DR. OREN K FIX (Orcid ID : 0000-0002-8570-9646) DR. DAVID C MULLIGAN (Orcid ID : 0000-0003-0901-2617) DR. HUGO R ROSEN (Orcid ID : 0000-0002-1110-6914)



AASLD Expert Panel Consensus Statement: Vaccines to Prevent COVID-19 Infection in Patients with Liver Disease

Oren K. Fix, MD, MSc, FAASLD^{1*}; Emily A. Blumberg, MD²; Kyong-Mi Chang, MD, FAASLD^{2,3}; Jaime Chu, MD⁴; Raymond T. Chung, MD, FAASLD⁵; Elizabeth K. Goacher, PA-C, AF-AASLD⁶; Bilal Hameed, MD⁷; Daniel R. Kaul, MD⁸; Laura M. Kulik, MD⁹; Ryan M. Kwok, MD¹⁰; Brendan M. McGuire, MD¹¹; David C. Mulligan, MD, FAASLD¹²; Jennifer C. Price, MD, PhD⁷; Nancy S. Reau, MD, FAASLD¹³; K. Rajender Reddy, MD, FAASLD²; Andrew Reynolds¹⁴; Hugo R. Rosen, MD, FAASLD¹⁵; Mark W. Russo, MD, MPH, FAASLD¹⁶; Michael L. Schilsky, MD, FAASLD¹²; Elizabeth C. Verna, MD, MS¹⁷; John W. Ward, MD¹⁸; Robert J. Fontana, MD, FAASLD^{8*} for the **AASLD COVID-19 Vaccine Working Group**

* Co-first authors with equal contributions

¹Elson S. Floyd College of Medicine, Washington State University, Spokane, WA; ²University of Pennsylvania, Philadelphia, PA; ³The Corporal Michael J. Crescenz VAMC; ⁴Icahn School of Medicine at Mt Sinai, New York, NY; ⁵Massachusetts General Hospital, Boston, MA; ⁶Duke University, Durham, NC; ⁷University of California, San Francisco, CA; ⁸University of Michigan, Ann Arbor, MI; ⁹Northwestern Medicine, Chicago, IL; ¹⁰Uniformed Services University, Bethesda, MD; ¹¹University of Alabama, Birmingham, AL; ¹²Yale University, New Haven, CT; ¹³Rush University, Chicago, IL; ¹⁴San Francisco AIDS

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the <u>Version of Record</u>. Please cite this article as <u>doi:</u> 10.1002/HEP.31751

Foundation, San Francisco, CA; ¹⁵Keck Medicine of USC, Los Angeles, CA; ¹⁶Atrium Health, Carolinas Medical Center, Charlotte, NC; ¹⁷Columbia University, New York, NY; ¹⁸Coalition for Global Hepatitis Elimination

| OKF: | oren.fix@wsu.edu |
|------|---------------------------------|
| EAB: | eblumber@pennmedicine.upenn.edu |
| KMC: | kmchang@pennmedicine.upenn.edu |
| JC: | jaime.chu@mssm.edu |
| RTC: | chung.raymond@mgh.harvard.edu |
| EKG: | elizabeth.goacher@duke.edu |
| BH: | bilal.hameed@ucsf.edu |
| DRK: | kauld@med.umich.edu |
| LMK: | laura.kulik@nm.org |
| RMK: | ryanmkwok@gmail.com |
| BMG: | bmcguire@uabmc.edu |
| DCM: | david.mulligan@yale.edu |
| JCP: | jennifer.price@ucsf.edu |
| NSR: | nancy_reau@rush.edu |
| KRR: | rajender.reddy@uphs.upenn.edu |
| AR: | areynoldshcv@gmail.com |
| HRR: | hugo.rosen@med.usc.edu |
| MWR: | mark.russo@atriumhealth.org |
| MLS: | michael.schilsky@yale.edu |
| ECV: | ev77@cumc.columbia.edu |
| JWW: | jward@taskforce.org |
| RJF: | rfontana@med.umich.edu |
| | |

Keywords: SARS-CoV-2, coronavirus, transplantation, cirrhosis, immunosuppression

Contact Information

Oren K. Fix, MD, MSc, FAASLD

Department of Medical Education and Clinical Sciences

Elson S. Floyd College of Medicine

Washington State University

412 E Spokane Falls Blvd

Spokane, WA 99202-2131

Tel: (509) 358-7944

oren.fix@wsu.edu

List of Abbreviations

1

| COVID-19 | coronavirus disease 2019 |
|------------|---|
| SARS-CoV-2 | severe acute respiratory syndrome coronavirus 2 |
| CLD | chronic liver disease |
| EUA | emergency use authorization |
| FDA | Food and Drug Administration |
| VE | vaccine efficacy |
| BNT162b2 | Pfizer-BioNTech mRNA vaccine |
| mRNA-1273 | Moderna mRNA vaccine |
| VAERS | Vaccine Adverse Event Reporting System |
| CDC | Centers for Disease Control and Prevention |
| SOT | solid organ transplant |
| ACR | acute cellular rejection |
| | |

Financial Support

| OKF: | None |
|------|--|
| EAB: | Research support: Merck, Takeda, Hologix; DSMB: Amplyx |
| KMC: | None |
| JC: | None |
| RTC: | Research support: Abbvie, BMS, Gilead, Janssen, Merck, Roche, Boehringer |
| EKG: | Consulting: AbbVie, Gilead, Intercept; Speaker: AbbVie, Gilead, Intercept |
| BH: | Consulting: Gilead; Research support: Conatus Pharmaceuticals, Genfit, Intercept |
| | Pharmaceuticals, Pliant Therapeutics, Valeant Pharmaceuticals; Advisory board: |
| | Mallinckrodt; Stock ownership: Intercept Pharmaceuticals, Pleiogenix |
| DRK: | Research support: AstraZeneca, Janssen, Merck, Shire, NIAID |
| LMK: | None |
| RMK: | None |

| BMG: | Research support: Gilead, Arrowhead | |
|------|--|---------------|
| DCM: | None | |
| JCP: | Research support: Gilead, Merck; Consulting: Theratechnologies; Ownershi | p interest |
| | (spouse): BMS, Johnson and Johnson, Abbvie, Merck | |
| NSR: | Consulting: AbbVie, Gilead, Salix, Intercept, Arbutus; Research support (pai | d to |
| | institution): AbbVie, Gilead | |
| KRR: | Advisory board: Mallinckrodt; Grant support (paid to the University of Pen | nsylvania): |
| | Mallinckrodt, Intercept, BMS, Gilead, Merck, Exact Sciences, Sequana, NAS | H-TARGET, |
| | HCC-TARGET; DSMB: Novartis | |
| AR: | None | |
| HRR: | None | |
| MWR: | None | |
| MLS: | Research support: Vivet, Alexion, Orphalan (GMPO), Wilson Disease Associ | ation |
| ECV: | Research support: Salix; Advisory board: Gilead | |
| JWW: | The Task Force for Global health received funds for the general support of | the Coalition |
| | for Global Hepatitis Elimination from Abbott, AbbVie, Cepheid, Gilead Scier | nces, Merck, |
| | Pharco, Roche Siemens, Zydus Cadila, philanthropic organizations, professi | onal |
| | associations, and US Government | |
| RJF: | Research support: Abbvie, BMS, and Gilead; Consulting: Sanofi | |

Author Contributions

| OKF: | Conceptualization, Project Administration, Visualization, Writing – original draft, Writing |
|------|---|
| | – review & editing |
| EAB: | Writing – original draft, Writing – review & editing |
| KMC: | Writing – original draft, Writing – review & editing |
| JC: | Writing – original draft, Writing – review & editing |
| RTC: | Conceptualization, Supervision, Writing – review & editing |
| EKG: | Writing – original draft, Writing – review & editing |
| BH: | Writing – original draft, Writing – review & editing |
| DRK: | Writing – original draft, Writing – review & editing |
| LMK: | Writing – original draft, Writing – review & editing |
| RMK: | Writing – original draft, Writing – review & editing |
| BMG: | Visualization, Writing – original draft, Writing – review & editing |

| DCM: | Writing – original draft, Writing – review & editing |
|------|---|
| JCP: | Writing – original draft, Writing – review & editing |
| NSR: | Writing – original draft, Writing – review & editing |
| KRR: | Conceptualization, Supervision, Writing – original draft, Writing – review & editing |
| AR: | Writing – original draft, Writing – review & editing |
| HRR: | Writing – original draft, Writing – review & editing |
| MWR: | Writing – original draft, Writing – review & editing |
| MLS: | Writing – original draft, Writing – review & editing |
| ECV: | Writing – original draft, Writing – review & editing |
| JWW: | Writing – review & editing |
| RJF: | Conceptualization, Project Administration, Visualization, Writing – original draft, Writing |
| | - review & editing |

