

Tocilizumab Prevents Progression of Early Systemic Sclerosis–Associated Interstitial Lung Disease

David Roofeh,¹ Celia J. F. Lin,² Jonathan Goldin,³ Grace Hyun Kim,³ Daniel E. Furst,⁴ Christopher P. Denton,⁵ Suiyuan Huang,¹ and Dinesh Khanna,¹ on behalf of the focuSSced Investigators

Objective. Tocilizumab (TCZ) has demonstrated lung function preservation in 2 randomized controlled trials in early systemic sclerosis (SSc). This effect has yet to be characterized in terms of radiographically evident quantitative lung involvement. We undertook this study to assess the impact of TCZ on lung function preservation in a post hoc analysis, stratifying treatment arms according to the degree of lung involvement.

Methods. The focuSSced trial was a phase III randomized placebo-controlled trial of TCZ in patients with SSc and progressive skin disease. Participants underwent baseline and serial spirometry along with high-resolution chest computed tomography at baseline and at week 48. Quantitative interstitial lung disease (QILD) and fibrosis scores were assessed by computer software. We classified QILD into the following categories of lung involvement: mild (>5–10%), moderate (>10–20%), and severe (>20%).

Results. Of 210 participants recruited for the trial, 136 patients (65%) had ILD. The majority of these patients (77%) had moderate-to-severe involvement (defined as >10% lung involvement). The TCZ arm demonstrated preservation of forced vital capacity percent predicted (FVC%) over 48 weeks (least squares mean change in FVC% = –0.1) compared to placebo (–6.3%). For mild, moderate, and severe QILD, the mean ± SD change in FVC% in the TCZ arm at 48 weeks were –4.1 ± 2.5% (n = 11), 0.7 ± 1.9% (n = 19), and 2.1 ± 1.6% (n = 26), respectively, and in the placebo group were –10.0 ± 2.6% (n = 11), –5.7 ± 1.6% (n = 26), and –6.7 ± 2.0% (n = 16), respectively. Similar treatment-related preservation findings were seen independent of fibrosis severity.

Conclusion. TCZ in early SSc–associated ILD with progressive skin disease stabilized FVC% over 48 weeks, independent of the extent of radiographically evident QILD.

INTRODUCTION

The majority of systemic sclerosis (SSc) patients will develop interstitial lung disease (ILD) (1,2). The disease process of SSc-associated ILD (SSc-ILD) usually proceeds through different phases. The initial phase is associated with findings from

high-resolution computed tomography (HRCT) of the chest that predominantly show ground-glass opacity with minimal fibrotic changes (considered by some to be immunoinflammatory), followed by more dense fibrotic changes with a nonspecific interstitial pneumonia pattern on HRCT scans; however, some patients may present with findings of usual interstitial pneumonitis (3). Those

ClinicalTrials.gov identifier: NCT02453256.

Dr. Roofeh's work was supported by the National Institute of Arthritis and Musculoskeletal and Skin Diseases, NIH (grant T32-AR-007080). Dr. Khanna's work was supported by the Immune Tolerance Network and the National Institute of Arthritis and Musculoskeletal and Skin Diseases, NIH (grants K24-AR-063129 and 1R01-AR-070470-01A1).

¹David Roofeh, MD, Suiyuan Huang, MPH, Dinesh Khanna, MD, MS: University of Michigan, Ann Arbor; ²Celia J. F. Lin, MD: Genentech, South San Francisco, California; ³Jonathan Goldin, MD, Grace Hyun Kim, PhD: University of California, Los Angeles; ⁴Daniel E. Furst, MD: University of California, Los Angeles, University of Washington, Seattle, and University of Florence, Florence, Italy; ⁵Christopher P. Denton, FRCP: University College London, London, UK.

Dr. Lin has received consulting fees, speaking fees, and/or honoraria from Genentech (less than \$10,000) and owns stock or stock options in Roche. Dr. Goldin has received consulting fees, speaking fees, and/or honoraria from MedQIA (less than \$10,000). Dr. Kim has received consulting fees, speaking fees, and/or honoraria from MedQIA (less than \$10,000) and has a patent application pending related to interstitial lung disease. Dr. Furst has received consulting fees, speaking fees, and/or honoraria from Corbus, CSL Behring, Galápagos, Gilead,

Pfizer, Talaris, AbbVie, Amgen, Novartis, Roche/Genentech, and Boehringer Ingelheim (less than \$10,000 each) and research grants from Corbus, CSL Behring, Galápagos, GlaxoSmithKline, Kadmon, PCORI, Pfizer, and Talaris. Dr. Denton has received consulting fees, speaking fees, and/or honoraria from Acceleron, Actelion, GlaxoSmithKline, Horizon, Sanofi, Inventiva, Boehringer Ingelheim, Roche, CSL Behring, and Corbus (less than \$10,000 each) and research grants from GlaxoSmithKline, Inventiva, and CSL Behring. Dr. Khanna has received consulting fees, speaking fees, and/or honoraria from Bristol Myers Squibb, Bayer, Acceleron, Actelion, Amgen, Blade Therapeutics, Boehringer Ingelheim, CSL Behring, Corbus, Cytori, Galápagos, Genentech/Roche, GlaxoSmithKline, Horizon, Merck, Mitsubishi Tanabe, Regeneron, Sanofi-Aventis, United Therapeutics, and Impact PH (less than \$10,000 each), research grants from Bristol Myers Squibb, Horizon, and Pfizer, and is the Chief Medical Officer of Eicos Sciences. No other disclosures relevant to this article were reported.

Address correspondence to Dinesh Khanna, MD, MS, University of Michigan, Division of Rheumatology, Department of Internal Medicine, 300 North Ingalls Street, Ann Arbor, MI 48109. Email: khannad@med.umich.edu.

Submitted for publication October 7, 2020; accepted in revised form January 26, 2021.

at risk of progressive disease have an archetype: early, diffuse cutaneous systemic sclerosis (dcSSc), with elevated acute-phase reactants such as C-reactive protein (CRP) level and topoisomerase I (topo I) antibody positivity (4–7). Patients with these high-risk features, especially those with disease in the initial phase of development, represent an important target for early intervention, as ILD is largely irreversible in SSc (4,8).

Tocilizumab (TCZ) is an anti-interleukin-6 (anti-IL-6) agent (IgG1 humanized anti-IL-6 receptor monoclonal antibody), approved for use in rheumatoid arthritis, giant cell arteritis, juvenile idiopathic arthritis, Castleman's disease, and other immune-mediated diseases. Two well-designed randomized controlled trials of TCZ in early dcSSc demonstrated a significant lung preservation effect in the treatment arm compared to placebo (9,10). This effect has yet to be characterized in terms of radiographically evident quantitative lung involvement.

