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7 **The feasibility of ultrasound-assisted endovascular laser thrombolysis in an acute**
8 **rabbit thrombosis model**

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10 Ultrasound-assisted laser thrombolysis

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25

26 **Abstract**

27 Purpose

28 The current study was aimed to test the feasibility of combined ultrasound and laser technique,
29 namely, ultrasound-assisted endovascular laser thrombolysis (USELT), for thrombolysis by
30 conducting in vivo tests in a rabbit thrombosis model.

31 Methods

32 An acute thrombus was created in the right jugular vein of rabbit and then was treated with
33 ultrasound only, laser only, and USELT to dissolve the blood clot. A total of 20 rabbits were
34 used. Out of which, the first three rabbits were used to titrate the laser and ultrasound parameters.
35 Then, five rabbits were treated with ultrasound only, five rabbits were treated with laser only,
36 and seven rabbits were treated with USELT. During USELT, 532-nm laser pulses were delivered
37 endovascularly directly to the clot through a fiber optic, and 0.5-MHz ultrasound pulses were
38 applied noninvasively to the same region. A laser fluence of 4 to 12 mJ/cm² and ultrasound
39 amplitude of 1 to 2 MPa were used. Recanalization of the jugular vein was assessed by
40 performing ultrasound doppler imaging immediately after the treatment. The maximum blood
41 flow speed after the treatment as compared to its value before the treatment was used to calculate
42 the blood flow recovery in vessel.

43 Results

44 The blood flow was fully recovered (100 %) in three rabbits, partially recovered in two rabbits
45 (more than 50 % and less than 100 %) with mean percentage recovery of 69.73 % and poorly

46 recovered in two rabbits (less than 50%) with mean percentage recovery of 6.2 % in USELT
47 group. On the contrast, the treatment group with ultrasound or laser alone did not show
48 recanalization of vein in any case, all the five rabbits were poorly/not recovered with a mean
49 percentage recovery of 0 %.

50 Conclusions

51 The USELT technology was shown to effectively dissolve the blood clots in an acute rabbit
52 jugular vein thrombosis model.

53 Keywords: Non-invasive ultrasound thrombolysis, endovascular laser thrombolysis, rabbit
54 thrombosis model, cavitation.

55 **Introduction**

56 Venous thromboembolism (VTE), a condition in which a blood clot (a thrombus) forms
57 in a vein, is a major disease affecting more than 10 million people worldwide each year ¹. VTE
58 can lead to a myriad of complications, including swelling, erythema, neurovascular compromise,
59 tissue necrosis or limb loss, acute respiratory symptoms, pulmonary hypertension, cardiovascular
60 collapse, thromboembolism, and death. A venous thrombus most commonly occurs in the deep
61 veins of the legs or pelvis; this is then called a deep vein thrombosis (DVT). A pulmonary
62 embolism (PE) occurs when a clot breaks loose and travels through the bloodstream to the lungs.
63 PE is an acute life-threatening complication, causing as many as 100,000 deaths annually in the
64 United States. Post-thrombotic syndrome, develops in 30% to 75 % of patients with DVT, which
65 is a costly chronic condition often causing lengthy disability ²⁻⁴. Overall, the annual medical
66 costs are \$7 to \$10 billion for VTE in the United States. Worldwide, the total cost can be as high
67 as \$69 billion annually ⁵.

68 Thrombosis is both an expensive and complicated condition to address. Treatment
69 requires dissolving the blood clots in the blood vessels, referred to as thrombolysis. The
70 treatment approaches are diverse. First-line therapy is to give patients direct oral anticoagulants
71 ^{6,7}. This medication prevents the formation of clotting factors. However, anticoagulant
72 medications are not efficient because they do not dissolve the clot and re-canalize vessels ⁸.

73 Another approach is thrombolytic therapy, which is undertaken by injecting a clot-dissolving
74 medication. The limitations of the treatment include the need for hospitalization due to risks of
75 bleeding and low effectiveness for totally occluded veins⁹.

76 Ultrasound-based treatment techniques have been evaluated as methods to induce
77 effective thrombolysis ¹⁰⁻¹⁶. The advantage of ultrasound-based techniques is that they can
78 dissolve blood clots quickly and re-canalize vessels noninvasively through cavitation. But, these
79 techniques require high acoustic peak negative pressure (as high as 19 MPa ¹⁷) at relatively low
80 ultrasound frequencies of 500 kHz or 1 MHz. In order to achieve high pressure and deliver
81 treatment to a blood clot, focused ultrasound must be employed. However, at low ultrasound
82 frequencies such as 0.5 MHz, the focal spot of the ultrasound field is usually larger than 10 mm
83 in length, which is larger than the diameters of most veins. As a result, damages can be induced
84 in the surrounding tissue and vessel walls ¹⁷. This could be especially problematic in areas with
85 delicate structures that have limited surgical options, such as retina vein occlusions, renal vein
86 thrombus, and stroke. Although the size of the focal zone could be reduced by using transducers
87 with small f-numbers, this has a tradeoff with the reduced treatment depth, and also is limited by
88 the available acoustic window ¹⁸. Alternatively, high intensity focused ultrasound (HIFU) with
89 higher frequency may be used to produce a smaller focal size, but it will reduce the efficiency of
90 thrombolysis because it is generally more difficult to produce cavitation at a higher frequency ¹⁹.
91 To increase the efficiency and safety of ultrasound-based thrombolysis, microbubbles can be
92 used ²⁰; however, requires an additional systemic injection of the microbubbles and may cause
93 unwanted vascular and tissue damages at high dosages ²¹⁻²³

94 Laser thrombolysis is an interventional procedure to re-canalize occluded vessels ²⁴⁻²⁹.
95 Laser thrombolysis utilizes a light wavelength that is highly absorptive to the blood clots. Laser
96 light generally is directed to the blood clot through a thin laser fiber, which induces heating of
97 the clot, and cavitation can occur in the blood clot through vaporization. Then, similar to
98 ultrasound thrombolysis, the expansion and collapse of a vapor cavity can also break up the
99 blood clot. The advantages of laser thrombolysis include low cost, a shorten recovery time, and
100 generally high safety. Laser thrombolysis can precisely induce cavitation in blood clots due to
101 the high optical absorption of blood clots at certain wavelength (like 308 nm ²⁶, 480 nm ²⁹ and

102 577 nm²⁸) compared to the surrounding tissue and structures. However, the produced cavitation
103 expansion and collapse cannot be controlled and often are not strong enough to efficiently break
104 up the clot. As a result, laser thrombolysis often cannot completely clear thrombotic occlusions
105 in the blood vessels, typically leaving residual thrombus on the blood vessel walls²⁸. Its
106 efficiency is also questionable in removing blood clots with high calcium contents.

