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Article type : Mini-Review

Analysis Approaches to Identify Pharmacogenetic Associations with Pharmacodynamics

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42 Abstract

43 Pharmacogenetics (PGx) seeks to enable selection of the right dose of the right drug for each patient to
44 optimize therapeutic outcomes. Most PGx focuses on pharmacokinetics (PK), due to our relatively
45 advanced understanding of the genes involved in PK and the causative effects of variants in those genes.
46 Genetic variants can also affect pharmacodynamics (PD), but relatively few PGx-PD associations have
47 been identified. This is partially due to a more limited understanding of the relevant genes and the
48 consequences of genetic variation, but is also due in part to the potential confounding of PK variability in
49 assessments of clinical outcomes that have a contribution from both PK and PD. For example, it is
50 challenging to confirm the effect of mu opioid receptor (*OPRM1*) genetic variation on opioid response
51 due to the contribution of *CYP2D6* genotype to bioactivation of some opioid drugs (i.e., codeine and
52 tramadol). The objectives of this mini-review are to describe several recent efforts to discover and
53 validate PGx-PD that disentangle the influence of PK variability and propose potential approaches that
54 could be used in future PGx-PD analyses. We use the effect of *OPRM1* genetics on opioid response to
55 illustrate how these analyses could be conducted and conclude by discussing how PGx-PD could be
56 translated into clinical practice to improve therapeutic outcomes.

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60 Pharmacogenetics of Pharmacodynamic Drug Response (PGx-PD)

61 The promise of pharmacogenetics (PGx) is the ability to select the right dose of the right drug for each
62 patient. This idea acknowledges that each patient is unique, and optimal treatment should incorporate
63 those factors that define the patient as a unique individual rather than assuming that the population
64 mean or median sufficiently represents the patient¹. Most of the work in PGx and personalized medicine
65 has focused on pharmacokinetics (PK) as the phenotype of interest, including individualized dosing to
66 achieve target drug exposures.

67 The focus on PGx of PK (PGx-PK) is due to several factors. PGx-PK builds upon substantial
68 understanding of drug PK, including knowledge of the specific enzymes and transporters primarily
69 responsible for the absorption, distribution, metabolism, and excretion of individual drugs. It also builds
70 on substantial work to identify functional variants in the genes coding for these enzymes and
71 transporters and the translation of genotypes to predicted activity phenotypes². PK data are relatively
72 easy to collect and measure, and provide a sensitive, quantitative phenotypic endpoint for PGx-PK
73 analyses, with the caveat that often systemic PK is measured, and this may not accurately reflect PK at
74 the target site. Clinical translation is relatively straightforward; adjustment of dosing reduces PK
75 variability across PGx-PK groups, or substitution of an alternate agent with a different metabolic
76 pathway may avoid inefficacy or toxicity. There are many examples of this approach, including recent
77 guidelines for dosing tacrolimus based on *CYP3A5* genotype³, or avoiding codeine in individuals with
78 extreme *CYP2D6* genetic phenotypes⁴, including poor or ultrarapid metabolizers.

79 Despite the potential for genetic variants to also affect drug sensitivity, or pharmacodynamics
80 (PD), there are relatively few established PGx-PD associations. Genetic variants may affect the
81 expression, function, occupancy, or activation of a drug target, among many other possible biological
82 mechanisms⁵. The clinical consequence of PGx-PD is that a systemic exposure within the desired range
83 may not necessarily elicit the desired response if genetic variation results in a drug target that is non-
84 functional or not expressed to an appreciable extent (putting the patient at risk for an off-target or
85 noxious on-target response). There are several reasons for the relative paucity of validated PGx-PD
86 effects, including incomplete understanding of candidate PD genes, incomplete knowledge of the
87 functional effects of variants within those genes, lack of well-phenotyped PD endpoints, variable efficacy

88 endpoints for different indications of the same drug, and perhaps smaller effects from many PD genes
89 and variants, similar to the genetics of complex diseases.

90 Genome-wide association studies (GWAS) have identified near monogenic PGx-PD associations
91 of genes that were unlikely to have been selected for candidate genetic studies⁶. To date, these striking
92 PGx-PD associations have been primarily observed in PK-independent outcomes, such as the
93 associations for *HLA* genes with drug-induced hypersensitivity⁶ or *CACNA1A/RYR1* with malignant
94 hyperthermia⁷. Unlike these strongly penetrant genetic associations, most clinical outcomes are
95 multifactorial, including a contribution from both PK and PD^{5,8} (**Figure 1**). There has been limited success
96 identifying these PGx-PD associations, partially due to the confounding effects of PK in the analysis. For
97 example, there is evidence that genetic variation in the mu opioid receptor (*OPRM1*) is associated with
98 response to opioid analgesics, but this association has been difficult to validate due to the confounding
99 of variability in morphine systemic exposure.⁴

100 The objective of this mini-review is to describe potential approaches to deconvolute the
101 confounding effects of PK to isolate PGx-PD effects. We illustrate these approaches using recent efforts
102 to identify PGx-PD and by returning to the example of the putative association of *OPRM1* genetics on
103 opioid response. We conclude by describing what is needed to advance PGx-PD research and integrate
104 PGx-PD into individualized treatment to improve therapeutic outcomes.