In this post hoc analysis, we comprehensively characterized the ILD participants in the focuSSced trial (10), assessed the relationship between degree of total lung involvement and fibrosis (using well-established quantitative HRCT measurements) and lung physiology, and evaluated the treatment effect of TCZ compared to placebo on forced vital capacity percent predicted (FVC%) and quantitative HRCT. Investigating the treatment effects in terms of radiographic changes in this cohort at high risk for progression of ILD provides important insight into disease pathophysiology and potential mechanisms of therapeutic benefit.

PATIENTS AND METHODS

Study design. This phase III trial (ClinicalTrials.gov identifier: NCT02453256) was a multicenter, randomized, double-blind placebo-controlled trial with 1:1 randomization to active treatment (1 subcutaneous injection of 162 mg TCZ per week) or placebo for 48 weeks (10). Background immunosuppressive therapy was not allowed in the trial, but escape therapy was allowed for prespecified skin and lung function progression and SSc-related complications.

Participants. All patients met the 2013 American College of Rheumatology/European Alliance of Associations for Rheumatology classification criteria (11), with disease onset <60 months from the onset of their first non-Raynaud's phenomenon sign or symptom, and had a modified Rodnan skin thickness score (MRSS) (12) between 10 and 35. All patients had early progressive skin disease with diffuse cutaneous distribution, because the main goal of the trial was to evaluate beneficial impact of TCZ on MRSS score. Participants also had elevated acute-phase reactants (≥ 1 of the following: CRP level >6 mg/liter, erythrocyte sedimentation rate >28 mm/hour, or platelet count $>330 \times 10^9$ /liter), and active disease was defined as having >1 of the following at screening: disease duration ≤ 18 months, MRSS increase of ≥ 3 , involvement of 1 new body area and MRSS increase of ≥ 2 , or involvement of 2 new body areas (each within the previous 6 months), or ≥ 1

tendon friction rub. The presence of lung disease was not required for enrollment. The study was approved by the institutional review boards of all participating sites, written informed consent was obtained from all participants, and the study was conducted in compliance with the Declaration of Helsinki.

Outcome measures. Serial spirometry plus diffusing capacity for carbon monoxide corrected for hemoglobin (DL_{CO}) was conducted at weeks 8, 16, 24, 36, and 48, based on the American Thoracic Society/European Respiratory Society Consensus Statement recommendations (13). Patients performed 3–8 exhalations into a spirometer, and the highest value was recorded. Patients received HRCT scans at baseline and week 48, completed at maximal inspiration. Images were acquired from 30 multidetector CT scanner models from 4 manufacturers, using a standardized procedure and following strict quality control protocols. HRCT quantification was performed on all scans based on previous publications (14–16).

The quantitative ILD (QILD) score refers to the summation of ground-glass opacities, honeycombing, and fibrotic reticulation, while the quantitative lung fibrosis (QLF) score refers to quantitative fibrosis (fibrotic reticulation) alone. Both scores range from 0% to 100% involvement of the whole lung (17). All scans had QILD and QLF measurements; ILD was identified visually by a thoracic radiologist (JG) based on the presence of ground-glass opacification and/or fibrosis with a basal predominance. Participants who had minimal interstitial changes without defined ILD were characterized as having no ILD; these cases were screened for factors other than SSc-ILD and were excluded (factors included body habitus, atelectasis, bronchitis, aspiration, and bronchiectasis). QILD cutoff points were set as minimal ($\leq 5\%$), mild (>5 – 10%), moderate (>10 – 20%), or severe ($>20\%$), based on the following: 1) classification by a chest radiologist (JG), and 2) findings from Goh et al that demonstrate total lung involvement of $>20\%$ was associated with higher mortality in a longitudinal cohort (18). Cutoff points for QLF were organized in tertiles according to the range (0.1–18.5%) of involvement.

Statistical analysis. Continuous and categorical variables were summarized using the mean \pm SD and percentages, respectively. We used *t*-tests to compare baseline FVC% according to baseline QILD and QLF cutoffs. Spearman's correlation coefficients were calculated for scatterplots of baseline FVC% according to numerical baseline QILD and QLF scores, separately. To assess how the baseline QILD or QLF score affects the change in FVC% over time, we fitted linear mixed-effect models, with change in FVC% as the outcome. Covariates included the following: 1) baseline FVC%, 2) treatment arm, 3) study time points, 4) baseline QILD/QLF group, 5) interaction of baseline FVC% and study time point, 6) interaction of treatment arm and study time point, 7) interaction of baseline QILD/QLF group and treatment arm, 8) interaction of baseline QILD/QLF group and study time

point, and 9) 3-way interaction of treatment arm, study time point, and baseline QILD/QLF group. We obtained least squares means (LSMs) from the models and plotted the LSM to show the FVC% change trend. Ninety-five percent confidence intervals (95% CIs) were calculated. No data were imputed. All analyses were done using SAS software (version 9.4).

RESULTS

Baseline characteristics of patients with ILD. The distribution of patients according to treatment arm and baseline radiographic assessments are shown in Supplementary Figure 1 (available on the *Arthritis & Rheumatology* website at <http://onlinelibrary.wiley.com/doi/10.1002/art.41668/abstract>). Two hundred ten participants were randomized and received treatment (placebo arm [$n = 106$], TCZ arm [$n = 104$]). Of these patients, 136 were confirmed by a thoracic radiologist to have ILD based on HRCT imaging performed at baseline. Table 1 shows the baseline characteristics of the overall population ($n = 210$) compared to the subset of patients with ILD ($n = 136$), which was further divided by treatment arm. Three participants were confirmed as having ILD based on baseline visual assessment of HRCT scans, but the quantitative measurements (including QILD and QLF scores)

were missing. Compared to those without ILD, the remaining 133 patients with ILD had numerically lower FVC% and DLco percent predicted (DLco%), a higher CRP level, and a greater proportion of anti-topo I antibody positivity. ILD patients had a mean \pm SD FVC% of $79.6 \pm 14.5\%$ and a mean \pm SD QILD of $18.7 \pm 11.1\%$; most of the QILD score was made up of ground-glass opacities (mean \pm SD $14.9 \pm 8.3\%$), with a mean \pm SD QLF of $3.0 \pm 3.6\%$. There were no significant differences between the TCZ and placebo arms in the ILD groups at baseline (Table 1).

Moderate-to-severe whole-lung involvement with limited fibrosis in majority of ILD patients.