107 We have developed a novel hybrid technology, based on the combination of light and
108 ultrasound, namely ultrasound-assisted endovascular laser thrombolysis (USELT), to safely and
109 efficiently dissolve the blood clots in the vein. The USELT system is based on our earlier
110 developed photo-mediated ultrasound therapy (PUT) technology³⁰⁻³³. Both technologies use
111 combination of ultrasound and laser to generate enhanced cavitation. In PUT, both laser and
112 ultrasound are applied non-invasively³⁴⁻³⁹, whereas in USELT the laser is delivered directly to
113 the clot using an endovascular laser catheter. Due to non-invasive laser in PUT, only certain
114 wavelength with good transparency in the intervening tissues can be used. Whereas in USELT,
115 any laser wavelength that maximize absorptive heating of the blood clot can be used. In USELT,
116 the laser is applied through catheter and noninvasive ultrasound energy is applied from outside
117 the body to the blood clot to drive the generated cavitation bubbles and achieve the best
118 thrombolysis outcomes. As a result, the advantages of laser and ultrasound treatment can be
119 combined, and blood clots can be dissolved rapidly. In a previous study⁴⁰, we initially
120 characterized the feasibility of the USELT system with an in vitro blood flow system, and
121 demonstrated the efficiency of thrombolysis as a function of ultrasound pressures and laser
122 fluences. In the current study, we tested the feasibility of the USELT system in an acute in vivo
123 rabbit thrombosis model. The translation potential of USELT is demonstrated, although further
124 development will be needed to optimize the treatment before clinical use.

125 **Materials and Methods**

126 A. Experiment Setup

127 A detailed schematic of the USELT system is shown in Fig 1. This system is a
128 combination of an endovascular laser thrombolysis system and a high-intensity focused
129 ultrasound system. The laser system uses a Q-switched diode pumped solid state laser (Elforlight

130 Model SPOT-10-200-532, Bozeman, MT) to produce 532-nm wavelength light. The laser pulse
131 duration and energy was 2-ns and 0-20 uJ, respectively. The laser light was delivered to the
132 desired treatment region using an optical fiber and fiber optic cannula. The produced laser light
133 was first passed to a long optical fiber to carry it near to the vein. Near the vein, the optical fiber
134 was connected to a fiber optic cannula of 400 μm (CFMLC14U-20, Thorlabs, Newton, NJ) to
135 produce a laser fluence of 4 to 12 mJ/cm^2 . The laser power was adjusted to the desired level
136 using an optical power meter (S425C, Thorlabs, Newton, NJ) before each treatment. The laser
137 system was triggered by a delay/pulse generator (DG535, Stanford Research Systems, Sunnyvale,
138 CA, USA) to give a pulse repetition frequency of 10 kHz. The same delay/pulse generator was
139 used to trigger ultrasound with a fixed delay in the ultrasound pulse. The delay was set such as to
140 provide desired synchronization between two systems such that the concurrent laser and
141 ultrasound pulses were applied on the treatment region.

142 For the ultrasound system, the delay/pulse generator triggered a function generator
143 (33250A, Agilent Technologies, Santa Clara, CA) to produce a 0.5 MHz signal. The signal was
144 first amplified by 50 dB in RF amplifier (2100L, ENI, Rochester, NY) and then passed to a
145 matching network (Impedance Matching Network H-107, Sonic Concepts) before being sent to
146 the transducer. The high-intensity focused ultrasound transducer (H-107, Sonic Concepts,
147 Bothell, WA) has a central frequency of 0.5 MHz with focal distance, focal depth and focal
148 width of 63.2 mm, 21.42 mm and 3.02 mm respectively. The front face of the transducer was
149 placed inside a conical plastic cone such that its focal point was present near the small opening
150 on the vertex of the cone. The focal peak negative pressure on the cone vertex opening was
151 measured using a standard needle hydrophone (SN-1462, 0.5 mm, Precision Acoustics Ltd, UK).
152 For each trigger, the function generator generated a 5-cycle ultrasound pulse, resulting in a 10%
153 duty cycle for an ultrasound burst.

154 B. Animal Model

155 All the animal handling procedures were carried out in compliance with a protocol
156 approved by the Institutional Animal Care and Use Committee (IACUC) at the University of
157 Kansas, protocol numbers AUS 188-11, PI Xinmai Yang, with strict adherence to the National
158 Institutes of Health guidelines. Specifically, a thrombus was induced in the right jugular vein of a

159 New Zealand rabbit (male or female, weight ranging from 2.2 to 2.8 kg). To induce the thrombus,
160 a rabbit was first anesthetized with a mixture of ketamine (40 mg/kg) and xylazine (5 mg/kg).
161 Once the rabbit was sufficiently anesthetized, the hair near the neck region was shaved with an
162 electric clipper. The rabbit was then shifted to the isoflurane anesthesia, and respiratory rate,
163 blood oxygenation and heart rate were monitored. After the condition of the rabbit became stable,
164 it was moved to the surgical room before inducing the blood clot. The right jugular vein was
165 exposed and isolated from nearby tissues by making an opening in the shaved neck region. In the
166 isolated jugular vein, blood flow was confirmed by ultrasound doppler imaging. Then two
167 vascular clips were placed on the jugular vein approximately 1 cm apart to create a segment of
168 blood vessel without flow. After restricting the blood flow, 0.02 ml of blood was drawn from the
169 clipped area and was mixed with 0.02 ml of thrombin solution. The thrombin solution was made
170 by dissolving 1000 units of thrombin (EMD Millipore Sigma, 605157-1KU) in 0.5 ml of
171 bacteriostatic water. 0.03 ± 0.005 ml of this mixture of thrombin solution and blood was injected
172 back to the restricted vein section. The upstream clip was removed 15 minutes after the thrombin
173 administration and the other clip was removed after an additional 5 minutes. The blood clot was
174 allowed to further mature over the next 30 minutes, and then ultrasound doppler imaging was
175 repeated to measure the blood flow and confirm the blood clot formation. Fig 2 shows a picture
176 of the region where the clot was created, and a doppler image of blood flow.