Acknowledgements

The authors would like to acknowledge the AASLD staff and the AASLD Governing Board for their contributions to and support of this paper: Raymond T. Chung, MD, FAASLD, Massachusetts General Hospital; Laurie D. DeLeve, MD, PhD, FAASLD, University of Southern California; Jorge A. Bezerra, MD, FAASLD, Cincinnati Children's Hospital; Meena G. Bansal, MD, FAASLD, Icahn School of Medicine at Mount Sinai; Vijay H. Shah, MD, FAASLD, Mayo Clinic Rochester; Norah Terrault, MD, MPH, FAASLD, Keck Medical Center of University of Southern California; W. Ray Kim, MD, FAASLD, Stanford University Medical Center; Grace L. Su, MD, FAASLD, University of Michigan; David C. Mulligan, MD, FAASLD, Yale University; Mary E. McCarthy Rinella, MD, FAASLD, Northwestern University Feinberg School of Medicine; Paul Martin, MD, FAASLD, University of Miami School of Medicine; Katie Duggan; John D. Lingerfelt

Abstract

Background and rationale: The aim of this document is to provide a concise scientific review of the currently available COVID-19 vaccines and those in development, including mRNA, adenoviral vectors, and recombinant protein approaches. The anticipated use of COVID-19 vaccines in patients with chronic liver disease and liver transplant recipients is reviewed and practical guidance is provided for health care providers involved in the care of patients with liver disease and liver transplantation. **Main results:** The Pfizer and Moderna mRNA COVID-19 vaccines are associated with a 94%-95% vaccine efficacy compared to placebo against COVID-19. Local site reactions

of pain and tenderness were reported in 70%-90% of clinical trial participants, and systemic reactions of fever and fatigue were reported in 40%-70% of participants, but these reactions were generally mild and self-limited and occurred more frequently in younger individuals. Severe hypersensitivity reactions related to the mRNA COVID-19 vaccines are rare and more commonly seen in women and individuals with a history of prior drug reactions for unclear reasons. Because patients with advanced liver disease and immunosuppressed patients were excluded from the vaccine licensing trials, additional data regarding the safety and efficacy of COVID-19 vaccines are eagerly awaited in these and other subgroups. **Conclusions:** Remarkably safe and highly effective mRNA COVID-19 vaccines are now available for widespread use and should be given to all adult patients with chronic liver disease and liver transplant recipients. The online companion document located at <u>https://www.aasld.org/about-aasld/covid-19-resources</u> will be updated as additional data become available regarding the safety and efficacy of development.

An earlier version of this document was first posted online on February 2, 2021 at <u>https://www.aasld.org/about-aasld/covid-19-resources</u> and will continue to be updated.

OVERVIEW & RATIONALE

Coronavirus disease 2019 (COVID-19) is the illness caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Multiple studies demonstrate that older individuals and those with certain comorbidities including chronic liver disease (CLD) (particularly cirrhosis), cardiac disease, obesity, and weakened immune systems from other diseases or medications may be at higher risk of death from COVID-19. Over the past year, over 60 vaccine candidates have been identified and are under development as a means to prevent COVID-19. As of the beginning of February 2021, two mRNA-based vaccines have received Emergency Use Authorization (EUA) from the US Food and Drug Administration (FDA). An EUA is a legal means for the FDA to provide preliminary authorization of new drugs, vaccines, or devices during a declared national emergency until a full review of the complete safety and efficacy data has been completed. Since persons with CLD and immunosuppressed transplant recipients are frequently hyporesponsive to licensed vaccines, additional studies regarding the safety and efficacy of COVID-19 vaccines are urgently needed in these patient subgroups.

The goal of this document is to provide concise safety and efficacy data regarding the commercially available COVID-19 vaccines and their use in CLD patients and liver transplant recipients. Our intent is to provide clinically useful information for all health care providers involved in the care of patients with liver disease, including hepatologists and liver transplant care providers, and their patients.

BACKGROUND ON NON-COVID-19 VACCINES IN PATIENTS WITH CHRONIC LIVER DISEASE AND IMMUNOSUPPRESSION

Patients with CLD display innate and adaptive immune dysregulation that is associated with vaccine hyporesponsiveness.(1) (**Supplemental Figure 1**) Since subjects with CLD have an increased risk of complications after infection with influenza, S. pneumoniae, HAV, and HBV,(2,3) vaccination against these pathogens is recommended. (**Supplemental Table 1**) Double dosing or booster dosing of the HAV and HBV vaccines can increase vaccine response rates in CLD patients.(4,5) Immunosuppressed liver transplant recipients are also known to have a lower response rate to many non-COVID-19 vaccines particularly when given early after transplant. Therefore, it is generally recommended that vaccines be given prior to transplant whenever possible or waiting until 3 to 6 months after transplant. The Advisory Committee on Immunization Practices (ACIP) recommends avoiding live virus vaccines in those receiving high-dose corticosteroids and other immunosuppressed individuals because of concerns of uncontrolled viral replication.(6) See **Supplemental Text** for additional information regarding non-COVID-19 vaccines.

GUIDANCE FOR DOUBLE DOSING OF COVID-19 VACCINES

• While double dosing is recommended for some non-COVID-19 vaccines in CLD patients, this approach is NOT recommended with COVID-19 vaccines.

BACKGROUND ON COVID-19 VACCINES

Types of COVID-19 Vaccines in Development

Entry of SARS-CoV-2 requires binding of the viral spike glycoprotein to the angiotensin-converting enzyme 2 (ACE2) receptor on human epithelial cells.(7) As a result, researchers have targeted the viral spike glycoprotein to induce vaccine-mediated immune response against SARS-CoV-2 using various delivery systems. The release of the 29,903-nucleotide sequence of the SARS-CoV-2 genome on January 10, 2020 led to diagnostic testing and the development of Operation Warp Speed in the US with the goal of developing safe and effective vaccines within one year.(8,9) Both Moderna and Pfizer-BioNTech developed a vaccine using synthetic nucleoside-modified mRNA that encodes the spike glycoprotein.(10,11) Other vaccines that are currently in development use DNA, protein subunits, inactivated SARS-CoV-2, viral vectors, and attenuated virus. (**Table 1, Figure 1**) All of the vaccines described below are not live SARS-CoV-2 and cannot replicate, even in immunocompromised persons.

mRNA Vaccines

mRNA-based vaccines involve the delivery of non-infectious synthetic mRNA encoding one or more target antigens (e.g., SARS CoV-2 spike protein) that can be taken up by host cells including antigen presenting cells (e.g., dendritic cells). (**Figure 1**) Upon cytoplasmic entry, the delivered mRNA uses the host ribosomal translational machinery to make the target antigens that can be processed for cell surface expression via class I and II major histocompatibility complex or be secreted. This induces protective immunity against a future attack (e.g., from SARS-CoV-2) by priming antigen-specific cytotoxic CD8 T-cells and helper T-cells and a neutralizing antibody response from B-cells.(12) A key challenge to the mRNA vaccine platform is its stability and efficiency, which is related to its susceptibility to enzymatic degradation, limited cellular uptake, and capacity for innate immune activation that can inhibit mRNA translation. In recent years, these challenges have been addressed by using lipid nanoparticles that protect the mRNA from enzymatic degradation and enhance their cellular uptake and degradation. Nevertheless, the mRNA-based vaccines degrade within a few hours at room temperature and require very cold temperatures during manufacturing, transportation, and storage.