105 Approaches to Identifying PGx-PD Associations

106 Drugs Dosed to Standardize Target Concentrations

107 For some drugs, particularly those with narrow therapeutic windows, clinical testing of drug
108 concentrations is pursued, and dosing is adjusted to achieve a target concentration, commonly referred
109 to as therapeutic drug monitoring (TDM). By adjusting the dose to achieve a target concentration, such
110 as a maximum or minimum concentration (e.g., C_{max} or C_{min}), TDM can enhance efficacy and/or reduce
111 toxicity. Tacrolimus, some antibiotics, and some anti-epileptics undergo TDM per routine. Effective use
112 of TDM minimizes variability in exposure (at least as indicated by sampling of blood) and reduces the
113 contribution of exposure variability in the analysis of treatment outcomes. This strategy was used to
114 identify the PGx-PD association between variants in the *KCNQ1* gene (encoding the pore-forming
115 subunit of the voltage-gated potassium channel, KvLQT1) and new-onset post-transplant diabetes
116 mellitus in patients treated with tacrolimus⁹.

117 There are several limitations to this approach. One is that the TDM metric used to individualize
118 dosing (i.e., C_{min}) is unlikely to capture all individual variability in PK and may not be the primary
119 determinant of the outcome of interest. For example, for PD outcomes such as nephrotoxicity or
120 efficacy, C_{max} or systemic exposure defined as area under the concentration-time curve (AUC) may be
121 more relevant than C_{min} (trough), but these data may not be available. Additionally, the steepness of the
122 concentration-response slope may be an important consideration. It is also required that the TDM
123 measurements are available for a time relevant to the PD outcome. For example, a GWAS of tacrolimus-
124 exposed individuals was unable to include drug levels as a covariate in the analysis of nephrotoxicity
125 because data were not available at the same time as renal function measurements¹².

126 Since TDM is only used for specific drugs, the use of clinically obtained drug concentrations is
127 relevant to a limited number of medications. There is evidence that TDM may be beneficial for a larger
128 number of medications, even including broad therapeutic index drugs. A *CYP2D6* genotype-stratified PK
129 study of atomoxetine revealed a 50-fold range in systemic exposure in patients receiving standard
130 dosing, and suggested that even maximal recommended dosing would fail to achieve target exposure
131 in a substantial proportion of *CYP2D6* normal metabolizers (NMs)¹⁰. A recently published CPIC guideline
132 recommends checking the atomoxetine concentration in patients exhibiting inadequate clinical
133 response to inform subsequent therapeutic decisions¹¹. Expanding TDM to more agents could have
134 direct clinical benefit while improving the identification of PGx-PD associations.

135 For the example of *OPRM1*, since TDM is not used to guide codeine dosing, codeine or morphine
136 metabolite data are not readily available to investigate PGx-PD effects for *OPRM1*. A concentration-
137 controlled clinical trial¹³, in which patients are randomly assigned to receive personalized codeine dosing
138 to achieve one of several pre-specified morphine exposure levels, could be a possible alternate source of
139 data that is similar to a TDM situation, with which PGx-PD analyses could be conducted without the
140 confounding of PK.

141 Investigate PGx-PD Associations by Adjusting for PK

142 For drugs that do not undergo TDM, there are several analytical approaches to reduce the contribution
143 of PK variability to investigate PGx-PD associations. One straightforward approach that does not require
144 measured drug concentrations is to conduct the PGx-PD analysis within a PGx-PK stratum. For instance,
145 investigating the association of *OPRM1* genotype with analgesic response to *CYP2D6* substrates, such as
146 codeine or tramadol, in only *CYP2D6* normal metabolizers. This approach will reduce the contribution of
147 PK variability but will not eliminate it completely as *CYP2D6* activity can vary several-fold within

148 individuals with the same genotype¹⁴. An extension of this approach is to conduct the analysis within
149 each CYP2D6 metabolic phenotype strata or adjust for metabolic phenotype.

150 When concentrations measurements are available, the simplest approach is to adjust for measured drug
151 concentrations in the analysis. For example, nephrotoxicity due to vancomycin is partially determined by
152 vancomycin trough concentrations. In order to identify PGx-PD associations, a GWAS of nephrotoxicity
153 adjusted for vancomycin trough concentrations, which enabled identification of a variant near the *GJA1*
154 gene (encoding connexin43)¹⁵. Applying this approach *OPRM1*, an analysis of the association of *OPRM1*
155 genotype with analgesia that adjusted for measured morphine concentrations during codeine treatment
156 could substantially reduce the contribution of PK variability.

