

van der Zee Sygrid (Orcid ID: 0000-0002-4244-369X)
Kanel Prabesh (Orcid ID: 0000-0003-1418-2960)
Boertien Jeffrey (Orcid ID: 0000-0002-9664-1771)
Muller Martijn L.T.M. (Orcid ID: 0000-0002-1133-7202)
Bohnen Nicolaas Ida (Orcid ID: 0000-0003-0352-6414)
Van Laar Teus (Orcid ID: 0000-0001-5088-480X)

Altered cholinergic innervation in de novo Parkinson's disease with and without cognitive impairment

Running title: Altered cholinergic innervation in de novo Parkinson

Sygrid van der Zee, PhD,^{1,2}, Prabesh Kanel, PhD,^{3,4}, Marleen J.J. Gerritsen, PhD,^{1,2}, Jeffrey M. Boertien, MD,¹, Anne C. Slomp, MSc,^{1,2}, Martijn L.T.M. Müller, PhD,^{3,4}, Nicolaas I. Bohnen, MD, PhD,^{3,4,5,6,7}, Jacoba M. Spikman, PhD,^{1,2}, Teus van Laar, MD, PhD,¹

¹ Department of Neurology, University of Groningen, University Medical Center Groningen, Groningen, the Netherlands

² Department of Neurology, Division of Clinical Neuropsychology, University of Groningen, University Medical Center, The Netherlands.

³ Department of Radiology, University of Michigan, Ann Arbor, MI, United States.

⁴ Morris K. Udall Center of Excellence for Parkinson's Disease Research, University of Michigan, Ann Arbor, MI, United States.

⁵ Department of Neurology, University of Michigan, Ann Arbor, MI, United States.

⁶ Neurology Service and GRECC, Veterans Administration Ann Arbor Healthcare System, Ann Arbor, MI, United States

⁷ University of Michigan Parkinson's Foundation Center of Excellent, Ann Arbor, MI, United States

Correspondence: dr. S. van der Zee Department of Neurology, University Medical Center Groningen, Hanzeplein 1, 9713 GZ Groningen, the Netherlands. +31 50361 2401. Email:

s.van.der.zee01@umcg.nl

Wordcount: 3665 words

Funding sources for the study: Weston Brain institute, 22 St. Clair Avenue East, Suite 2001, Toronto, ON Canada M4T 2S3, Email: info@westonbrain.org, Telephone: +1-416-967-7979

Trial registration: NCT04180865

No conflicts of interest related to the manuscript

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: [10.1002/mds.28913](https://doi.org/10.1002/mds.28913)

This article is protected by copyright. All rights reserved.

Abstract

Background: Altered cholinergic innervation plays a putative role in cognitive impairment in Parkinson's disease (PD), at least in advanced stages. Identification of the relationship between cognitive impairment and cholinergic innervation early in the disease will provide better insight in disease prognosis and possible early intervention.

Objective: To assess regional cholinergic innervation status in de novo patients with PD, with and without cognitive impairment.

Methods: 57 newly diagnosed, treatment-naïve, PD patients (32 males, mean age 64.6 ± 8.2 years) and 10 healthy control subjects (5 males, mean age of 54.6 ± 6.0 years) were included. All participants underwent cholinergic [^{18}F]Fluoroethoxybenzovesamicol PET, and detailed neuropsychological assessment. PD patients were classified as either cognitively normal (PD-NC) or mild cognitive impairment (PD-MCI). Whole brain voxel-based group comparisons were performed.

Results: Results show bidirectional cholinergic innervation changes in PD. Both PD-NC and PD-MCI groups showed significant cortical cholinergic denervation compared to controls ($p < 0.05$, FDR corrected), primarily in the posterior cortical regions. Higher than normal binding was most prominently present in PD-NC in both cortical and sub-cortical regions, including the cerebellum, cingulate cortex, putamen, gyrus rectus, hippocampus and amygdala.

Conclusion: Altered cholinergic innervation is already present in de novo patients with PD. Posterior cortical cholinergic losses were present in all patients independent of cognitive status. Higher than normal binding in cerebellar, frontal and subcortical regions in cognitively intact patients may reflect compensatory cholinergic upregulation in early-stage PD. Limited or failing cholinergic upregulation may play an important role in early, clinically evident cognitive impairment in PD.

