant lineages in these communities, facilitating their diversification.

- 27 The special ecological circumstances of the Pebas are thought to have also spurred
- 28 diversification in crocodylians (Salas-Gismondi et al. 2015) and mollusks (Nuttall, 1990).

29 Supporting our arguments, Chabain et al. (2017) show that proposed potamotrygonid fossil teeth

30 recovered from the Oligocene- late Miocene from the Contamana region in Peru show

31 morphological disparities that relate to ecological adaptations and diversification.

1 The estimate for the age of origin of Potamotrygoninae (26 Ma) is older than the 23 Ma 2 proposed by Lovejoy et al. (1998), but younger than the 38.2 (30.1 - 47.7) Ma proposed by 3 Bloom and Lovejoy (2017). However, all three of these estimates roughly coincide with the age 4 of Miocene marine incursions and the Pebas wetlands, and the error estimate for the age proposed here (21 to 32 Ma) overlaps with these earlier estimates. Our age estimates for the 5 Potamotrygoninae origin contrasts with older ages proposed by some authors based on fossil 6 7 evidence. Based on the phylogenetic position of non-potamotrygonine fossil stingrays from the Green River and Monte Bolca formations, Carvalho et al. (2004) suggest that the origin of the 8 9 potamotrygonines was older than the Oligocene-Miocene. Fossil evidence for Neotropical 10 freshwater stingrays is scarce, mostly consisting of disarticulated teeth (Adnet et al. 2014; 11 Chabain et al. 2017). Adnet et al. (2014) studied stingray teeth from freshwater Eocene deposits 12 from Contamana in the Peruvian Amazon (CTA-27) that they attributed to potamotrygonines; however, these teeth do not exhibit phylogenetically diagnostic characters for potamotrygonines 13 14 and their classification depends on their freshwater provenance. Using ecological setting to 15 determine the phylogenetic position of fossils is weakened by the fact that different lineages of 16 stingrays have invaded freshwater habitats on multiple occasions. More recently, Chabain et al. (2017) describe additional fossil stingray teeth from Oligocene-Miocene deposits also from 17 18 Contamana (CTA-43) that they ascribe to potamotrygonines, and the age of these fossils matches 19 the molecular results presented here. Ideally, fossils with potamotrygonine synapomorphies will 20 eventually be found, enabling an independent test of the age estimates proposed here.

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22 4.2 Dispersal of stingrays from the upper Amazon

The dynamics of the Pebas wetlands during the Miocene caused physical reorganizations of river basins and likely created connections between drainages that made fish faunal exchanges possible (Albert et al. 2018; Dagosta and Pinna, 2017, 2019). During this period, the extension and contraction of headwater boundaries played an important role in connecting river basins (Hoorn, Wesselingh, Hovikoski, et al. 2010; Wesselingh and Hoorn, 2011; Carvalho and Albert, 2011). The Pebas system occupied most of the current upper Amazon region, extending to the Caribbean to the north, the Orinoco Basin to the northeast, the middle Amazon (Purus arch region) to the east, the Andes to the west, and possibly as far as Bolivia to the south (Albert et al.
 2018, figure 4; Bernal et al. 2019, figures 2, 3).

We propose that the extensive area and connectivity of the Pebas wetlands made it possible 3 for freshwater fishes to disperse from the western Amazon to adjacent basins. Our results show 4 evidence of these dispersals in stingrays: 'BioGeoBears' estimates the ancestral ranges for early 5 nodes (Early/Middle Miocene) of the subfamily Potamotrygoninae as upper Amazon, with 6 7 dispersals to adjacent drainages in the Negro/Branco, the Parana-Paraguay, and the trans-Andean 8 basins (Figs. 3, 3a). We propose that during the Miocene, the northeastern reaches of the Pebas, marginal to the Purus arch and Guiana Shield (Hoorn, Wesselingh, Ter Steege, et al. 2010), 9 allowed connections between the upper Amazon and Negro/Branco, and the southern-most 10 11 reaches of the Pebas allowed connections between the Amazon and the Parana-Paraguay.

12

13 4.3 Colonization of the Brazilian Shield and lower Amazon

14 Geological evidence associates the modern channelization of the Amazon River with the 15 formation of the Amazon Fan, when Andean sediments were deposited at the Atlantic mouth of the Amazon River (Figueiredo et al. 2009; Hoorn at al. 2017). Ongoing uplift of the Andes 16 17 during the middle and late Miocene (around 10 Ma), combined with weathering of the Purus 18 arch, caused drastic changes in Amazon Basin physiognomy, allowing rivers that once drained to 19 the Pebas system to overcome paleogeographic barriers such as the Purus and Gurupá arches and 20 begin draining to the Atlantic (Figueiredo et al. 2009; Hoorn, Wesselingh, Ter Steege, et al. 21 2010; Shephard et al. 2010; Wesselingh and Hoorn, 2011; Hoorn et al. 2017). Before this point, 22 the upper and middle Amazon regions were geographically separated from what is now the lower 23 Amazon, and aquatic faunas in these two areas were likely isolated from one another. The 24 modern west to east transcontinental drainage of the Amazon, and the establishment of the lower 25 course of the Amazon, coincides with the end of the Pebas wetlands around 10 Ma (Hoorn et al. 26 2017; Albert et al. 2018). Our study provides evidence that the lower Amazon main channel was 27 colonized during the late Miocene/Pliocene after the connections between these areas and the 28 rest of the Amazon were established. We do not recover any lower Amazon lineages older than 5 29 Ma, and most Brazilian Shield associated lineages (Xingu, Tapajós areas) are no older than 7 Ma

(Figs. 3, 4b, 4c, S3). We recover dispersal from the upper Amazon and Orinoco to the Brazilian
Shield regions, and from the upper Amazon to lower Amazon, corroborating the idea that the
lower Amazon and Brazilian Shield regions were colonized by lineages moving from sources in
central and western Amazon once the modern Amazon was established (Figs. 3, 4d). The lack of
freshwater stingray lineages in the lower Amazon before 10 Ma provides additional evidence
that the marine-freshwater transition for this group was not a result of invasion from the western
Atlantic via the mouth of the Amazon.

8

9 4.4 Paleogeographic connections across the South American landscape

Biogeographic patterns in Potamotrygoninae are directly related to paleogeographic changes
in the connections and structure of South American river drainages. Below, we show how our
analyses corroborate other biogeographic patterns and paleogeographic events in South
American freshwaters.