157 Demonstrate PK-outcome Association Stratified by PGx-PD

158 PK-outcomes associations can be detected using standard statistical approaches such as regression
159 models. However, these associations can be confounded by PGx-PD effects, which can lead to an
160 inability to detect the PK-outcome association in pooled patients. If the PK-outcome association is
161 revealed by stratifying the cohort by the PGx-PD genotype, this provides evidence that the genotype is
162 contributing to the outcome of interest. Importantly, this type of stratified analysis can accommodate
163 PGx-PD effects that invert the PK-outcome association (**Figure 2**), as has been recently reported for
164 paroxetine and genotype of the paroxetine drug target *SLC6A4*¹⁶. In patients with genetic variants
165 associated with low *SLC6A4* expression, patients with lower plasma concentrations had better clinical
166 improvement than patients with higher concentrations, potentially due to target saturation and off-
167 target effects. In patients with variants associated with high *SLC6A4* expression, the response improved
168 with higher blood concentration. This inverse association within each genotype group would be difficult
169 to detect using any of the other strategies proposed in this commentary. In terms of *OPRM1*, at least
170 one study has analyzed the association between opioid concentration and analgesic response within
171 *OPRM1* genotype groups¹⁷.

172 Introduce PGx-PD into PK-outcomes Model

173 Another somewhat related approach is to first establish the PK-outcomes model and then introduce the
174 PGx-PD variable. Multivariable models retain only variables that explain residual variability in the
175 outcome of interest. Including measured PK in a model accounts for the contribution of PK variability,
176 and the residual variability will be predominantly contributed by PD (**Figure 3**). This approach was
177 recently used in a PGx-PD analysis of paclitaxel-induced peripheral neuropathy. First, a model was
178 created that included systemic paclitaxel PK and other clinical variables that were associated with the

179 risk of neuropathy¹⁸. Then genes involved in hereditary neuropathy that had previously been reported
180 to increase risk of paclitaxel-induced neuropathy were investigated, including variants in *EPHA5*.
181 Introducing those variants into the neuropathy-prediction model demonstrated that these genotypes
182 affect a patient's neuropathy sensitivity after accounting for variability in cumulative paclitaxel
183 exposure¹⁹. Importantly, a post-hoc analysis confirmed that this association for *EPHA5* would not have
184 been detected without including measured paclitaxel PK in the multivariable model. In the case of
185 *OPRM1*, this approach would be attempted by first modeling the morphine-analgesia association and
186 then adding *OPRM1* genotype as a covariate in the model to see if it explains residual variability in the
187 resulting analgesic effect.

188 Incorporation of Genetics in Pharmacometric Models

189 The previously described multivariable statistical approaches are empirical, simpler approaches for
190 investigators who do not have expertise in population pharmacokinetic-pharmacodynamic (popPKPD)
191 modeling. The ideal methods to investigate PGx-PD effects are likely to develop popPKPD or possibly
192 physiologically-based PKPD (PBPKPD) models. PopPKPD models²⁰ are typically used to understand the
193 relationship between drug concentration and PD response by accounting for the variability in PK and PD
194 parameters from covariates. Traditionally, these covariates are clinical variables that affect PK or PD
195 parameters, though it has become increasingly common to investigate PGx factors affecting PK
196 parameters (i.e., drug clearance). Similarly, genetic factors of PD response could be explored in a
197 popPKPD model to understand variability in PD parameters, including maximal drug effect (E_{max}) or
198 potency (EC_{50}). This approach was recently used within a study of the effectiveness of buprenorphine for
199 reducing illicit opioid use²¹. The base PKPD model identified the buprenorphine EC_{50} for successful opioid
200 abstinence. One of the covariates associated with this EC_{50} was the rs678849 genotype of the delta-
201 opioid receptor (*OPRD1*). A very similar approach could be used for *OPRM1* by building a popPKPD
202 model that relates morphine exposure to analgesia, and then investigating the *OPRM1* genotype as a
203 covariate on the analgesic response parameters E_{max} or EC_{50} .

204 On the other hand, PBPKPD models²² provide a mechanistic representation of the drug in the biological
205 system by explicitly considering the organs and tissues to estimate drug concentrations within each
206 tissue. Thus, PBPKPD models can potentially link target site concentrations to the PD response and any
207 PGx factors affecting the PD response, for instance the binding of the drug to its target site, could be
208 modeled. In the *OPRM1* example, the PBPKPD model could be built in which the concentrations of

209 morphine in the brain could be estimated, followed by modeling of the binding of morphine to OPRM1
210 and then investigating the effects of *OPRM1* genotype on this binding and the resulting analgesic effect.

