

1 **ARTICLE TITLE** Bioregions are predominantly climatic for fishes of northern lakes

2 **RUNNING HEAD** Bioregionalization of inland fishes

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21 **BIOSKETCH** Charlie Loewen is a community and macroecologist. His research takes a  
22 quantitative approach to understanding key processes and dynamics of freshwater ecosystems,  
23 with a focus on their response to compounding environmental changes.

24 **AUTHOR CONTRIBUTIONS** All authors contributed to data compilation and manuscript  
25 revision. CJGL led study conception and design, conducted the analyses, and composed the  
26 initial draft manuscript.

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9 **ARTICLE TITLE**

10 Bioregions are predominantly climatic for fishes of northern lakes

11 **RUNNING HEAD**

12 Bioregionalization of inland fishes

13 **ABSTRACT**

14 **Aim:** Recurrent species assemblages integrate important biotic interactions and joint responses  
15 to environmental and spatial filters that enable local coexistence. Here, we applied a bipartite  
16 (site-species) network approach to develop a natural typology of lakes sharing distinct fish  
17 faunas and provide a detailed, hierarchical view of their bioregions. We then compared the roles  
18 of key biogeographic factors to evaluate alternative hypotheses about how fish communities are  
19 assembled from the regional species pool.

20 **Location:** Ontario, Canada and the Upper Midwest, USA.

21 **Time period:** 1957–2017.

22 **Major taxa studied:** Freshwater fishes.

23 **Methods:** Bipartite modularity analysis was performed on 90 taxa from 10,016 inland lakes in  
24 the Southwestern Hudson Bay, Mississippi River, and St. Lawrence River drainages, uncovering

25 bioregionalization of North American fishes at a large, subcontinental scale. We then used a  
26 latent variable approach, pairing non-metric partial least-squares structural equation modelling  
27 with multiple logistic regression, to show differences in the biogeographic templates of each  
28 community type. Indicators of contemporary and historical connectivity, climate, and habitat  
29 constructs were estimated using a geographic information system.

30 **Results:** Fish assemblages reflected broad, overlapping patterns of postglacial colonization,  
31 climate, and geological setting, but community differentiation was most linked to temperature,  
32 precipitation, and, for certain groups, lake area and water quality. Bioregions were also marked  
33 by non-native species, showing broad-scale impacts of introductions to the Great Lakes and  
34 surrounding basins.

35 **Main conclusions:** The dominant effects of climate across broad spatial gradients indicate  
36 differing sensitivities of fish communities to rapidly accelerating climate change and  
37 opportunities for targeted conservation strategies. By assessing biological variation at the level of  
38 recurrent assemblages, we accounted for the non-stationarity of macroecological processes  
39 structuring different sets of species on the landscape, and thus offer novel inference on the  
40 assembly of inland fish communities.

#### 41 **KEYWORDS**

42 Bioregionalization, climate change adaptation, community assembly, conservation  
43 biogeography, environmental filtering, freshwater fishes, lake connectivity, latent variable  
44 approach, network modularity, species sorting

#### 45 **INTRODUCTION**

46 Species distributions, and their assembly into communities, reflect an array of contemporary and  
47 historical filters operating at multiple spatial scales (Jackson, Peres-Neto, & Olden, 2001; Dias et  
48 al., 2014; Mantyka-Pringle, Martin, Moffatt, Linker, & Rhodes, 2014; Ficetola, Mazel, &  
49 Thuiller, 2017). Studies seeking to understand sources of variation across ecological  
50 communities have often tested their compositional turnover or aggregate properties, such as  
51 species richness (e.g. Legendre & Legendre, 1984; Mandrak, 1995; Oikonomou, Leprieur, &  
52 Leonardos, 2014). More recently, tools drawn from graph theory have been applied to reveal  
53 community typologies and natural boundaries of bioregions (i.e. geographic areas with similar

54 biota) based on multispecies distribution patterns (e.g. Carstensen & Olesen, 2009; Bloomfield,  
55 Knerr, & Encinas-Viso, 2018; McGarvey & Veech, 2018). Because co-occurrence implies  
56 broadly shared niche characteristics and colonization history, repeated assemblages provide  
57 insight into formative ecological processes and a basis for setting habitat management units for  
58 conservation planning and targeted interventions (Brooks et al., 2006; Olden et al. 2010; Wehrly,  
59 Breck, Wang, & Szabo-Kraft, 2012; Montalvo-Mancheno, Ondei, Brook, & Buettel, 2020).  
60 Community differentiation is especially informative for freshwater organisms, such as fishes,  
61 which face strong dispersal constraints imposed by the hydrology of the watersheds they inhabit.  
62 Freshwater ecosystems exhibit high biodiversity and rates of endemism globally, as well as  
63 generally greater threats and species imperilment than terrestrial or marine habitats (Reid et al.,  
64 2019). Yet, compared to other realms, inland waters have received little attention paid to  
65 delineating recurrent assemblages and their underlying drivers at the broad spatial scales required  
66 to develop regional, cross-jurisdictional conservation and management strategies (Brooks et al.,  
67 2006; Wehrly et al., 2012; Oikonomou et al., 2014; Heino et al., 2021).

68 Our ability to predict changes in species diversity is challenged by the multitude of contributing  
69 spatial and environmental factors (e.g. Dormann et al., 2013; Mantyka-Pringle et al., 2014;  
70 Loewen, Strecker, Gilbert, & Jackson, 2020). For instance, spatial patterns of northern fish  
71 communities may reflect historical imprints of glaciation (Bailey & Smith, 1981; Legendre &  
72 Legendre, 1984; Dias et al., 2014), as well as present-day dispersal networks and regional habitat  
73 availability (e.g. Hitt & Angermeier, 2008). As most freshwater organisms are ectothermic, they  
74 are also intrinsically sensitive to temperature (Magnuson, Crowder, & Medvick, 1979). Thus,  
75 while fish dependence on hydrologic connections to reach new habitats limits their ability to  
76 move in response to interannual climate variations, longer-term temperature conditions should  
77 constrain the distributional boundaries of species by restricting the fitness of edge populations  
78 (Shuter & Post, 1990; Alofs, Jackson, & Lester, 2014). Persistence of species arriving at a new  
79 location further depends on habitat quantity and quality. Larger waterbodies may offer more  
80 space to more species (MacArthur & Wilson, 1963) and a greater diversity of habitats, including  
81 competitive or predatory refugia (Tonn & Magnuson, 1982). Aspects of lake basin morphometry,  
82 along with terrestrial (Heino et al., 2021) and geological setting (Conroy & Keller, 1976), also  
83 drive gradients in surface water quality and biological production (D'Arcy & Carignan, 1997).  
84 Thus, local habitat conditions combine with regional connectivity and climate filters to provide a

85 multilayered template for fish species sorting along geographic gradients (Smith & Powell,  
86 1971).

87 Our objectives were two-fold. First, we leveraged an unique dataset of fish records across the  
88 Laurentian Shield and surrounding Lowlands of Ontario, Canada and the Upper Midwest, USA,  
89 to develop a site-species network graph. We then applied bipartite modularity analysis to detect  
90 community typology and provide a hierarchical view of freshwater fish bioregionalization.

91 Second, we tested alternative hypotheses about the relative importance of biogeographic factors  
92 driving community differentiation, predicting varying roles of connectivity, climate, and habitat  
93 filters across groups. For instance, we hypothesized a key role of climate for northern fishes,

94 with contrasting responses to temperature and precipitation (owing to influence on lake thermal,  
95 chemical, and hydrological regimes, including timing of ice-out and major run-off events)

96 revealing sensitive regions and community types to target in future climate change adaptation

97 strategies. We expected connectivity to constrain communities both within drainages and in

98 relation to deglaciation patterns, as glacial lakes provided an important means of dispersal and

99 southern sites may have had more time for postglacial colonization. Habitat filters were predicted

100 to be of lesser importance at broad spatial scales, though we anticipated potential differentiation

101 of communities linked to contrasting shield (metamorphic and igneous) and sedimentary

102 lithologies as mediated by their effects on water chemistry (e.g. alkalinity and nutrient levels).

## 103 **MATERIALS AND METHODS**

### 104 **Study system**

105 We conducted a large data synthesis to assess fish assemblages and their potential driving factors

106 across a region of inland lakes in northeastern North America spanning over 13.4° of latitude

107 (1,500 km), 21.9° of longitude (1,700 km), and 580 m in elevation (Figure 1a). Fish occurrence

108 records and associated environmental measurements in portions of the Southwestern Hudson

109 Bay (n = 3,637), St. Lawrence (n = 5,757), and Mississippi River (n = 622) drainages were

110 compiled from existing datasets in Ontario (Aquatic Habitat Inventory; Dodge, Goodchild, Tilt,

111 Waldriff, & MacRitchie, 1987), Michigan (Status and Trends; Wehrly, Carter, & Breck, 2021),

112 and Minnesota (Index of Biotic Integrity; Drake & Pereira, 2002). To facilitate visualization of

113 bioregions and certain other geospatial analyses, major drainage areas were further divided into

114 secondary (Figure 1b) and smaller tertiary drainages (equivalent to sub-sub-drainages in Canada  
115 and hydrologic unit code 8 drainages in USA; Figure 1c).