14 The distribution of closely related taxa across cis/trans-Andean basins is a repeated pattern 15 among many groups of Neotropical fishes (Albert et al. 2006; 2018). The uplift of the Andes 16 isolated western draining (trans-Andean) basins and their associated biotas from eastern draining 17 (cis-Andean) basins (Albert et al., 2006, 2018; Rodríguez-Olarte et al., 2011). Specifically, the 18 uplift of the Eastern Cordillera in Colombia separated the trans-Andean Magdalena Basin from 19 cis-Andean rivers approximately 12-10 Ma, and the uplift of the Merida Andes isolated the trans-20 Andean Maracaibo Basin from cis-Andean rivers around 8 Ma (Hoorn et al. 1995; Lundberg et 21 al. 1998; Albert et al. 2006, 2018; Hoorn, Wesselingh, Ter Steege, et al. 2010). Our data indicate 22 that the divergence of the trans-Andean potamotrygonine lineage (~ 14 Ma) occurred slightly 23 before the uplift of the Eastern Cordillera. However, while paleogeographic reconstruction 24 predicts a ((Maracaibo, cis-Andean), Magdalena) relationship, we instead find that Maracaibo 25 and Magdalena lineages are sister taxa. This is a pattern observed in many other fish clades 26 (Albert et al., 2006), and could be explained by extinction of ancestral cis-Andean lineages 27 and/or invasion of the Maracaibo from the Magdalena through coastal stream captures (Lundberg et al. 1998, Albert et al., 2006). The findings of this study add to growing evidence for the mixed 28 29 biogeographical origins of the Maracaibo fauna.

1 Our analyses provide insight regarding the relationship between the Amazon and Orinoco 2 drainages. Prior to the late Miocene (10 - 8 Ma), a north-flowing proto-Amazon/Orinoco 3 connected the current Amazon and Orinoco regions before these basins assumed their modern 4 arrangement (Hoorn et al. 1995; Gregory-Wodnicki 2000; Mora et al. 2010). Subsequent orogeny in the Andean region resulted in sediment accumulation in parts of the sub-Andean 5 6 foreland and the uplift of the Vaupes arch, which ultimately disrupted the connection between 7 the western Amazon and the Orinoco basins (Hoorn et al. 1995; Mora et al. 2010; Winemiller and Willis, 2011; Albert et al. 2018). These drainages may then have been isolated until a later 8 9 connection, the Casiquiare, was formed, possibly as late the Pleistocene (Lujan and Armbruster, 10 2011). For freshwater stingrays, our analyses date the earliest entry to the Orinoco to 8 Ma (from 11 the Negro/Branco; Figs. 3, S3), which suggests a role of the proto-Amazon/Orinoco in this 12 distribution. After that, the next reconstructed dispersals from the Amazon (Negro/Branco) to 13 Orinoco occurred at 2 and 1.5 Ma. These more recent dispersals are likely related to the 14 formation of the Casiquiare River, and our findings thus contribute additional evidence for the 15 timing of this important connection between the Amazon and Orinoco.

16 Our results also shed light on the colonization of the Essequibo and other Atlantic-draining 17 Guiana Shield drainages. We found that stingray lineages in the Essequibo and other Atlantic-18 draining Guiana Shield rivers are relatively young, originating from both the Orinoco and 19 Negro/Branco during the late Pliocene. The close relationships between Branco, Essequibo and 20 Guiana Shield taxa recovered for the Pliocene and Pleistocene can be explained by intermittent 21 river connections between the lower portions of these drainages (as discussed in Lima, 2017); 22 alternatively, these patterns could be related to headwater capture events between the Branco and 23 the proto-Berbice (a large drainage system from the central Guiana Shield that drained most of 24 the Guyana and parts of Brazil, Venezuela and Surinam from the Cenozoic to the Pleistocene see Sinha, 1968, Schaefer and do Vale, 1997; Lujan and Armbruster, 2011). Pleistocene and 25 26 younger interactions between the Branco and Essequibo basins can be attributed to the Rupununi 27 Portal, a flooded savanna area that allows for seasonal faunal interchange between these basins 28 (Lovejoy and Araújo, 2000; Hubert and Renno, 2006; De Souza et al. 2012; 2020).

We recover three independent interchange events between the Amazon and the ParanaParaguay basins - two during the Miocene and one during the Pliocene/Pleistocene. The oldest

1 recovered Parana-Paraguay lineage (Po. brachyura) dates to ~12 Ma and resulted from a 2 colonization from the upper Amazon region (Figs. 3, 4a, S3a). A second independent 3 colonization event from the Tapajós to the Parana-Paraguay occurred ~8 Ma (Fig. S3f). Finally, 4 we find evidence that the upper Amazon species *Po. tatianae* is the product of an invasion from the Parana-Paraguay ~5 Ma (Fig. S3h). We propose that connections between the Amazon and 5 Parana-Paraguay basins resulted from headwater captures between upper Amazon tributaries 6 7 (e.g., the Madeira and the Madre de Díos) and the Bolivian Chaco. A link between these basins during the Miocene has been supported both by geological and fossil evidence (Lundberg et al. 8 1998, Uba et al. 2006; Latrubesse et al. 2007, Albert et al. 2018), and has been observed in other 9 10 fish taxa (Montova-Burgos, 2003; Carvalho and Albert, 2011). Similarly, more recent 11 associations between the Parana-Paraguay and upper portions of Brazilian Shield drainages have 12 been documented in other fishes (Lima et al. 2007; Lima and Ribeiro, 2011; Ribeiro et al. 2013). 13 We speculate that a series of Pliocene or younger river captures and reorganizations might have allowed for exchanges between these areas. 14

15

16 4.5 Upper Amazon as source of diversity and dispersal bias to adjacent basins

17 As the upper Amazon is the region of origin of Neotropical freshwater stingrays, it is 18 expected that it would host a high diversity of species. The Pebas system, in its complexity of 19 habitats and environmental gradients, would have provided ideal conditions for stingray 20 diversification, and the upper Amazon would have inherited this product (Lundberg et al. 1998; 21 Vermeij and Wesselingh, 2002). In fact, 14 of 38 potamotrygonine species occur in the upper 22 Amazon (Table S3). After 10 Ma, a direct connection between the upper and lower Amazon 23 resulted in the modern trans-continental Amazon River basin. This connection between upper 24 and lower Amazon likely contributed to the high diversity of potamotrygonine species observed 25 in the lower Amazon (nine in total). Compared to this main axis of diversity, adjacent basins 26 (e.g., the Orinoco, Atlantic-draining rivers of the Guianas, and the Parana-Paraguay) have fewer 27 species (Table S3). The observed differences in levels of diversity among basins is likely related 28 to evolutionary time (Rabosky, 2009; Gehrke and Linder, 2011): areas that were inhabited earlier 29 are occupied by older lineages that have had more time to speciate and diversify, resulting in

high species richness, while areas farther from the western Amazon are less species rich and
 composed of younger lineages (Figs. 3, 4, S1).