211 Clinical Translation and Recommendations for Future Work

212 Integrating PGx-PD into Precision Treatment

213 As mentioned earlier, the clinical translation of PGx-PK by adjusting doses according to PGx-PK genotype
214 to standardize drug concentration is relatively straightforward. In this sense, “individualized treatment”
215 means stratifying dosing so all patients achieve the same exposure. PGx-PD is somewhat more complex
216 to translate into clinical practice since it implies that the optimal exposure level for each patient is
217 distinct. The appropriate dosing patients with each PD-PGx genotype is a function of the direction of
218 effect (i.e., sensitive vs. resistant) of that genotype and the relevant clinical outcome (i.e., efficacy
219 and/or toxicity) (**Figure 4**). Patients who are “sensitive” to the therapeutic effects of a drug may achieve
220 greater benefit at typical levels of exposure; this may enable downward titration of the exposure to
221 reduce risk of toxicity (or maintaining typical exposure to enhance efficacy without increasing toxicity).
222 Patients who are “sensitive” to drug toxicity cannot tolerate typical exposure and require reduced
223 exposure, which may reduce efficacy. Toxicity “resistant” patients can tolerate higher exposure, which
224 could allow for upward titration of exposure to enhance efficacy or maintaining exposure to maintain
225 efficacy, potentially with less toxicity. Finally, patients “resistant” to therapeutic effects will require
226 higher exposure or may not be able to achieve therapeutic response at any tolerable exposure level. The
227 sensitivity/resistance to therapeutic effects and toxicity may be linked, in which case proper titration
228 could yield the typical balance of efficacy and toxicity, or may be independent, based on the biologic
229 mechanism. Importantly, these situations demonstrate the complexity of translating PGx-PD into clinical
230 practice by individualizing dosing so each patient achieves the exposure that optimizes their clinical
231 outcomes.

232 Conclusion and Future Directions for Research and Practice

233 We have described several approaches for reducing confounding by PK to assist with identifying PGx-PD
234 effects for multi-factorial clinical outcomes. Other challenges mentioned earlier, such as the limited
235 understanding of the genes responsible for PD effects and the consequences of genetic variation in
236 those genes and the lack of well-phenotyped PD endpoints, require additional consideration and
237 investigation but are beyond the scope of this mini-review. In addition to a general recognition of the
238 challenges with PGx-PD, there are several other initiatives that would improve our ability to conduct the

239 analyses described within this mini-review. First, a greater effort is needed to collect samples for PK
240 analysis, as this is the most direct way to account for PK variability in PGx-PD studies. One highly
241 efficient potential approach is to collect scavenged samples, which reduces the cost and some of the
242 regulatory issues around PK sampling and is especially beneficial in patients who are difficult to sample
243 including neonates, children, and the elderly²³. PGx-PD analyses would also benefit from development of
244 more precise biomarkers defining clinically relevant outcomes. These analyses also require further
245 development of modeling approaches that integrate PGx-PD analyses, perhaps including simulation
246 approaches to determine the optimal exposure and necessary dosing for patients based on PD
247 genotype. Finally, clinical translational researchers will likely need to develop prospective study designs
248 to demonstrate the clinical utility of individualized treatment based on PGx-PD effects²⁴. One possible
249 study design would be a variation of concentration-controlled clinical trials, PGx-PD stratified studies, in
250 which participants are genotyped for the drug target of interest. In such a study design a
251 pharmacometric model is used to individualize drug doses to achieve a target exposure (C_{max} or AUC);
252 inadequate therapeutic response at the initial exposure level can be followed with an increase in
253 exposure and re-assessment of therapeutic response, allowing for exposure-response relationships to
254 be established for each drug target genotype. This approach is analogous to genetics-stratified dose
255 escalation studies that have been used to validate PGx-PK effects in oncology²⁵. A more concerted effort
256 to discover and validate PGx-PD effects will someday usher in a new era of personalized treatment in
257 which patients are dosed to achieve their personalized target concentration, improving therapeutic
258 outcomes, and realizing the promise of PGx.

259 References:

- 260 **1.** Leeder JS. Who Believes They Are "Just Average": Informing the Treatment of Individual Patients
261 Using Population Data. *Clin Pharmacol Ther.* 2019;106:939-941. doi: 910.1002/cpt.1612. Epub
262 2019 Sep 1011.
- 263 **2.** Caudle KE, Sangkuhl K, Whirl-Carrillo M, et al. Standardizing CYP2D6 Genotype to Phenotype
264 Translation: Consensus Recommendations from the Clinical Pharmacogenetics Implementation
265 Consortium and Dutch Pharmacogenetics Working Group. *Clin Transl Sci.* 2020;13:116-124. doi:
266 110.1111/cts.12692. Epub 12019 Oct 12624.
- 267 **3.** Birdwell KA, Decker B, Barbarino JM, et al. Clinical Pharmacogenetics Implementation
268 Consortium (CPIC) Guidelines for CYP3A5 Genotype and Tacrolimus Dosing. *Clin Pharmacol Ther.*
269 2015;98:19-24. doi: 10.1002/cpt.1113. Epub 2015 Jun 1003.