Baseline QILD scores of 133 patients were stratified into 4 groups corresponding to minimal ($\leq 5\%$), mild ($>5\text{--}10\%$), moderate ($>10\text{--}20\%$), and severe ($>20\%$) lung involvement. The majority of patients with ILD ($n = 102$; 77%) had moderate or severe lung involvement, as defined by a QILD of $>10\%$ (range 10.2–52.6) (Table 2). Higher degrees of QILD scores were associated with increasing MRSS scores, percentages of anti-topo I antibody positivity, lower baseline FVC% and DLco%, and higher QLF scores. Table 2 also shows ILD patients stratified according to QLF tertiles (0.1–1.0%, 1.1–2.7%, or 2.8–18.5%), with approximately two-thirds of patients ($n = 89$; 67%) having $<2.8\%$ fibrosis. Similar to QILD, increasing QLF scores were associated with

Table 1. Baseline characteristics of the overall focuSSc population and those with ILD detected on HRCT scans*

	Total patients ($n = 210$)	ILD patients ($n = 136$)	TCZ group with ILD ($n = 68$)	Placebo group with ILD ($n = 68$)
Demographics				
Female, %	81.4	79.4	77.9	80.9
Age, years	48.2 ± 12.4	48.1 ± 12.9	47.6 ± 12.5	48.7 ± 13.3
SSc duration, months	22.6 ± 16.5	22.8 ± 16.8	23.0 ± 17.2	22.6 ± 16.6
Disease features†				
Total MRSS	20.3 ± 6.8	20.8 ± 7.0	20.7 ± 6.8	20.9 ± 7.2
CRP, mg/liter	7.9 ± 13.1	9.6 ± 15.4	11.2 ± 17.4	8.0 ± 13.1
ANA positive, no. (%)	183 (92.4)	124 (96.9)	65 (98.5)	59 (95.2)
Anti-topo I positive, no. (%)	103 (51.0)	90 (68.7)	46 (68.7)	44 (68.8)
Anti-RNAP positive, no. (%)	35 (17.3)	19 (14.5)	13 (19.4)	6 (9.4)
ACA positive, no. (%)	17 (8.4)	2 (1.5)	1 (1.5)	1 (1.6)
PFTs				
FVC, ml	$2,996.7 \pm 836.8$	$2,885.4 \pm 835.8$	$2,826.8 \pm 873.7$	$2,944.1 \pm 798.3$
FVC%	82.1 ± 14.8	79.6 ± 14.5	77.7 ± 13.9	81.5 ± 14.9
DLco%‡	75.6 ± 18.9	70.4 ± 16.9	68.7 ± 16.8	72.1 ± 17.0
QILD measurements, whole lung %§				
HRCT total QILD	15.9 ± 11.4	18.7 ± 11.1	20.5 ± 12.8	16.8 ± 8.8
Ground-glass opacity	13.0 ± 8.8	14.9 ± 8.3	16.2 ± 9.5	13.6 ± 6.7
QLF	2.3 ± 3.3	3.0 ± 3.6	3.5 ± 4.2	2.5 ± 3.0
Honeycombing	0.4 ± 1.2	0.4 ± 1.3	0.5 ± 1.5	0.3 ± 1.2

* Except where indicated otherwise, values are the mean \pm SD. None of the differences between the tocilizumab (TCZ) and placebo groups were significant. SSc = systemic sclerosis; MRSS = modified Rodnan skin thickness score; CRP = C-reactive protein; PFTs = pulmonary function tests; FVC% = forced vital capacity percent predicted; DLco% = diffusing capacity for carbon monoxide corrected for hemoglobin percent predicted.

† Data were not available for all patients, as follows: for antinuclear antibody (ANA) positivity (total patients, $n = 198$; interstitial lung disease [ILD] patients, $n = 128$; TCZ arm, $n = 66$; placebo arm, $n = 62$); for anti-topoisomerase I (anti-topo I), anti-RNA polymerase (anti-RNAP), and anticentromere antibody (ACA) positivity (total patients, $n = 202$; ILD patients, $n = 131$; TCZ arm, $n = 67$; placebo arm, $n = 64$).

‡ Data were not available for all patients, as follows: total patients, $n = 208$; ILD patients, $n = 135$; TCZ arm, $n = 68$; placebo arm, $n = 67$.

§ Three patients were confirmed to have ILD based on baseline visual assessment of high-resolution computed tomography (HRCT), but data on quantitative measurements (including quantitative ILD [QILD] and quantitative lung fibrosis [QLF] scores) were missing. For these parameters, data were not available for all patients, as follows: total patients, $n = 202$; ILD patients, $n = 133$; TCZ arm, $n = 67$; placebo arm, $n = 66$.

Table 2. Baseline characteristics of ILD patients stratified by QILD and QLF involving the whole lung*

	QILD severity			QLF severity		
	Minimal, ≤5% (n = 6)	Mild, >5–10% (n = 25)	Moderate, >10–20% (n = 54)	Severe, >20% (n = 48)	First tertile, 0.1–1.0% (n = 45)	Second tertile, 1.1–2.7% (n = 44)
Demographics						
Female, %	79.0	76.0	83.3	77.1	77.8	77.3
Age, years	48.0 ± 13.0	45.5 ± 11.3	45.9 ± 13.3	52.1 ± 12.3	43.2 ± 12.7	52.5 ± 12.3
SSc duration, months	22.9 ± 16.9	22.3 ± 16.6	27.0 ± 17.2	18.5 ± 16.5	22.3 ± 13.5	19.9 ± 16.9
Disease features†						
Total MRSS	20.8 ± 7.1	18.8 ± 5.8	20.9 ± 7.6	22.3 ± 6.7	19.7 ± 6.9	20.9 ± 7.4
CRP, mg/liter	9.8 ± 15.5	5.4 ± 8.3	11.4 ± 16.8	7.5 ± 9.0	10.9 ± 18.7	11.5 ± 17.0
ANA positive, no. (%)	121 (96.8)	24 (100)	48 (96.0)	43 (95.6)	42 (97.7)	40 (97.6)
Anti-topo I positive, no. (%)	88 (68.8)	15 (62.5)	33 (66.0)	36 (75.0)	28 (65.1)	26 (63.4)
Anti-RNAP positive, no. (%)	19 (14.8)	3 (12.5)	5 (10.0)	10 (20.8)	4 (9.3)	9 (22.0)
ACA positive, no. (%)	2 (1.6)	1 (4.2)	1 (2.0)	0	1 (2.3)	1 (2.4)
PFTs						
FVC, ml	2,881.4 ± 833.6	3,483.3 ± 1,079.0	2,945.7 ± 672.4	2,532.1 ± 720.8	3,216.4 ± 908.1	2,817.5 ± 656.4
FVC%	79.5 ± 14.5	88.4 ± 18.3	81.1 ± 14.4	73.6 ± 12.9	84.8 ± 14.6	79.6 ± 13.9
DLco%‡	70.4 ± 17.1	88.5 ± 19.7	67.3 ± 12.9	63.6 ± 14.9	75.9 ± 17.4	70.9 ± 17.0
QLD measurements, whole lung %§						
HRCT total QILD	18.7 ± 11.1	4.0 ± 0.9	14.5 ± 2.8	30.8 ± 8.6	9.8 ± 4.2	16.4 ± 5.6
Ground-glass opacity	14.9 ± 8.3	3.7 ± 0.8	12.3 ± 2.7	23.5 ± 7.2	8.9 ± 3.9	14.0 ± 5.4
QLF	3.0 ± 3.6	0.3 ± 0.1	1.5 ± 1.0	6.1 ± 4.5	0.6 ± 0.3	1.7 ± 0.4
Honeycombing	0.43 ± 1.3	0	0.3 ± 0.9	0.8 ± 2.0	0.1 ± 0.5	0.3 ± 0.9

* Except where indicated otherwise, values are the mean ± SD. See Table 1 for definitions.