116 Fishes were sampled from lakes, ponds, reservoirs, and embayments (herein referred to as lakes  
117 or sites) during the open-water season over a 60-year period (1957–2017). Although we were  
118 unable to explicitly consider species turnover during this time (e.g., Lynch et al., 2016; Cazelles  
119 et al., 2019; Keller, Heneberry, & Edwards, 2019), we found that community differentiation was  
120 robust to minor changes in species occurrence (see our sensitivity analysis for non-native species  
121 below) and temporal shifts in climate were generally small compared to spatial differences. For  
122 example, the mean annual air temperatures of sampled lakes (averaged across the study period)  
123 ranged from -4.1 to 9.7 °C, while changes in mean temperatures between the 1950s and 2010s  
124 ranged from -0.4 to 1.1 °C across sites (Wang, Hamann, Spittlehouse, & Carroll, 2016). While  
125 potentially contributing to shifts in distribution or relative abundance for some populations (e.g.  
126 Finigan, Mandrak, & Tufts, 2018), warming is unlikely to have confounded multispecies  
127 associations and the delineation of bioregions at our scale of analysis. Samples were collected  
128 using a combination of netting, trapping, and electrofishing, and quantitative survey results were  
129 collapsed to species presence/absence to reduce biases associated with different methods  
130 (Jackson & Harvey, 1997). Records of 132 fish taxa were initially compiled across 11,112 lakes,  
131 but several species and lakes were excluded from analysis. We first removed lakes lacking  
132 maximum depth or surface area measurements. We further omitted records for fishes with  
133 riverine life histories because they were likely transient when captured and therefore infrequently  
134 sampled in lake ecosystems. These undersampled species were identified based on expert  
135 opinion informed by comparisons to other sampling programs in the region (e.g. Lester,  
136 Marshall, Armstrong, Dunlop, & Ritchie, 2003; Drake, & Valley, 2005; Sandstrom, Rawson, &  
137 Lester, 2013) and aspects of organismal biology and encounter rates (relating to sampling  
138 methods/design; see Table S1.1 in Supporting Information for list of sampled fishes). Finally,  
139 because we were interested in patterns of interacting species, we removed sites with only a single  
140 species as well as species with only a single occurrence. Our final dataset included 90 taxa across  
141 10,016 lakes (mean species richness = 8).

#### 142 **Bipartite network modularity analysis**

143 We modelled an unweighted network graph comprised of lakes and fishes as separate sets of  
144 nodes, with edges linking species to the sites at which they were observed. While there are  
145 several approaches to grouping communities, most fail to fully exploit the bipartite nature of site-  
146 species networks to reveal both sites with similar species and species tending to form  
147 assemblages. Distance-based methods have conventionally been used to cluster sites based on  
148 their species composition and/or spatial configurations (e.g. Legendre & Legendre, 1984;  
149 Oikonomou et al., 2014); however, these procedures flatten bipartite networks, reducing species  
150 information to one of many site dissimilarity indices. Modularity analysis of site-species  
151 networks has proven an effective alternative to uncovering concise bioregions and their  
152 transitional zones (see participation coefficients; Guimerà & Amaral 2005) across spatial scales  
153 (e.g. Bloomfield et al., 2018; McGarvey & Veech, 2018; Bernardo-Madrid et al. 2019).

154 We used the DIRTLPA<sup>+</sup> algorithm of Beckett (2016), which is a label propagation approach to  
155 identifying groups of sites and species that are linked together more densely than others by  
156 maximizing Barber's modularity (see Appendix 1 for further details). Barber's index performs  
157 well for defining groups in bipartite networks (Thébault 2013) and classifies both sets of nodes  
158 (sites and species) simultaneously. We applied the algorithm using the computeModules function  
159 in the 'bipartite' package (Dormann, Gruber, & Fründ, 2008) with default settings for random  
160 initializations (implemented with R 3.6.1; R Development Core Team). This internally repeating  
161 procedure was itself replicated ten times (with different seeds) to check consistency and we  
162 recorded site and species assignments of the trial with the highest modularity (Table S1.2).  
163 Significance of resultant modularity was evaluated by one-sided randomization tests based on  
164 100 null networks obtained from the efficient, sequentially swapping curveball algorithm  
165 (Strona, Nappo, Boccacci, Farrorini, & San-Miguel-Ayanz, 2014). The curveball approach  
166 produces uniformly distributed null matrices maintaining row and column sums and was  
167 implemented with 1,000 burn-in and 1,000 thinning steps across 20 unique chains using the  
168 nullmodel and oecosimu functions in the 'vegan' package (Oksanen et al., 2020). Simulations  
169 were checked for convergence prior to use. Participation coefficients measuring the among-  
170 module connectivity of each site were calculated using the czvalues function (Dormann et al.,  
171 2008). To reveal finer-scale structures, we invoked a second round of analysis restricted to each  
172 primary module. This multiscale bioregionalization was summarized spatially as the dominant



173 primary (or sub) module across tertiary drainages (with ties adopting the most dominant type  
174 among neighbours).

### 175 **Environmental and spatial factors**

176 We estimated environmental and spatial variables to capture key dimensions of biogeographic  
177 constructs (connectivity, climate, and habitat; Table S1.3) hypothesized to determine natural site  
178 typology (see statistical analysis section below for details of latent variable approach). A  
179 connectivity construct was formed by regional habitat density, river network dispersal distance,  
180 glacial lake/marine distance, and time since glaciation. Variables were estimated from remote-  
181 sensing data using a geographic information system (ArcGIS Pro 2.6.2; Esri, Redlands,  
182 California, USA). Regional habitat density was calculated by summarizing all lake, pond, and  
183 reservoir features within tertiary drainages and dividing the area of this combined habitat by the  
184 total area of the watershed (excluding the Great Lakes; 1:50,000 scale or better). River network  
185 dispersal distances estimated the lengths of potential contemporary dispersal routes to secondary  
186 drainage features in either the Southwestern Hudson Bay drainage area (Lake Winnipeg or  
187 Hudson Bay), St. Lawrence drainage area (Lake Superior, Lake Michigan, Lake Huron, Lake  
188 Erie, Lake Ontario, or St. Lawrence River) or Mississippi River (Figure 1d; see Appendix 1 for  
189 further details). Historical glacial lake/marine distance was calculated as the geodesic distance  
190 from each lake to the nearest glacial lake or marine extent polygon (Figure S1.1). Time since  
191 glaciation (i.e. age of lake) was calculated by spatially joining sampling points near the center of  
192 each lake with a time-series of ice coverage polygons and determining the most recent overlap.  
193 Estimated historical ice, lake, and marine extents were obtained at approximately 500-year  
194 resolution and 1:7,000,000 scale (Dyke, 2004).

195 A climate construct was formed by mean warmest month temperature, mean coldest month  
196 temperature, May to September total precipitation, and total precipitation as snow. Climate  
197 variables were elevation-adjusted point estimates of average annual conditions at each lake over  
198 the entire study period. We extracted and downscaled estimates from moderate resolution  
199 Climate Research Unit data (4 km gridded; Mitchell & Jones, 2005) using ClimateNA v6.40  
200 (Wang et al., 2016; Figure S1.2).

201 Finally, a habitat construct was formed by surface area, maximum depth, Secchi depth, and  
202 proportion shield lithology. Physical size measurements were retrieved from existing records.

203 Secchi depth, which measures water clarity, provided a proxy for basin productivity given our  
204 lack of consistent water chemistry data. Secchi depths were measured at the majority (98%) of  
205 sites. For the 191 sites without direct measures, we estimated Secchi depths from their regional  
206 relationships to maximum depth. Here, separate linear regression models were constructed for  
207 each secondary drainage area stratified into depth categories (0–20 m, 20–40 m, and >40 m) and  
208 used to predict missing values. We estimated proportion shield lithology to capture the dominant  
209 geological gradient in our study region (i.e. surface exposed portions of igneous and  
210 metamorphic Laurentian Shield contrasting the sedimentary rocks of the Interior and Hudson  
211 Bay Lowlands). Surficial weathering of rocks provides a natural source of variation in water  
212 quality (e.g. ion chemistry) and biological production, especially between major lithological  
213 units (Conroy & Keller, 1976). Lake polygons were overlain with detailed lithological maps  
214 compiled by Harmann & Moosdoft (2012) at an average scale of 1:3,750,000 and used to  
215 calculate the proportions of each basin comprised of either shield or sedimentary rock (Figure  
216 S1.3).

## 217 **Statistical analysis**

218 To evaluate the roles of spatial and environmental constraints on fish communities, we applied  
219 non-metric partial least-squares structural equation modelling (NM-PLS-SEM, also called PLS  
220 path modelling) paired with multiple logistic regression (Petrarca, Russolillo, & Trinchera,  
221 2017). PLS-SEM is a variance-based approach to estimating unobserved latent variables and  
222 their relations by a series of simple or multiple ordinary least-squares (OLS) regressions  
223 (Sanchez, 2013). A key application of PLS-SEM is for building latent variables as formative  
224 constructs using a composite measurement model. Here, latent variables (or constructs) are  
225 formed as linear combinations of their manifest variables (or observed indicators) by an iterative  
226 process of optimizing weights and paths in the outer measurement and inner structural equation  
227 models (Sanchez, 2013). Thus, the algorithm offers an intuitive inferential framework while also  
228 applying dimension reduction to develop proxies for multidimensional concepts, such as  
229 connectivity, climate, and habitat (e.g. Loewen et al., 2020). PLS methods can also be performed  
230 with (non-metric) nominal or ordinal indicators by introducing an optimal scaling step (Petrarca  
231 et al., 2017). Here, non-metric variables are assigned values on an interval scale by optimizing  
232 additional scaling parameters as part of the iterative PLS-SEM process.