Abbreviations:

[¹⁸F]FEOBV: [¹⁸F]Fluoroethoxybenzovesamicol

FDR: False Discovery Rate

HC: healthy controls

LED: Levodopa Equivalent Dose

MDS-UPDRS-III: Movement Disorder Society - Revised Unified Parkinson's Disease Rating Scale part III

PD: Parkinson's disease

PD-MCI: Parkinson's disease Mild Cognitive Impairment

PC-NC: Parkinson's disease Normal Cognition

PET: Positron Emission Tomography

VACHT: Vesicular Acetylcholine Transporter

Introduction

Cognitive impairment is common in Parkinson's disease (PD) and a major source of disability and lower quality of life (1,2). Mild cognitive impairment (PD-MCI) is already present in 25-30% of newly diagnosed patients and is a major risk factor for the development of PD dementia (PDD) (3–5). Cognitive impairment in PD is heterogeneous with multiple domains affected and great variability in onset and progression (6).

It has been increasingly recognized that comorbid cholinergic dysfunction is a major contributor to the pathophysiology of cognitive impairment in PD (7,8). The four major human brain cholinergic systems are the basal forebrain corticopetal (BF) projection system, cholinergic efferents of the peduncopontine-laterodorsal tegmental complex (PPN/LDTC), medial vestibular nucleus (MVN) cholinergic neurons projecting to the cerebellum, and striatal cholinergic interneurons (9–12).

Previous *in vivo* neuroimaging assessment of cholinergic innervation, using acetylcholinesterase PET, has demonstrated predominantly posterior cholinergic denervation in PD, with more severe cholinergic degeneration in PDD (13–16). There is additional evidence that the cholinergic system is a major driver of cognitive impairment in PD even in the absence of dementia (17). We previously showed that deficits in attention, executive functioning and memory correlated with loss of cholinergic activity on both a global cortical level (16,18) as well as on a regional (sub)cortical level (19). Although cholinergic denervation has been shown in cognitively impaired patients with PD, less is known about cholinergic innervation changes in early-stage disease and before the onset of cognitive changes. Elucidation of the relationship between cognitive impairment and cholinergic integrity in a very early stage of the disease may provide new clues that may inform novel therapeutic strategies.

The goal of this study was to characterize cholinergic innervation status in newly diagnosed, treatment-naïve PD patients with and without cognitive deficits. We compared vesicular acetylcholine transporter (VAChT) [¹⁸F]Fluoroethoxybenzovesamicol ([¹⁸F]FEOBV) PET imaging between *de novo* patients (with and without mild cognitive impairment) and healthy control (HC) subjects. Unlike previously used acetylcholinesterase PET ligands, [¹⁸F]FEOBV PET allows for detailed assessment of not only low level cortical but also high binding subcortical structures, such as the basal ganglia and the cerebellum (20–22).

Methods

Participants

57 Newly diagnosed patients with PD and 10 healthy control (HC) subjects were included in this cross-sectional study. Patients were enrolled in the Dutch Parkinson Cohort (DUPARC) study between 2017 and 2019 (For details: Boertien et al., 2020) (23). Inclusion criteria for patients consisted of PD diagnosis by a movement disorders specialist according to Movement Disorder Society (MDS) Clinical Diagnosis Criteria for PD (24) and with a confirmed dopaminergic striatal deficit on 3,4-dihydroxy-6-18F-fluoro-1-phenylalanine (¹⁸F-FDOPA) PET. HC had a normal neurological examination and did not have a history of neurological or psychiatric disorders. Exclusion criteria for both PD and HC subjects included the inability to provide written informed consent, the use of dopaminergic and (anti-)cholinergic medication and an estimated low premorbid intelligence level (estimated IQ <70, on the Dutch Adult Reading test (25)). All subjects gave written informed consent and the study was approved by the local ethics committee.