- 270 **4.** Crews KR, Monte AA, Huddart R, et al. Clinical Pharmacogenetics Implementation Consortium
271 Guideline for CYP2D6, OPRM1, and COMT Genotypes and Select Opioid Therapy. *Clin Pharmacol*
272 *Ther.* 2021;2.
- 273 **5.** Hertz DL, McLeod HL. Using pharmacogene polymorphism panels to detect germline
274 pharmacodynamic markers in oncology. *Clinical cancer research : an official journal of the*
275 *American Association for Cancer Research.* 2014;20:2530-2540.
- 276 **6.** Daly AK. Using genome-wide association studies to identify genes important in serious adverse
277 drug reactions. *Annual Review of Pharmacology and Toxicology.* 2012;52:21-35.
- 278 **7.** Gonsalves SG, Dirksen RT, Sangkuhl K, et al. Clinical Pharmacogenetics Implementation
279 Consortium (CPIC) Guideline for the Use of Potent Volatile Anesthetic Agents and
280 Succinylcholine in the Context of RYR1 or CACNA1S Genotypes. *Clin Pharmacol Ther.*
281 2019;105:1338-1344. doi: 1310.1002/cpt.1319. Epub 2019 Jan 1324.
- 282 **8.** McLaughlin MJ, Wagner J, Shakhnovich V, Carleton B, Leeder JS. Considerations for
283 Implementing Precision Therapeutics for Children. *Clin Transl Sci.* 2019;12:140-150. doi:
284 110.1111/cts.12607. Epub 12019 Jan 12625.
- 285 **9.** Tavira B, Coto E, Díaz-Corte C, et al. KCNQ1 gene variants and risk of new-onset diabetes in
286 tacrolimus-treated renal-transplanted patients. *Clin Transplant.* 2011;25:E284-291. doi:
287 210.1111/j.1399-0012.2011.01417.x. Epub 02011 Mar 01411.
- 288 **10.** Brown JT, Abdel-Rahman SM, van Haandel L, Gaedigk A, Lin YS, Leeder JS. Single dose, CYP2D6
289 genotype-stratified pharmacokinetic study of atomoxetine in children with ADHD. *Clin*
290 *Pharmacol Ther.* 2016;99:642-650. doi: 610.1002/cpt.1319. Epub 2016 Jan 1012.
- 291 **11.** Brown JT, Bishop JR, Sangkuhl K, et al. Clinical Pharmacogenetics Implementation Consortium
292 Guideline for Cytochrome P450 (CYP)2D6 Genotype and Atomoxetine Therapy. *Clin Pharmacol*
293 *Ther.* 2019;106:94-102. doi: 110.1002/cpt.1409. Epub 2019 Apr 1013.
- 294 **12.** Oetting WS, Wu B, Schladt DP, et al. Genetic Variants Associated With Immunosuppressant
295 Pharmacokinetics and Adverse Effects in the DeKAF Genomics Genome-wide Association
296 Studies. *Transplantation.* 2019;103:1131-1139. doi: 1110.1097/TP.0000000000002625.

- 297 **13.** Peck C. The randomized concentration-controlled clinical trial (CCR) : an informationrich
298 alternative to the randomized placebo controlled clinical trial (PCT). *Clin. Pharmacol. Ther.*
299 1990;47:148.
- 300 **14.** Gaedigk A, Dinh JC, Jeong H, Prasad B, Leeder JS. Ten Years' Experience with the CYP2D6 Activity
301 Score: A Perspective on Future Investigations to Improve Clinical Predictions for Precision
302 Therapeutics. *J Pers Med.* 2018;8.
- 303 **15.** Van Driest SL, McGregor TL, Velez Edwards DR, et al. Genome-Wide Association Study of Serum
304 Creatinine Levels during Vancomycin Therapy. *PLoS One.* 2015;10:e0127791. doi:
305 0127710.0121371/journal.pone.0127791. eCollection 0122015.
- 306 **16.** Tomita T, Yasui-Furukori N, Nakagami T, et al. The influence of 5-HTTLPR genotype on the
307 association between the plasma concentration and therapeutic effect of paroxetine in patients
308 with major depressive disorder. *PLoS One.* 2014;9:e98099. doi:
309 98010.91371/journal.pone.0098099. eCollection 0092014.
- 310 **17.** Boswell MV, Stauble ME, Loyd GE, et al. The role of hydromorphone and OPRM1 in
311 postoperative pain relief with hydrocodone. *Pain Physician.* 2013;16:E227-235.
- 312 **18.** Hertz DL, Kidwell KM, Vangipuram K, et al. Paclitaxel Plasma Concentration After the First
313 Infusion Predicts Treatment-Limiting Peripheral Neuropathy. *Clin Cancer Res.* 2018.
- 314 **19.** Marcath LA, Kidwell KM, Vangipuram K, et al. Genetic variation in EPHA contributes to sensitivity
315 to paclitaxel-induced peripheral neuropathy. *Br J Clin Pharmacol.* 2020;86:880-890. doi:
316 810.1111/bcp.14192. Epub 12020 Feb 14194.
- 317 **20.** Upton RN, Mould DR. Basic concepts in population modeling, simulation, and model-based drug
318 development: part 3-introduction to pharmacodynamic modeling methods. *CPT*
319 *Pharmacometrics Syst Pharmacol.* 2014;3:e88. doi: 10.1038/psp.2013.1071.
- 320 **21.** Ngaimisi E, Gopalakrishnan M, Ivaturi V, Zhang W, Young M, Laffont CM. Exposure-response
321 analyses to support dosing recommendations for RBP-6000 buprenorphine monthly formulation
322 in subjects with opioid use disorder. *Journal of Pharmacokinetics and Pharmacodynamics.*
323 2017;44:S50.