233 We performed NM-PLS-SEM to assess the roles of biogeographic factors using the ‘path  
234 scheme’ method and formative measurement (mode B) with the `plspm` function in the ‘`plspm`’  
235 package (Sanchez et al., 2017). Indicator variables were natural log- or logit-transformed (for  
236 proportional data; adjusting by minimum observed values where necessary) and standardized  
237 (centered at zero and scaled to unit variance) prior to analysis. Connectivity, climate, and habitat  
238 constructs were formed as linear combinations of their respective numeric indicators and used to  
239 predict individual modules. The approach was also applied to multilevel categorical outcomes of  
240 our modularity analysis. However, because estimating a binomial response by OLS is less than  
241 ideal, even with the optimal scaling algorithm, we used a modified approach (Petrarca et al.,  
242 2017) first obtaining latent variable scores from NM-PLS-SEM and then re-estimating inner  
243 model path coefficients by multiple logistic regression (generalized linear regression with  
244 binomial error distribution and a logit-link function; `glm` function in base R). The explanatory  
245 power of logistic models was measured by Tjur’s  $R^2$  using the `RsqGLM` function in the  
246 ‘`modEVA`’ package (Barbosa, Real, Muñoz, & Brown, 2015). Tjur’s coefficient of discrimination  
247 is an analogue to the coefficient of determination in linear regression but calculated as the  
248 difference in mean fitted values for successful and failed predictions (Tjur, 2009). Finally,  
249 because we anticipated potential collinearity among constructs, we used a variation-partitioning  
250 framework to reveal their unique, shared, and total explanatory powers. Variation partitioning  
251 based on Tjur’s  $R^2$  from logistic regression was performed using the `varPart` function (Barbosa  
252 et al., 2015) and results were visualized by approximately area-proportional Euler diagrams  
253 using the `euler` function in the ‘`eulerr`’ package (Larsson, 2020). To emphasize broad,  
254 biogeographic variation in fish assemblages, we limited our statistical analysis to primary  
255 modules. To test the influence of non-native species on bioregionalization patterns (Table S1.1),  
256 we conducted sensitivity analyses to determine changes to site and species assignments, as well  
257 as interpretations of formative processes, when either non-native species were excluded from the  
258 species pool or sites with non-natives were excluded from the site pool.

## 259 **RESULTS**

260 We identified clear spatial patterns of fish community differentiation, which we summarized by  
261 tertiary watershed to identify tractable northern bioregions (Figures 2, S1.4, and S1.5). Sites from  
262 module 1 ( $n = 1,258$  lakes; referred to as the southern group) clustered in southern portions of the

263 St. Lawrence and the Mississippi River drainages. Module 2 (n = 3,010; the trout group) and  
264 module 3 (n = 1,851; the central Great Lakes group) were situated in more northern and east-  
265 central portions of the St. Lawrence drainage, respectively. Module 4 lakes (n = 3,897; the  
266 northern group) were dominant in northern areas draining into the Southwestern Hudson Bay.  
267 However, none of the assemblages was restricted to any single drainage. Primary modules were  
268 internally consistent and had significant non-random structure with a modularity value (Q) of  
269 0.332 (P <0.01; Table S1.2). Participation coefficients showed that among-module connections  
270 were generally denser in the southern half of the study region, specifying a bioregional transition  
271 zone stretching from northern portions of the Mississippi River drainage to the centre of the  
272 Great Lakes Basin, as well as adjacent to the St. Lawrence River (Figure 2b). Sub-modularity  
273 analysis, where procedures were repeated with restricted site and species pools, revealed further  
274 variation within primary modules (Figures 2c–f). Submodule assignments were weaker and more  
275 variable across trials but significantly non-random (P <0.01; Table S1.2). Module 1 was shown  
276 to possess finer-scale north-south and east-west structure (Figure 2c) while module 4 revealed  
277 two central community types (in the north-west and south-east portions of the Southwestern  
278 Hudson Bay drainage) with greater heterogeneity to the south-west (Figure 2f).

279 We similarly found groups of associated fishes (Figure 3). Module 1 (n = 44; the southern group)  
280 contained the most species, including several warmwater fishes with limited occurrence near  
281 their northern range limits in our dataset (especially submodule 1.4). Module 2 (n = 15; the trout  
282 group) was characterized by several salmonids, i.e. lake trout (*Salvelinus namaycush*), brook  
283 trout (*Salvelinus fontinalis*), round whitefish (*Prosopium cylindraceum*), and rainbow trout  
284 (*Oncorhynchus mykiss*), and a subset of leuciscid minnows. Module 3 (n = 8; the central Great  
285 Lakes group) contained species with known associations to the St. Lawrence River and Lake  
286 Ontario, such as American eel (*Anguilla rostrata*) and rainbow smelt (*Osmerus mordax*), along  
287 with certain warm and coolwater sportfishes, including muskellunge (*Esox masquinongy*),  
288 smallmouth bass (*Micropterus dolomieu*), and rock bass (*Ambloplites rupestris*). Module 4 (n =  
289 23; the northern group) consisted of several species common to northern lakes, including yellow  
290 perch (*Perca flavescens*), white sucker (*Catostomus commersonii*), and the sportfishes walleye  
291 (*Sander vitreus*), lake whitefish (*Coregonus clupeaformis*), cisco (*C. artedi*), and northern pike  
292 (*Esox lucius*).

293 Climate was the dominant factor underlying community typology, while the importance of  
294 connectivity and habitat filters varied across groups. The influence of biogeographic constructs  
295 (modelled as latent variables; see meta-model in Figure 4) on the occurrence of each assemblage  
296 and the roles of individual environmental and spatial indicators can be interpreted from the  
297 model coefficients presented in Figure 5. Here, indicator weights and loadings describe the  
298 contributions and correlations of indicators to their respective latent variables in the outer  
299 measurement model (Figure 5a), while logistic path coefficients from the inner structural model  
300 show the magnitude and directionality (negative/positive) of relationships between latent  
301 variables and each of the primary modules (Figure 5b). For example, connectivity constructs  
302 were mostly driven by time since glaciation, which contributed positively to module 1 (the  
303 southern group), negatively to module 4 (the northern group), and negligibly to modules 2 and 3  
304 (the trout and central Great Lakes groups). River network dispersal distance had a stronger  
305 negative association to the connectivity construct for module 3, but the small path coefficient for  
306 connectivity indicated that its effect was negligible. Similarly, error bars were inflated for  
307 indicators of connectivity contributing to module 2 because the construct had a minor effect and  
308 coefficients flipped between positive and negative across bootstrapped samples. Climate  
309 indicators were generally correlated with one another (and time since glaciation; Table S1.4) and  
310 their individual contributions varied across modules. Precipitation as snow had a relatively  
311 strong negative correlation with module 1 and positive correlation with module 2, while modules  
312 3 and 4 were more associated with higher and lower temperatures, respectively. Habitat  
313 indicators also had variable contributions and were most relevant to modules 2 and 4.  
314 Specifically, module 2 involved smaller lakes with greater Secchi depth, while module 4 was  
315 linked to greater surface area and proportion shield lithology. Categorical analysis indicated that  
316 climate was the principal driver of community differentiation overall (Tables S1.5 and S1.6).

317 The predominance of climate in the bioregionalization of northern fishes was further supported  
318 by our partitioning of latent variable explanatory power (Figure 6). Here, we found that climate  
319 was the most predictive construct for each assemblage except module 2 (the trout group), which  
320 was driven by lake area and water clarity. Modules 1 and 4 (the southern and northern groups)  
321 had the greatest classification success, with strong explanatory overlap between climate and  
322 connectivity (the latter associated with time since glaciation). Variation in connectivity and  
323 habitat were of lesser importance to module 3 (the Great Lakes group), contributing almost no

324 additional prediction after accounting for temperature and precipitation effects (see Tables S1.7  
325 and S1.8 for detailed results).

326 The most frequent non-natives were common carp (*Cyprinus carpio*; module 1), rainbow trout  
327 (module 2), and rainbow smelt (module 3; Table S1.1). We found that only two native species,  
328 spoonhead sculpin (*Cottus ricei*) and white sucker, changed modules when non-native species ( $n$   
329 = 11) were excluded from the species pool (both from module 4 to 2). Assignments changed for  
330 646 lakes (6.4%), the majority changing from module 4 to 2 (261) or 3 to 2 (134; Table S1.9).  
331 Changes to bioregions were few and mostly in transitional regions of southern Ontario (Figure  
332 S1.6b). However, when sites with non-native species were excluded ( $n = 944$ ; 474 from module  
333 1, 243 from module 2, 197 from module 3, and 30 from module 4), assignments changed for nine  
334 native species (one from module 1 to 3, one from module 1 to 4, and seven from module 3 to 1)  
335 and 1,766 lakes (19.4%). Most site differences involved module 3 (94%), which largely merged  
336 with the southern group (Figure S1.6c), but interpretations of how communities were shaped by  
337 environmental and spatial processes were generally robust to the inclusion or exclusion of non-  
338 native species (Figures S1.7–S1.10).

## 339 **DISCUSSION**

340 We leveraged exceptional sampling effort across our study region to present a detailed view of  
341 freshwater bioregions and their underlying drivers at a broad, subcontinental scale. As  
342 bioregionalization is inherently taxa- and scale-dependent (McGarvey & Veech, 2018), we  
343 provide a novel classification of northern fish communities and advance understanding of their  
344 assembly from the regional species pool. While global maps of fish biogeographical zones have  
345 been produced (Abell et al., 2008; Leroy et al., 2019), they have aggregated disparate surveys  
346 across larger watersheds and thus have limited use for differentiating local communities or  
347 tractable management units. Alternative indices and module detection algorithms also exist (e.g.  
348 Guimerà & Amaral, 2005; Rosvall & Bergstrom, 2008), which may reveal different patterns (see  
349 Thébault, 2013; Bloomfield et al., 2018). Similar to the flow-based Infomap method (Edler,  
350 Guedes, Zizka, Rosvall, & Antonelli, 2017), Beckett's (2016) label propagation identifies groups  
351 of highly associated sites and species simultaneously (without unipartite projection). Thus, our  
352 application of site-species network analysis builds on existing knowledge, revealing both lakes  
353 with similar fish assemblages and fishes tending to co-occur. By partitioning our network

354 independently of spatial and environmental contexts, we were able to test the relative importance  
355 of key formative processes determining different sets of species and generate several hypotheses  
356 for future study.