Clinical examination

All patients underwent comprehensive neuropsychological assessment covering all cognitive domains (23). A selection of outcome measures of tests and subtests of the cognitive test battery was made a priori, meeting level II criteria for PD-MCI (26,27), listed in table 1. Subject scores for each of the cognitive tests were compared to established test-specific normative data generated by age, gender and education. A performance of >1.5 standard deviation (SD) below normative values was considered abnormal. Patients were categorized as either PD with normal cognition (PD-NC) or PD-MCI. PD-MCI was based on level II criteria for PD-MCI and required below-threshold performance on at least two neuropsychological tests (26). Any patient or clinical characteristics possibly influencing performance on the neuropsychological assessments and MCI grouping, including visual difficulties, color blindness, speech problems and significant mood disorders were taken into account at the time of assessment and prior to data analysis, and if necessary excluded. HC subjects underwent cognitive testing using the Montreal Cognitive Assessment (MoCA) test.

Additional clinical assessment included the subjective duration of motor complaints before PD diagnosis and the Hospital Anxiety and Depression scale (HADS) (28). All PD subjects underwent motor examination using the MDS-revised Unified Parkinson's Disease Rating Scale part III (MDS-UPDRS-III). In addition, specific items from the MDS-UPDRS parts II and III were used for the classification of motor phenotypetype, using criteria previously formulated by Stebbins et al. (29). Patients were classified into tremor dominant (TD), postural instability and gait difficulty (PIGD), and indeterminate motor phenotypes

Image acquisition

All subjects underwent brain MRI and VAcH PET imaging, using (^{18}F)FEObv. MR imaging of PD subjects was acquired using Siemens Magnetom Prisma 3-Tesla magnetic resonance imaging scanners (Best, Netherlands) equipped with SENSE-8 channel head coils. For each subject, anatomical T1-weighted images were obtained using a sagittal 3-dimensional gradient-echo T1-weighted sequence with 0.9 x 0.9 x 0.9 mm acquisition. HC subjects underwent a T1-weighted MRI scan (3T Inera, Philips, The Netherlands) with 1.0 x 1.0 x 1.0 mm acquisition. (^{18}F)FEObv imaging was performed on the same day as MR imaging. After a low-dose CT for attenuation and scatter correction, participants were scanned using either a Biograph 40-mCT or 64-mCT (Siemens Healthcare, USA). Both these systems are EARL certified and identical in software version, acquisition- and reconstruction-protocols and PET detectors, and only differ in the number of CT slices. Patient and control subjects were randomly divided over the two PET scanners. (^{18}F)FEObv was injected using an intravenous bolus and delayed imaging was performed over 30 minutes (in six 5-minute frames) starting 210 minutes after injection.

Image processing was performed using Statistical Parametric Mapping (SPM) software (30). (^{18}F)FEObv PET imaging frames were spatially coregistered within subjects with a rigid-body transformation. The cropped T1-weighted MR scan was coregistered with the subject PET image. Freesurfer software package (Laboratory for Computational Neuroimaging, Athinoula A. Martinos Center for Biomedical Imaging, Boston, MA, USA) was used to segment MRI into cortical and subcortical brain regions. We calculated the distributed volume ratio (DVR) of each grey matter target region using the summed six delayed imaging frames and the white matter reference region as previously described (21,31). A parametric image for individual subjects was created by using the average of six delayed imaging frames divided by the mean of the white matter reference region. Partial volume correction on our parametric images was done using the Muller-Gartner method (32).

Imaging analysis

Voxel-based analysis was performed as previously described (33). Parametric PET images were spatially normalized to the Montreal Neurological Institute stereotactic template and smoothed with a Gaussian kernel of 8 mm full width half maximum. The relevant brain areas were displayed in Montreal Neurological Institute atlas coordinates (in millimeters) in the stereotactic space using the automated anatomical labeling toolbox.

Statistical analyses

To evaluate brain cholinergic innervation in *de novo* patients with PD, we first compared HC, PD-NC and PD-MCI groups on baseline demographic and clinical characteristics. Comparisons between all

three groups were performed using ANOVA for parametric variables and χ^2 testing for dichotomous variables. Comparisons between the two PD groups were performed using an independent samples t-test or a Mann-Whitney U test, for either normal and skewed distributed data, respectively. Statistical analysis was considered significant for $\alpha < 0.05$. Statistical analyses were performed using SPSS Statistics for Windows, Version 24.0 (IBM Statistics, USA) and SPM software.