- 324 **22.** Kuepfer L, Niederal C, Wendl T, et al. Applied Concepts in PBPK Modeling: How to Build a
325 PBPK/PD Model. *CPT Pharmacometrics Syst Pharmacol*. 2016;5:516-531. doi:
326 10.1002/psp1004.12134. Epub 2016 Oct 12119.
- 327 **23.** Van Driest SL, Marshall MD, Hachey B, et al. Pragmatic pharmacology: population
328 pharmacokinetic analysis of fentanyl using remnant samples from children after cardiac surgery.
329 *Br J Clin Pharmacol*. 2016;81:1165-1174. doi: 1110.1111/bcp.12903. Epub 2016 Apr 12915.
- 330 **24.** Diouf B, Evans WE. Pharmacogenomics of Vincristine-Induced Peripheral Neuropathy: Progress
331 Continues. *Clin Pharmacol Ther*. 2019;105:315-317.
- 332 **25.** Sharma MR, Joshi SS, Karrison TG, et al. A UGT1A1 genotype-guided dosing study of modified
333 FOLFIRINOX in previously untreated patients with advanced gastrointestinal malignancies.
334 *Cancer*. 2019;125:1629-1636.

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336 Figure Legends:

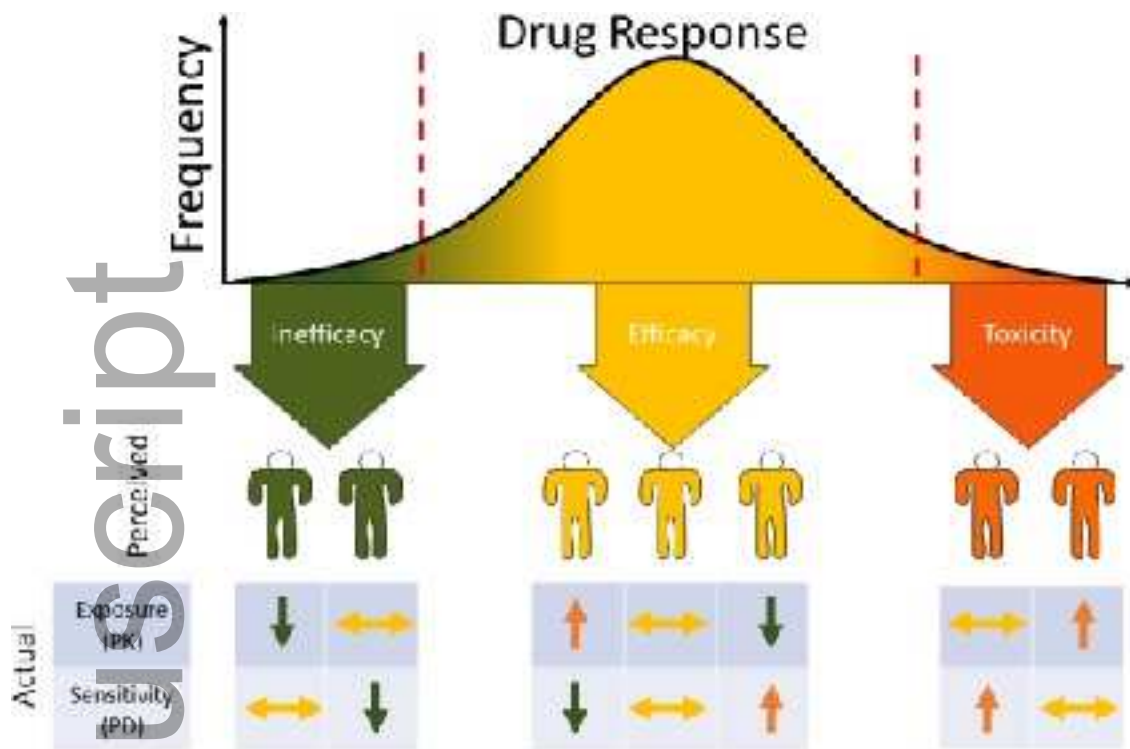
337 **Figure 1:** Drug response is a consequence of drug exposure (pharmacokinetics, PK) and sensitivity
338 (pharmacodynamics, PD). A patient who has ineffective treatment could be due to inadequate exposure
339 or reduced sensitivity (i.e., resistance). Similarly, a patient who experiences toxicity could be due to
340 supra-therapeutic exposure or enhanced sensitivity. Patients who experience efficacy without toxicity
341 could have normal exposure and sensitivity, or off-setting increased exposure and decreased sensitivity,
342 or vice-versa, that produce a typical response.