357 As species sorting along geographic gradients involves multiple, often highly correlated factors,  
358 we evaluated drivers using a latent variable approach to capture broadly defined climate,  
359 connectivity, and habitat filters (Dormann et al., 2013; Loewen et al., 2020). We discovered that  
360 climate had the greatest effect in differentiating fish communities, similar to recent reports for  
361 subcontinental patterns of terrestrial fauna (Ficetola et al., 2017). Thermal or hydrological  
362 regimes may also account for finer-scale differences, such as the sub-modular structure observed  
363 within the southern and northern groups (modules 1 and 4; Figure 2) or variation among lakes  
364 within tertiary catchments (e.g. Wehrly et al., 2012; Figures S1.4 and S1.5). We found that  
365 historical connectivity had a related, albeit lesser, influence on modules 1 and 4, pointing to  
366 differences in postglacial colonization opportunities between lakes in more southern and  
367 northern areas of our study region (Bailey & Smith, 1981; Legendre & Legendre, 1984; Dias et  
368 al., 2014). However, distances to glacial lakes do not necessarily capture dispersal routes and  
369 given the degree of overlap between glacial recession and present temperature gradients (Figures  
370 S1.1 and S1.2), analysis of shifting recolonization pathways from glacial lakes and their outflows  
371 may offer clearer support (e.g. Mandrak & Crossman, 1992). Although habitat factors are often  
372 linked to species sorting at smaller scales (Jackson et al., 2001; Lansac-Tôha et al., 2021), we  
373 found evidence of broad-scale filtering by lake surface area, Secchi depth, and shield lithology.  
374 For instance, the trout group (module 2) showed affinity to smaller lakes with greater Secchi  
375 depth, which serves as an integrative proxy for multiple terrestrial factors (e.g. sediment loading)  
376 and internal productivity (Tilzer, 1988). Smaller lakes with greater water clarity may thus  
377 provide greater open-water foraging success for salmonids (Ortega, Figueiredo, da Graça,  
378 Agostinho, & Bini, 2020). In contrast, the northern group (module 4) was linked to larger lakes  
379 with greater influence of shield lithology as well as those with reduced water clarity (as  
380 encountered in areas overlaying glacial lake beds). Here, species such as walleye may experience  
381 optimal foraging in habitats with lower light intensity (Lester, Dextrase, Kushneriuk, Rawson, &  
382 Ryan, 2004). Bioregional transition zones also matched southern boundaries of the Laurentian  
383 Shield (Figures 2b and S1.3), showing potential influence of geological setting via effects on  
384 water chemistry (Conroy & Keller, 1976; Neff & Jackson, 2012).

385 In addition to site typology, we revealed recurrent groups of fishes, implying broadly shared  
386 natural histories and niche space. We highlight several sportfishes of interest to fisheries  
387 management but provide only a cursory examination of fish module composition. Future  
388 research should test for associations among species within and between groups (e.g. Cordero &  
389 Jackson, 2019), including their functional trait and phylogenetic similarities. For instance, the  
390 predominance of climate filters should sort species into thermal guilds (Magnuson et al., 1979),  
391 while connectivity constraints may produce assemblages related by their dispersal ability (i.e.  
392 swimming proficiency and tolerance to flow conditions) or common ancestry (Comte & Olden,  
393 2018). Habitat filters should further aggregate species with similar life-history strategies,  
394 including requirements for forage, reproduction, and overwintering (e.g. Tonn & Magnuson,  
395 1982). Explicit consideration of competition and predation among co-occurring species may  
396 yield additional mechanistic insights; however, as with habitat filters, we anticipate that species  
397 interactions will be most important at smaller scales of inquiry (Jackson et al., 2001).

398 Our findings have applications in conservation biogeography, revealing opportunities for  
399 targeted climate adaptation strategies. Setting conservation priorities for regional planning (e.g.  
400 protected areas or resource extraction) requires defining management units and understanding  
401 their formative processes (Brooks et al., 2006; Oikonomou et al., 2014; Montalvo-Mancheno et  
402 al., 2020). Applying space-for-time substitution, the central roles of temperature and  
403 precipitation provide evidence that fish communities are sensitive to climatic changes (Alofs et  
404 al., 2014; Lynch et al., 2016), but future impacts will depend on community type. We also found  
405 that connectivity does not appear to have been a major factor preventing lacustrine fishes from  
406 matching their temperature preferences within catchments. For instance, the southern group  
407 (module 1) was much more associated with warmer temperatures and less precipitation as snow  
408 than contemporary dispersal indicators, suggesting potential to expand northward as  
409 temperatures rise and connectivity permits. Indeed, there is evidence that certain warm and  
410 coolwater sportfishes are already expanding their northern ranges via multiple mechanisms  
411 (Alofs et al., 2014), including anthropogenic introductions. Coldwater fishes in the trout group  
412 (module 2) were linked to cooler summers and more snow, but habitat size and quality had a  
413 greater influence. Thus, this group may be relatively sensitive to hydrological changes and  
414 increasing water column turbidity or brownification. In contrast, the Great Lakes group (module  
415 3) was clustered in warmer and snowier areas around the Great Lakes in south-central Ontario.



416 These results suggest that Great Lakes effects create conditions for warm and coolwater fishes  
417 that may offer partial buffering against climatic changes, though future climates are uncertain  
418 and will be regionally variable (Notaro, Bennington, & Vavrus, 2015). Finally, the northern  
419 group (module 4) was most sensitive to warming (especially in winter months) and potentially  
420 more dispersal limited (based on the greater importance of river network dispersal distance),  
421 indicating that these communities may require extra attention for climate change adaptation.  
422 However, our estimates of dispersal limitation reflect only the natural connectivity of sites within  
423 their respective drainages.

424 Establishment of introduced fishes outside of historical ranges indicates that natural colonization  
425 either lags changing conditions or is impeded by other environmental, connectivity, or biotic  
426 factors (e.g. Allee effects). Fish movements are constrained by physical barriers (e.g. dams,  
427 culverts, and falls) and limited hydrological connections between catchments (especially along  
428 continental divides) may induce critical bottlenecks for migration (Jackson et al., 2001; Melles,  
429 Chu, Alofs, & Jackson, 2015). Species also have thermal and depth requirements for dispersal,  
430 possibly limiting movement by coldwater species as climates warm. While assemblages  
431 generally reflect shared colonization histories of dispersal along lake and river networks, effects  
432 of increasing fragmentation on future range shifts and persistence may vary (e.g. Herrera-R et al.,  
433 2020). Actions such as human-assisted migration or construction of fishways warrant  
434 consideration, weighing potential benefits with risks of unintended consequences from  
435 introducing harmful species (Olden et al., 2010), including pathogens.

436 Human introductions have already impacted the spatial structure of fish communities globally  
437 (Olden et al., 2010; Alofs et al., 2014). For instance, regional translocations contribute to biotic  
438 homogenization among lakes with higher angling pressure or nearer urban centres (Olden,  
439 Kennard, & Pusey, 2008). Stocking programs and movement of baitfishes may have also  
440 contributed to a minor role of connectivity for the trout group (module 2), as introductions  
441 obscure how species are naturally filtered from—or serve as additions to—the regional pool  
442 (Alofs et al., 2014; Bernardo-Madrid et al., 2019; Cazelles et al., 2019). Although we were  
443 unable to account for regional translocations, the small effect of excluding records for non-native  
444 species (from outside the study region) on our understanding of community differentiation  
445 highlights the robustness of our results to potential sampling errors. While biases associated with

446 site selection (e.g. targeting lakes with fishes of greater management interest) or imperfect  
447 detection can confound species distribution patterns, errors were mitigated by our large sample  
448 size and the relative insensitivity of our analytical approach to missing data. However, removing  
449 a species record does not remove its community impact, and our sensitivity analysis showed a  
450 major role of invaded communities in the differentiation of the southern and Great Lakes groups  
451 (modules 1 and 3). Whether driven by species interactions, loss of coverage, or some other  
452 aspect of invaded sites, results excluding lakes with non-native species both revealed the imprint  
453 of introductions on fish communities in the region and upheld our interpretations of key  
454 formative processes and climatic sensitivities. Despite non-native species typically establishing  
455 by anthropogenic means (such as shipping, recreational stocking, or escape from  
456 aquaculture/aquaria), they contribute to contemporary bioregions and are often management  
457 priorities as fisheries or biotic stressors on native communities (e.g. Loewen et al., 2020).

458 Finally, as we focus on natural drivers of community differentiation, important questions remain  
459 about how anthropogenic factors overlay observed patterns. For instance, regional land-use can  
460 cause changes in downstream water quality and fish community structure (e.g. Mantyka-Pringle  
461 et al., 2014). In our study region, most urban and agricultural developments are concentrated at  
462 southern latitudes where fishes generally contend with heightened angling pressure (Lester et al.,  
463 2003). Many thousands of lakes in central Ontario have also experienced anthropogenic  
464 acidification, leaving lasting impacts on aquatic food webs (Keller et al., 2019) and contributing  
465 to a suite of waterscape stressors affecting freshwater ecosystems at multiple scales (Heino et al.,  
466 2021). Further integration of these and other human activities will be needed to reconcile natural  
467 patterns of freshwater biogeography with ongoing global changes and spread of invasive species.

## 468 **REFERENCES**

- 469 Abell, R.A., Thieme, M.L., Revenga, C., Bryer, M., Kottelat, M., Bogutskaya, N., ... Petry, P.  
470 (2008). Freshwater ecoregions of the world: a new map of biogeographic units for freshwater  
471 biodiversity conservation. *BioScience*, 58, 403–414.
- 472 Alofs, K.M., Jackson, D.A., & Lester, N.P. (2014). Ontario freshwater fishes demonstrate  
473 differing range-boundary shifts in a warming climate. *Diversity and Distributions*, 20, 123–136.

474 Bailey, R.M., & Smith, G.R. (1981). Origin and geography of the fish fauna of the Laurentian  
475 Great Lakes Basin. *Canadian Journal of Fisheries and Aquatic Sciences*, 38, 1539–1561.

476 Barbosa, A.M., Real, R., Muñoz, A.-R., & Brown, J.A. (2015). New measures for assessing  
477 model equilibrium and prediction mismatch in species distribution models. *Diversity and*  
478 *Distributions*, 19, 1333–1338.