Adenoviral Vectors

Adenovirus-based vaccines use a harmless, genetically modified exogenous virus as the carrier to bring DNA that encodes the SARS-CoV-2 spike protein into the recipient's cells. Once the adenovirus enters a cell, it delivers the DNA for the SARS-CoV-2 spike protein into the nucleus and the corresponding mRNA is transcribed. Using the host cellular machinery, the mRNA is then translated into SARS-CoV-2 spike protein, which triggers the host immune response after being expressed on cell surface membranes or secreted into the serum.

There are hundreds of known adenoviruses and most do not cause disease in humans, while others cause a range of symptoms depending on the tropism of the strains. The adenovirus vector is modified to prevent it from replicating in host cells. Adenovirus-vector vaccines are stable at room temperature for prolonged periods. Prior studies have shown that replication defective chimp adenoviral vector vaccines can effectively deliver viral genes to the liver, induce a host immune response, and are safe to use in both healthy volunteers and patients with CLD.(15,16)

Early safety data were favorable in the Phase 3 clinical trial of Oxford/AstraZeneca's AZD1222 adenovirus vectored COVID-19 vaccine,(17) leading to its authorization for emergency use in the United Kingdom (UK) on December 29, 2020.(18) Phase 3 clinical trials from Johnson & Johnson (Ad26) demonstrate vaccine efficacy (VE) rates of 57%-72% following a single dose.(19) Early clinical trials with

CanSino's non-replicating adenovirus (Ad5) vectored COVID-19 vaccine showed mild to moderate increases in total bilirubin (8% of recipients) and serum alanine aminotransferase levels (9% of recipients).(20) While these observations were not considered clinically significant, more data and experience with this and other replication defective adenovirus-based vaccines are needed before their widespread use can be recommended in patients with CLD. Replication defective adenovirus-based vaccines are not live or attenuated SARS-CoV-2 and are not expected to pose a risk to immunocompromised patients. Further guidance from regulatory and public health authorities is eagerly awaited following their authorization, and further information regarding any elevations in liver function tests (although not in patients with advanced liver disease) will be available from ongoing or recently completed large phase 3 trials.

Other Vaccines

Inactivated whole virus vaccines are made by treating the virus with heat and/or chemicals (usually formalin) to prevent its capacity to replicate but are typically given in multiple doses to induce a more robust host immune response. This technology has been used for vaccines against rabies, polio, and HAV. The inactivated whole virus Sinovac vaccine against SARS-CoV-2 has completed small phase 3 trials with mixed efficacy.

Protein subunit vaccines (e.g., Herpes Zoster vaccine) isolate immunogenic portions of the pathogen of interest that are often combined with an adjuvant (e.g., Alum, MF59, AS01, AS03, AS04).(21) The Novavax product is a recombinant spike protein subunit and adjuvant vaccine using nanoparticle technology that has started phase 3 trials in the US. Novavax recently reported a VE of 89% in the UK and a VE of 49.4% in South Africa, where the majority of COVID-19 cases are caused by an escape variant (B.1.351).(22) A Sanofi/GlaxoSmithKline protein subunit vaccine failed to generate adequate immune response in older adults and is being reengineered.

Live-attenuated viral vaccines are also being developed for COVID-19. One of these uses a virus of limited pathogenicity in humans, vesicular stomatitis virus (VSV), where VSV genes are replaced with the SARS-CoV-2 spike protein gene to generate a host anti-spike glycoprotein response. An effective Ebola virus vaccine using VSV in this manner has been approved in the US. While there are no live-attenuated virus COVID-19 vaccines nearing FDA authorization, in general, live-attenuated vaccines are not recommended for use in immunocompromised patients due to concerns of excessive viral replication.

See **Supplemental Text** for background on assays to detect immunity to COVID-19.

Safety and Efficacy of FDA EUA mRNA COVID-19 Vaccines

In December 2020, the FDA granted EUA to two mRNA vaccines to prevent COVID-19: BNT162b2 manufactured by Pfizer-BioNTech and mRNA-1273 by Moderna, respectively. (**Table 1**) Both vaccines are based on the SARS-CoV-2 spike glycoprotein antigen encoded by mRNA in lipid nanoparticles. The spike glycoprotein antigen mediates binding of the virus to the ACE2 receptor on host cells to enable viral entry and replication. In both vaccines, the mRNA encodes the spike glycoprotein antigen stabilized in its pre-fusion form, which more closely resembles the intact virus.(23)

VE for the Pfizer-BioNTech primary endpoint (confirmed COVID-19 occurring at least 7 days after the second dose in participants without serological or virological evidence of past SARS-CoV-2 infection) was 95.0%, while VE for the Moderna primary endpoint (COVID-19 occurring at least 14 days after the second dose in participants who were negative for SARS-CoV-2 at baseline) was 94.1%. (Figure 2) In both vaccines, reactogenicity and adverse events (AEs) were generally milder and less frequent in older than in younger participants and more frequent and more severe after the second dose. (Figure 3) See **Supplemental Text** for additional information regarding these currently authorized vaccines. At the time of writing, Johnson and Johnson/Janssen have submitted an application to the FDA for EUA of their single-shot COVID-19 vaccine candidate.(24)

Post-Marketing Reports of Anaphylactic Reactions to mRNA COVID-19 Vaccines

During December 14-23, 2020, monitoring by the Vaccine Adverse Event Reporting System (VAERS) detected 21 cases of anaphylaxis after administration of a reported 1,893,360 first doses of the Pfizer-BioNTech mRNA COVID-19 vaccine (11.1 cases per million doses).(25) 71% of these occurred within 15 minutes of vaccination, 17 (81%) had a documented history of allergies or allergic reactions, and 90% were female. Nineteen (90%) were treated with epinephrine, 4 (19%) were hospitalized (including three in intensive care), and 17 (81%) were treated in an emergency department. No deaths from anaphylaxis were reported after receiving the Pfizer-BioNTech COVID-19 vaccine.

Similarly, during December 21, 2020-January 10, 2021, monitoring by VAERS detected 10 cases of anaphylaxis after administration of a reported 4,041,396 first doses of the Moderna mRNA COVID-19 vaccine (2.5 cases per million doses).(26) 90% of these occurred within 15 minutes of vaccination, 9 (90%) had a documented history of allergies or allergic reactions, and 100% were female. All patients were treated with epinephrine, 6 (60%) were hospitalized (including five in intensive care), and 17 (81%) were treated in an emergency department. No deaths from anaphylaxis were reported after receiving the Moderna COVID-19 vaccine. It is unclear why the vast majority of cases of anaphylaxis occurred in women; however, more women than men received the first doses of the mRNA COVID-19 vaccines during the analytic period.(25,26)

The incidence of anaphylaxis associated with the Pfizer-BioNTech vaccine may be 10 times as high (1 in 100,000) as the incidence reported with previously approved vaccines (1 in 1,000,000).(27) Polyethylene glycol (PEG), used to stabilize the lipid nanoparticles and prolong their half-life in both mRNA vaccines, has been implicated as a potential cause for anaphylaxis.

GUIDANCE FOR ALLERGIC REACTIONS TO mRNA COVID-19 VACCINES

- Anyone with a history of severe or immediate allergic reaction to any vaccine components, including PEG, should NOT receive either mRNA COVID-19 vaccine.
- Anyone with a severe or immediate allergic reaction to the first dose of an mRNA COVID-19 vaccine should NOT receive additional doses of either mRNA COVID-19 vaccine.