177 C. Treatment Procedure

178 After the successful creation of a blood clot in the right jugular vein, the USELT system
179 shown in Fig 1 was used to recanalize the jugular vein. A 22G catheter was inserted into the vein
180 from an upstream location to the blood clot region and the laser optic cannula was further passed
181 through the catheter, such that its tip reached the blood clot inside the vein. The cone containing
182 transducer was placed directly on the vein surface such that its focal zone covered the entire vein
183 cross-section. Ultrasound gel was used for providing coupling between the cone tip and tissue.
184 The cannula tip placed inside vein was directly under the transducer focal zone. For treatment,
185 the ultrasound cone and cannula were placed at one position for 1 minute and then were moved
186 to the next position. The entire blood clot was treated for 3.5 to 4.5 minutes depending upon the
187 length of the blood clot (5-7 mm). The image of the treatment has been shown in Fig 1 (b). After

188 completing the treatment, ultrasound doppler imaging was performed at an upstream location to
189 observe the blood flow and confirm recanalization of the vessel. During the entire procedure, the
190 respiratory rate and heart rate of rabbit were noted after every 15 minutes to ensure the normal
191 physiological functioning. The laser catheter was then removed, and rabbit was observed for
192 over 30 minutes for any unusual changes to respiratory rate and heart rate due to treatment. The
193 rabbit was then euthanized by intravenously injecting pentobarbital, and the treated jugular vein
194 was collected. Histology on the collected veins was performed with hematoxylin and eosin
195 (H&E) stain to assess the vessel for damage and to observe the amount of blood clot dissolved.
196 For control groups, the exact same procedure was performed, but only therapeutic ultrasound
197 energy was applied for ultrasound-only group, and only laser energy was applied for laser-only
198 group.

199 The blood flow in the vein was observed using the doppler mode of an ultrasound
200 imaging unit (Z.One PRO, Mindray, Mahwah, NJ, USA) connected to a linear probe (L14-5W,
201 Mindray). It was used with a pulse repetition frequency of 1500 Hz and a continuous doppler
202 frequency of 5.5 MHz. The ultrasound doppler imaging was performed three times during the
203 experiment. It was performed first time to confirm the blood flow in jugular vein right after it
204 was exposed. The US doppler imaging was done second time after the clot formation in vein. A
205 final US doppler imaging was performed on vein after the completion of the treatment to observe
206 if blood flow was resumed or not.

207 D. Treatment and Control Group

208 The study included 20 rabbits with body weights ranging from 2.2 to 2.8 kg. The first
209 rabbit was used to test the experimental procedure and was not included in the results data. The
210 second and third rabbits were used to titrate the applied ultrasound pressure and laser fluence,
211 wherein one rabbit was treated with a peak negative ultrasound pressure (P-) of 1 MPa and laser
212 fluence of 12 mJ/cm² and the other was treated with a P- of 2 MPa and laser fluence of 4 mJ/cm²
213 (Table S-1). The remaining of 17 rabbits were divided into three treatment groups. In the first
214 group, the rabbits were treated with USELT using a P- of 1.3 MPa and laser fluence of 8 mJ/cm²
215 respectively. In the second group, the rabbits were treated with only ultrasound using a P- of 1.3
216 or 2 MPa. In the third group, the rabbits were treated with only laser using a laser fluence of 8

217 mJ/cm². Seven, five and five rabbits were treated in each group, respectively. The earlier studies
218 have also used ultrasound only and laser only for thrombolysis, but a very high peak negative
219 ultrasound pressure (as high as 19 MPa ¹⁷) and very high laser fluence (as high as 4500 mJ/cm².
220 ²⁶) was used. The laser assisted thrombolysis ^{26,29} has been used in clinics but ultrasound
221 thrombolysis is not yet used in clinics.

222 **Results**

223 Based on the maximum blood flow speed in the rabbit's vein before the clot formation
224 and after the treatment, the outcomes were divided into three categories. In the first category, the
225 rabbit's vein which has 100 percent maximum blood flow speed after treatment as compared to
226 its value before clot formation were considered as full recovery. Similarly, in the second and
227 third category, the rabbit's vein with maximum blood flow speed recovery in range of 50-99 %
228 and less than 50 % are considered as partial recovery and poor/no recovery, respectively. Fig 3
229 shows the total number of rabbits with full recovery, partial recovery, and poor/no recovery in
230 the maximum blood flow speed in the jugular vein after treatment with USELT (group I),
231 ultrasound-only (group II) and laser-only (group III). Ultrasound-only and laser-only control
232 groups were included because thrombolysis may be achieved by ultrasound-only or laser-only.
233 However, due to the low energy levels used during USELT, ultrasound-only and laser -only
234 control groups did not show recanalization. Fig 4 shows the mean percentage recovery for the
235 full recovery, partial recovery, and poor/no recovery veins within each treatment group.

236 In group I treated with USELT, out of the seven treated rabbits, three rabbits (rabbit No.
237 4, 8 and 11) were fully recovered, two (rabbit No. 9 and 12) were partially recovered and other
238 two (rabbit No. 7 and 10) were poorly recovered. Whereas, in group II and group III, none of the
239 five rabbits were fully or partially recovered. All the veins in group II and group III have a mean
240 percentage recovery of 0 (no recovery) as shown in Fig 4, which means no recovery took place
241 with ultrasound-only and laser-only treatment. Whereas, in group I, the poorly recovered vein
242 has a mean percentage recovery of 6.2 %. Also, in group I, the mean percentage recovery was
243 100 % and 69.73 % in full recovery and partial recovery group.