Whole brain voxel-based analyses were performed to assess difference in cholinergic innervation between groups. Parametric [^{18}F]FEOBV DVR images were used for a two-sample voxel-wise statistical comparison between groups, in both positive and negative directions, controlling for age. The minimum cluster size was set to 50. The false discovery rate (FDR) approach was used for correction for multiple testing effects in the voxel-based analyses. Additional post-hoc Freesurfer-based volume-of-interest (VOI) analysis was performed on regions selected based on the voxel-based analysis. The percentage change of these regions was calculated by dividing the difference between the mean PD and HC group by the mean HC group.

Results

Demographic and clinical characteristics

The 57 *de novo* PD patients (32 males) had a mean (SD) age of 64.6 (8.2) years and a mean (SD) MDS-UPDRS-III score of 30.8 (11.4). 17 (29.8%) patients were classified as PD-MCI. Demographic and clinical characteristics of the different groups are listed in table 2. HC subjects had a significantly lower age and higher MoCA score compared to both PD groups ($p < 0.001$). The PD-MCI group had higher MDS-UPDRS-III scores ($p < 0.01$) and Hoehn and Yahr stage ($p = 0.05$) compared to the PD-NC group, reflecting more severe motor impairment in the PD-MCI group. No significant difference was found between the PD groups on the motor phenotype (TD, PIGD or indeterminate). Information on duration of motor complaints was missing for 11 patients. Available data showed no significant difference in the duration of motor symptom complaints prior to PD diagnosis.

Cognitive functioning

Overall, 15 out of 17 patients classified as PD-MCI showed multi-domain cognitive impairment, reflecting impaired scores in two or more cognitive domains. The most frequently affected domains were memory and executive functions, with 12 and 10 patients presenting impairments in these domains, respectively. The attention and visuospatial domains were affected in 7 out of 17 PD-MCI patients. Language was the least affected domain with 4 PD-MCI patients showing impaired scores. Detailed cognitive performance on the neuropsychological tests for both PD groups is listed in the supplementary materials.

Cholinergic innervation compared to controls.

Whole brain voxel-based analyses were performed to compare regional brain VAcHt binding between the PD groups (PD-MCI and PD-NC) and the HC subjects in positive and negative directions, while controlling for age. The PD groups showed evidence of bidirectional changes in different topographic patterns compared to the control subjects.

Significantly lower VAcHt binding was found in both PD groups, primarily in the temporal and posterior cortical regions, compared to HC (Figure 1; $p < 0.05$, FDR corrected). The topographic profiles for PD-NC and PD-MCI were comparable, with main affected regions including the parietal, parieto-occipital junction and lateral temporal cortices. PD-NC presented with additional lower binding in the frontal cortex, insula and caudate (tail), and PD-MCI with additional lower binding in the pons-medulla regions. A detailed overview of significant clusters is provided in the supplemental data.

Reverse direction voxel-based comparison between HC and both PD groups showed significantly higher VAcHt binding in PD subjects compared to HC ($p < 0.05$, FDR corrected). This higher cholinergic innervation was most prominently apparent in the PD-NC group (Figure 2A), including the cerebellum, the cingulate cortex, the gyrus rectus, the anteroventral striatum, putamen, the dorsal tegmentum, thalamus, metathalamus (lateral geniculate nucleus (LGN) and medial geniculate nucleus (MGN)), pulvinar, the amygdala, the hippocampus and parahippocampal regions. The PD-MCI groups showed a more limited topography of higher VAcHt binding, including the cerebellar vermis, the right gyrus rectus, amygdala, hippocampus and metathalamus, dorsal tegmentum, right inferior thalamus/pulvinarthalamus, and the left precentral gyrus (figure 2B). A detailed overview of significant clusters is provided in the supplemental data.

Most prominent representative examples of the magnitude of absolute binding changes with HC based on VOI comparison include reduced binding in the lateral occipital gyri (-13.3%), the pericalcarine (-19.0%), the transverse temporal gyri (-23.1%), and increases in the cerebellar cortex (+14.2%), the medial and lateral orbitofrontal region (resp +8.8% and +6.5%), frontal pole (+6.1%), the bilateral amygdala (+4.8%), and anterior cingulate (+3.8%) in the PD-NC group. For the PD-MCI group, representative examples include the lateral occipital gyri (-15.7%), the cuneus (-19.6%), pericalcarine (-23.9%), the middle temporal gyri (-14.2%) and the inferior parietal gyri (-15.4%) with lower VAcHt binding in PD-MCI, and increases in the cerebellar cortex (+13.2%), the right medial and lateral orbitofrontal region (resp +2.6% and +3.9%), and the right amygdala (+2.9%).