SARS-CoV-2 Viral Variants

Rapidly spreading variants of SARS-CoV-2 have been described from the UK (B.1.1.7)(28) and South Africa (501Y.V2 or B.1.351)(29) that share the spike N501Y substitution located in the viral spike protein receptor binding domain for cell entry. Another variant from Brazil (P.1) also contains mutations in the receptor binding domain of the spike protein.(30) All of these variants have been found in the US,(31) are more transmissible,(28) and may be associated with higher morbidity and mortality.(31,32) Nonpeer reviewed studies suggest that both Pfizer-BioNTech and Moderna mRNA COVID-19 vaccines may provide protection against the UK B.1.1.7 and South African B.1.351 variants.(33–35)

GUIDANCE FOR VACCINATION IN SETTING OF SARS-COV-2 VIRAL VARIANTS

 Withholding COVID-19 vaccination due to concerns about current or future SARS-CoV-2 viral variants is NOT recommended

Pediatric Considerations in COVID-19 Vaccination

Only the Pfizer-BioNTech mRNA vaccine has been authorized for children <18 years of age (specifically ≥16 years). However, there are multiple vaccine trials underway for children ≥12 years. While a small subset of children have had severe COVID-19 symptoms and/or developed complications such as multisystem inflammatory syndrome in children (MIS-C),(36) the vast majority of children with

COVID-19 have had mild illness. Data from a North American pediatric registry suggest that children with liver disease and those post-liver transplant have outcomes similar to the general pediatric population.(37) The differences in COVID-19 presentations and disease course from adults underscores the importance of continued pediatric clinical trials to establish vaccine efficacy, dosing, and safety in children. Co-administration of different vaccines is usually safe; however, administration of the COVID-19 vaccine with other childhood immunizations has not yet been tested.

GUIDANCE FOR ADMINSTRATION INTERVAL OF COVID-19 AND OTHER VACCINES

• The Centers for Disease Control and Prevention (CDC) recommends that COVID-19 vaccines be administered alone with a minimum interval of 14 days before or after administration of other vaccines.

COVID-19 VACCINES IN PATIENTS WITH CHRONIC LIVER DISEASE AND IMMUNOSUPPRESSION Patients with Liver Disease in COVID-19 Vaccine Clinical Trials

Patients with stable chronic medical conditions such as compensated CLD, HIV, HBV, or HCV were eligible to participate in both the Pfizer-BioNTech and Moderna Phase 3 trials. However, those on immunosuppressive therapy were excluded.

In the Pfizer-BioNTech Phase 2/3 trial, 20.5% of study participants had a comorbidity defined by the Charlson Comorbidity Index categories, which include liver disease (8030 with a comorbidity received BNT162b2 and 8029 received placebo). VE was 95.3% in participants with comorbidities and was similar to that seen in patients without comorbidities (94.7%). Among the 214 participants (0.6%) with liver disease, 124 received BNT162b2 and 90 received placebo, but safety and efficacy data in this subgroup have not yet been reported.

In the Moderna Phase 3 trial, at least one high risk condition was present in 22.3% of the participants. Among the 196 (0.6%) participants with liver disease, 100 received mRNA-1273 and 96 received placebo. Since no participants with liver disease developed COVID-19, VE cannot be determined for this subgroup.

Prioritization During Limited Supply of COVID-19 Vaccines

The COVID-19 vaccines are currently a limited resource that requires rational selection of the highest risk candidates for priority access. Providers must administer COVID-19 vaccines in accordance with prioritization groups determined by appropriate public health authorities.(38) The CDC has

published a dynamic document that ranks groups at high risk for exposure or poor outcome from COVID-19 (Phases 1a, 1b, 1c, 2).(39) Healthcare workers are prioritized by the CDC (Phase 1a) to receive the COVID-19 vaccines because of their high risk of exposure to SARS-CoV-2, the need to protect patients from infection, and the need to preserve the capacity to care for patients.(40) Patients with underlying medical conditions, including liver disease (e.g., compensated and decompensated cirrhosis, liver cancer), solid organ transplantation (SOT), and immunosuppression, are at risk for severe COVID-19 and are included in Phase 1c.(41–45)

Because of the scarcity of COVID-19 vaccines and the observation that SARS-CoV-2 reinfection is uncommon within 90 days of first infection, the CDC recognizes that individuals with recent SARS-CoV-2 infection may want to defer vaccination for up to 90 days. In addition, early work suggests that COVID-19 vaccine-related side effects may be more common in those with previous SARS-CoV-2 infection, particularly when vaccinated soon after infection.(46)

PRINCIPLES REGARDING PRIORITIZATION OF PATIENTS FOR COVID-19 VACCINATION

- All healthcare workers should be prioritized for the COVID-19 vaccine (Phase 1a).
- Patients with comorbidities identified as high risk by the CDC, including CLD, should be prioritized for vaccination (Phase 1c).(47)
- For liver transplant candidates, vaccination against COVID-19 should proceed even if liver transplant is likely to occur before the second dose can be administered. The second dose of vaccine should be given at the earliest appropriate interval after transplant (e.g., 6 weeks posttransplant).
- Data are insufficient to determine the risk of severe COVID-19 in patients with immunemediated liver disease on chronic immunosuppression and posttransplant patients relative to patients with cirrhosis; therefore, they should be prioritized for vaccination (Phase 1c).(48–50)
- Data are lacking to determine if a prior diagnosis of COVID-19 or the presence of antibodies to SARS-CoV-2 should be used to determine the need for vaccination; therefore, in the absence of contraindications (hypersensitivity to any vaccine components), all patients with CLD and SOT recipients should be encouraged to get vaccinated.
- Healthcare providers should be knowledgeable of the local criteria for vaccination, know where vaccine is available, and actively inform patients of this information.
- Vaccinated healthcare providers are encouraged to volunteer to assist with their local vaccination efforts.

• See Supplemental Text for additional guidance for COVID-19 vaccination.

COVID-19 Vaccination in Patients with Chronic Liver Disease

Due to the increased mortality with COVID-19 infection in adult CLD patients and particularly those with cirrhosis, it is recommended that these patients be prioritized for COVID-19 vaccination (Phase 1c). Although safety and efficacy data with the two available mRNA vaccines in CLD patients are limited, adverse events are not anticipated to be more frequent nor is efficacy expected to be lower than the general population; however, additional prospective studies are needed.

If the supply of COVID-19 vaccine is limited, it is reasonable to prioritize patients with higher Model for End-stage Liver Disease (MELD) or Child-Turcotte-Pugh scores for vaccination or those who are anticipated to undergo imminent liver transplantation, but all CLD patients should be vaccinated whenever possible.

GUIDANCE FOR COVID-19 VACCINATION IN PATIENTS WITH CLD

- Patients with CLD who are receiving antiviral therapy for HBV or HCV or medical therapy for primary biliary cholangitis or autoimmune hepatitis should NOT withhold their medications while receiving the COVID-19 vaccines.
- Patients with hepatocellular carcinoma undergoing locoregional or systemic therapy should also be considered for vaccination without interruption of their treatment. However, patients with recent infections or fever should NOT receive the COVID-19 vaccine until they are medically stable.
- mRNA COVID-19 vaccines are expected to have a favorable efficacy and safety profile in immunosuppressed patients and should be administered according to their standard dose and schedule.
- Liver transplant candidates with CLD should receive the mRNA COVID-19 vaccine prior to transplantation whenever possible to help ensure an adequate immune response.
- CLD patients receiving the mRNA COVID-19 vaccine may have local and systemic reactions (fever, myalgias, headache) in the first 48 hours after vaccination. However, respiratory symptoms or systemic symptoms may be indicative of COVID-19 and warrant further investigation.
- All patients with CLD, including vaccine recipients, should continue to mitigate their risk of SARS-CoV-2 exposure, e.g., masking, social distancing, hand washing, etc.

COVID-19 Vaccination in Immunosuppressed Liver Transplant Recipients

As immunocompromised patients and SOT recipients were not included in the clinical trials of vaccines against SARS-CoV-2, there is a lack of data regarding the safety and efficacy of the available vaccines in this population. Other unknowns regarding vaccination of liver transplant recipients include:

- 1. Efficacy of the immune response to the vaccine to prevent SARS-CoV-2 infection in SOT recipients.
- 2. Whether the duration of vaccine-conferred immunity differs from immunocompetent hosts.
- 3. Whether intensified immunosuppression in the immediate posttransplant period and following treatment of acute cellular rejection (ACR) reduces VE.
- 4. The best timing and safety of vaccine administration for patients who had COVID-19.
- 5. The frequency of elevation of liver tests or ACR following vaccination.
- 6. The best choice of vaccine in this population.

Despite these uncertainties, the available mRNA COVID-19 vaccines do not contain live or attenuated virus and therefore are unlikely to pose a safety concern for immunosuppressed patients.(51) Since replication defective or non-replicating vaccines have also not yet been tested in SOT recipients or other immunosuppressed patients, additional data are needed before use of these COVID-19 vaccines can be recommended in these patients.