We also find evidence for a bias in the directionality of dispersal-- lineages often disperse 3 from the upper Amazon to peripheral basins but not the converse. For example, although both 4 Orinoco and Guiana Shield drainages have been invaded by Amazonian lineages several times, 5 none of the Orinoco or Guyana endemics have re-invaded Amazonian waters. What causes this 6 7 directionality of dispersal? One possible explanation is evolutionary priority effects, where lineages that are already present in an area prevent the subsequent establishment of competing 8 lineages, ultimately affecting the dispersal success of the latter (Belyea and Lancaster, 1999; 9 Leopold et al. 2015). Potamotrygonine lineages that successfully colonized peripheral basins 10 would have faced competition upon return to the Amazon, as ecological niches in this species-11 12 rich area would already be occupied by incumbent species. In this way, macroecological effects 13 could produce dispersal bias that correlates with comparative levels of species richness among the areas involved. 14

Regardless of age, both lower Amazon lineages and lineages from the Guianas, Orinoco, and 15 Parana-Paraguay are often the sister groups to upper Amazon lineages. Considering the patterns 16 described above, this suggests that the upper Amazon may act as a species pump for adjacent 17 18 basins and eastern lowlands. Areas classified as species pumps are diversity rich, due to higher 19 speciation rates and persistence of older lineages, and contribute lineage diversity to adjacent, 20 more species-poor regions (Stebbins, 1974; Jetz et al. 2004; Rangel et al. 2018). Another 21 proposed species pump in South America is the Andes, a region of high diversification that 22 contributed to the diversity of adjacent lowland habitats (e.g., Antonelli and Sanmartín, 2011; Ledo and Colli, 2017; Rangel et al. 2018). We propose that, for freshwater stingrays and possibly 23 other fishes, the upper Amazon has acted as a species pump, injecting diversity to adjacent 24 basins over the past 10 million years. 25

The diversification process is regulated by many different biotic and abiotic factors. Here, we provide phylogenetic evidence for the influence of time, biogeography, and ecology on diversification and distribution patterns in a diverse group of Neotropical freshwater fishes. A combination of lineage age, biogeographical connectivity, and evolutionary priority effects likely

play a role in the observed patterns of endemism, distribution, and diversity, highlighting the
 macroevolutionary complexity of the Neotropics.

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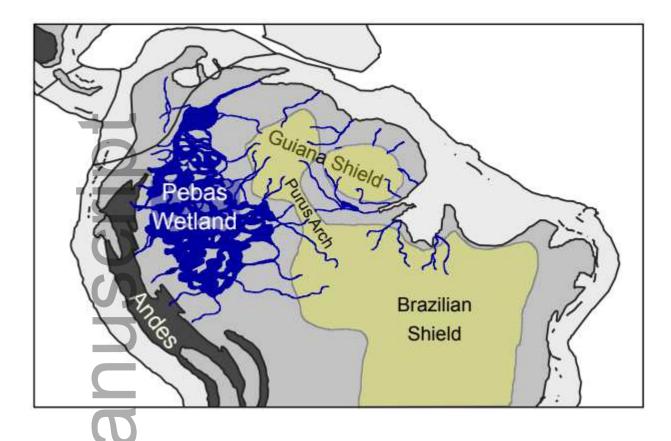
14

Figures



- 6 TABLE 1: Models estimated in 'BioGeoBEARS' for ancestral range estimation of
- 7 potamotrygonine stingrays. For each model, we provide the literature reference, log-likelihood
- 8 value (lnL), number of parameters (numparams), rate of range expansion (d), rate of range
- 9 contraction (e), relative weight of jump dispersal at cladogenesis (j), and corrected Akaike's
- 10 information criteria (AICc). Models are ordered from best to worst fit based on AICc values.

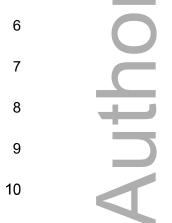
| Model | Reference | lnL | numparams | d | e | j | AICc |
|---------------|---------------------------------|-----------|-----------|----------|----------|--------|----------|
| DEC+J | Matzke, 2014 | -260.7868 | 3 | 1.00E-12 | 1.00E-12 | 0.0229 | 527.7381 |
| DIVALIKE+J | Matzke, 2014 | -262.1620 | 3 | 1.00E-12 | 1.00E-12 | 0.0233 | 530.4883 |
| BAYAREALIKE+J | Matzke, 2014 | -262.2082 | 3 | 1.00E-07 | 1.00E-07 | 0.0230 | 530.5809 |
| DEC | Ree and Smith, 2008 | -358.4968 | 2 | 9.90E-02 | 0 | 0 | 721.0752 |
| DIVALIKE | Ronquist and Sanmartín, 2011 | -473.9102 | 2 | 1.20E-01 | 8.10E-01 | 0 | 951.9020 |
| BAYAREALIKE | Yu et al. 2013 | -473.9102 | 2 | 1.20E-01 | 8.10E-01 | 0 | 951.9020 |
| + | | | | | | | |
| _ | | | | | | | |



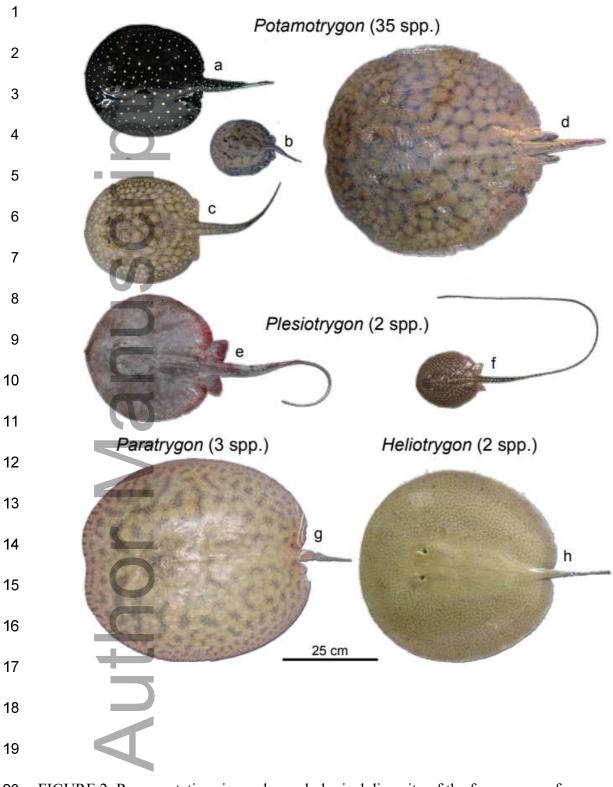
- 1
- 2 FIGURE 1: Map of South America during the middle Miocene showing position of the Pebas
- 3 Wetlands (adapted from Hoorn et al. 2010. using shapefiles from Cao et al. 2017). Shield

4 formations indicated in yellow, lowlands in gray, continental shelf in light gray, and the Andes in

5 dark gray. Map in EPSG:4326 (WGS84) projection.