343 **Figure 2:** In the combined cohort there is no apparent association between drug exposure (PK) and
344 response. However, when the cohort is stratified by pharmacogenetic variant that impacts
345 pharmacodynamics (PGx-PD), there are inverse associations between exposure and response within
346 each PGx-PD genotype, as per the example in the text of paroxetine and *SLC6A4*.

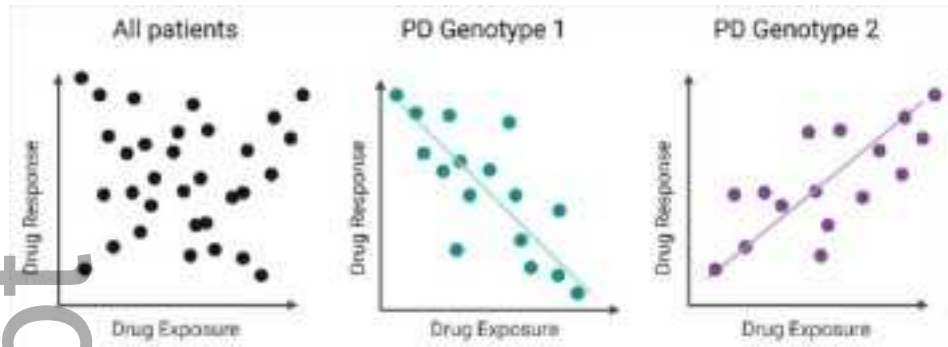
347 **Figure 3:** Pharmacogenetics of pharmacodynamics (PGx-PD) affects the patient's drug response at a
348 given exposure level. In this example, at a given drug exposure (solid vertical line) a patient with wild-
349 type PGx-PD would have near complete drug response (solid horizontal line). At that same exposure, a
350 patient carrying a single resistance allele would have a small response (dashed horizontal line) and a
351 patient with homozygous resistant genotype would have almost no drug response (dotted horizontal
352 line). The corollary is that achieving the same drug response in patients with different PGx-PD genotypes
353 requires different drug exposures.

354 **Figure 4:** Integrating pharmacogenetics of pharmacodynamics (PGx-PD) into clinical practice requires
355 adjusting dosing so that patients achieve the exposure that is consistent with their optimal treatment
356 outcomes. Patients can be “sensitive” (orange bodies) or “resistant” (dark green bodies) to efficacy
357 and/or toxicity. In each case, treating that patient with standard dosing will result in higher or lower
358 efficacy and/or toxicity than is typical. Depending on the PGx-PD genotype, a dose decrease or increase
359 could result in superior (blue), inferior (red), or similar (yellow) treatment outcomes.