† Data were not available for all patients, as follows: for ANA positivity (total ILD patients, n = 125; mild QILD, n = 24; moderate QILD, n = 50; severe QILD, n = 45; first tertile QLF, n = 43; second tertile QLF, n = 41; third tertile QLF, n = 41); for anti-topo I, anti-RNAP, and ACA positivity (total ILD patients, n = 128; mild QILD, n = 24; moderate QILD, n = 50; severe QILD, n = 48; first tertile QLF, n = 43; second tertile QLF, n = 41).

‡ Data were not available for all patients, as follows: total ILD patients, n = 132; severe QILD, n = 47; third tertile QLF, n = 43.

§ Three patients were confirmed to have ILD based on baseline visual assessment of HRCT, but data on quantitative measurements (including QILD and QLF scores) were missing.

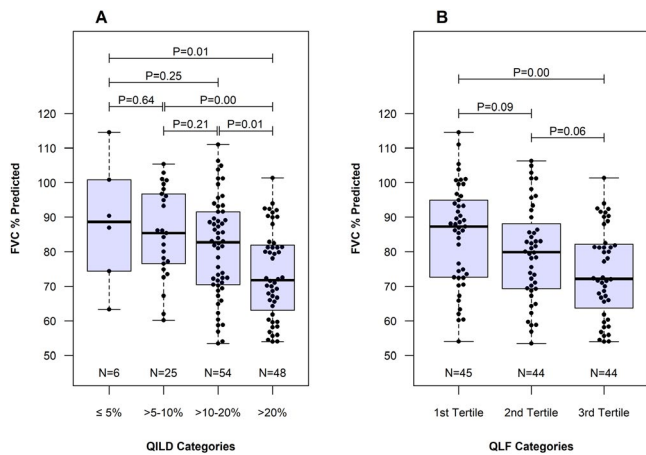


Figure 1. Relationship of forced vital capacity percent predicted (FVC%) with increasing severity of baseline quantitative interstitial lung disease (QILD) (A) and with increasing severity of baseline quantitative lung fibrosis (QLF) (B). Data are shown as box plots. Each box represents the upper and lower interquartile range. Lines inside the boxes represent the median. Whiskers represent the minimum and maximum values. Each symbol represents an individual subject.

higher percentages of anti-topo I antibody positivity and QILD, and lower baseline FVC% and DLco%.

Inverse correlation of QILD and QLF with FVC%.

Figure 1 demonstrates an inverse relationship between baseline FVC% and degree of QILD; baseline FVC% significantly declined with each escalating QILD cutoff point. The mean baseline FVC% in patients with severe QILD was significantly lower (mean ± SD 73.6 ± 12.9%) compared to those with minimal QILD (mean ± SD 88.4 ± 18.3%; *P* = 0.01), mild QILD (mean ± SD 85.4 ± 13.1%;

P = 0.00), and moderate QILD (mean ± SD 81.1 ± 14.4%; *P* = 0.01). There is an inverse correlation between baseline FVC% and QILD, with a correlation coefficient of -0.36 (*P* = 0.00). Figure 1 also demonstrates a similar inverse relationship between baseline FVC% and QLF, with the mean baseline FVC% significantly higher in the first tertile compared to the third tertile (*P* = 0.00). The correlation coefficient was also -0.36 (*P* = 0.00).

Stabilization by TCZ of FVC% over 48 weeks for mild-to-severe baseline QILD and all ranges of baseline QLF scores.

The TCZ arm demonstrated preserved FVC% over 48 weeks: the LSM of FVC% change was -0.1% for TCZ and -6.3% for placebo (Figure 2). The difference between treatment groups was 6.2% (*P* < 0.0001). Figure 2 shows the mean trend over 48 weeks of FVC% change, accounting for the covariates listed in Methods; the results are separated by treatment arm and stratified according to the extent of QILD. As there were only 2 and 4 evaluable patients with ≤5% QILD in the placebo and TCZ groups, respectively, they were excluded from what is depicted in Figure 2. Specifically, those with >5% QILD in the TCZ group showed FVC% stabilization over 48 weeks; this preservation was not influenced by the escalating degree of QILD involvement. For mild, moderate, and severe QILD, the mean ± SD change in FVC% in the TCZ arm at 48 weeks were $-4.1 ± 2.5\%$ (*n* = 11), $0.7 ± 1.9\%$ (*n* = 19), and $2.1 ± 1.6\%$ (*n* = 26), respectively, and in the placebo group were $-10.0 ± 2.6\%$ (*n* = 11), $-5.7 ± 1.6\%$ (*n* = 26), and $-6.7 ± 2.0\%$ (*n* = 16), respectively. A pairwise comparison at week 48 in the TCZ arm showed no significant differences between the mild, moderate, and severe QILD strata. Those with >5% QILD in the placebo arm showed worsening

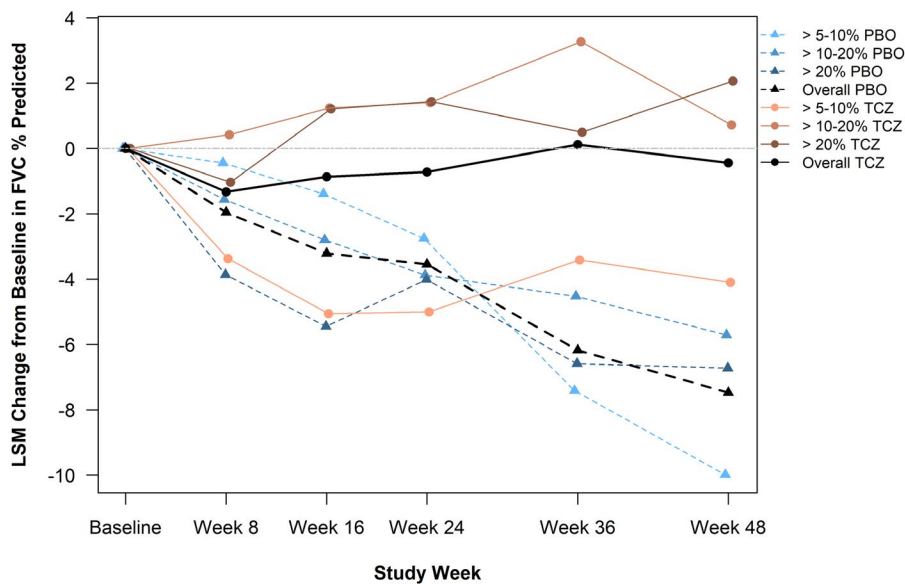


Figure 2. Mean trend over time of change in forced vital capacity percent predicted (FVC%) in the interstitial lung disease (ILD) patients, according to treatment group and quantitative ILD (QILD) score of the whole lung. The QILD severity category of ≤5% was removed from this model, as there were only 2 evaluable patients in the placebo (PBO) group and 4 evaluable patients in the tocilizumab (TCZ) treatment group with ≤5% QILD over 48 weeks. LSM = least squares mean.