GUIDANCE FOR COVID-19 VACCINATION IN LIVER TRANSPLANT RECIPIENTS

- COVID-19 vaccination is recommended for all SOT recipients including liver transplant recipients.
- The best time to administer the COVID-19 vaccine in liver transplant recipients is likely at least 3 months post liver transplantation when immunosuppression is lower and other prophylactic medications are stopped or minimized. However, given the ongoing community spread of SARS-CoV-2, immunization may be given as early as 6 weeks posttransplant, especially for the highest risk individuals with other comorbid factors associated with severe COVID-19.
- A reduction in immunosuppression is NOT RECOMMENDED in liver transplant recipients solely to elicit an immune response to immunization against SARS-CoV-2 as there is a risk of acute cellular rejection (ACR) with lower immunosuppression.

- Avoid COVID-19 vaccination in liver transplant recipients with active ACR, those being treated for ACR, or those on high daily doses of corticosteroids until the episode is resolved and their baseline immunosuppression re-established.
- In patients whose liver tests increase after vaccination and do not immediately return to baseline on repeat testing, a thorough evaluation should follow to exclude ACR or viral infection of the liver.
- Given the life-saving nature of liver transplantation, deceased donor transplantation should NOT be delayed in a patient who received a COVID-19 vaccine.
- If the patient is due for a second dose of vaccine in the immediate posttransplant period, this may be delayed 6 weeks to elicit a better immune response.
- Potential live liver donors and recipients of live donor livers should be prioritized for COVID-19 vaccination and preferably receive the second dose of the COVID-19 vaccine at least two weeks before transplantation when feasible based upon vaccine availability. A lack of COVID-19 vaccination should NOT delay lifesaving living donor liver transplantation.
- Family members and caregivers of liver transplant recipients should be vaccinated against SARS-CoV-2 whenever possible.

COVID-19 Vaccination Knowledge Gaps

Patients with advanced CLD and liver transplant recipients have not been included in the mRNA vaccine studies and as such data on effectiveness and safety are lacking in these populations. Post-marketing research is being conducted on antibody response to COVID-19 vaccines in patients with chronic conditions, including cirrhosis and autoimmune diseases. Acute and chronic liver diseases encompass a wide spectrum of etiologies and severity of disease and thus represent a heterogeneous population. Further, there are known racial and ethnic differences in prevalence and incidence of various liver diseases.(52,53) Several confounders such as obesity, diabetes mellitus, hypertension, and alcohol use may impact immune regulation, liver disease progression and severity that are relevant in the context of vaccination.(53,54) Cirrhosis is inherently a state of qualitative and quantitative immune dysregulation,(1) while some patients may be further immunosuppressed with medications such as transplant recipients and those with autoimmune hepatitis. Increasing liver disease severity has been associated with lower non-COVID-19 vaccine responsiveness.(55,56) These large knowledge gaps related to liver disease and transplantation require special attention in further studies. (**Table 2**)

CONCLUSION

Since the identification of the SARS-CoV-2 genome in January 2020, remarkable progress has been made in the development of highly effective and generally safe mRNA vaccines for COVID-19. The CDC currently recommends that all adults over the age of 18 should receive the 2-dose mRNA vaccines according to the manufacturers' recommendations to prevent future COVID-19. Pre- and post-vaccination serological testing is not recommended because of the absence of studies regarding their impact on outcomes. Both mRNA COVID-19 vaccines are recommended for all patients with CLD (compensated or decompensated) and immunosuppressed SOT recipients. The AASLD recommends that providers advocate for prioritizing patients with compensated or decompensated cirrhosis or liver cancer, immunosuppressed patients such as SOT recipients, and living liver donors for COVID-19 vaccination based upon local health policies, protocols, and vaccine availability.

The clinical impact of SARS-CoV-2 viral variants is rapidly evolving, and until further studies are available, COVID-19 vaccination should not be withheld or deferred in any patient because of efficacy or safety concerns aside from severe allergic reaction to any vaccine components. All COVID-19 vaccine recipients are recommended to continue social distancing, masking, frequent hand washing, and follow other exposure-mitigating behaviors. The online companion document located at https://www.aasld.org/about-aasld/covid-19-resources will be updated as additional data become available regarding the safety and efficacy of other COVID-19 vaccines in development.

References

- 1. Bonnel AR, Bunchorntavakul C, Reddy KR. Immune dysfunction and infections in patients with cirrhosis. Clin Gastroenterol Hepatol 2011 September;9:727–738.
- Van Kerkhove MD, Vandemaele KAH, Shinde V, Jaramillo-Gutierrez G, Koukounari A, Donnelly CA, et al. Risk factors for severe outcomes following 2009 influenza A (H1N1) infection: a global pooled analysis. PLoS Med 2011 July;8:e1001053.
- van Hoek AJ, Andrews N, Waight PA, Stowe J, Gates P, George R, et al. The effect of underlying clinical conditions on the risk of developing invasive pneumococcal disease in England. J Infect 2012 July;65:17–24.
- 4. Gutierrez Domingo I, Pascasio Acevedo JM, Alcalde Vargas A, Ramos Cuadra A, Ferrer Ríos MT, Sousa Martin JM, et al. Response to vaccination against hepatitis B virus with a schedule of four 40-

μg doses in cirrhotic patients evaluated for liver transplantation: factors associated with a response. Transplant Proc 2012 August;44:1499–1501.

- Bonazzi PR, Bacchella T, Freitas AC, Osaki KT, Lopes MH, Freire MP, et al. Double-dose hepatitis B vaccination in cirrhotic patients on a liver transplant waiting list. Braz J Infect Dis 2008 August;12:306–309.
- CDC. ACIP general best practice guidelines for immunization. Published November 20, 2020. https://www.cdc.gov/vaccines/hcp/acip-recs/general-recs/index.html. Accessed February 2021.
- Belouzard S, Millet JK, Licitra BN, Whittaker GR. Mechanisms of coronavirus cell entry mediated by the viral spike protein. Viruses 2012 June;4:1011–1033.
- 8. Wu F, Zhao S, Yu B, Chen Y-M, Wang W, Song Z-G, et al. A new coronavirus associated with human respiratory disease in China. Nature 2020 March;579:265–269.
- US Department of Defense. Coronavirus: Operation Warp Speed timeline. https://www.defense.gov/Explore/Spotlight/Coronavirus/Operation-Warp-Speed/Operation-Warp-Speed/Operation-Warp-Speed-Timeline. Accessed February 2021.
- 10. Jackson LA, Anderson EJ, Rouphael NG, Roberts PC, Makhene M, Coler RN, et al. An mRNA vaccine against SARS-CoV-2 preliminary report. N Engl J Med 2020 November 12;383:1920–1931.
- 11. Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, et al. Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. N Engl J Med 2020 December 31;383:2603–2615.
- Pardi N, Hogan MJ, Naradikian MS, Parkhouse K, Cain DW, Jones L, et al. Nucleoside-modified mRNA vaccines induce potent T follicular helper and germinal center B cell responses. J Exp Med 2018 June 4;215:1571–1588.
- 13. Reichmuth AM, Oberli MA, Jaklenec A, Langer R, Blankschtein D. mRNA vaccine delivery using lipid nanoparticles. Ther Deliv 2016;7:319–334.
- 14. Buschmann MD, Carrasco MJ, Alishetty S, Paige M, Alameh MG, Weissman D. Nanomaterial delivery systems for mRNA vaccines. Vaccines (Basel) 2021 January 19;9:65.