PD-MCI vs PD-NC group comparison

Lower VAcHt binding was found in the PD-MCI group in the fusiform and hippocampal region, the anterior insula and the left prefrontal cortex (uncorrected, $p < 0.001$). No significant differences were found after correction for multiple comparisons.

Post hoc: controlling for motor impairment

Clinical characteristics comparing both PD groups demonstrated a significant higher MDS-UPDRS-III score in patients with PD-MCI compared to PD-NC (table 2), indicating more severe motor impairment in the PD-MCI group. Bidirectional whole brain voxel-based analysis comparing both PD groups with controls was therefore repeated, controlling for both age and MDS-UPDRS-III.

Limited lower VAcHt binding was found in PD-NC and PD-MCI compared to HC ($p < 0.001$, uncorrected), including small regional differences in occipital and temporal regions.

Reverse direction analysis showed higher VAcHt binding in both PD-NC and PD-MCI compared to controls (figure 3; $p < 0.001$, uncorrected). PD-NC showed most extensive higher binding when compared to controls, with a cholinergic topography including the brainstem, gyrus rectus, posterior cingulate cortex, amygdala, and hippocampus and right parahippocampal region. The PD-MCI group demonstrated less widespread higher VAcHt binding, limited to the right superior temporal region, the left pulvinar, left optic radiation/occipital region and precentral gyrus.

Discussion

We investigated the topography of regional cholinergic innervation status in newly diagnosed, treatment-naïve patients with PD, with and without cognitive impairment, compared to healthy control subjects. We found altered cholinergic innervation, demonstrated by both higher and lower binding cholinergic brain regions. First, we demonstrate that cortical cholinergic denervation is already present at the start of the disease in both cognitively impaired and cognitively unaffected patients with PD. Second, we also found evidence of increased cholinergic binding in the PD groups. Cognitively unimpaired PD patients showed most robust evidence of higher-than-normal VAcHT binding compared to the PD-MCI group.

Previous studies have established the importance of the cholinergic system in cognitive impairment in PD, even in the absence of dementia (13,34). However, the majority of previous in vivo imaging studies are based on PET imaging using acetylcholinesterase tracers, which limit accurate estimates of high binding brain areas. The use of a VAcHT PET tracer, like [¹⁸F]FEOBV, allows for more reliable and regionally specific cholinergic assessment, especially in high binding areas such as the striatum and cerebellum (20–22). To our knowledge, this is the first assessment of regional cholinergic innervation in de novo patients with PD, with and without cognitive impairment, on a detailed cortical and subcortical level.

29.8% of patients in our study met PD-MCI criteria, which is in line with previous research on the cognitive impairment in early PD (3). In addition, the PD-MCI group presented with more severe motor impairment. However, there was no difference between PD-NC and PD-MCI in terms of motor phenotype (TD, PIGD or indeterminate) or duration of subjective motor complaints prior to PD diagnosis.

A striking finding in our study was the higher-than-normal cholinergic binding in de novo patients with PD compared to controls, most prominent in the PD-NC group. The brain topography represented by higher VAcHT binding included the cerebellum, parts of the thalamic complex, putamen, hippocampus and parahippocampal region, amygdala, gyrus rectus and cingulate cortex. Higher VAcHT binding in the cerebellum and thalamus indicate a substantial role of the cholinergic projections from the medial vestibular nucleus and PPN/LDTC, respectively (10,11). In addition, the involvement of the hippocampus, parahippocampal region, amygdala, gyrus rectus and cingulate cortex suggests involvement of projections originating from the basal forebrain, including Ch1-2 and Ch4 (Nucleus basalis of Meynert) groups (12). From the Ch4 cell group two major pathways can be identified; the medial pathway joining the white matter of the gyrus rectus and projecting to the

cingulum, and the lateral pathway of which the capsular division projects to the amygdala and temporal lobe (11). Finally, the altered [^{18}F]FEOBV uptake in the putamen and caudate nucleus indicate a role of intrinsic cholinergic striatal interneurons or PPN/LDTG projections (35). A possible role of the dopaminergic system should also be considered, as previous studies have shown an intricate interaction between the dopaminergic and cholinergic system; it is the multiplicative and interacting effects of the two systems that lead to cognitive deficits (17,36).