11

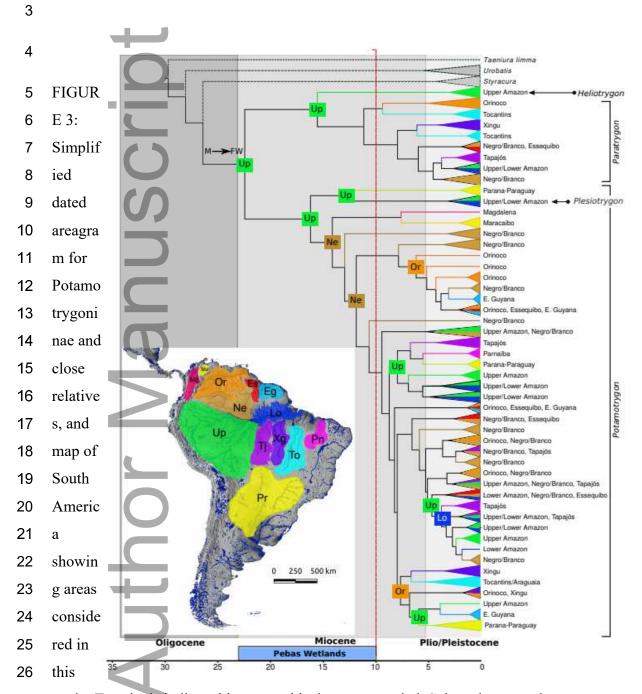


20 FIGURE 2: Representative size and morphological diversity of the four genera of

- 21 Potamotrygoninae, with currently recognized numbers of species for each genus. a:
- 22 Potamotrygon albimaculata; **b**: Potamotrygon wallacei; **c**: Potamotrygon orbignyi; **d**:

1 Potamotrygon brachyura; e: Plesiotrygon iwamae; f: Plesiotrygon nana; g: Paratrygon aiereba;



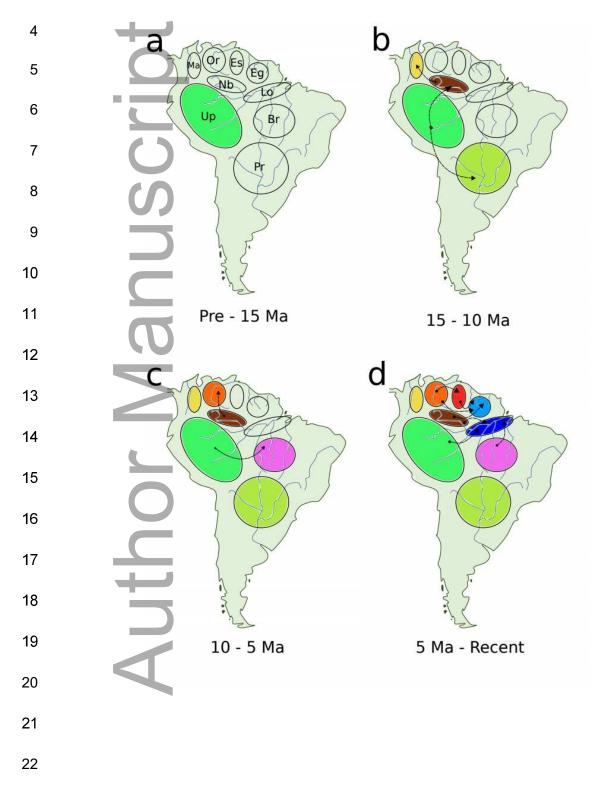


study. Terminals indicate biogeographical areas occupied. Selected ancestral area reconstructions
are shown (most probable estimated ancestral ranges according to 'BioGeoBEARS'), and
reconstructed position of marine to freshwater transition is indicated. Approximate time of Pebas
Wetlands is shown, with vertical line showing conclusion. Abbreviations as follows: Eg: Eastern

1 Guiana; Es: Essequibo; Lo: lower Amazon; Ma: Maracaibo; Mg: Magdalena; Nb: Negro-Branco;

2 Or: Orinoco; Pn: Parnaíba; Pr: Parana-Paraguay; Tj: Tapajós; Xg: Xingu; To: Tocantins-

3 Araguaia; Up: upper Amazon. Map in EPSG:4326 (WGS84) projection



2 FIGURE 4: Graphical summary of overall changes in the distribution of Potamotrygoninae in South America over time, based on 'BioGeoBEARS' analysis. Only the first colonization(s) of 3 each new area is shown, with arrows indicating the direction of colonization. Abbreviations 4 (shown in panel a) as follows: Br: Brazilian Shield; Eg: Eastern Guiana; Es: Essequibo; Lo: 5 lower Amazon; Ma: Maracaibo; Nb: Negro-Branco; Or: Orinoco; Pr: Parana-Paraguay; Up: 6 7 upper Amazon. a. Before 15 Ma; b. Early / middle Miocene (Approx. 15 – 10 Ma); c. Late Miocene/Pliocene (Approx. 10 - 5 Ma); d. Pliocene/Recent (Approx. 5 Ma -Recent). Map in 8 EPSG:4326 (WGS84) projection. 9

10 DATA AVAILABILITY STATEMENT:

Sequences used in this study are available on GenBank under accession codes MW475721 –
MW476061 for *coI*, MW480896 – MW481205 for *its1*, MW481727 – MW482078 for *cytb* and
MW482079 - MW482433 for *atpase*.

14 REFERENCES

Adnet, S., Salas Gismondi, R. and Antoine, P. O. (2014). Comparisons of dental morphology in
river stingrays (Chondrichthyes: Potamotrygonidae) with new fossils from the middle
Eocene of Peruvian Amazonia rekindle debate on their evolution. *Naturwissenschaften*101(1), 33–45.

19 Albert, J. S.; Petry, P. and Reis, R. E. (2011). Major biogeographic and phylogenetic patterns.

Historical biogeography of Neotropical freshwater fishes, University of California Press,
Berkeley, California, 21-57.

Albert, J. S., Val, P., and Hoorn, C. (2018). The changing course of the Amazon River in the
 Neogene: center stage for Neotropical diversification. *Neotropical Ichthyology*, *16*(3).

Aleman, A., and Ramos, V. A. (2000). Northern Andes. *Tectonic evolution of South America*, 31,
453-480.

Antonelli, A., and Sanmartín, I. (2011). Why are there so many plant species in the Neotropics? *Taxon*, 60(2), 403-414.