244 A Fisher's exact test was performed using a two by two contingency table to observe the
245 association between the treatment (USELT, ultrasound-only/laser-only) and the result of the
246 treatment (vein recovered or not). All the full and partial recovery veins were consolidated into
247 the recovery group and poor/no recovery veins were placed in no recovery group. Accordingly,
248 in group I, five veins were considered as recovery and two as no recovery, while in group II and
249 III, all five veins were considered as no recovery. A p-value of 0.0278 was obtained, which
250 shows the statically significance between the treatments and its result. A relative risk of 0.2857
251 for treatment failure was obtained, which means USELT has a failure chance of only 28.57% as
252 compared to ultrasound-only or laser-only treatment (100 % failure).

253 Figure 5 shows the ultrasound doppler imaging of three rabbit's jugular veins treated with
254 USELT, ultrasound only, and laser only. The ultrasound doppler imaging of the right jugular
255 vein of rabbit No. 8 treated with USELT is shown in Fig 5 (a)-(c). Blood flow is normal
256 immediately after isolating the vein (Fig. 5a), but no flow was observed due to total occlusion of
257 vein after injecting thrombin (Fig. 5b). Fig 5 (c) shows the ultrasound doppler imaging of the
258 same jugular vein after the blood clot was treated with USELT using P- of 1.3 MPa and laser
259 fluence of 8 mJ/cm². The red dot in Fig 5 (c), at a depth of around 4-5 mm from the scanner
260 surface, is at the same location as in Fig 5 (a), confirming resumption of blood flow in the vein.
261 Similarly, Figs 5 (d), (e) and (f) show the ultrasound doppler imaging of rabbit No. 5 treated with
262 ultrasound-only using a P- of 1.3 MPa and Fig 5 (g), (h) and (i) show rabbit No. 6 treated with
263 laser-only using a laser fluence of 8 mJ/cm². Of note, blood flow recovered completely in the
264 USELT-treated vein, Fig. 5 (c), but there was no recovery of blood flow in the ultrasound-only
265 or laser-only treated veins, Figs. 5 (f) and 5 (i), respectively, indicating the mono-therapies were
266 unable to dissolve the clots.

267 Fig 6 shows the maximum blood flow velocity in the jugular vein of rabbits (Group I, II
268 and III) obtained using ultrasound doppler imaging before and after the formation of a blood clot,
269 and after the treatment (USELT, ultrasound-only, laser-only). Before formation of a blood clot,
270 normal blood flow was observed in all the seventeen rabbit's veins with a mean maximum blood
271 flow velocity of 2.85 cm/s (standard deviation of 1.2 cm/s). In all cases, thrombin treatment
272 successfully formed a clot that completely occluded the vein and stopped blood flow. In group I,

273 which was treated with USELT using a P- of 1.3 MPa and laser fluence of 8 mJ/cm², the mean
274 maximum blood flow velocity from all seven rabbits was 2.26 cm/s with a standard deviation of
275 1.72 cm/s. In group II, there was no restoration of blood flow (0 cm/s) in the five rabbits treated
276 with ultrasound only using a P- of 1.3 or 2 MPa. Likewise, treatment with the laser only using a
277 laser fluence of 8 mJ/cm² failed to restore blood flow (0 cm/s) in any of the 5 rabbits in group III.
278 The maximum blood flow velocity after treatment between group I (USELT) and group II/III
279 (ultrasound-only/laser-only) was statistically significant (p=0.028), and before treatment and
280 after treatment with USELT was not statistically significant (p=0.33). Also, the maximum blood
281 flow velocity, before blood clot formation and after blood clot formation was statistically
282 significant (p<0.001), after blood clot formation and after treatment with USELT was
283 statistically significant (p<0.001).

284 Fig 7 shows the sections of a treated rabbit vein, stained with hematoxylin and eosin
285 (H&E). The Fig 7 (a) is an H&E stained vessel cross-section from a rabbit which was treated
286 with ultrasound only using P- of 1.3 MPa. The entire blood clot is intact and clearly visible in Fig
287 7 (a). The Fig 7 (b)-(e) shows different H&E stained vessel cross-sections of a partially
288 recovered vein that was treated with USELT using a P- of 1 MPa and laser fluence of 12 mJ/cm².
289 A large part of the clot is dissolved due to USELT treatment in the sections shown in Fig 7 (b)-
290 (e), Image processing was also done on the sections shown in Fig 7 (b), (c) and (d) using
291 MATLAB 2019a. It was found that around 58%, 49% and 54% of the clot area was dissolved in
292 the sections shown in Fig 7 (b), (c) and (d) respectively with the treatment of combined
293 ultrasound and laser. In this calculation, the entire vessel lumen area in the imaging was used as
294 the initial blood clot size because there was no blood flow after the blood clot formation. The
295 vessel wall adjacent to the dissolved clot area can be seen intact in all the sections in Fig 7 (b)-(e).
296 Fig 7 (e) is magnified image of the vessel wall highlighted in Fig 7 (c). Some residues of the
297 blood clot can be seen adjacent to the vessel wall, however there is no visible damage to the
298 vessel wall after treatment. Moreover, no major changes in heart rate (HR) and respiratory rate
299 (RR) were observed during and after the treatment indicating the normal physiological
300 functioning of rabbit. The HR and RR data for each rabbit in group I, which was treated with
301 USELT, are attached as Figs S-1 and S-2 in supplementary materials. The blood oxygenation
302 remained 98% or above throughout the experiment for all rabbits.