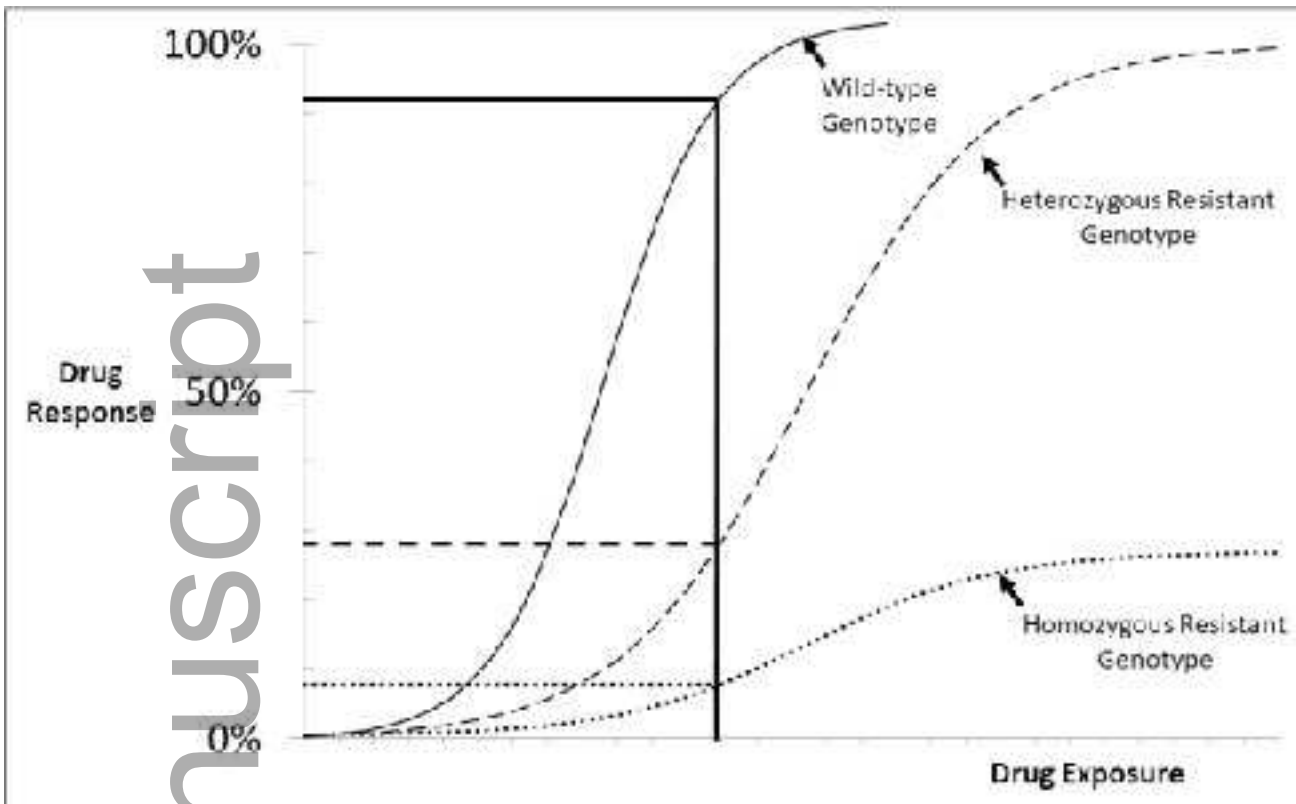
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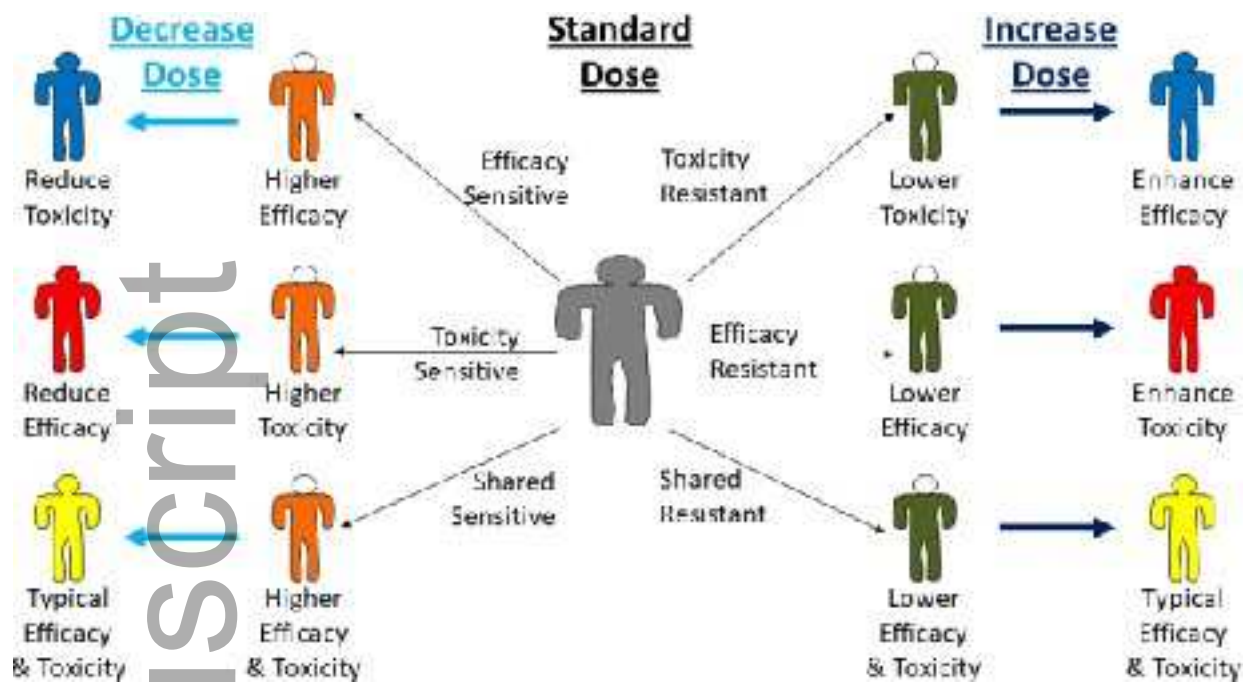
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