The higher cholinergic innervation in de novo PD suggests a compensatory cholinergic upregulation in the early phase of the disease. A compensatory role of the cholinergic system has previously been suggested in Alzheimer's disease (AD) where cholinergic upregulation was found in prodromal patients with MCI (37,38). Bohnen et al. (2015) and Kim et al. (2019) expanded on these findings by showing both independent and interactive roles of the cholinergic and dopaminergic system in cognitive functioning in PD (17,39). In the context of dopaminergic losses, preservation or even upregulation of cholinergic innervation may help preserve cognitive functioning (40). In addition, Liu et al (2018) reported increased cortical acetylcholinesterase activity related to LRRK2 gene mutation in (premanifest) PD (41). Bedard et al. (2019) recently showed significantly higher [^{18}F]FEOBV uptake in patients with idiopathic REM sleep behavior disorder (RBD) (42). RBD is considered an important marker of prodromal PD (43) and associated with cognitive impairment conversion (44). Our study is the first to demonstrate evidence of higher-than-normal cholinergic innervation in PD patients, substantiating the hypothesis of a compensatory cholinergic upregulation in very early PD.

Interestingly, the higher cholinergic VAcHT binding was less profound in the PD-MCI subgroup than in the cognitively unimpaired PD group, even though both groups consisted of newly diagnosed, treatment-naïve patients with PD with a similar duration of motor complaints prior to PD diagnosis. These findings suggest that a possible lack of compensatory cholinergic upregulation may be related to clinically evident cognitive impairment in early PD patients. In addition, the demonstrated denervation pattern in both PD-NC and PD-MCI groups provides further support for this hypothesis, as the relative extent of the denervation changes in the two PD subgroups had remarkable overlap and PD-MCI did not show more profound cholinergic denervation than PD-NC when compared to controls. In contrast, the PD-NC group showed slightly more extent denervation changes, possibly related to the higher number of subjects in this group providing more statistical power.

However, it should be noted that an alternative explanation for the difference in higher-than-normal VAcHT binding between PD-NC and PD-MCI may be related to the severity of motor impairment. The PD-MCI group presented with a higher MDS-UPDRS-III score than PD-NC, indicating more severe motor impairment. Therefore, a post-hoc analysis was performed, controlling for motor

impairment. Our data showed that the higher cholinergic binding was still present in PD-NC, but less widespread and robust. The upregulation in the cerebellum and thalamic complex was less profound, as well as the striatal regions, suggesting an important role of increased PPN cholinergic innervation and cholinergic striatal interneurons in motor performance in early PD. This is in line with previous research showing cholinergic PPN projections and striatal interneurons strongly contributing to motor symptoms in PD, especially in the motor subtype presenting with postural instability and gait disorder (31,45). In contrast, regions with basal forebrain cholinergic projections, including the posterior cingulate cortex, gyrus rectus, amygdala, hippocampus and parahippocampal region remained present after correction for this confounder variable. Interestingly, these regions show partial overlap with the regional cholinergic topography we previously found to be related to cognitive functioning in PD on the level of multiple cognitive domains (19), substantiating the cholinergic role of these regions in cognitive functioning in PD. On the other hand, controlling for MDS-UPDRS-III scores might cause overcorrection, as more severe motor and cognitive impairment often coincide (46,47) and MDS-UPDRS-III scores are significantly correlated with the majority of included cognitive tests (supplementary). The post-hoc analysis controlling for motor performance may therefore give an underestimation of the regional topography of higher cholinergic binding related to cognitive status.