303 Discussion

304 The USELT device combines the advantages of both ultrasound-based and laser-based
305 thrombolysis. The first potential advantage is that USELT can dissolve blood clots based on
306 optical absorption at low ultrasound and laser energy levels. By taking advantage of the high
307 intrinsic contrast in optical absorption between blood clots and other tissues, the treatment effect
308 is limited to blood clots, and unwanted damage to the surrounding tissues can be minimized
309 (shown in Fig. 7). The blood vessel wall, which has a significantly lower optical absorption than
310 the blood clot at the laser's wavelength, is unlikely to be harmed during USELT treatment. Note
311 that USELT is based on the synergistic effect between the light pulse and the ultrasound burst.
312 Strong cavitation will only be induced at the location where laser and ultrasound energy overlap
313 and are properly synchronized³⁰. Due to the low applied ultrasound energy intensity, ultrasound-
314 alone is not capable of producing cavitation on the vessel wall and surrounding tissues (shown in
315 Fig 3, US only; Fig 4, US only; Fig 5, (d), (e) and (f); Fig 6, US only; Fig 7, (a)). On the other
316 hand, due to the high optical absorption and scattering of blood, laser energy cannot effectively
317 penetrate the entire blood clots to damage the vessel wall or surrounding tissues when the
318 catheter is properly placed. As a result, the induced cavitation will be limited to the blood clots.
319 It gives USELT the potential to be highly selective, precise, and safe. The USELT is based on
320 our earlier developed PUT technology. The high selectivity, precision and safety of PUT
321 technology has been demonstrated in our previous studies³⁰⁻³³. Our current in vivo study
322 demonstrated that USELT utilizing laser pulses with a fluence of 8 mJ/cm² at 532-nm
323 wavelength and ultrasound bursts with a peak negative pressure of 1.3 MPa at 0.5 MHz were
324 effective for thrombolysis (shown in Fig. 3; Fig. 5; Fig. 6). The 8 mJ/cm² fluence is extremely
325 low compared to the laser fluence used in traditional laser therapy which generally requires laser
326 fluence greater than 1 J/cm²⁴¹. The 1.3 MPa ultrasound at 0.5 MHz is also far below the
327 cavitation threshold (~4 MPa) reported in the literature⁴², and results in a Mechanical Index of
328 1.8, which is below the FDA safety limit of 1.9 for ultrasound imaging.

329 The second potential advantage is that USELT dissolves blood clots through the
330 mechanical effect of cavitation with minimal temperature rise. The cavitation in USELT is from
331 the pulsed laser induced photoacoustic (PA) effect^{33,43-45}. We have selected short-duration

332 ultrasound pulses (5 cycles) and laser pulses (2-ns) to minimized temperature increase and
333 maximize the mechanical effect of cavitation. Unlike thermal-based therapy, which can produce
334 damage in surrounding tissue due to thermal diffusion, the mechanical effect of cavitation is
335 precisely localized. Only the tissues next to the produced cavitation are affected, while no
336 surrounding tissue is damaged (shown in Fig. 7) ⁴⁶⁻⁵⁰.

337 The third potential advantage is that USELT is highly efficient by combining the
338 advantages of ultrasound and laser thrombolysis techniques. Laser thrombolysis can easily
339 induce cavitation but the collapse of cavitation is not sufficiently strong because of the lack of
340 driving force, whereas ultrasound can induce strong collapse of cavitation but requires strong
341 power to initiate cavitation. Combination of laser and ultrasound can easily produce cavitation in
342 the blood clot and the collapse of cavitation will be driven by ultrasound to achieve highly
343 efficient thrombolysis.

344 Technically, USELT also dissolves a blood clot through mechanical force. The
345 mechanical force is produced by the induced micro or nano-size bubbles in the blood clot. A big
346 advantage of USELT is that the produced mechanical force is not necessary to be exerted on the
347 blood vessel wall. Hence the damage to the blood vessel wall is minimized. Many traditional
348 mechanical thrombectomy devices generally exert a force on the inside of the blood vessel wall
349 to “scrape” a blood clot off. The inside surface of a vein has venous valves to prevent the
350 backflow of blood. Scaping off a blood clot inside a vein always has the potential to damage the
351 venous valvular function. With USELT, the mechanical force produced by cavitation is based on
352 optical absorption of the blood clot. One can always select an optical wavelength that is highly
353 absorbed by the blood clot, but less absorbed by the venous valve and vessel wall during USELT
354 to minimize the unwanted damage.

355 A serious limitation of noninvasive PUT was that laser wavelengths were limited to those
356 with adequate transparency in the tissues between the emitter and the clot, and due to high
357 scattering and non-targeted absorbance, treatments were limited to tissue depths of a few
358 millimeters. USELT retains the advantages of endovascular laser thrombolysis. During USELT,
359 laser light can be delivered to the blood clot using an optical fiber as the same matter for
360 endovascular laser therapy, while ultrasound can be applied noninvasively. One major advantage

361 of endovascular laser thrombolysis is that the size of a laser fiber can be very small, for example,
362 100- μ m in diameter (as shown in Fig 1 (c), (d)). This small size provides great flexibility for
363 endovascular laser therapy and allow it to be used to recanalize small blood vessels, a huge
364 advantage when it is used to dissolve blood clots during stroke therapy. On the other side, if
365 needed, multiple laser fibers can be bundled together to treat blood clots in large size vessels.
366 Since laser energy is delivered via the laser fiber and not by transmission through intervening
367 tissues, USELT treatment sites are limited only by endovascular catheter access.

368 Although the initial outcome is promising, the USELT technology needs further
369 improvement. One important next step is to incorporate an imaging technique with USELT for
370 image-guided intervention. Our current study showed blood clot residuals in blood vessels. The
371 reason for these residuals is more likely because the blood clot was not treated completely during
372 USELT. An imaging technique that can precisely locate blood clot and assess the size of blood
373 clot residuals after USELT could significantly improve the effectiveness of USELT. Potential
374 imaging modalities for guiding USELT include ultrasound imaging and photoacoustic imaging,
375 two imaging modalities that are complementary to each other and can potentially share same
376 equipment with USELT ³⁴.

377 In addition, as a key step toward future commercialization success, detailed efficacy and
378 safety studies should be performed beyond the current feasibility study. Particularly, vessel
379 injury, persisting thrombotic attachments to the wall and possible complications need to be
380 further investigated. We believe a large animal model such as a porcine model will be better
381 suitable for such future study given its similar size to human. Upon the completion of safety and
382 efficacy studies, comparisons can be made between USELT and established thrombectomy
383 devices such as Angio-Jet to demonstrate the pros and cons of USELT's potential in the clinics.

384 In summary, the combination of different but complementary therapeutic techniques
385 represents a major trend in recent biomedical research. The combined treatment is likely to
386 overcome the limitations associated with individual techniques and, therefore, has a better
387 chance to achieve improved treatment outcomes. The current study is the first attempt to
388 combine laser and ultrasound for removing blood clots in vivo. It is an excellent example of
389 combined therapy, considering that the two energy types are different but complementary in the

390 cavitation mechanism. By synergistically combining laser and ultrasound, optimized ablation
391 with reduced side effects becomes possible, shedding new light on clinical management of DVT
392 and stroke.