- Kelly C, Swadling L, Capone S, Brown A, Richardson R, Halliday J, et al. Chronic hepatitis C viral infection subverts vaccine-induced T-cell immunity in humans. Hepatology 2016 May;63:1455– 1470.
- Swadling L, Halliday J, Kelly C, Brown A, Capone S, Ansari MA, et al. Highly-immunogenic virallyvectored T-cell vaccines cannot overcome subversion of the T-cell response by HCV during chronic infection. Vaccines (Basel) 2016 August 2;4:27.
- Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, et al. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. Lancet 2021 January 9;397:99–111.
- AstraZeneca. AstraZeneca's COVID-19 vaccine authorised for emergency supply in the UK. https://www.astrazeneca.com/media-centre/press-releases/2020/astrazenecas-covid-19-vaccineauthorised-in-uk.html. Accessed February 2021.
- Johnson & Johnson Press Release. Johnson & Johnson announces single-shot Janssen COVID-19 vaccine candidate met primary endpoints in interim analysis of its phase 3 ENSEMBLE trial. Published January 29, 2021. https://www.jnj.com/johnson-johnson-announces-single-shot-janssencovid-19-vaccine-candidate-met-primary-endpoints-in-interim-analysis-of-its-phase-3-ensembletrial. Accessed February 2021.
- 20. Zhu F-C, Li Y-H, Guan X-H, Hou L-H, Wang W-J, Li J-X, et al. Safety, tolerability, and immunogenicity of a recombinant adenovirus type-5 vectored COVID-19 vaccine: a dose-escalation, open-label, non-randomised, first-in-human trial. The Lancet 2020 June 13;395:1845–1854.
- 21. Cao Y, Zhu X, Hossen MN, Kakar P, Zhao Y, Chen X. Augmentation of vaccine-induced humoral and cellular immunity by a physical radiofrequency adjuvant. Nat Commun 2018 September 12;9:3695.
- 22. Novavax Press Release. Novavax COVID-19 vaccine demonstrates 89.3% efficacy in UK phase 3 trial. Published January 28, 2021. https://ir.novavax.com/news-releases/news-release-details/novavax-covid-19-vaccine-demonstrates-893-efficacy-uk-phase-3. Accessed February 2021.
- 23. COVID-19 Real-Time Learning Network. Vaccines & Immunity. https://www.idsociety.org/covid-19real-time-learning-network/vaccines. Accessed February 2021.

- Janssen Press Release. Johnson & Johnson announces submission of application to the U.S. FDA for Emergency Use Authorization of its investigational single-shot Janssen COVID-19 vaccine candidate. Published February 4, 2021. https://www.janssen.com/johnson-johnson-announces-submissionapplication-us-fda-emergency-use-authorization-its. Accessed February 2021.
- CDCMMWR. Allergic reactions including anaphylaxis after receipt of the first dose of Pfizer-BioNTech COVID-19 vaccine — United States, December 14–23, 2020. MMWR Morb Mortal Wkly Rep 2021;70:46–51.
- CDCMMWR. Allergic reactions including anaphylaxis after receipt of the first dose of Moderna COVID-19 vaccine — United States, December 21, 2020–January 10, 2021. MMWR Morb Mortal Wkly Rep 2021 January 29;70:125–129.
- Castells MC, Phillips EJ. Maintaining safety with SARS-CoV-2 vaccines. N Engl J Med 2020 December
 doi: 10.1056/NEJMra2035343. [Online ahead of print]
- 28. Volz E, Mishra S, Chand M, Barrett JC, Johnson R, Geidelberg L, et al. Transmission of SARS-CoV-2 lineage B.1.1.7 in England: Insights from linking epidemiological and genetic data. MedRxiv 2021 January 4. doi: 10.1101/2020.12.30.20249034. [Preprint article that has not been peer-reviewed]
- 29. Tegally H, Wilkinson E, Giovanetti M, Iranzadeh A, Fonseca V, Giandhari J, et al. Emergence and rapid spread of a new severe acute respiratory syndrome-related coronavirus 2 (SARS-CoV-2) lineage with multiple spike mutations in South Africa. MedRxiv 2020 December 22. doi: 10.1101/2020.12.21.20248640. [Preprint article that has not been peer-reviewed]
- Faria NR, Claro IM, Candido D, Moyses Franco LA, Andrade PS, Coletti TM, et al. Genomic characterisation of an emergent SARS-CoV-2 lineage in Manaus: preliminary findings - SARS-CoV-2 coronavirus / nCoV-2019 Genomic Epidemiology. Virological. Published January 12, 2021. https://virological.org/t/genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-manauspreliminary-findings/586. Accessed February 2021.
- Galloway SE, Paul P, MacCannell DR, Johansson MA, Brooks JT, MacNeil A, et al. Emergence of SARS-CoV-2 B.1.1.7 lineage - United States, December 29, 2020-January 12, 2021. MMWR Morb Mortal Wkly Rep 2021 January 22;70:95–99.

- Gallagher J. Coronavirus: UK variant "may be more deadly." Published January 22, 2021. https://www.bbc.com/news/health-55768627. Accessed February 2021.
- Xie X, Zou J, Fontes-Garfias CR, Xia H, Swanson KA, Cutler M, et al. Neutralization of N501Y mutant SARS-CoV-2 by BNT162b2 vaccine-elicited sera. BioRxiv 2021 January 7. doi: 10.1101/2021.01.07.425740. [Preprint article that has not been peer-reviewed]
- Muik A, Wallisch A-K, Sänger B, Swanson KA, Mühl J, Chen W, et al. Neutralization of SARS-CoV-2 lineage B.1.1.7 pseudovirus by BNT162b2 vaccine-elicited human sera. BioRxiv 2021 January 19. doi: 10.1101/2021.01.18.426984. [Preprint article that has not been peer-reviewed]
- Wu K, Werner AP, Moliva JI, Koch M, Choi A, Stewart-Jones GBE, et al. mRNA-1273 vaccine induces neutralizing antibodies against spike mutants from global SARS-CoV-2 variants. BioRxiv 2021 January 25. doi: 10.1101/2021.01.25.427948. [Preprint article that has not been peer-reviewed]
- Feldstein LR, Rose EB, Horwitz SM, Collins JP, Newhams MM, Son MBF, et al. Multisystem inflammatory syndrome in U.S. children and adolescents. N Engl J Med 2020 July 23;383:334–346.
- Lobritto S, Danziger-Isakov L, Michaels MG, Mazariegos GV. Impact of COVID-19 Pandemic on Pediatrics and pediatric transplantation programs. Front Pediatr 2020;8:612627.
- CDC. COVID-19 vaccination provider requirements and support. Published January 7, 2021. https://www.cdc.gov/vaccines/covid-19/vaccination-provider-support.html. Accessed February 2021.
- CDC. COVID-19: When vaccine is limited, who gets vaccinated first. Published December 31, 2020. https://www.cdc.gov/coronavirus/2019-ncov/vaccines/recommendations.html. Accessed February 2021.
- CDC. The importance of COVID-19 vaccination for healthcare personnel. Published February 11, 2020. https://www.cdc.gov/coronavirus/2019-ncov/vaccines/recommendations/hcp.html. Accessed February 2021.
- 41. Iavarone M, D'Ambrosio R, Soria A, Triolo M, Pugliese N, Del Poggio P, et al. High rates of 30-day mortality in patients with cirrhosis and COVID-19. J Hepatol 2020 November;73:1063–1071.

- 42. Williamson EJ, Walker AJ, Bhaskaran K, Bacon S, Bates C, Morton CE, et al. Factors associated with COVID-19-related death using OpenSAFELY. Nature 2020;584:430–436.
- 43. Moon AM, Webb GJ, Aloman C, Armstrong MJ, Cargill T, Dhanasekaran R, et al. High mortality rates for SARS-CoV-2 infection in patients with pre-existing chronic liver disease and cirrhosis: preliminary results from an international registry. J Hepatol 2020 September;73:705–708.
- Singh S, Khan A. Clinical characteristics and outcomes of COVID-19 among patients with pre-existing liver disease in United States: a multi-center research network study. Gastroenterology 2020 August;159:768–771.
- 45. Bajaj JS, Garcia-Tsao G, Biggins SW, Kamath PS, Wong F, McGeorge S, et al. Comparison of mortality risk in patients with cirrhosis and COVID-19 compared with patients with cirrhosis alone and COVID-19 alone: multicentre matched cohort. Gut 2020 July 13. doi: 10.1136/gutjnl-2020-322118. [Online ahead of print]
- 46. Krammer F, Srivastava K, the PARIS Team, Simon V. Robust spike antibody responses and increased reactogenicity in seropositive individuals after a single dose of SARS-CoV-2 mRNA vaccine. MedRxiv 2021 February 1. doi: 10.1101/2021.01.29.21250653. [Preprint article that has not been peer-reviewed]
- CDC. COVID-19: People with certain medical conditions. Published December 29, 2020. https://www.cdc.gov/coronavirus/2019-ncov/need-extra-precautions/people-with-medicalconditions.html. Accessed February 2021.
- Gerussi A, Rigamonti C, Elia C, Cazzagon N, Floreani A, Pozzi R, et al. Coronavirus Disease 2019 (COVID-19) in autoimmune hepatitis: a lesson from immunosuppressed patients. Hepatology Communications 2020 June 9;4:1257–1262.
- 49. Colmenero J, Rodríguez-Perálvarez M, Salcedo M, Arias-Milla A, Muñoz-Serrano A, Graus J, et al. Epidemiological pattern, incidence and outcomes of COVID-19 in liver transplant patients. J Hepatol 2021 January;74:148–155.
- Webb GJ, Marjot T, Cook JA, Aloman C, Armstrong MJ, Brenner EJ, et al. Outcomes following SARS-CoV-2 infection in liver transplant recipients: An international registry study. Lancet Gastroenterol Hepatol 2020 November;5:1008–1016.