1

- 1 Aschliman, N. C., Nishida, M., Miya, M., Inoue, J. G., Rosana, K. M., and Naylor, G. J. P.
- 2 (2012). Body plan convergence in the evolution of skates and rays (Chondrichthyes:
- 3 Batoidea). *Molecular Phylogenetics and Evolution*, 63(1), 28–42.
- 4 <u>https://doi.org/10.1016/j.ympev.2011.12.012</u>
- 5 Belyea, L. R., and Lancaster, J. (1999). Assembly rules within a contingent ecology. *Oikos*, 4026 416.
- 7 Bernal, R., Bacon, C.D., Balslev, H., Hoorn, C., Bourlat, S.J., Tuomisto, H., Salamanca, S., van
- Manen, M.T., Romero, I., Sepulchre, P. and Antonelli, A. (2019). Could coastal plants in
 western Amazonia be relicts of past marine incursions? *Journal of Biogeography*, 46(8),
- 10 1749-1759.
- 11 Bloom, D. D., and Lovejoy, N. R. (2011). The biogeography of marine incursions in South
- 12 America. *Historical biogeography of Neotropical freshwater fishes*, University of
- 13 California Press, Berkeley, California, 137-144.
 - Bloom, D. D., and Lovejoy, N. R. (2017). On the origins of marine-derived freshwater fishes in South America. *Journal of Biogeography*, 44(9), 1927–1938. <u>https://doi.org/10.1111/jbi.12954</u>
- 14 Brea, M., and Zucol, A. F. (2011). The Parana-Paraguay basin: geology and paleoenvironments.
- 15 *Historical biogeography of Neotropical freshwater fishes*, University of California Press,
- 16 Berkeley, California, 69-87.
- 17 Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard, M.A., Rambaut,
- A. and Drummond, A.J., (2014). BEAST 2: a software platform for Bayesian evolutionary
 analysis. *PLoS computational biology*, *10*(4), p.e1003537.
- 20 Cao, W., Zahirovic, S., Flament, N., Williams, S., Golonka, J., and Müller, R. D. (2017).
- 21 Improving global paleogeography since the late Paleozoic using paleobiology.
- 22 Biogeosciences, 14(23), 5425–5439. <u>https://doi.org/10.5194/bg-14-5425-2017</u>
- 23 Carvalho, T. and Albert, J. (2011). The Amazon-paraguay divide. *Historical Biogeography of*
- 24 Neotropical Freshwater Fishes, University of California Press, Berkeley, California, 193–
- 25 202.

| 1 | Carvalho, M. R.; Lovejoy, N. and Rosa, R. (2003). Family Potamotrygonidae (river stingrays). |
|----|--|
| 2 | <i>Check list of the freshwater fishes of South and Central America</i> , Porto Alegre, Brazil: |
| 3 | EDIPUCRS, 22–28. |
| 4 | Carvalho, M. R.; Maisey, J. G. and Grande, L. (2004). Freshwater stingrays of the Green River |
| 5 | Formation of Wyoming (Early Eocene), with the description of a new genus and species |
| 6 | and an analysis of its phylogenetic relationships (Chondrichthyes: Myliobatiformes). |
| 7 | Bulletin of the American Museum of Natural History, 1–136. |
| 8 | Carvalho, M. R., Loboda, T. S., and Da Silva, J. P. C. B. (2016). A new subfamily, Styracurinae, |
| 9 | and new genus, Styracura, for Himantura schmardae (Werner, 1904) and Himantura |
| 10 | pacifica (Beebe and Tee-Van, 1941) (Chondrichthyes: Myliobatiformes). Zootaxa, |
| 11 | 4175(3), 201-221. |
| 12 | Cassens, I., Vicario, S., Waddell, V.G., Balchowsky, H., Van Belle, D., Ding, W., Fan, C., |
| 13 | Mohan, R.L., Simões-Lopes, P.C., Bastida, R. and Meyer, A., (2000). Independent |
| 14 | adaptation to riverine habitats allowed survival of ancient cetacean lineages. Proceedings |
| 15 | of the National Academy of Sciences, 97(21), 11343-11347. |
| 16 | Castello, H., Lasso, C. A., Fontenelle, J. P., Loboda, T., da Silva, J. P. C. B., Carvalho, M. R., |
| 17 | Rosa, R. S. (2016). Potamotrygon brachyura. XV. Rayas de Agua Dulce |
| 18 | (Potamotrygonidae) de Suramérica. Parte II. Colombia, Brasil, Perú, Bolivia, Paraguay, |
| 19 | Uruguay y Argentina, Instituto Humboldt, Colombia, 137-141. |
| 20 | Cavalcanti, M. J., dos Santos-Silva, E. N., and Primeiro, L. J. D. O. G. (2019). Database of the |
| 21 | marine-derived aquatic biota of the Amazon Basin. Tropical Diversity, 1(1), 12-25. |
| 22 | Chabain, J., Antoine, P. O., Altamirano-Sierra, A. J., Marivaux, L., Pujos, F., Gismondi, R. S., |
| 23 | and Adnet, S. (2017). Cenozoic batoids from Contamana (Peruvian Amazonia) with focus |
| 24 | on freshwater potamotrygonins and their paleoenvironmental significance. Geobios, 50(5- |
| 25 | 6), 389-400. |
| 26 | Dagosta, F. C., and de Pinna, M. C. (2017). Biogeography of Amazonian fishes: deconstructing |
| 27 | river basins as biogeographic units. Neotropical Ichthyology, 15(3), 1-24. |
| 28 | Dagosta, F. C., and de Pinna, M. C. (2019). The fishes of the Amazon: distribution and |
| 29 | biogeographical patterns, with a comprehensive list of species. Bulletin of the American |
| 30 | Museum of Natural History, 431, 1-163. |
| | |