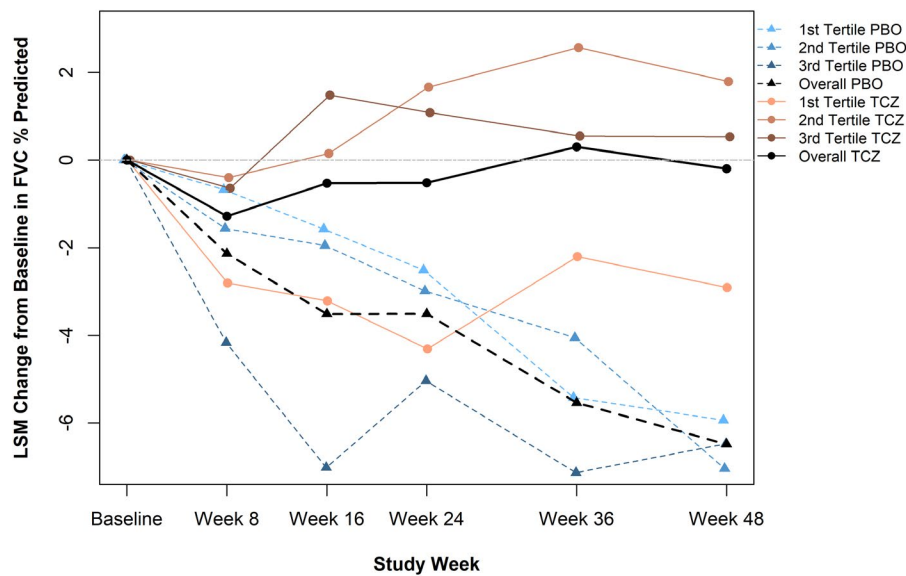


Figure 3. Mean trend over time of FVC% in theILD patients, according to treatment group and quantitative lung fibrosis score of the whole lung. See Figure 2 for other definitions.

FVC% decline, also with no significant pairwise differences in the trajectory of decline based on QILD severity.

Figure 3 shows a similar preservation effect in the TCZ arm, which was not present in the placebo arm when stratified according to QLF severity. The mean trend over time of FVC% change, accounting for the covariates listed in Methods, did not differ based on the extent of QLF for either the TCZ or placebo arm.

Stabilization by TCZ of QILD and QLF over 48 weeks, for all ranges of baseline QILD and QLF scores. Table 3 shows QILD and QLF scores at baseline and at 48 weeks, separated by treatment arm and stratified according to baseline QILD and QLF cutoff points. As expected, higher baseline QILD and QLF scores reflected higher QILD and QLF scores at 48 weeks. At 48 weeks, the overall QILD scores for the TCZ arm showed significant improvement (mean -1.8% [95% CI $-3.8, 0.09$]; $P = 0.02$). This benefit appears to be largely driven by high degrees of QILD at baseline; patients with $>20\%$ QILD showed the largest improvement of any of the subsets (mean -4.9 [95% CI $-8.5, -1.2$]; $P = 0.01$). In terms of fibrosis, there was a significant increase in QLF scores at 48 weeks in the placebo arm (mean 0.7 [95% CI $0.3, 1.2$]; $P = 0.00$) that was not seen in the TCZ arm (mean -0.5 [95% CI $-1.3, 0.3$]; $P = 0.12$). This decline in the placebo arm appears to be driven by worsening of QLF scores in the first and second tertiles.

DISCUSSION

In an earlier phase II trial, TCZ showed preservation of FVC% compared to the placebo group in a population of patients with early dcSSc; fewer patients in the TCZ arm showed a decline in FVC% (10% in the TCZ group versus 23% in the placebo group had $\geq 10\%$ absolute decrease in FVC%) (9). Based on these

preliminary findings, the focuSSced trial was designed showing that, in patients with early dcSSc, the effect of lung function preservation was replicated over 48 weeks (mean decline in the TCZ group -0.6% versus -4.0% in the placebo group; $P = 0.002$) (10). In the present study, we performed a post hoc analysis using individual patient data from the focuSSced trial and demonstrated that $\sim 65\%$ of patients with early dcSSc had HRCT-defined ILD, with 77% of participants having $>10\%$ total lung involvement (as assessed by QILD). The preservation of FVC in the TCZ arm did not vary according to baseline QILD or QLF score, emphasizing the importance of early intervention to retard progression for those with even mild lung involvement. In addition, the placebo arm showed worsening lung fibrosis on HRCT scans at 48 weeks, whereas the TCZ arm showed attenuation of development of progressive fibrosis.

Our population in the focuSSced trial included an at-risk group for progressive ILD: early dcSSc patients with progressive skin disease and elevated acute-phase reactants. This cohort may represent an immunoinflammatory phase, rather than advanced-stage fibrotic ILD studied in previous SSc-ILD trials. Four large prior studies (e.g., the Scleroderma Lung Study I [SLS I] [19] and SLS II [20], the FAST trial [21], and the SENSICIS trial [22]) included patients with both limited cutaneous SSc and dcSSc, with a median disease duration of ≤ 7 years, and included patients who were categorized as having clinical ILD based on respiratory symptoms (grade ≥ 2 exertional dyspnea according to baseline Mahler Dyspnea Index [23] in SLS I and SLS II) and fibrosis ($\geq 10\%$ of the lungs in the SENSICIS trial) (4,24,25). Participants in these trials had moderate-to-severe fibrotic disease: subjects in SLS II had a mean \pm SD QLF score of $8.6 \pm 6.9\%$, and subjects in the SENSICIS trial had a mean \pm SD visual fibrosis score of 36.8 ± 21.8 in the treatment arm and of 35.2 ± 20.7