479 Beckett, S.J. (2016). Improved community detection in weighted bipartite networks. *Royal*  
480 *Society Open Science*, 3, 140536.

481 Bernardo-Madrid, R., Calatayud, J., González-Suárez, M., Rosvall, M., Lucas, P.-M., Rueda, M.,  
482 ... Revlla, E. (2019). Human activity is altering the world's zoogeographical regions. *Ecology*  
483 *Letters*, 22, 1297–1305.

484 Bloomfield, N.J., Knerr, N., & Encinas-Viso, F. (2018). A comparison of network and clustering  
485 methods to detect biogeographical regions. *Ecography*, 41, 1–10.

486 Brooks, T.M., Mittermeier, R.A., da Fonseca, G.A.B., Gerlach, J., Hoffmann, M., Lamoreux,  
487 J.F., ... Rodrigues, A.S.L. (2006). Global biodiversity conservation priorities. *Science*, 313, 58–  
488 61.

489 Carstensen, D.W., & Olesen, J.M. (2009). Wallacea and its nectarivorous birds: nestedness and  
490 modules. *Journal of Biogeography*, 36, 1540–1550.

491 Cazelles, K., Bartley, T., Guzzo, M.M., Brice, M.-H., MacDougall, A.S., Bennett, J.R., ...  
492 McCann, K.S. (2019). Homogenization of freshwater lakes: recent compositional shifts in fish  
493 communities are explained by gamefish movement and not climate change. *Global Change*  
494 *Biology*, 25, 4222–4233.

495 Comte, L., & Olden, J.D. (2018). Evidence for dispersal syndromes in freshwater fishes.  
496 *Proceedings of the Royal Society B*, 285, 20172214.

497 Conroy, N., & Keller, W. (1976). Geological factors affecting biological activity in Precambrian  
498 shield lakes. *The Canadian Mineralogist*, 14, 62–72.

499 Cordero, R.D., & Jackson, D.A. (2019). Species-pair associations, null models, and test of  
500 mechanisms structuring ecological communities. *Ecosphere*, 10, e02797.

501 D'Arcy, P., & Carignan, R. (1997). Influence of catchment topography on water chemistry in  
502 southeastern Québec Shield lakes. *Canadian Journal of Fisheries and Aquatic Sciences*, 54,  
503 2215–2227.

504 Dias, M.S., Oberdorff, T., Hugueny, B., Leprieur, F., Jézéquel, C., Cornu, J.-F., ... Tedesco, P.A.  
505 (2014). Global imprint of historical connectivity on freshwater fish biodiversity. *Ecology Letters*,  
506 17, 1130–1140.

507 Dodge, D.P., Goodchild, G.A., Tilt, J.C., Waldriff, D.G., & MacRitchie, I. (1987). *Manual of*  
508 *instructions: Aquatic habitat inventory surveys*. Ontario Ministry of Natural Resources, Toronto,  
509 Canada.

510 Dormann, C.F., Gruber, B., & Fründ, J. (2008). Introducing the bipartite package: analysing  
511 ecological networks. *R News*, 8, 8–11.

512 Dormann, C.F., Elith, J., Bacher, S., Buchmann, C., Carl, G., Carré, G, ... Lautenbach, S. (2013).  
513 Collinearity: a review of methods to deal with it and a simulation study evaluating their  
514 performance. *Ecography*, 36, 27–46.

515 Drake, M.T., & Pereira, D.L. (2002). Development of a fish-based index of biotic integrity for  
516 small inland lakes in Central Minnesota. *North American Journal of Fisheries Management*, 22,  
517 1105–1123.

518 Drake, M.T., & Valley, R.D. (2005). Validation and application of a fish-based index of biotic  
519 integrity for small central Minnesota lakes. *North American Journal of Fisheries Management*,  
520 25, 1095–1111.

521 Dyke, A.S. (2004). *An outline of North American deglaciation with emphasis on central and*  
522 *northern Canada*. Geological Survey of Canada, Ottawa, Canada.

523 Edler, D., Guedes, T., Zizka, A., Rosvall, M., & Antonelli, A. (2017). Infomap bioregions:  
524 interactive mapping of biogeographical regions from species distributions. *Systematic Biology*,  
525 66, 197–204.

526 Ficetola, G.F., Mazel, F., & Thuiller, W. (2017). Global determinants of zoogeographical  
527 boundaries. *Nature Ecology and Evolution*, 1, 0089.

528 Finigan, P.A., Mandrak, N.E., & Tufts, B.L. (2018). Large-scale changes in the littoral fish  
529 communities of lakes in southeastern Ontario, Canada. *Canadian Journal of Zoology*, 96, 753–  
530 759.

531 Guimerà, R., & Amaral, L.A.N. (2005). Functional cartography of complex metabolic networks.  
532 *Nature*, 433, 895–900.

533 Harmann, J., & Moosdoft, N. (2012). The new global lithological map database GLiM: a  
534 representation of rock properties at the Earth surface. *Geochemistry, Geophysics, Geosystems*,  
535 13, Q12004.

536 Heino, J., Alahuhta, J., Bini, L.M., Cai, Y., Heiskanen, A.-S., Hellsten, S., ... Angeler, D.G.  
537 (2021). Lakes in the era of global change: moving beyond single-lake thinking in maintaining  
538 biodiversity and ecosystem services. *Biological Reviews*, 96, 89–106.

539 Herrera-R, G.A., Oberdorff, T., Anderson, E.P., Brosse, S., Carvajal-Vallejos, F.M., ... Tedesco,  
540 P.A. (2020). The combined effects of climate change and river fragmentation on the distribution  
541 of Andean Amazon fishes. *Global Change Biology*, 26, 5509–5523.

542 Hitt, N.P., & Angermeier, P.L. (2008). Evidence for fish dispersal from spatial analysis of stream  
543 network topology. *Journal of the North American Benthological Society*, 27, 304–320.

544 Jackson, D.A., & Harvey, H.H. (1997). Qualitative and quantitative sampling of lake fish  
545 communities. *Canadian Journal of Fisheries and Aquatic Sciences*, 54, 2807–2813.

546 Jackson, D.A., Peres-Neto, P.R., & Olden, J.D. (2001). What controls who is where in freshwater  
547 fish communities—the roles of biotic, abiotic, and spatial factors. *Canadian Journal of Fisheries*  
548 *and Aquatic Sciences*, 58, 157–170.

549 Keller, W.B., Heneberry, J., & Edwards, B.A. (2019). Recovery of acidified Sudbury, Ontario,  
550 Canada, lakes: a multi-decade synthesis and update. *Environmental Reviews*, 27, 1–16.

551 Lansac-Tôha, F.M., Bini, L.M., Heino, J., Meira, B.R., Segovia, B.T., Pavanelli, C.S., ... Velho,  
552 L.F.M. (2021). Scale-dependent patterns of metacommunity structuring in aquatic organisms  
553 across floodplain systems. *Journal of Biogeography*, 48, 872–885.

554 Larsson, J. (2020). eulerr: area-proportional Euler and Venn diagrams with ellipses. R package  
555 version 6.1.0.

556 Legendre, P., & Legendre, V. (1984). Postglacial dispersal of freshwater fishes in the Québec  
557 Peninsula. *Canadian Journal of Fisheries and Aquatic Sciences*, 41, 1781–1802.

558 Lester, N.P., Marshall, T.R., Armstrong, K., Dunlop, W.I., & Ritchie, B. (2003). A broad-scale  
559 approach to management of Ontario’s recreational fisheries. *North American Journal of*  
560 *Fisheries Management*, 23, 1312–1328.

561 Lester, N.P., Dextrase, A.J., Kushneriuk, R.S., Rawson, M.R., & Ryan, P.A. (2004). Light and  
562 temperature: key factors affecting walleye abundance and production. *Transactions of the*  
563 *American Fisheries Society*, 133, 588–605.

564 Leroy, B., Dias, M.S., Giraud, E., Hugueny, B., Jézéquel, C., Leprieur, F., Oberdorff, T., &  
565 Tedesco, P.A. (2019). Global biogeographical regions of freshwater fish species. *Journal of*  
566 *Biogeography*, 46, 2407–2419.

567 Loewen, C.J.G., Strecker, A.L., Gilbert, B., & Jackson, D.A. (2020). Climate warming  
568 moderates the impacts of introduced sportfish on multiple dimensions of prey biodiversity.  
569 *Global Change Biology*, 26, 4937–4951.

570 Lynch, A.J., Myers, B.J.E., Chu, C., Eby, L.A., Falke, J.A., Kovach, R.P., ... Whitney, J.E.  
571 (2016). Climate change effects on North American inland fish populations and assemblages.  
572 *Fisheries*, 41, 346–361.

573 MacArthur, R.H., & Wilson, E.O. (1963). An equilibrium theory of insular zoogeography.  
574 *Evolution*, 17, 373–387.

575 Magnuson, J.J., Crowder, L.B., & Medvick, P.A. (1979). Temperature as an ecological resource.  
576 *American Zoology*, 19, 331–343.

577 Mandrak, N.E. (1995). Biogeographic patterns of fish species richness in Ontario lakes in  
578 relation to historical and environmental factors. *Canadian Journal of Fisheries and Aquatic*  
579 *Sciences*, 52, 1462–1474.

580 Mandrak, N.E., & Crossman, E.J. (1992). Postglacial dispersal of freshwater fishes into Ontario.  
581 *Canadian Journal of Zoology*, 70, 2247–2259.

582 Mantyka-Pringle, C.S., Martin, T.G., Moffatt, D.B., Linke, S., & Rhodes, J.R. (2014).  
583 Understanding and predicting the combined effects of climate change and land-use change on  
584 freshwater macroinvertebrates and fish. *Journal of Applied Ecology*, 51, 572–581.

585 McGarvey, D.J., & Veech, J.A. (2018). Modular structure in fish co-occurrence networks: a  
586 comparison across spatial scales and grouping methodologies. *PLoS One*, 13, e0208720.