393 **Conclusion**

394 This current study demonstrated that USELT was effective in treating thrombolysis,
395 whereas the low-fluence laser alone or low-pressure ultrasound alone was not able to dissolve
396 blood clots in an in vivo rabbit model. Hence the feasibility of the USELT technology was
397 demonstrated. In short, USELT have the potential to treat the DVT with high efficiency and
398 with minimal effect on nearby tissues. In future, image-guided USELT should be developed to
399 further improve the efficiency and facilitate clinical translation for thrombolysis.

400

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402

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405 University of Kansas for sectioning vein tissue.

406

407 **Conflict of Statement**

408 We have no conflict of statement to disclose.

409

410 **Data Availability**

411 The data that support the findings of this study are available from the corresponding author upon
412 reasonable request.

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Figure Legends:

- (1.) Fig. 1. (a) Detailed schematic of the integrated endovascular laser thrombolysis system with high-intensity focused ultrasound system for the treatment. (b) Photograph of the rabbit's jugular vein being treated. (c) Schematic of the rabbit vein irradiation with laser (fiber optic cannula) and ultrasound (HIFU transducer), which is the boxed region in (a). (d) Detailed schematic of the combined ultrasound and laser irradiation inside rabbit vein resulting in blood clot dissolution, which is boxed region in (c).
- (2.) Fig. 2. (a) Location of the blood clot in the isolated rabbit's right jugular vein. (b) Ultrasound doppler imaging before formation of blood clot in jugular vein. (c) Ultrasound doppler imaging after formation of blood clot in jugular vein.
- (3.) Fig. 3. Total number of rabbits for full recovery, partial recovery and poorly/no recovery within each treatment group. Number placed above the column bar shows the total number of rabbits. This figure includes data from all the three groups (I, II and III). USELT = ultrasound-assisted endovascular laser thrombolysis; US = ultrasound; L = laser.
- (4.) Fig. 4. Percentage of the maximum blood flow speed recovered in full recovery, partial recovery, and poor/no recovery groups that were shown in Fig. 3 within each treatment group. Number placed above the column bar shows the mean blood flow recovery after the treatment. This figure includes data from all the three groups (I, II and III). USELT = ultrasound-assisted endovascular laser thrombolysis; US = ultrasound; L = laser.
- (5.) Fig. 5. Ultrasound doppler imaging of the treated jugular vein, jugular vein is marked by arrow. (a), (b), (c) are from rabbit No. 8, which was treated by USELT, (d), (e), (f) are from rabbit No. 5, which was treated by ultrasound-only, and (g), (h), (i) are from rabbit No. 6, which was treated by laser-only. (a), (d), (g) Ultrasound doppler imaging before formation of blood clot in jugular vein. (b), (e), (h) Ultrasound doppler imaging after formation of blood clot. (c) Ultrasound doppler imaging after treatment with USELT using P- of 1.3 MPa and laser fluence of 8 mJ/cm². (f) Ultrasound doppler imaging after treatment with ultrasound only using P- of 1.3 MPa. (i) Ultrasound doppler imaging after treatment with laser only using laser fluence of 8 mJ/cm². P- = peak negative ultrasound pressure. Scale bar = 5mm
- (6.) Fig. 6. Maximum blood flow speed in rabbit jugular vein measured with ultrasound doppler imaging before blood clot formation, after blood clot formation, after treatment with USELT using P- of 1.3 MPa and laser fluence of 8 mJ/cm², after treatment with ultrasound-only using P- of 1.3 MPa or 2 MPa and after treatment with laser-only using laser fluence of 8 mJ/cm². Maximum blood flow speed Before BC & After BC was significant (p<0.001); After BC & USELT was significant (p<0.001); USELT & US only was significant

($p < 0.05$); USELT & Laser only was significant ($p < 0.05$); USELT & Before BC was not significant ($p < 0.33$). P- = peak negative ultrasound pressure; BC = blood clot; US = ultrasound.

(7.) Fig. 7. (a) Histology image (hematoxylin and eosin stain) of vein section treated with ultrasound-only using P- of 1.3 MPa. (b), (c), (d) Histology images (hematoxylin and eosin stain) of vein sections treated with USELT using P- of 1 MPa and laser fluence of 12 mJ/cm². (e) Magnified image of the highlighted area in the red box in (c). P- = peak negative ultrasound pressure.

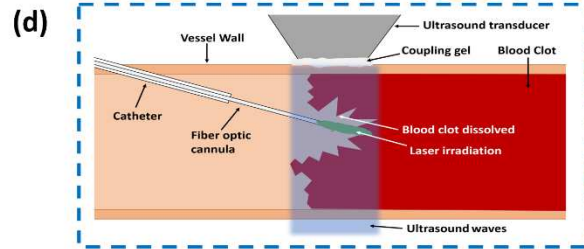
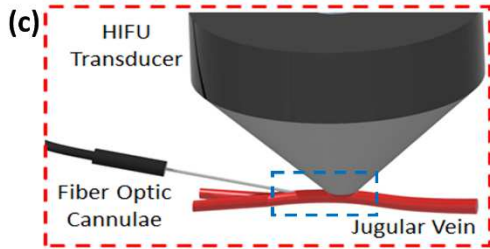
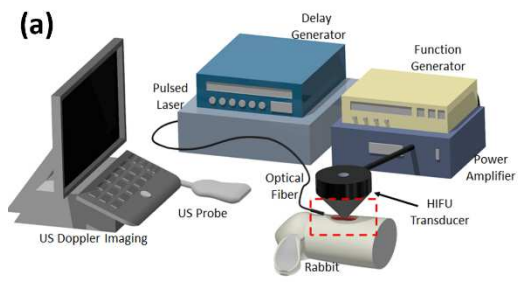
(8.) Fig S1. Heart Rate of rabbits in group I treated with USELT. HR = heart rate, R = rabbit number, ◆ = treatment started, ■ = treatment completed.

(9.) Fig S2. Respiratory Rate of rabbits in group I treated with USELT. HR = heart rate, R = rabbit number, ◆ = treatment started, ■ = treatment completed.