Table 3. Patient QILD and QLF scores at baseline and at week 48*

	Baseline, mean (95% CI) [n]		Week 48, mean (95% CI) [n]		Change from baseline, mean (95% CI) [n]	
	TCZ group	Placebo group	TCZ group	Placebo group	TCZ group	Placebo group
Whole-lung QILD						
Overall (n = 103)	21.1 (17.6, 24.5) [55]	16.0 (13.8, 18.1) [48]	19.2 (16.2, 22.3) [55]	17.4 (14.9, 19.9) [48]	-1.8 (-3.8, 0.09) [55]†	1.5 (-0.3, 3.3) [48]
≤5% (n = 4)	3.9 (1.5, 6.3) [3]	4.3 (NA) [1]	3.6 (-1.2, 8.4) [3]	5.9 (NA) [1]	-0.3 (-3.8, 3.2) [3]	1.6 (NA) [1]
>5-10% (n = 19)	7.2 (6.0, 8.4) [9]	8.0 (7.0, 9.0) [10]	7.8 (5.0, 10.6) [9]	11.6 (6.6, 16.7) [10]	0.6 (-1.6, 2.7) [9]	3.6 (-0.9, 8.1) [10]
>10-20% (n = 43)	15.1 (13.8, 16.4) [19]	14.4 (13.2, 15.6) [24]	15.7 (12.6, 18.7) [19]	16.8 (14.9, 19.9) [24]	0.6 (-2.3, 3.4) [19]	2.4 (0.06, 4.7) [24]
>20% (n = 37)	33.2 (29.5, 36.8) [24]	25.8 (22.6, 29.0) [13]	28.3 (24.5, 32.1) [24]	24.0 (18.2, 29.7) [13]	-4.9 (-8.5, -1.2) [24]‡	-1.9 (-6.0, 2.3) [13]
Whole-lung QLF						
Overall (n = 104)	3.7 (2.6, 4.9) [55]	2.3 (1.4, 3.2) [49]	3.3 (2.3, 4.2) [55]	3.0 (2.0, 4.1) [49]	-0.5 (-1.3, 0.3) [55]	0.7 (0.3, 1.2) [49]§
First tertile (n = 35)	0.6 (0.4, 0.7) [15]	0.6 (0.5, 0.8) [20]	0.7 (0.3, 1.0) [15]	1.1 (0.7, 1.5) [20]	0.09 (-0.2, 0.4) [15]	0.5 (0.06, 0.8) [20]§
Second tertile (n = 36)	1.7 (1.5, 1.9) [18]	1.7 (1.4, 1.9) [18]	1.7 (1.1, 2.3) [18]	3.1 (2.0, 4.1) [18]	0.01 (-0.7, 0.7) [18]	1.4 (0.5, 2.3) [18]§
Third tertile (n = 33)	7.6 (5.4, 9.7) [22]	6.3 (3.2, 9.3) [22]	6.3 (4.6, 8.0) [22]	6.4 (2.4, 10.4) [11]	-1.3 (-3.3, 0.7) [22]	0.1 (-1.3, 1.6) [11]

* Negative score denotes improvement. QILD data are missing for 33 patients: 19 patients dropped out between weeks 0 and 48 (7 in the TCZ arm and 12 in the placebo arm), and 14 were active through week 48 but had missing data at week 48 (6 in the TCZ arm and 8 in the placebo arm). QLF data are missing for 32 patients: 19 dropped out between weeks 0 and 48 (7 in the TCZ arm and 12 in the placebo arm), and 13 were active through week 48 but had missing data at week 48 (6 in the TCZ arm and 7 in the placebo arm). NA = not applicable (i.e., the 95% confidence interval [95% CI] could not be calculated, as n = 1) (see Table 1 for other definitions).

† P = 0.02 versus baseline, by Wilcoxon's signed rank test.

‡ P = 0.01 versus baseline, by Wilcoxon's signed rank test.

§ P = 0.00 versus baseline, by Wilcoxon's signed rank test.

in the placebo arm (24,26). With the exception of the FAST trial (FVC% 80.1% and 81.0% in the treatment and placebo arms, respectively), participants in these studies demonstrated FVC% impairment: 68.1% in SLS I, 66.5% in SLS II, and 72% in SENCIS (19,20,22).

Placebo-controlled trials and observational cohort studies inform our understanding of the natural progression of SSc-ILD; these data play an important role in illuminating the pathogenesis of SSc-ILD progression in our group with additional clinical ILD patients (26–30). The resulting mean \pm SD rate of decline of FVC in the focuSSced placebo group was 228.2 ± 394.2 ml over 48 weeks, or an FVC% of $\sim 6.5\%$, which was considerably higher than those previously reported. For instance, the FAST trial demonstrated a mean decline of 3.0% (21), which was similar to that of the SLS I trial (2.6%) (19), and the SENCIS cohort showed a decline of 2.6%, or mean \pm SD 93.3 ± 13.5 ml, over 52 weeks (22). As such, our current analysis may influence trial design by providing a template to target early ILD, in which the participants have no or minimal respiratory symptoms, and include more patients with progressive fibrotic ILD, where treatment impact may be easier to detect (31).

Considerable variability in screening for SSc-ILD with HRCT still exists (32). There is increasing consensus that all patients with SSc should receive screening with HRCT (33). Our data demonstrate the value of obtaining HRCT scans at the time of diagnosis: pulmonary function tests (PFTs) are not sensitive enough to accurately assess the presence of ILD, and delays in treatment initiation may lead to irreversible disease (25,34). Recently, the Fleischner Society published a consensus statement on interstitial lung abnormalities (35). They acknowledged that abnormalities identified during screening for ILD in high-risk groups (e.g., those with SSc) are not considered to be interstitial lung abnormalities because they are not incidental (35). Data analysis shows that QILD involvement of $>5\%$ (with a majority of patients having involvement in their lower body areas) was associated with a large decline in FVC% in the placebo group over 48 weeks, which was mirrored in those with $>10\%$ QILD in the placebo group, highlighting the need for universal screening with HRCT in early dcSSc.

A unified treatment algorithm does not yet exist for SSc-ILD. Recent published work has established evidence-based consensus statements on medical management of SSc-ILD; however, these do not address the varying subsets of SSc-ILD severity that impact clinical treatment decisions in practice (4,24,25). Our treatment algorithm classified patients as having either subclinical ILD (those with minimal ILD and preserved lung function) or clinical ILD (those with moderate-to-severe ILD and/or decline in PFTs). Based on the current data, we propose to treat those with subclinical ILD with at-risk features (4,24,25). As evidence accumulates for treatment effects in subsets of SSc-ILD, practice guidelines may favor targeted immunomodulatory therapies in early disease versus antifibrotic therapy in later disease.

Strengths of our analysis include well-characterized data from a clinical trial and utilization of a well-established quantitative lung disease program to provide finer granularity for understanding the lung preservation effect of TCZ. This study serves as an example of the use of quantitative HRCT measurements in understanding SSc-ILD pathophysiology and its response to treatment (14,36).