- 51. Marjot T, Webb GJ, Barritt AS, Ginès P, Lohse AW, Moon AM, et al. SARS-CoV-2 vaccination in patients with liver disease: responding to the next big question. The Lancet Gastroenterology & Hepatology 2021 January 11. doi: 10.1016/S2468-1253(21)00008-X. [Online ahead of print]
- 52. Forde KA, Tanapanpanit O, Reddy KR. Hepatitis B and C in African Americans: current status and continued challenges. Clin Gastroenterol Hepatol 2014 May;12:738–748.
- 53. Sheka AC, Adeyi O, Thompson J, Hameed B, Crawford PA, Ikramuddin S. Nonalcoholic steatohepatitis: a review. JAMA 2020 March 24;323:1175–1183.
- 54. Mendenhall C, Roselle GA, Lybecker LA, Marshall LE, Grossman CJ, Myre SA, et al. Hepatitis B vaccination. Response of alcoholic with and without liver injury. Dig Dis Sci 1988 March;33:263–269.
- 55. Dumot JA, Barnes DS, Younossi Z, Gordon SM, Avery RK, Domen RE, et al. Immunogenicity of hepatitis A vaccine in decompensated liver disease. Am J Gastroenterol 1999 June;94:1601–1604.
- Carey W, Pimentel R, Westveer MK, Vogt D, Broughan T. Failure of hepatitis B immunization in liver transplant recipients: results of a prospective trial. Am J Gastroenterol 1990 December;85:1590– 1592.
- 57. Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, et al. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. N Engl J Med 2021 February 4;384:403–416.
- Sadoff J, Le Gars M, Shukarev G, Heerwegh D, Truyers C, de Groot AM, et al. Interim results of a Phase 1-2a trial of Ad26.COV2.S Covid-19 vaccine. N Engl J Med 2021 January 13. doi: 10.1056/NEJMoa2034201. [Online ahead of print]
- Zhu F-C, Guan X-H, Li Y-H, Huang J-Y, Jiang T, Hou L-H, et al. Immunogenicity and safety of a recombinant adenovirus type-5-vectored COVID-19 vaccine in healthy adults aged 18 years or older: a randomised, double-blind, placebo-controlled, phase 2 trial. Lancet 2020 August 15;396:479–488.
- Simões E. New Brazil data shows disappointing 50.4% efficacy for China's CoronaVac vaccine. Reuters. Published January 13, 2021. https://www.reuters.com/article/us-health-coronavirus-brazilcoronavirus-idUSKBN29H2CE. Accessed February 2021.

 Xia S, Zhang Y, Wang Y, Wang H, Yang Y, Gao GF, et al. Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBIBP-CorV: a randomised, double-blind, placebo-controlled, phase 1/2 trial. Lancet Infect Dis 2021 January;21:39–51.

Author Manuscr

| | Vaccine | Dosing | Efficacy | Safety issues | Storage issues |
|--------------------|---------------|-----------------|---------------------------|--------------------|---------------------|
| | | | Vaccines with FD | A EUA | |
| | mRNA | 30 µg (0.3 mL) | 95%(11) | Synthetic lipid | Store between |
| | BNT162b2 | IM x 2 doses 21 | (95.3% in those | nanoparticle | -80 °C to -60 °C |
| | (Pfizer- | days apart | with | Contraindicated if | Once thawed and |
| | BioNTech) | | comorbidities | history of severe | diluted, multi- |
| | | EUA for ages 16 | including CLD) | or immediate | dose vials must |
| | \mathbf{O} | and older | | allergic reaction | be stored |
| | () | | | to any vaccine | between 2 °C to |
| | | | | components, | 25 °C and used |
| 10 | | | | including PEG* | within 6 hours |
| mRNA vaccines | mRNA-1273 | 100 μg (0.5 mL) | 94.1%(57) | Synthetic lipid | Store between |
| vac | (Moderna) | IM x 2 doses 28 | (Unknown in | nanoparticle | -25 °C to -15 °C |
| RNA | | days apart | CLD patients | Contraindicated if | Thawed vials |
| 2 | M | | because no | history of severe | stored at 2 °C to 8 |
| | | EUA for ages 18 | vaccine or | or immediate | °C for up to 30 |
| | | and older | placebo pts | allergic reaction | days or between |
| | | | developed | to any vaccine | 8 °C to 25 °C for |
| | | | COVID-19 in | components, | up to 12 hours |
| | | | clinical trials) | including PEG* | Once first dose is |
| | | | | | withdrawn, vial |
| | | | | | must be used |
| | | | | | within 6 hours |
| | | Vac | cines in Phase 3 de | velopment | |
| | AZD1222 | 1 or 2 IM doses | 70.4% (pooled) | Replication- | Stored and |
| tors | (AstraZeneca) | 28 day apart | after the 2 nd | defective | distributed at 2 °C |
| Adenoviral vectors | | | dose | chimpanzee | to 8 °C for up to 6 |
| ovira | | EUA in UK, | 62% standard | adenovirus vector | months |
| Vden | | Europe, and | dose (SD)/SD | | |
| < | | South America | | | |

| | | for ages ≥18 | 90% low | 2 cases of | Easier to scale up |
|-------------|--------------|-----------------|------------------|-------------------|---------------------|
| | | years | dose/SD(17) | transverse | production vs |
| | | | Unknown in | myelitis reported | mRNA |
| | | | CLD patients | | |
| | Ad26.COV2.S | 1 or 2 IM doses | 72% in US with | Replication- | Stored at 2 °C to 8 |
| | (Johnson and | are being | single dose | defective | °C for up to 3 |
| | Johnson/ | tested | 66% in Latin | adenovirus 26 | months |
| | Janssen) | | America | vector (used in | |
| | \mathbf{O} | | 57% in S | Ebola vaccine) | |
| | () | | Africa(58) | | |
| | | | | Low | |
| | | | | seroprevalence of | |
| | | | | antibodies in N | |
| | | | | America | |
| | Ad5-NCoV | | 96%-97% | Replication- | |
| | (CanSino | | antibody | defective | |
| | biologics) | | induction at day | adenovirus type 5 | |
| | | | 28(59) | vector | |
| | NCX- | 2 IM doses 3 | 89.3% in UK | Recombinant | 2 °C to 8 °C |
| tein | CoV2373 | weeks apart | study | spike protein | |
| ant protein | (Novavax) | | 49.4% in S | nanoparticles | |
| nant | | | Africa(22) | | |
| Recombin | | | | Adjuvant of M- | |
| Reco | | | | matrix which may | |
| | | | | be allergenic | |
| σ | CoronaVac | | 50.4% | Inactivated SARS- | |
| Inactivated | (Sinovac) | | protection in | CoV-2 with alum | |
| activ. | | | Brazilian | hydroxide | |
| 5 | | | study(60) | adjuvant | |

| BBIBP-Cor | / | 100% antibody | Inactivated whole | |
|-------------|---|------------------|-------------------|--|
| Inactivated | I | induction at day | virion SARS-CoV-2 | |
| COVID-19 | | 42(61) | | |
| (Wuhan) | | | | |