In contrast to earlier findings (14,15,48), we found only limited cortical cholinergic denervation in patients with PD-MCI, without a substantial VAcHT binding differences versus PD-NC. A possible explanation for this lack of difference may be the heterogeneity of cognitive performance in patients with PD. In our study, the majority of patients with PD-MCI showed multidomain cognitive impairment, with a variety of domains affected. In addition, not only heterogeneity in cognitive functioning, but also in other PD symptoms, including motor profile and non-motor symptoms, may result in cholinergic heterogeneity (49). The finding of a limited cholinergic denervation pattern may also contribute to the understanding why previous studies have found limited effectiveness of cholinesterase inhibitors in PD-MCI (50,51). We suggest that early cognitive decline is the result of failing cholinergic compensation, rather than cholinergic denervation per se. A better stratification, including the specific profile of dopaminergic and cholinergic innervation, in addition to clinical cognitive performance, might therefore improve the effectiveness of cholinergic treatment in PD.

Limitations

One limitation of this study is the relatively small sample size of the PD subgroups. In addition, the PD-MCI group presented with a heterogeneous cognitive profile, affecting multiple domains. The clinical heterogeneity of this group and the relatively small sample size of the subgroups may have

contributed to the limited difference found between PD-MCI and PD-NC. Future studies with a larger sample size can allow for more detailed stratification and improve our understanding on possible compensatory mechanisms specific to cognitive functioning across different domains. Second, even though MDS guidelines were followed, the grouping of patients into PD-NC and PD-MCI is an arbitrary process, as cognitive changes occur along a continuous spectrum. This might add to the heterogeneity. Furthermore, the cross-sectional design of our study does not allow assessment of temporal changes. More detailed data on cognitive performance over time and the progression to PD-MCI and PDD will enhance our understanding of the cholinergic role in the progression of the disease and the suggested compensatory mechanism. Finally, the HC group had a significantly lower age than both PD groups. We previously demonstrated an important role of age in the relationship between cognition and cholinergic innervation (19). Although the analyses we corrected for age, a possible role of the age difference can not be ruled out.

Conclusion

This study demonstrates evidence of bidirectional changes in cholinergic innervation in de novo patients with PD, with and without cognitive impairment, compared to healthy controls. Increased cholinergic binding in early PD, especially in the cognitively intact patients, suggests a compensatory cholinergic upregulation in this group. Taken together, we postulate that in early, treatment-naïve patients with PD, the clinical syndrome of PD-MCI may be related to limited or failing cholinergic upregulation, instead of a more progressed (posterior cortical) cholinergic denervation.

Author contributions

Study concept and design: van der Zee, Boertien, Gerritsen, Spikman, van Laar

Acquisition, analysis or interpretation of the data: van der Zee, Kanel, Slomp, Boertien, van Laar

Drafting of the manuscript: van der Zee

Critical revision of the manuscript: all authors

Statistical analysis: van der Zee, Kanel

Obtained funding: van der Zee, van Laar

Conflict of Interest Disclosures

Dr. van der Zee, dr. Kanel, ms Slomp, dr. Gerritsen and dr. Spikman have nothing to disclose. JM Boertien has received a writer's fee from Britannia Pharmaceuticals (Kinetic Magazine) and owns exchange traded funds (ETFs) that might include stocks from medically related fields. Dr. Müller has received research funding from the NIH, Department of Veteran Affairs and the Michael J. Fox Foundation. Dr. Bohnen has received research funding from the NIH, Department of Veterans Affairs, Parkinson's Foundation, Farmer Family Foundation, the Michael J. Fox Foundation, Eisai, and EIP Pharma. He has participated in an Eisai advisory board and received in kind research support from Scion Neurostim, Expansion Therapeutics and Innovative Health Solutions. Dr. van Laar has received research funding from Weston Brain Institute, Lysosomal Therapeutics, the Michael J Fox Foundation, the Dutch Brain Institute and the UMCG. He received lecture- and consultancy fees from AbbVie, Britannia and PureIMS.

Additional contributors: The authors thank all patients, caregivers, health care professionals and students who have contributed and collaborated in this project. The inclusion of patients was established with the help of the collaborative Parkinson Platform Northern Netherlands (PPNN).