| 1 | Darriba, D., Taboada, G. L., Doallo, R., and Posada, D. (2012). jModelTest 2: more models, new |
|----|---|
| 2 | heuristics and parallel computing. Nature methods, 9(8), 772-772. |
| 3 | De Souza, L. S.; Armbruster, J. W. and Werneke, D. C. (2012). The influence of the Rupununi |
| 4 | portal on distribution of freshwater fish in the Rupununi district, Guyana. CYBIUM 36(1), |
| 5 | 31–43. |
| 6 | De Souza, L.S., J.W. Armbruster, and P. Willink. (2020). Connectivity of neotropical river |
| 7 | basins in the central Guiana Shield based on fish distributions. Frontiers in Forests and |
| 8 | Global Change 3(8): 1-15. doi: 10.3389/ffgc.2020.00008.Drummond, A. J.; Ho, S. Y.; |
| 9 | Phillips, M. J. and Rambaut, A. (2006). Relaxed phylogenetics and dating with confidence. |
| 10 | PLoS biology 4(5), e88. |
| 11 | Drummond, A. J., and Rambaut, A. (2007). BEAST: Bayesian evolutionary analysis by sampling |
| 12 | trees. BMC Evolutionary Biology, 7(1), 214. https://doi.org/10.1186/1471-2148-7-214. |
| 13 | Figueiredo, J. J. J. P., Hoorn, C., Van der Ven, P., and Soares, E. (2009). Late Miocene onset of |
| 14 | the Amazon River and the Amazon deep-sea fan: Evidence from the Foz do Amazonas |
| 15 | Basin. Geology, 37(7), 619-622. |
| 16 | Fontenelle, J. P. and Carvalho, M. (2016). Systematic Implications of Brain Morphology in |
| 17 | Potamotrygonidae (Chondrichthyes: Myliobatiformes). Journal of Morphology 277(2), |
| 18 | 252–263. |
| 19 | Fontenelle, J. P., Loboda, T. S., Kolmann, M., and de Carvalho, M. R. (2017). Angular cartilage |
| 20 | structure and variation in Neotropical freshwater stingrays (Chondrichthyes: |
| 21 | Myliobatiformes: Potamotrygonidae), with comments on their function and evolution. |
| 22 | Zoological Journal of the Linnean Society, 183(1), 121–142. |
| 23 | https://doi.org/10.1093/zoolinnean/zlx054 |
| 24 | Frailey, C. D. (1986). Late Miocene and Holocene mammals, exclusive of the Notoungulata, of |
| 25 | the Rio Acre region, western Amazonia. |
| 26 | Gehrke, B., and Linder, H. P. (2011). Time, space and ecology: why some clades have more |
| 27 | species than others. Journal of Biogeography, 38(10), 1948-1962. |
| 28 | Gregory-Wodzicki, K. M. (2000). Uplift history of the Central and Northern Andes: a review. |
| 29 | Geological Society of America Bulletin 112(7), 1091–1105. |

| 1 | Hamilton, H., Caballero, S., Collins, A. G., and Brownell Jr, R. L. (2001). Evolution of river |
|----|---|
| 2 | dolphins. Proceedings of the Royal Society of London. Series B: Biological Sciences, |
| 3 | 268(1466), 549-556. |
| 4 | Ho, S. Y. and Phillips, M. J. (2009). Accounting for calibration uncertainty in phylogenetic |
| 5 | estimation of evolutionary divergence times. Systematic Biology 58(3), 367-380. |
| 6 | Hoorn, C.; Guerrero, J.; Sarmiento, G. A. and Lorente, M. A. (1995). Andean tectonics as a |
| 7 | cause for changing drainage patterns in Miocene northern South America. Geology 23(3), |
| 8 | 237–240. |
| 9 | Hoorn, C. (2006). Mangrove forests and marine incursions in Neogene Amazonia (lower |
| 10 | Apaporis River, Colombia). Palaios, 21(2), 197-209. |
| 11 | Hoorn, C., Wesselingh, F.P., Ter Steege, H., Bermudez, M.A., Mora, A., Sevink, J., Sanmartín, |
| 12 | I., Sanchez-Meseguer, A., Anderson, C.L., Figueiredo, J.P. and Jaramillo, C., (2010). |
| 13 | Amazonia through time: Andean uplift, climate change, landscape evolution, and |
| 14 | biodiversity. Science, 330(6006), 927-931. |
| 15 | Hoorn, C., Wesselingh, F. P., Hovikoski, J., and Guerrero, J. (2010). The development of the |
| 16 | amazonian mega-wetland (Miocene; Brazil, Colombia, Peru, Bolivia). Amazonia, |
| 17 | landscape and species evolution: a look into the past, Blackwell-Wiley, Hoboken, 123- |
| 18 | 142. |
| 19 | Hoorn, C., Bogotá-A, G.R., Romero-Baez, M., Lammertsma, E.I., Flantua, S.G., Dantas, E.L., |
| 20 | Dino, R., do Carmo, D.A. and Chemale Jr, F., (2017). The Amazon at sea: onset and stages |
| 21 | of the Amazon River from a marine record, with special reference to Neogene plant |
| 22 | turnover in the drainage basin. Global and Planetary Change, 153, 51-65. |
| 23 | Hovikoski, J., Wesselingh, F. P., Räsänen, M., Gingras, M., and Vonhof, H. B. (2010). Marine |
| 24 | influence in Amazonia: evidence from the geological record. Amazonia, landscape and |
| 25 | species evolution: a look into the past, Blackwell-Wiley, Hoboken, 143-160. |
| 26 | Hubert, N., and Renno, J. F. (2006). Historical biogeography of South American freshwater |
| 27 | fishes. Journal of Biogeography, 33(8), 1414-1436. |
| 28 | Jetz, W., Rahbek, C., and Colwell, R. K. (2004). The coincidence of rarity and richness and the |
| 29 | potential signature of history in centres of endemism. Ecology Letters, 7(12), 1180-1191. |
| | |

| 1 | Kolmann, M. A., Welch, K. C., Summers, A. P., and Lovejoy, N. R. (2016). Always chew your |
|----|---|
| 2 | food: freshwater stingrays use mastication to process tough insect prey. Proceedings of the |
| 3 | Royal Society B: Biological Sciences, 283(1838), 20161392. |
| 4 | https://doi.org/10.1098/rspb.2016.1392 |
| 5 | Landis, M. J., Matzke, N. J., Moore, B. R., and Huelsenbeck, J. P. (2013). Bayesian Analysis of |
| 6 | Biogeography when the Number of Areas is Large. Systematic Biology, 62(6), 789-804. |
| 7 | https://doi.org/10.1093/sysbio/syt040 |
| 8 | Latrubesse, E. M.; da Silva, S. A.; Cozzuol, M. and Absy, M. L. (2007). Late Miocene |
| 9 | continental sedimentation in southwestern Amazonia and its regional significance: Biotic |
| 10 | and geological evidence. Journal of South American Earth Sciences 23(1), 61-80. |
| 11 | Ledo, R. M. D., and Colli, G. R. (2017). The historical connections between the Amazon and the |
| 12 | Atlantic Forest revisited. Journal of Biogeography, 44(11), 2551-2563. |
| 13 | Leigh, E. G.; O'dea, A. and Vermeij, G. J. (2014). Historical biogeography of the Isthmus of |
| 14 | Panama. Biological Reviews 89(1), 148–172. |
| 15 | Leopold, D. R., Tanentzap, A. J., Lee, W. G., Heenan, P. B., and Fukami, T. (2015). |
| 16 | Evolutionary priority effects in New Zealand alpine plants across environmental gradients. |
| 17 | Journal of Biogeography, 42(4), 729-737. |
| 18 | Lima, F.C.T. (2017). A revision of the cis-andean species of the genus Brycon Müller and |
| 19 | Troschel (Characiformes: Characidae). Zootaxa, 4222(1), 1-189. |
| 20 | https://doi.org/10.11646/zootaxa.4222.1.1 |
| 21 | Lima, F.C.T., H.A. Britski and F.A. Machado. (2007). A new Moenkhausia (Characiformes: |
| 22 | Characidae) from central Brazil, with comments on the area relationships between the |
| 23 | upper rio Tapajós and upper rio Paraguai systems. Aqua, International Journal of |
| 24 | Ichthyology, 13(2), 45-54. |
| 25 | Lima, F. C. and Ribeiro, A. C. (2011). Continental-scale tectonic controls of biogeography and |
| 26 | ecology. Historical Biogeography of Neotropical freshwater fishes, University of |
| 27 | California Press, Berkeley, California, 145–164. |
| 28 | Lima, S. M., Berbel-Filho, W. M., Araújo, T. F., Lazzarotto, H., Tatarenkov, A., and Avise, J. C. |
| 29 | (2017). Headwater capture evidenced by paleo-rivers reconstruction and population genetic |
| | |