PEG: polyethylene glycol

* Ingredients include mRNA, lipids, polyethylene glycol, cholesterol, potassium chloride, potassium

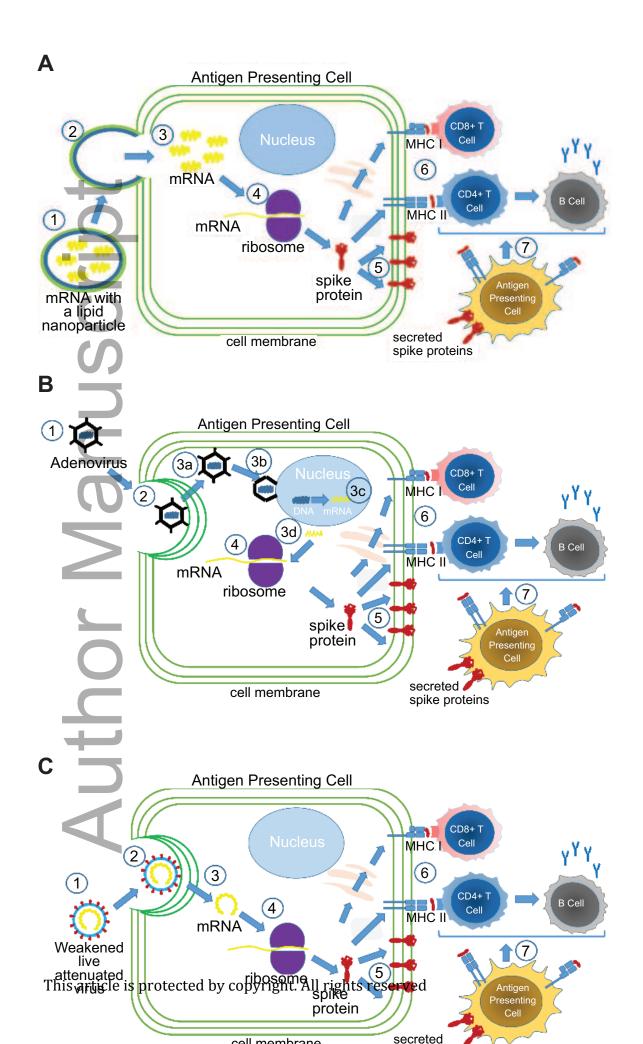
phosphate, sodium chloride, sodium phosphate, sucrose

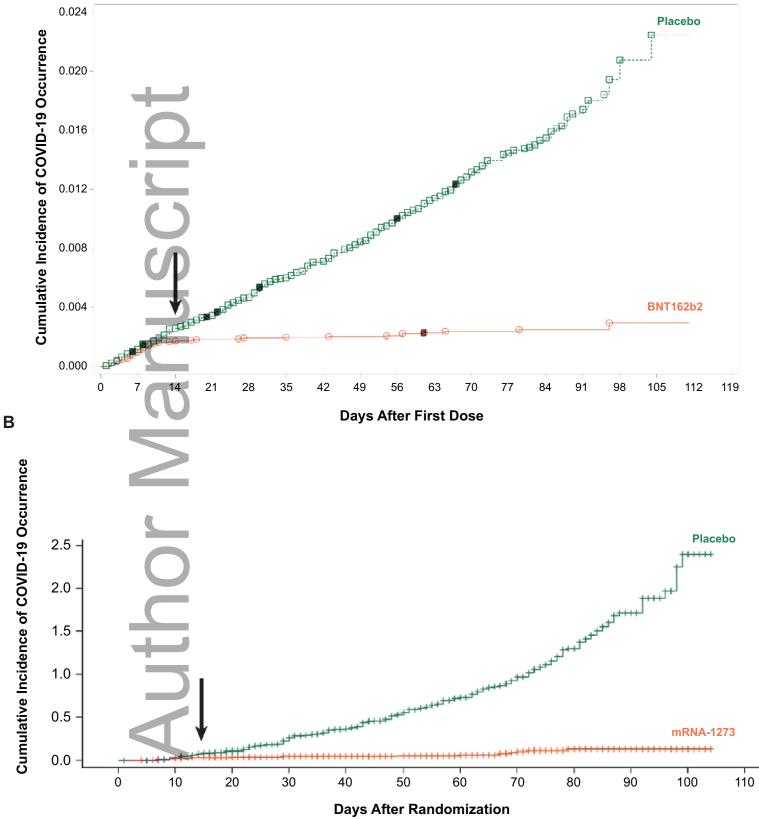
Author Manuso

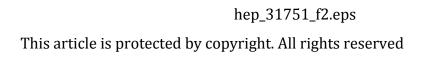
Table 2. COVID-19 Vaccination Knowledge Gaps

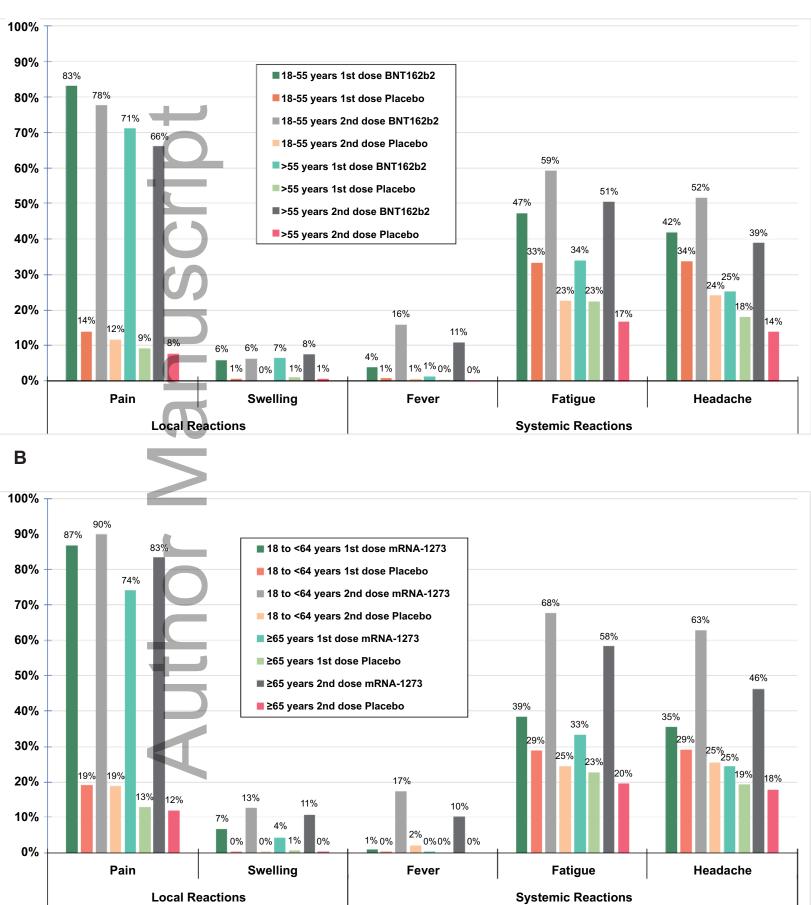
- Effectiveness and safety in patients with CLD based on liver disease etiology, comorbidities, CTP class, and MELD score
- Effectiveness and safety in immunocompromised/immunosuppressed individuals including transplant recipients
- Effectiveness and safety in pediatric populations (adolescents and children)
- Effectiveness and safety in pregnant and lactating women
- Effectiveness and safety in individuals previously infected with SARS-CoV-2
- Effectiveness against SARS-CoV-2 viral variants (e.g., B.1.1.7, B.1.351, P.1)
- Effectiveness against asymptomatic infection
- Effectiveness against SARS-CoV-2 transmission
- Effectiveness against long-term effects of COVID-19
- Effectiveness and safety in a diverse population including different racial and ethnic backgrounds
- Effectiveness and safety of vaccination with a different vaccine following a prior allergic/anaphylactic reaction to an mRNA COVID-19 vaccine
- Duration of protective immunity against SARS-CoV-2 infection
- Mechanisms of vaccine failure

Author









Α