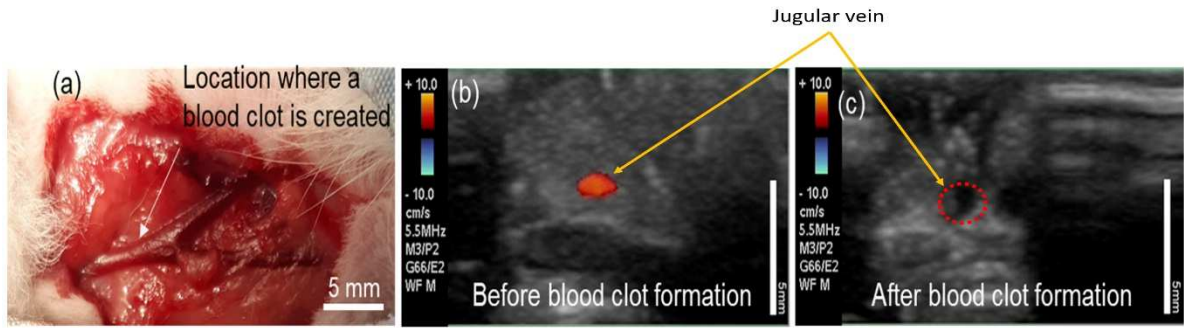
Table 1: Treatment data of entire experiment containing seventeen rabbits using different ultrasound and

Group	No. of Rabbits	Rabbit Number	Peak Negative Ultrasound Pressure (MPa)	Laser Fluence (mJ/cm²)
I	7	4, 7, 8, 9, 10, 11, 12	1.3	8
II	5	5, 17, 19, 13, 14	1.3 or 2	0
III	5	6, 15, 16, 18, 20	0	8