- structure of the armored catfish (*Pareiorhaphis garbei*) in the Serra do Mar mountains of
 southeastern Brazil. *Frontiers in genetics*, 8, 199.
- Loboda, T., Lasso, C. A., Rosa, R., de Carvalho, M. *In press*. Two new species of freshwater
 stingrays of the genus *Paratrygon* (Chondrichthyes: Potamotrygonidae) from the Orinoco
- 5 basin, with comments on the taxonomy of *Paratrygon aiereba*. *Neotropical Ichthyology*.
- 6 Louterbach, M., Roddaz, M., Bailleul, J., Antoine, P.O., Adnet, S., Kim, J.H., van Soelen, E.,
- 7 Parra, F., Gérard, J., Calderon, Y. and Gagnaison, C.,(2014). Evidence for a paleocene
- 8 marine incursion in southern Amazonia (Madre de Dios sub-Andean zone, Peru).
- 9 *Palaeogeography, palaeoclimatology, palaeoecology, 414, 451-471.*
- 10 Lovejoy, N.R. (1996). Systematics of myliobatoid elasmobranchs: with emphasis on the
- 11 phylogeny and historical biogeography of neotropical freshwater stingrays
- 12 (Potamotrygonidae: Rajiformes). *Zoological Journal of the Linnaen Society*, 117, 207–257.
- 13 Lovejoy, N. R., Bermingham, E., and Martin, A. P. (1998). Marine incursion into South
- 14 America. *Nature*, *396*(6710), 421-422.
 - Lovejoy, N. and De Araújo, M. (2000). Molecular systematics, biogeography and population structure of Neotropical freshwater needlefishes of the genus *Potamorrhaphis*. *Molecular Ecology* 9(3), 259–268.
 - Lovejoy, Nathan R., Albert, J. S., and Crampton, W. G. R. (2006). Miocene marine incursions and marine/freshwater transitions: Evidence from Neotropical fishes. *Journal of South American Earth Sciences*, 21(1–2), 5–13. <u>https://doi.org/10.1016/j.jsames.2005.07.009</u>
- 15 Lovejoy, N. R.; Willis, S. C. and Albert, J. S. (2010). Molecular signatures of Neogene
- biogeographical events in the Amazon fish fauna. *Amazonia: Landscape and Species Evolution: A look into the past.* Hoboken: Wiley-Blackwell: 405–417.
- Lujan, N. K. and Armbruster, J. W. (2011). The Guiana Shield. *Historical biogeography of Neotropical freshwater fishes*, University of California Press, Berkeley, California, 211 224.
- 21 Lundberg, J. G., Marshall, L. G., Guerrero, J., Horton, B., Malabarba, M. C. S. L., and
- 22 Wesselingh, F. (1998). The stage for Neotropical fish diversification: a history of tropical
- 23 South American rivers. *Phylogeny and classification of Neotropical fishes*, *603*, 14-48.

Marques, F. P. L. (2001). Evolution of neotropical freshwater stingrays and their parasites,
 taking into account space and time. PhD thesis (University of Toronto, Toronto-ON,
 Canada., 2001).

Matzke, N. J. (2013). BioGeoBEARS: BioGeography with Bayesian (and Likelihood)
Evolutionary Analysis in R Scripts. University of California, Berkeley, Berkeley, CA.
Matzke, N. J. (2014). Model selection in historical biogeography reveals that founder-event
speciation is a crucial process in island clades. *Systematic Biology 63*(6), 951–970.

- 8 Montoya-Burgos, J.-I. (2003). Historical biogeography of the catfish genus *Hypostomus*9 (Siluriformes: Loricariidae), with implications on the diversification of Neotropical
 10 ichthyofauna. *Molecular Ecology 12*(7), 1855–1867.
- 11 Mora, A., Reyes-Harker, A., Rodriguez, G., Tesón, E., Ramirez-Arias, J. C., Parra, M., .. and

12 Ibañez, M. (2013). Inversion tectonics under increasing rates of shortening and

13 sedimentation: Cenozoic example from the Eastern Cordillera of Colombia. *Geological*

14 Society, London, Special Publications, 377(1), 411-442.

Nuttall, C. (1990). A review of the tertiary non-marine molluscan faunas of the Pabasian and
other inland basins of north-western South America. *Bulletin of the British Museum Natural History*, 45, 165-372.

Rabosky, D. L. (2009). Ecological limits and diversification rate: alternative paradigms to
explain the variation in species richness among clades and regions. *Ecology letters*, 12(8),
735-743.

Rangel, T. F., Edwards, N. R., Holden, P. B., Diniz-Filho, J. A. F., Gosling, W. D., Coelho, M.
T. P., and Colwell, R. K. (2018). Modeling the ecology and evolution of biodiversity:

23 Biogeographical cradles, museums, and graves. *Science*, *361*(6399).

- Reis, R. E., Albert, J. S., Di Dario, F., Mincarone, M. M., Petry, P., and Rocha, L. A. (2016).
 Fish biodiversity and conservation in South America. *Journal of fish biology*, 89(1), 12-47.
- 26 Ribeiro, A.C., R.M. Jacob, R.R.S.R. Silva, F.C.T. Lima, D.C. Ferreira, K.M. Ferreira, T.C.
- 27 Mariguela, L.H.G. Pereira and C. Oliveira. (2013). Distributions and phylogeographic data
- of rheophilic freshwater fishes provide evidences on the geographic extension of a central-
- brazilian amazonian palaeoplateau in the area of the present day Pantanal Wetland.
- 30 *Neotropical Ichthyology 11(2)*, 319-326.