This analysis is not without limitations. First, the analysis is post hoc and should be considered as hypothesis-generating. Second, while the reduction in FVC reflects fewer functional alveolar units (37), it is an indirect measurement of the flow-resistive properties of the lung (38), and other factors in early SSc may confound the results (e.g., hide-bound chest thickness can cause thoracic restriction, poor patient effort, an inability to form a tight seal around the mouthpiece). This was addressed by standardizing spirometry in the clinical trial. Finally, the minimal ($\leq 5\%$) QILD group had too few patients to establish any meaningful assumptions. Nevertheless, as the field of quantitative radiomics advances its ability to reliably identify interstitial disease changes this small, even this low percentage of lung involvement may prove to have clinical implications.

In conclusion, early dcSSc is associated with high prevalence of ILD, with 77% having moderate-to-severe ILD. TCZ was effective in preserving the lung function, irrespective of the degree of QILD and QLF at baseline. This likely represents targeting of the immunoinflammatory, early fibrotic phase of the disease (39) and may be a window of therapeutic opportunity to preserve lung function in early dcSSc. We also highlight the natural history of early ILD that may serve as a template for other fibrotic diseases.

ACKNOWLEDGMENTS

We thank the sites and the patients who participated in the trial. We also thank the following focuSSced investigators from around the world: Eleonora Lucero, Bernardo Pons-Estel, Mariano Rivero, and Guillermo Tate (Argentina); Vanessa Smith, Ellen De Langhe (Belgium); Rasho Rashkov, Anastas Batalov, Ivan Goranov, and Rumen Stoilov (Bulgaria); James Dunne, Sindhu R. Johnson, and Janet E. Pope (Canada); Dušanka Martinović Kaliterna (Croatia); Mette Mogensen and Anne Braae Olesen (Denmark); Yannick Allanore (France); Joerg Christoph Henes, Ulf Müller-Ladner, Gabriela Riemekasten, and Alla Skapenko (Germany); Panayiotis Vlachoyiannopoulos (Greece); Emese Kiss and Tünde Minier (Hungary); Lorenzo Beretta, Elisa Gremese, Marco Matucci-Cerinic, and Gabriele Valentini (Italy); Yoshihide Asano, Tatsuya Atsumi, Hironobu Ihn, Tomonori Ishii, Osamu Ishikawa, Masataka Kuwana, Yoshihito Shima, Hiroki Takahashi, Kazuhiko Takehara, Yoshiya Tanaka, and Yoshioki Yamasaki (Japan); Loreta Bukauskiene and Irena Butrimiene (Lithuania); Gabriel Medrano Ramirez, Cesar Ramos-Remus, and Tatiana Sofia Rodriguez Reyna (Mexico); Jeska de Vries-Bouwstra and Jacob M. van Laar (The Netherlands); Bogdan Batko, Slawomir Jeka, Eugeniusz Kucharz, Maria Majdan, Marzena Olesinska, and Zaneta Smolenska (Poland); Jose Alves and Maria Santos (Portugal); Carmen Marina Mihai and Simona Rednic (Romania); Ivan Castellvi Barranco, Francisco Javier Lopez Longo, Carmen Simeon Aznar, and Patricia Carreira (Spain); Oliver Distler and Ulrich A. Walker (Switzerland); Emma Derrett-Smith, Bridget Griffiths, and Neil McKay (UK); Jacob Aelion, Michael Borofsky, Roy Fleischmann, Joseph Z. Forstot, Suzanne Kafaja, M. Faisal Khan, Michael D. Kohen,

Richard W. Martin, Fabian Mendoza-Ballesteros, Alireza Nami, Shirley Pang, Grissel Rios, Robert Simms, Keith Michael Sullivan, and Virginia D. Steen (US).

AUTHOR CONTRIBUTIONS

All authors were involved in drafting the article or revising it critically for important intellectual content, and all authors approved the final version to be published. Dr. Khanna had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study conception and design. Roofeh, Lin, Goldin, Kim, Furst, Denton, Huang, Khanna.

Acquisition of data. Roofeh, Lin, Goldin, Kim, Furst, Denton, Khanna.

Analysis and interpretation of data. Roofeh, Lin, Goldin, Kim, Furst, Denton, Huang, Khanna.

ADDITIONAL DISCLOSURES

Author Lin is an employee of Genentech.