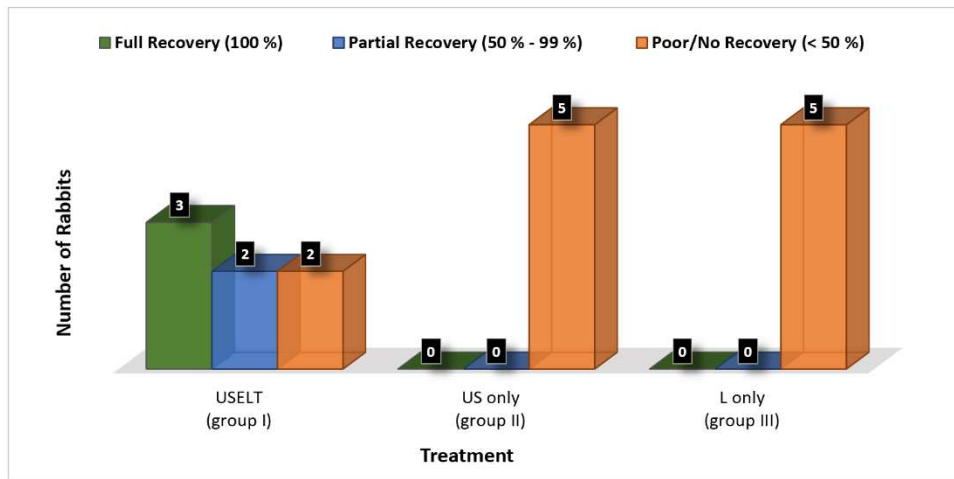
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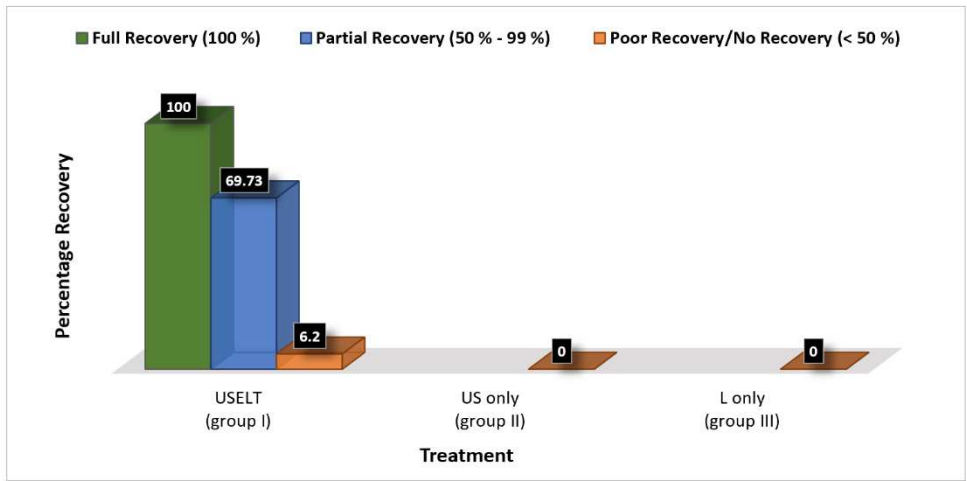
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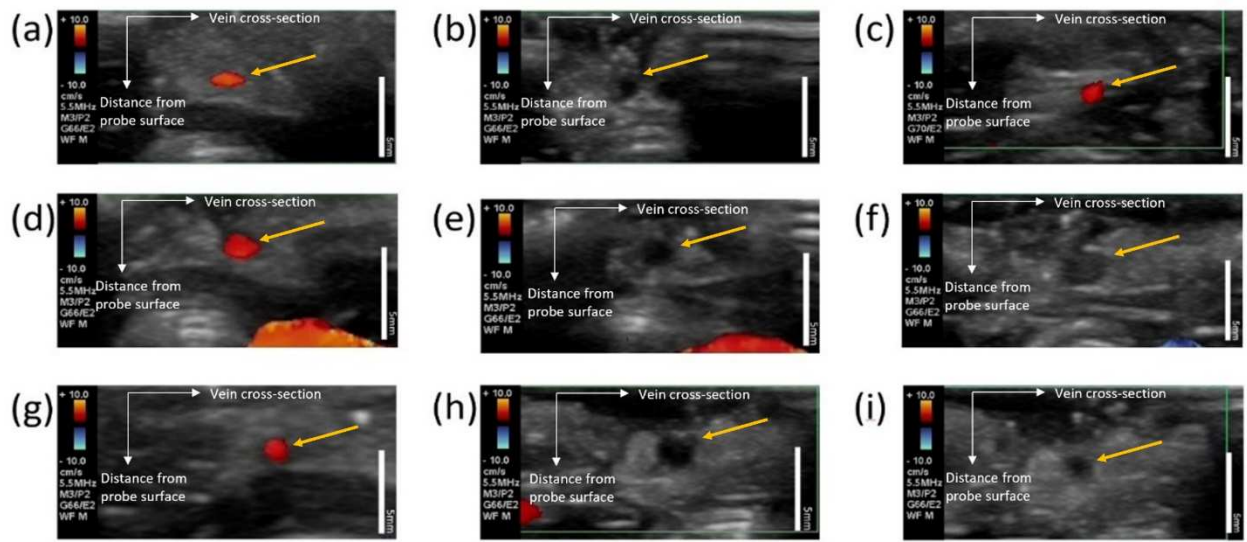
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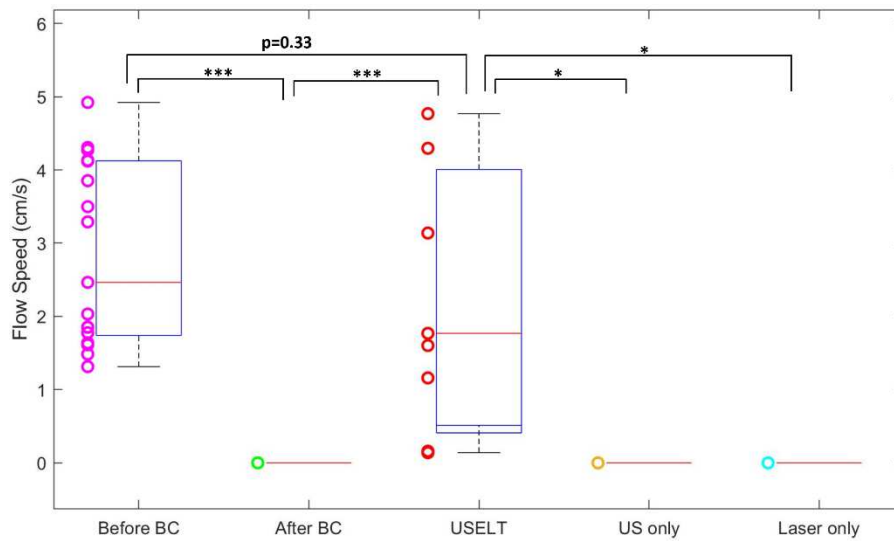
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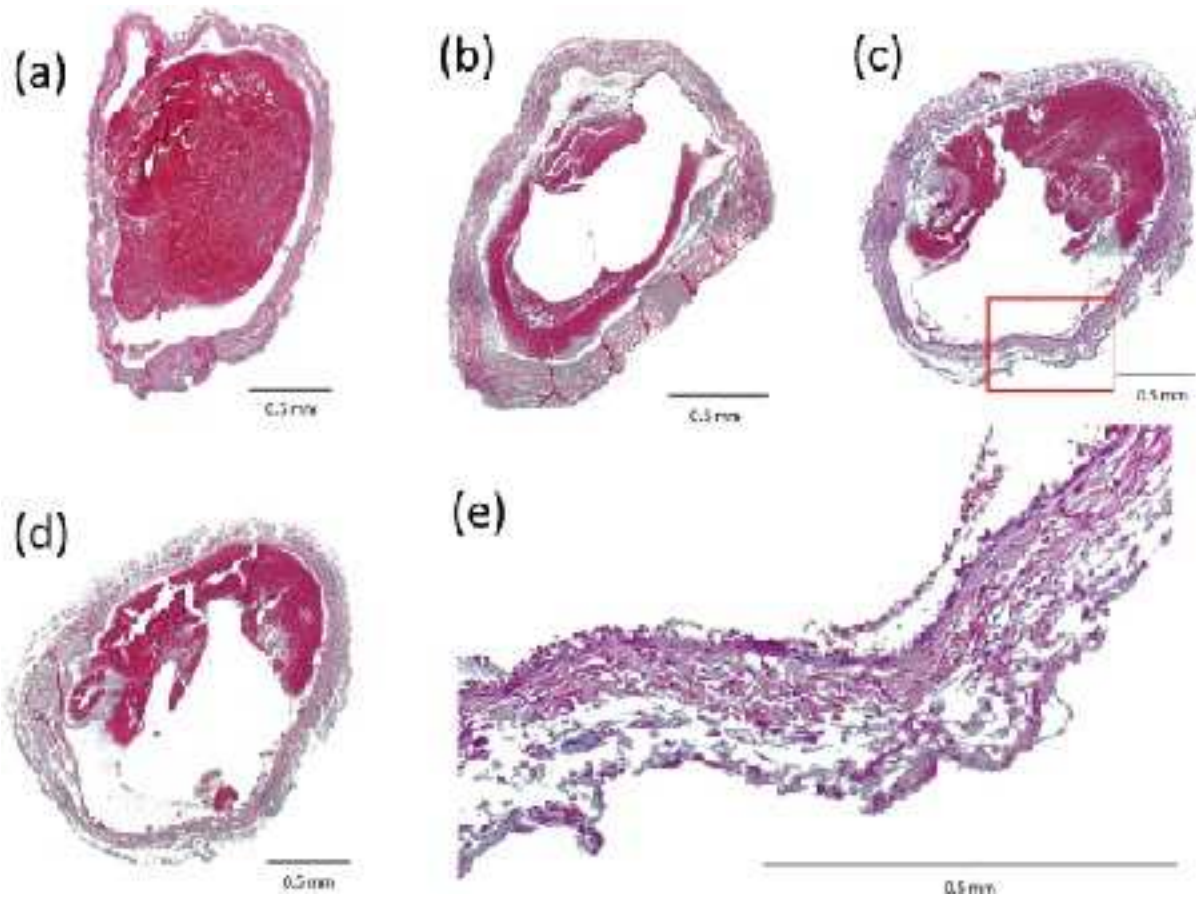
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