1 Roberts, T. R. (1972). Ecology of fishes in the Amazon and Congo basins. Bull Mus Comp Zool, 2 143(2), 117-147.

Rodríguez-Olarte, D., Taphorn, D. C., and Lobón-Cerviá, J. (2011). Do protected areas conserve 3 4 neotropical freshwater fishes? A case study of a biogeographic province in Venezuela.

- Animal Biodiversity and Conservation, 34(2), 273-285. 5
- Ronquist, F., and Sanmartín, I. (2011). Phylogenetic Methods in Biogeography. Annual Review 6 7 of Ecology, Evolution, and Systematics, 42(1), 441-464. https://doi.org/10.1146/annurevecolsys-102209-144710 8
- 9 Rosa, R. S., Araújo, M. L. G., Carvalho, M. R. (2016). Potamotrygon wallacei. XV. Rayas de

Agua Dulce (Potamotrygonidae) de Suramérica. Parte II. Colombia, Brasil, Perú, Bolivia, 10 11 Paraguay, Uruguay y Argentina. Instituto Humboldt, Colombia, 167-172.

Rutledge, K.M., Summers, A.P. and Kolmann, M.A., (2019). Killing them softly: Ontogeny of 12

13 jaw mechanics and stiffness in mollusk-feeding freshwater stingrays. Journal of

Morphology, 280(6), 796-808. 14

Salas-Gismondi, R., Flynn, J. J., Baby, P., Tejada-Lara, J. V., Wesselingh, F. P., and Antoine, P. 15

16 O. (2015). A Miocene hyperdiverse crocodylian community reveals peculiar trophic

17 dynamics in proto-Amazonian mega-wetlands. Proceedings of the Royal Society B:

Biological Sciences, 282(1804), 20142490. 18

19 Schaefer, C. E. R., and Vale Júnior, J. F. (1997). Mudanças climáticas e evolução da paisagem

- 20 em Roraima: Uma resenha do Cretáceo ao recente. Homem, Ambiente e Ecologia em 21 Roraima, Instituto Nacional de Pesquisas da Amazônia, Manaus, 231-265.
- 22 Shephard, G.; Müller, R.; Liu, L and Gurnis, M. (2010). Miocene drainage reversal of the 23

Amazon River driven by plate-mantle interaction. *Nature Geoscience* 3(12), 870.

24 Silva, J. P. C. B., and Loboda, T. S. (2019). Potamotrygon marguesi, a new species of

- neotropical freshwater stingray (Potamotrygonidae) from the Brazilian Amazon Basin. 25
- Journal of Fish Biology, 95(2), 594–612. https://doi.org/10.1111/jfb.14050 26
- 27 Sinha, N. K. (1968). Geomorphic Evolution of The Northern Rupununi Basin, Guyana (No. Tr-
- 28 12). Mcgill Univ Montreal (Quebec) Dept Of Geography.

| 1 | Solomon, S. E., Bacci Jr, M., Martins Jr, J., Vinha, G. G., and Mueller, U. G. (2008). |
|----|---|
| 2 | Paleodistributions and comparative molecular phylogeography of leafcutter ants (Atta spp.) |
| 3 | provide new insight into the origins of Amazonian diversity. <i>PloS one</i> , <i>3</i> (7), e2738. |
| 4 | Stebbins, G. L. (1974). Flowering plants. Evolution above the species level. The Belknap Press of |
| 5 | Harvard University Press. Cambridge, MA, USA. |
| 6 | Uba, C. E.; Heubeck, C. and Hulka, C. (2006). Evolution of the late Cenozoic Chaco foreland |
| 7 | basin, southern Bolivia. Basin Research 18(2), 145–170. |
| 8 | Vermeij, G. J., and Wesselingh, F. P. (2002). Neogastropod molluscs from the Miocene of |
| 9 | western Amazonia, with comments on marine to freshwater transitions in molluscs. |
| 10 | Journal of Paleontology, 76(2), 265-270. |
| 11 | Wesselingh, F. P., Hoorn, M. C., Guerrero, J., Rasanen, M. E., Romero Pittman, L., and Salo, J. |
| 12 | S. (2006). The stratigraphy and regional structure of Miocene deposits in western |
| 13 | Amazonia (Peru, Colombia and Brazil), with implications for late Neogene landscape |
| 14 | evolution. Scripta Geologica, 133, 291-322. |
| 15 | Wesselingh, F. P., and Ramos, M. I. F. (2010). Amazonian aquatic invertebrate faunas |
| 16 | (Mollusca, Ostracoda) and their development over the past 30 million years. Amazonia: |
| 17 | Landscape and Species Evolution: A look into the past, Blackwell-Wiley, Hoboken, 302- |
| 18 | 316. |
| 19 | Wesselingh, F. P. and Hoorn, C. (2011). Geological Development of Amazon and Orinoco |
| 20 | Basins. Historical Biogeography of Neotropical Freshwater Fishes, University of |
| 21 | California Press, Berkeley, 59–68. |
| 22 | Winemiller, K. O. and Willis, S. C. (2011). The Vaupes arch and Casiquiare canal. Historical |
| 23 | Biogeography of Neotropical Freshwater Fishes, University of California Press, Berkeley, 225- |
| 24 | 242. |
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| | |
| 26 | BIOSKETCH |

27 João Pedro Fontenelle is interested in the biogeography of the Neotropical region. This work is a

28 component of his PhD work at UTSC on the evolution and diversification of the stingrays of the

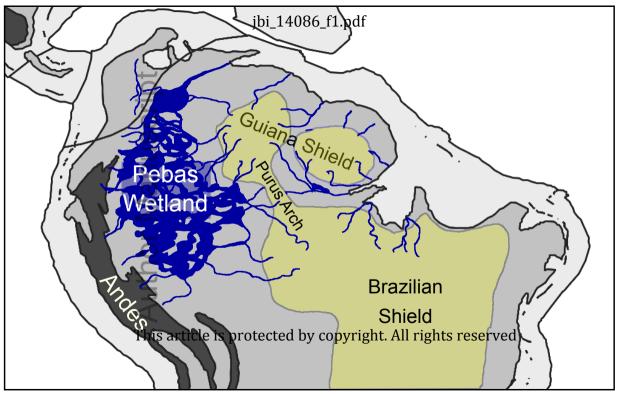
1 subfamily Potamotrygoninae. He and the other authors collaborate on questions pertaining

2 freshwater fish diversity and evolution.

6

- 3 Author contributions: JPF and NRL conceived the ideas; JPF, NRL, MAK and FPLM all
- 4 conducted field work and collected the data with additional material from museum collections;
- 5 JPF, MAK and FPLM analyzed the data; JPF led the writing with assistance from NRL, MAK

and FPLM. Author Manuscr



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