587 Melles, S.J., Chu, C., Alofs, K.M., & Jackson, D.A. (2015). Potential spread of Great Lakes  
588 fishes given climate change and proposed dams: an approach using circuit theory to evaluate  
589 invasion risk. *Landscape Ecology*, 30, 919–935.

590 Mitchell, T.D., & Jones, P.D. (2005). An improved method of constructing a database of  
591 monthly climate observations and associated high-resolution grids. *International Journal of*  
592 *Climatology*, 25, 693–712.

593 Montalvo-Mancheno, C.S., Ondeï, S., Brook, B.W., & Buettel, J.C. (2020). Bioregionalization  
594 approaches for conservation: methods, biases, and their implications for Australian biodiversity.  
595 *Biodiversity and Conservation*, 29, 1–17.

596 Neff, M.R., & Jackson, D.A. (2012). Geology as a structuring mechanism of stream fish  
597 communities. *Transactions of the American Fisheries Society*, 141, 962–974.

598 Notaro, M., Bennington, V., & Vavrus, S. (2015). Dynamically downscaled projections of lake-  
599 effect snow in the Great Lakes Basin. *Journal of Climate*, 28, 1661–1684.

600 Oikonomou, A., Leprieur, F., & Leonardos, I.D. (2014). Biogeography of freshwater fishes of  
601 the Balkan Peninsula. *Hydrobiologia*, 738, 205–220.

602 Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., ... Wagner, H.  
603 (2020). *vegan*: community ecology package. R package version 2.5-7.

604 Olden, J.D., Kennard, M.J., & Pusey, B.J. (2008). Species invasions and the changing  
605 biogeography of Australian freshwater fishes. *Global Ecology and Biogeography*, 17, 25–37.

606 Olden, J.D., Kennard, M.J., Leprieur, F., Tedesco, P.A., Winemiller, K.O., & García-Berthou, E.  
607 (2010). Conservation biogeography of freshwater fishes: recent progress and future challenges.  
608 *Diversity and Distributions*, 16, 496–513.

609 Ortega, J.C.G., Figueiredo, B.R.S., da Graça, W.J., Agostinho, A.A., & Bini, L.M. (2020).  
610 Negative effect of turbidity on prey capture for both visual and non-visual aquatic predators.  
611 *Journal of Animal Ecology*, 89, 2427–2439.

612 Petrarca, F., Russolillo, G., & Trinchera, L. (2017). Integrating non-metric data in partial least  
613 squares path models: methods and application. In H. Latan & R. Noonan (Eds.), *Partial Least  
614 Squares Path Modeling* (pp. 259–279). Cham, Switzerland: Springer International Publishing.

615 Reid, A.J., Carlson, A.K., Creed, I.F., Eliason, E.J., Gell, P.A., Johnson, P.T.J., ... Cooke, S.J..  
616 (2019). Emerging threats and persistent conservation challenges for freshwater biodiversity.  
617 *Biological Reviews*, 94, 849–873.

618 Rosvall, M., & Bergstrom, C.T. (2008). Maps of random walks on complex networks reveal  
619 community structure. *Proceedings of the National Academy of Sciences of the United States of  
620 America*, 105, 1118–1123.

621 Sanchez, G. (2013). *PLS Path Modeling with R*. Berkeley, CA: Trowchez Editions.

622 Sanchez, G., Trinchera, L., & Russolillo, G. (2017). *plsmpm: tools for partial least squares path  
623 modeling (PLS-PM)*. R package version 0.4.9.

624 Sandstrom, S., Rawson, M., & Lester, N. P. (2013). *Manual of instructions for broad-scale fish  
625 community monitoring using North American (NA1) and Ontario small mesh (ON2) gillnets*.  
626 Peterborough, ON: Ontario Ministry of Natural Resources.

627 Ministry of Natural Resources. Shuter, B.J., & Post, J.R. (1990). Climate, population viability,  
628 and the zoogeography of temperate fishes. *Transactions of the American Fisheries Society*, 119,  
629 314–336.

630 Smith, C.L., & Powell, C.R. (1971). The summer fish communities of Brier Creek, Marshall  
631 County, Oklahoma. No. 2458. New York, NY: American Museum of Natural History.



632 Strona, G., Nappo, D., Boccacci, F., Fattorini, S., & San-Miguel-Ayanz, J. (2014). A fast and  
633 unbiased procedure to randomize ecological binary matrices with fixed row and column totals.  
634 Nature Communications, 5, 4114.

635 Thébault, E. (2013). Identifying compartments in presence-absence matrices and bipartite  
636 networks: insights into modularity measures. Journal of Biogeography, 40, 759–768.

637 Tilzer, M.M. (1988). Secchi disk – chlorophyll relationships in a lake with highly variable  
638 phytoplankton biomass. Hydrobiologia, 162, 163–171.

639 Tjur, T. (2009). Coefficients of determination in logistic regression models—a new proposal: the  
640 coefficient of discrimination. The American Statistician, 63, 366–372.

641 Tonn, W.M., & Magnuson, J.J. (1982). Patterns in the species composition and richness of fish  
642 assemblages in northern Wisconsin lakes. Ecology, 63, 1149–1166.

643 Wang, T., Hamann, A., Spittlehouse, D., & Carroll, C. (2016). Locally downscaled and spatially  
644 customizable climate data for historical and future periods for North America. PLoS One, 11,  
645 e0156720.

646 Wehrly, K.E., Breck, J.E., Wang, L. & Szabo-Kraft, L. (2012). A landscape-based classification  
647 of fish assemblages in sampled and unsampled lakes. Transactions of the American Fisheries  
648 Society, 141, 414–425.

649 Wehrly, K.E., Carter, G.S., & Breck, J.E. (2021). Standardized sampling methods for the inland  
650 lakes status and trends program. Fisheries Special Report. Lansing, MI: Michigan Department  
651 of Natural Resources.

## 652 **DATA AVAILABILITY STATEMENT**

653 Processed data (including site and species modules and derived spatial and environmental  
654 indicator variables) and code supporting our results are archived in a dedicated GitHub  
655 repository (<https://github.com/loewenecology/Fish-modularity-and-nmplssem>). Raw fish data  
656 (with species at risk removed for their protection) are available from the Dryad Digital  
657 Repository (<https://doi.org/10.5061/dryad.44j0zpcfc>). As contractual requirements prevent us  
658 from releasing detailed sampling records for Ontario directly, these data are provided with  
659 approximate location coordinates. The full set of fish data (including species at risk) are readily

660 available for non-commercial purposes from the Ministry of Northern Development, Mines,  
661 Natural Resources, and Forestry in Ontario ([https://geohub.lio.gov.on.ca/datasets/aquatic-](https://geohub.lio.gov.on.ca/datasets/aquatic-resource-area-polygon-segment-)  
662 [resource-area-polygon-segment-](https://geohub.lio.gov.on.ca/datasets/aquatic-resource-area-polygon-segment-)) and Departments of Natural Resources in Michigan  
663 ([https://www.michigan.gov/dnr/0,4570,7-350-79137\\_79765\\_84005\\_84013---,00.html](https://www.michigan.gov/dnr/0,4570,7-350-79137_79765_84005_84013---,00.html)) and  
664 Minnesota ([https://www.dnr.state.mn.us/waters/surfacewater\\_section/lake\\_ibi/index.html](https://www.dnr.state.mn.us/waters/surfacewater_section/lake_ibi/index.html))  
665 through their standard data-sharing agreements.

## 666 **FIGURE LEGENDS**

667 **Figure 1.** Maps presenting (a) lake sampling locations across Ontario, Minnesota, and  
668 Michigan; (b) portions of the major Southwestern Hudson Bay (green), Mississippi River  
669 (yellow), and St. Lawrence River (orange) drainage areas and their secondary drainages within  
670 our study region; (c) tertiary drainage areas (sub-sub-drainage or United States Geological  
671 Survey hydrologic unit code 8) in the Lake Huron watershed showing regional habitat density;  
672 and (d) a single tertiary drainage area showing river network dispersal distance between a lake  
673 sampling location (green point) and its nearest secondary drainage feature (Lake Huron; red  
674 point).

675 **Figure 2.** Maps presenting network modularity results for site nodes summarized by tertiary  
676 drainages to identify bioregions. Bioregions were assigned to the dominant primary module (a)  
677 and submodule (c-f) across sites in each drainage. Participation coefficients measure diversity of  
678 connections between primary modules and were assigned to the mean across sites, with greater  
679 values indicating bioregional transition zones (b).

680 **Figure 3.** Hierarchical edge bundling plot showing network modularity results for fish nodes to  
681 identify recurrent assemblages. Connections (edges) denote fish co-occurrences, with darker  
682 lines indicating more observations. Around the outside of the plot, node sizes indicate relative  
683 frequencies of fish occurrence, node colours indicate fish submodule assignment, and label  
684 colours indicate fish primary module assignment. Colours correspond to bioregions in Figure 2.

685 **Figure 4.** Non-metric partial least-squares structural equation (NM-PLS-SEM) meta-model.  
686 Major biogeographic constructs (connectivity, climate, and habitat) were formed as linear  
687 combinations of observed spatial and environmental factors to explain variation in primary site  
688 module assignment. Biogeographic constructs and site modules were treated as composite and

689 single-indicator latent variables (circles), respectively, for the inner structural model. Spatial and  
690 environmental factors (and nominal indicators of fish modules) were treated as formative  
691 indicators (rectangles) of their respective latent variables.