REFERENCES

- Denton CP, Khanna D. Systemic sclerosis [review]. *Lancet* 2017;390:1685–99.
- Allanore Y, Simms R, Distler O, Trojanowska M, Pope J, Denton CP, et al. Systemic sclerosis [review]. *Nat Rev Dis Prim* 2015;1:1–21.
- De Lauretis A, Veeraraghavan S, Renzoni E. Connective tissue disease-associated interstitial lung disease: how does it differ from IPF? How should the clinical approach differ? *Chron Respir Dis* 2011;8:53–82.
- Roofeh D, Distler O, Allanore Y, Denton CP, Khanna D. Treatment of systemic sclerosis-associated interstitial lung disease: lessons from clinical trials. *J Scleroderma Relat Disord* 2020;5:61–71.
- Nihtyanova SI, Schreiber BE, Ong VH, Rosenberg D, Moizadeh P, Coghlan JG, et al. Prediction of pulmonary complications and long-term survival in systemic sclerosis. *Arthritis Rheumatol* 2014;66:1625–35.
- Liu X, Mayes MD, Pedroza C, Draeger HT, Gonzalez EB, Harper BE, et al. Does C-reactive protein predict the long-term progression of interstitial lung disease and survival in patients with early systemic sclerosis? *Arthritis Care Res (Hoboken)* 2013;65:1375–80.
- Assassi S, Leyva AL, Mayes MD, Sharif R, Nair DK, Fischbach M, et al. Predictors of interstitial lung disease in early systemic sclerosis: a prospective longitudinal study of the GENISOS cohort. *Arthritis Res Ther* 2010;12:R166.
- Distler O, Volkman ER, Hoffmann-Vold AM, Maher TM. Current and future perspectives on management of systemic sclerosis-associated interstitial lung disease [review]. *Expert Rev Clin Immunol* 2019;15:1009–17.
- Khanna D, Denton CP, Jahreis A, van Laar JM, Frech TM, Anderson ME, et al. Safety and efficacy of subcutaneous tocilizumab in adults with systemic sclerosis (faSScinate): a phase 2, randomised, controlled trial. *Lancet* 2016;387:2630–40.
- Khanna D, Lin CJ, Furst DE, Goldin J, Kim G, Kuwana M, et al. Tocilizumab in systemic sclerosis: a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Respir Med* 2020;8:963–74.
- Van den Hoogen F, Khanna D, Fransen J, Johnson SR, Baron M, Tyndall A, et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum* 2013;65:2737–47.
- Clements PJ, Lachenbruch PA, Ng SC, Simmons M, Sterz M, Furst DE. Skin score: a semiquantitative measure of cutaneous involvement that improves prediction of prognosis in systemic sclerosis. *Arthritis Rheum* 1990;33:1256–63.
- Graham BL, Steenbruggen I, Miller MR, Barjaktarevic IZ, Cooper BG, Hall GL, et al. Standardization of spirometry 2019 update. An official American Thoracic Society and European Respiratory Society technical statement. *Am J Respir Crit Care Med* 2019;200:e70–88.
- Kim GH, Tashkin DP, Lo P, Brown MS, Volkman ER, Gjertson DW, et al. Using transitional changes on high-resolution computed tomography to monitor the impact of cyclophosphamide or mycophenolate mofetil on systemic sclerosis-related interstitial lung disease. *Arthritis Rheumatol* 2020;72:316–25.
- Goldin JG, Kim GH, Tseng CH, Volkman E, Furst D, Clements P, et al. Longitudinal changes in quantitative interstitial lung disease on computed tomography after immunosuppression in the Scleroderma Lung Study II. *Ann Am Thorac Soc* 2018;15:1286–95.
- Volkman ER, Tashkin DP, Sim M, Li N, Khanna D, Roth MD, et al. Cyclophosphamide for systemic sclerosis-related interstitial lung disease: a comparison of Scleroderma Lung Study I and II. *J Rheumatol* 2019;46:1316–25.
- Kim HJ, Tashkin DP, Clements P, Li G, Brown MS, Elashoff R, et al. A computer-aided diagnosis system for quantitative scoring of extent of lung fibrosis in scleroderma patients. *Clin Exp Rheumatol* 2010;28 Suppl 62:S26–35.
- Goh NS, Desai SR, Veeraraghavan S, Hansell DM, Copley SJ, Maher TM, et al. Interstitial lung disease in systemic sclerosis: a simple staging system. *Am J Respir Crit Care Med* 2008;177:1248–54.
- Tashkin DP, Elashoff R, Clements PJ, Goldin J, Roth MD, Furst DE, et al. Cyclophosphamide versus placebo in scleroderma lung disease. *N Engl J Med* 2006;354:2655–66.
- Tashkin DP, Roth MD, Clements PJ, Furst DE, Khanna D, Kleerup EC, et al. Mycophenolate mofetil versus oral cyclophosphamide in scleroderma-related interstitial lung disease (SLS II): a randomised controlled, double-blind, parallel group trial. *Lancet Respir Med* 2016;4:708–19.
- Hoyles RK, Ellis RW, Wellsbury J, Lees B, Newlands P, Goh NS, et al. A multicenter, prospective, randomized, double-blind, placebo-controlled trial of corticosteroids and intravenous cyclophosphamide followed by oral azathioprine for the treatment of pulmonary fibrosis in scleroderma. *Arthritis Rheum* 2006;54:3962–70.
- Distler O, Highland KB, Gahlemann M, Azuma A, Fischer A, Mayes MD, et al. Nintedanib for systemic sclerosis-associated interstitial lung disease. *N Engl J Med* 2019;380:2518–28.
- Mahler DA, Weinberg DH, Wells CK, Feinstein AR. The measurement of dyspnea: contents, interobserver agreement, and physiologic correlates of two new clinical indexes. *Chest* 1984;85:751–8.
- Roofeh D, Khanna D. Management of systemic sclerosis: the first five years [review]. *Curr Opin Rheumatol* 2020;32:228–37.
- Roofeh D, Jaafar S, Vummidi D, Khanna D. Management of systemic sclerosis-associated interstitial lung disease [review]. *Curr Opin Rheumatol* 2019;31:241–9.
- Khanna D, Tashkin DP, Denton CP, Renzoni EA, Desai SR, Varga J. Aetiology, risk factors, and biomarkers in systemic sclerosis with interstitial lung disease. *Am J Respir Crit Care Med* 2020;201:650–60.
- Khanna D, Nagaraja V, Tseng CH, Abtin F, Suh R, Kim G, et al. Predictors of lung function decline in scleroderma-related interstitial lung disease based on high-resolution computed tomography: implications for cohort enrichment in systemic sclerosis-associated interstitial lung disease trials. *Arthritis Res Ther* 2015;17:1–10.
- Steen VD, Conte C, Owens GR, Medsger TA Jr. Severe restrictive lung disease in systemic sclerosis. *Arthritis Rheum* 1994;37:1283–9.
- Steen V. Predictors of end stage lung disease in systemic sclerosis. *Ann Rheum Dis* 2003;62:97–9.

30. Winstone TA, Assayag D, Wilcox PG, Dunne JV, Hague CJ, Leipsic J, et al. Predictors of mortality and progression in scleroderma-associated interstitial lung disease: a systematic review. *Chest* 2014;146:422–36.
31. Silver RM. Systemic sclerosis: choosing patients wisely when treating interstitial lung disease [review]. *Nat Rev Rheumatol* 2017;13:455–6.
32. Bernstein EJ, Khanna D, Lederer DJ. Screening high-resolution computed tomography of the chest to detect interstitial lung disease in systemic sclerosis: a global survey of rheumatologists. *Arthritis Rheumatol* 2018;70:971–2.
33. Hoffmann-Vold AM, Maher TM, Philpot EE, Ashrafzadeh A, Barake R, Barsotti S, et al. The identification and management of interstitial lung disease in systemic sclerosis: evidence-based European consensus statements. *Lancet Rheumatol* 2020;2:E71–83.
34. Suliman YA, Dobrota R, Huscher D, Nguyen-Kim TD, Maurer B, Jordan S, et al. Pulmonary function tests: high rate of false-negative results in the early detection and screening of scleroderma-related interstitial lung disease. *Arthritis Rheumatol* 2015;67:3256–61.
35. Hatabu H, Hunninghake GM, Richeldi L, Brown KK, Wells AU, Remy-Jardin M, et al. Interstitial lung abnormalities detected incidentally on CT: a Position Paper from the Fleischner Society. *Lancet Respir Med* 2020;8:726–37.
36. Montesi SB, Caravan P. Novel imaging approaches in systemic sclerosis-associated interstitial lung disease [review]. *Curr Rheumatol Rep* 2019;21:25.
37. O'Donnell D. Physiology of interstitial lung disease. In: Schwartz M, King T, editors. *Interstitial lung disease*. Hamilton: Marcel Dekker; 1998. p. 51–70.
38. Grippi MA, Tino G. Pulmonary function testing. In: Grippi MA, Elias JA, Fishman JA, Kotloff RM, Pack AI, Senior RM, Siegel MD, editors. *Fishman's pulmonary diseases and disorders*, 5th ed. New York: McGraw-Hill; 2015. p. 1–75.
39. Denton CP, Ong VH, Xu S, Chen-Harris H, Modrusan Z, Lafyatis R, et al. Therapeutic interleukin-6 blockade reverses transforming growth factor- β pathway activation in dermal fibroblasts: insights from the faSScinate clinical trial in systemic sclerosis. *Ann Rheum Dis* 2018;77:1362–71.