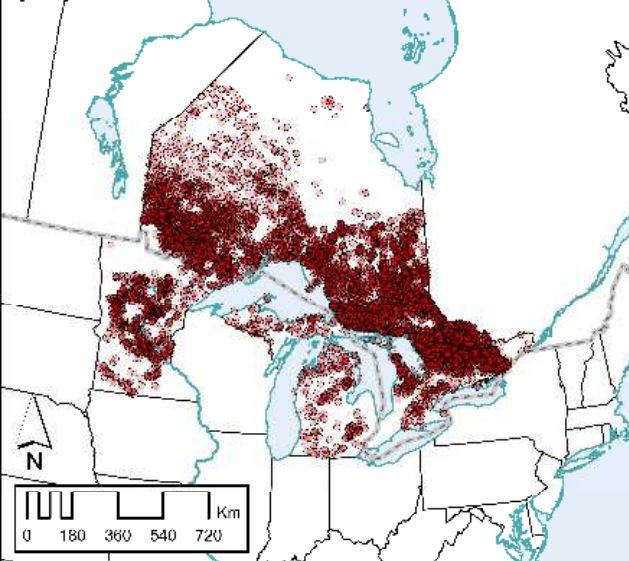
692 **Figure 5.** Results of NM-PLS-SEM and multiple logistic regression of primary site modules on  
693 biogeographic constructs. Outer measurement model results (**a**) are presented as weights (darker  
694 bars; representing multiple ordinary least-squares regression coefficients) and loadings (lighter  
695 bars; representing Pearson correlation coefficients) for standardized indicators. Inner structural  
696 model results (**b**) are presented as path coefficients from multiple logistic regression for latent  
697 variables. Error bars show the lower 2.5<sup>th</sup> and upper 97.5<sup>th</sup> percentiles of bootstrap and profile  
698 likelihood confidence intervals for outer and inner model coefficients, respectively. Detailed  
699 results are available in Supplementary Tables S1.5-S1.8. Colours and model structure correspond  
700 to the meta-model presented in Figure 4.

701 **Figure 6.** Approximately area-proportional Euler plots showing the total, shared, and unique  
702 explanatory power of biogeographic constructs (connectivity, climate, and habitat) in logistic  
703 regression. Fractions are expressed as Tjur's  $R^2$  (coefficients of discrimination) from models of  
704 primary site modules (binary responses) predicted by latent variable scores (calculated by NM-  
705 PLS-SEM). Colours correspond to the meta-model presented in Figure 4.

## 706 **SUPPORTING INFORMATION**

707 Additional supporting information may be found online in the Supporting Information section at  
708 the end of the article.

(a)



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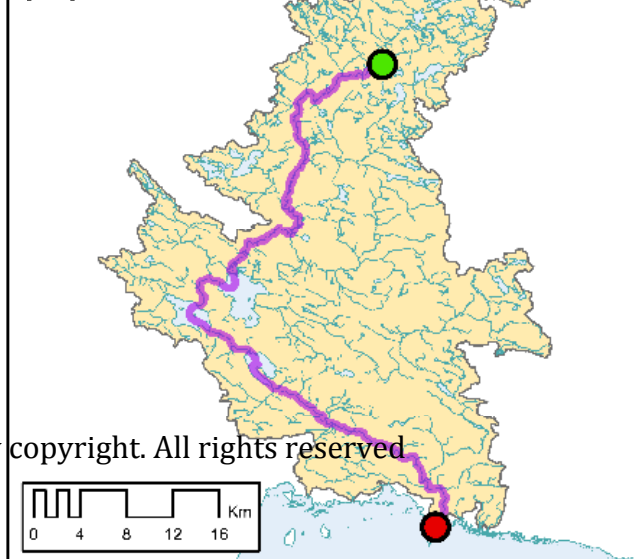
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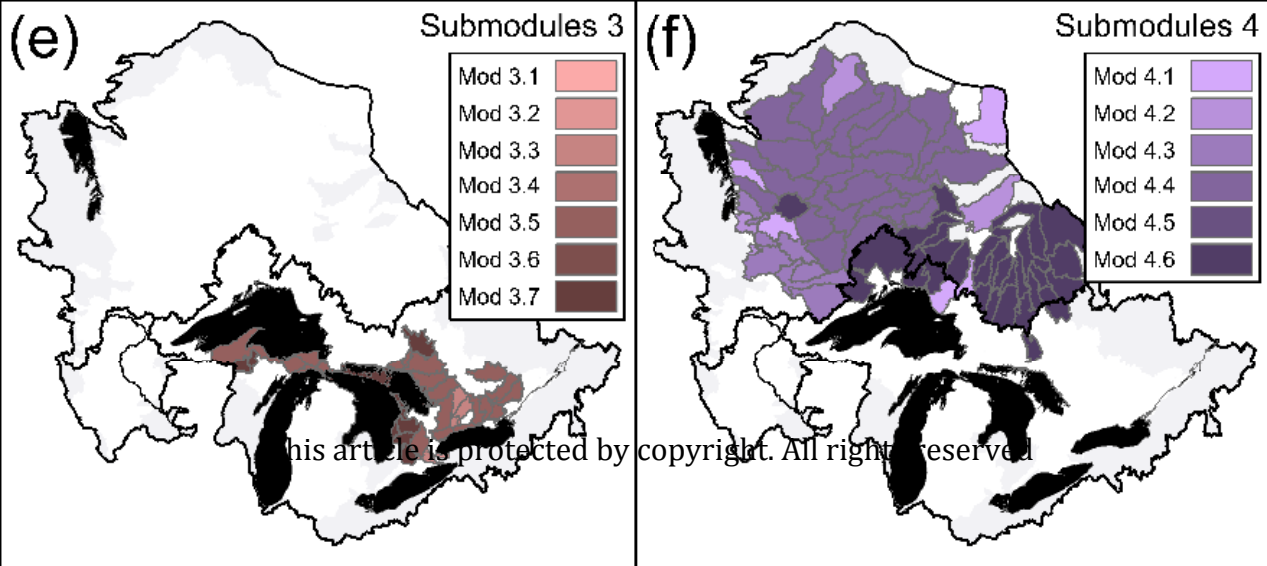
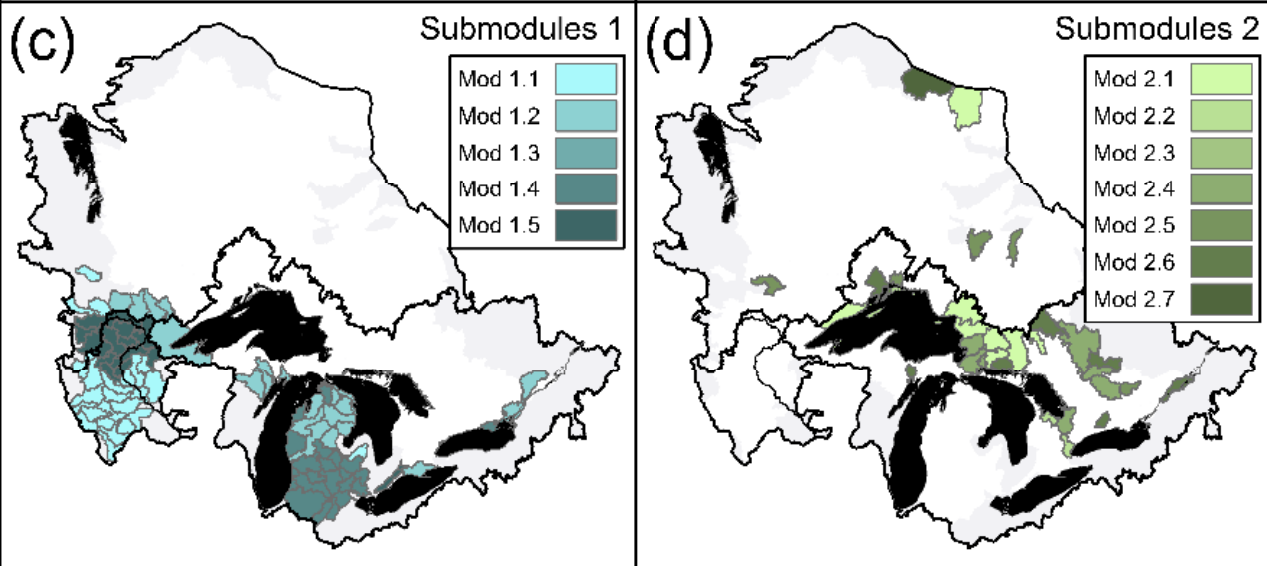
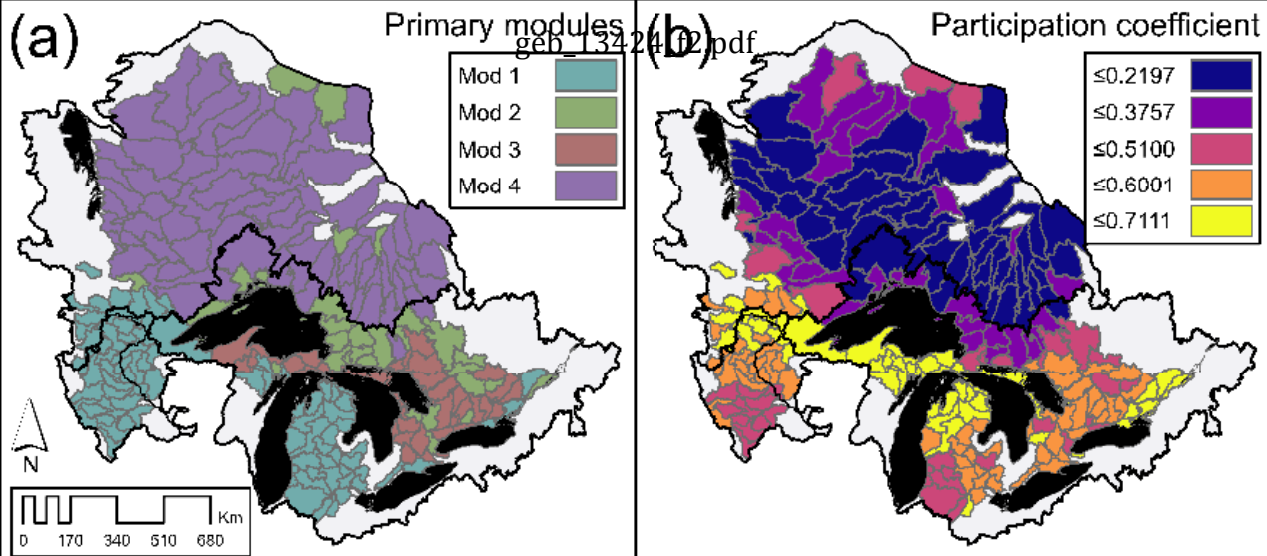
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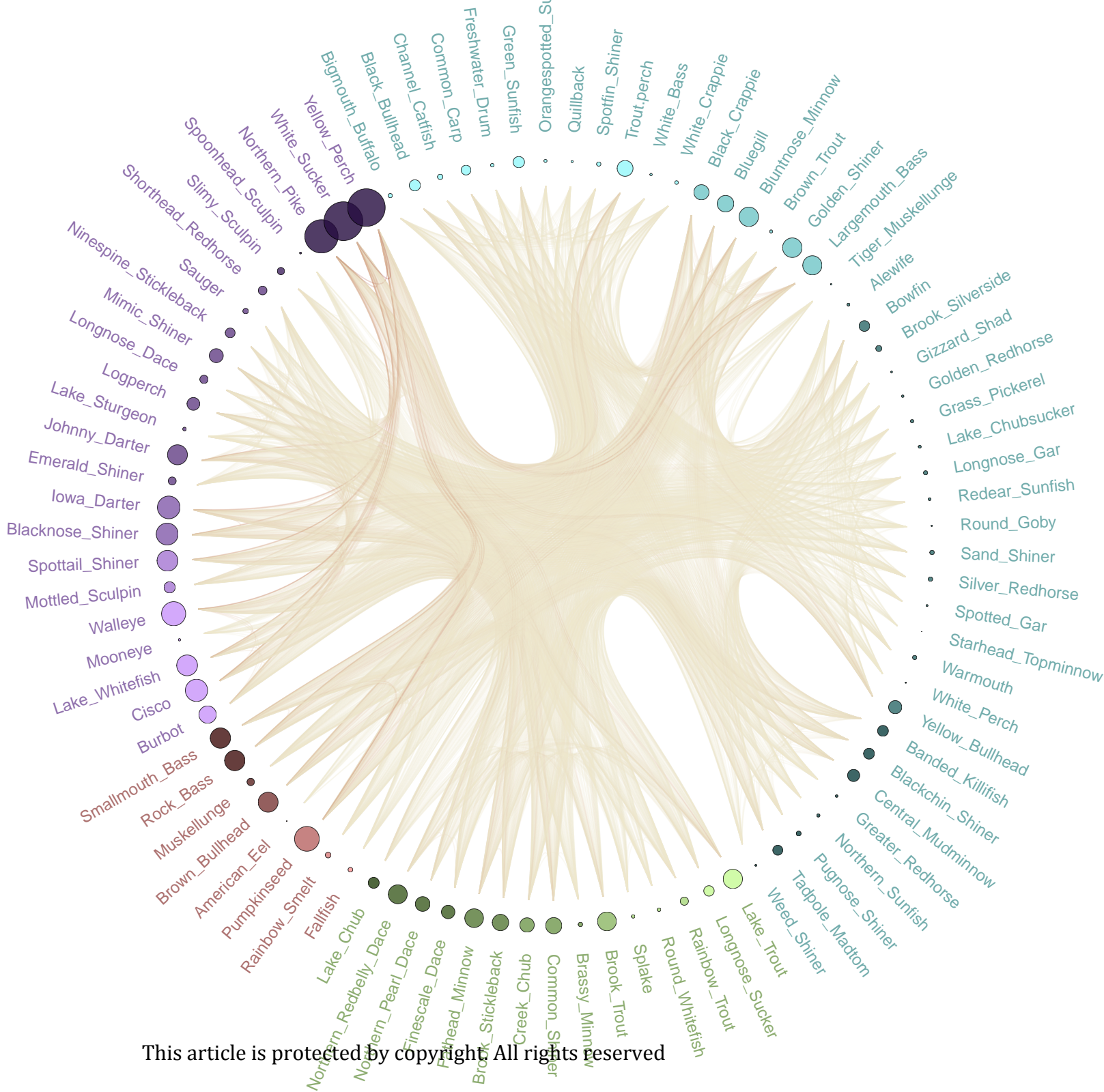


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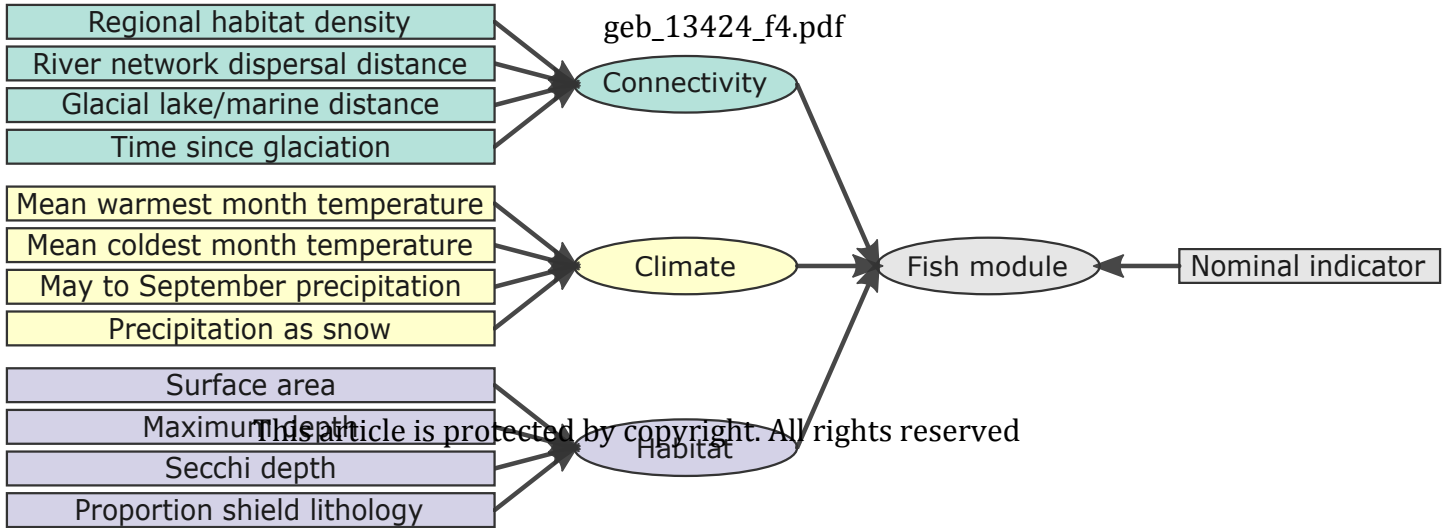


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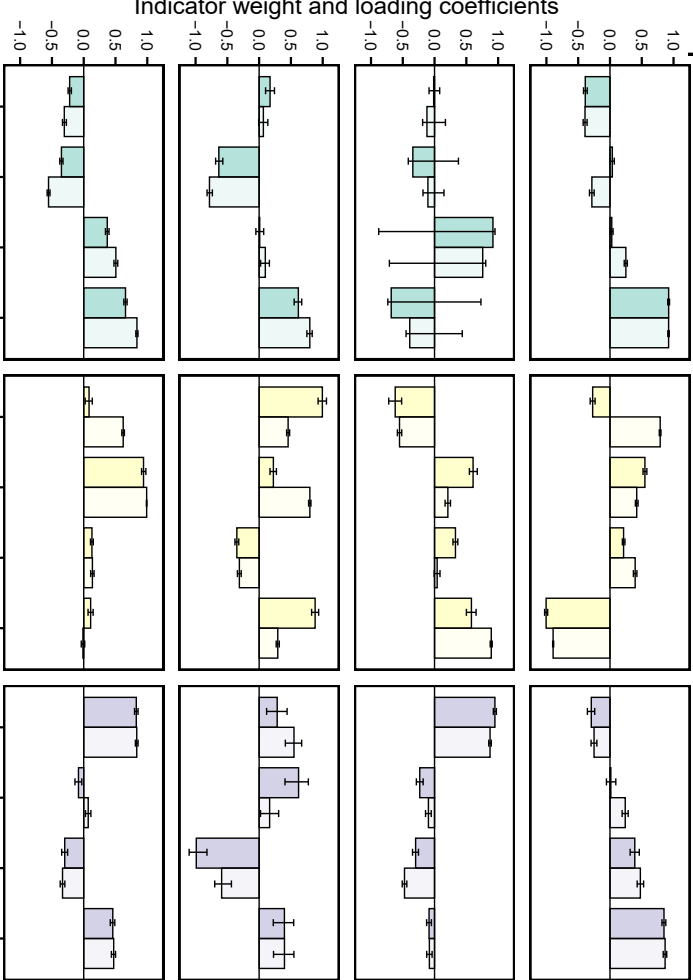


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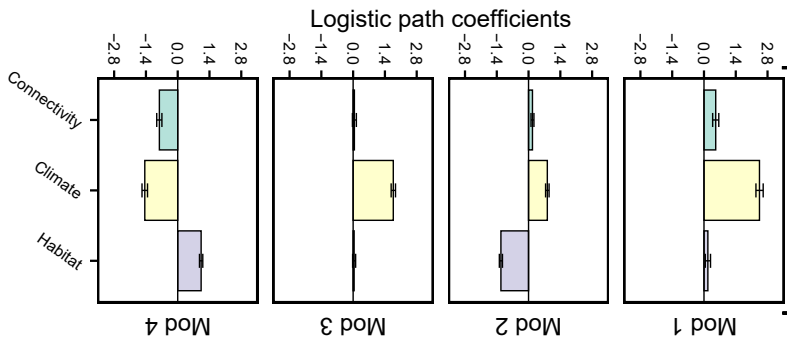


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(a)



(b)



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Outer measurement (indicator) and inner structural (latent) variables



Mod 1

Mod 2

Mod 3

Mod 4

Connectivity

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Climate

Habitat