References

1. Post B, Muslimovic D, van Geloven N, Speelman JD, Schmand B, de Haan RJ, et al. Progression and prognostic factors of motor impairment, disability and quality of life in newly diagnosed Parkinson's disease. *Mov Disord*. 2011 Feb 15;26(3):449–56.
2. Lawson RA, Yarnall AJ, Duncan GW, Khoo TK, Breen DP, Barker RA, et al. Severity of mild cognitive impairment in early Parkinson's disease contributes to poorer quality of life. *Parkinsonism Relat Disord*. 2014;20(10):1071–5.
3. Aarsland D, Bronnick K, Williams-Gray C, Weintraub D, Marder K, Kulisevsky J, et al. Mild cognitive impairment in Parkinson disease: a multicenter pooled analysis. *Neurology*. 2010 Sep 21;75(12):1062–9.
4. Domellöf ME, Ekman U, Forsgren L, Elgh E. Cognitive function in the early phase of Parkinson's disease, a five-year follow-up. *Acta Neurol Scand*. 2015;132(2):79–88.
5. Aarsland D, Andersen K, Larsen JP, Lolk A, Kragh-Sorensen P. Prevalence and characteristics of dementia in Parkinson disease: an 8-year prospective study. *Arch Neurol*. 2003;60(3):387–92.
6. Palavra NC, Naismith SL, Lewis SJG. Mild cognitive impairment in Parkinson's disease: a review of current concepts. *Neurol Res Int*. 2013;2013:576091.
7. Halliday GM, Leverenz JB, Schneider JS, Adler CH. The neurobiological basis of cognitive impairment in Parkinson's disease. *Mov Disord*. 2014 Apr 15;29(5):634–50.
8. Bohnen NI, Albin RL. The cholinergic system and Parkinson disease. *Behav Brain Res*. 2011 Aug 10;221(2):564–73.
9. Ballinger EC, Ananth M, Talmage DA, Role LW. Basal Forebrain Cholinergic Circuits and Signaling in Cognition and Cognitive Decline. *Neuron* [Internet]. 2016;91(6):1199–218. Available from: <http://dx.doi.org/10.1016/j.neuron.2016.09.006>
10. Zhang C, Zhou P, Yuan T. The cholinergic system in the cerebellum: from structure to function. *Rev Neurosci*. 2016 Dec;27(8):769–76.
11. Selden NR, Gitelman DR, Salamon-Murayama N, Parrish TB, Mesulam MM. Trajectories of

- cholinergic pathways within the cerebral hemispheres of the human brain. *Brain* [Internet]. 1998;121 (Pt 1:2249–57. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/9874478>
12. Mesulam MM. The systems-level organization of cholinergic innervation in the human cerebral cortex and its alterations in Alzheimer's disease. *Prog Brain Res*. 1996;109:285–97.
 13. Hilker R, Thomas A V., Klein JC, Weisenbach S, Kalbe E, Burghaus L, et al. Dementia in Parkinson disease: Functional imaging of cholinergic and dopaminergic pathways. *Neurology*. 2005;65(11):1716–22.
 14. Klein JC, Eggers C, Kalbe E, Weisenbach S, Hohmann C, Vollmar S, et al. Neurotransmitter changes in dementia with Lewy bodies and Parkinson disease dementia in vivo. *Neurology*. 2010;74(11):885–92.
 15. Shimada H, Hirano S, Shinotoh H, Aotsuka a., Sato K, Tanaka N, et al. Mapping of brain acetylcholinesterase alterations in Lewy body disease by PET. *Neurology*. 2009;73(4):273–8.
 16. Bohnen NI, Müller MLTM, Kotagal V, Koeppe RA, Kilbourn MR, Gilman S, et al. Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *J Cereb Blood Flow Metab*. 2012;32(8):1609–17.
 17. Bohnen NI, Albin RL, Müller MLTM, Petrou M, Kotagal V, Koeppe R a, et al. Frequency of Cholinergic and Caudate Nucleus Dopaminergic Deficits Across the Predemented Cognitive Spectrum of Parkinson Disease and Evidence of Interaction Effects. *JAMA Neurol* [Internet]. 2015;72(2):194–200. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/25506674>
 18. Bohnen NI, Kaufer DI, Hendrickson R, Ivanko LS, Lopresti BJ, Constantine GM, et al. Cognitive correlates of cortical cholinergic denervation in Parkinson's disease and parkinsonian dementia. *J Neurol*. 2006;253(2):242–7.
 19. van der Zee S, Müller MLTM, Kanel P, van Laar T, Bohnen NI. Cholinergic Denervation Patterns Across Cognitive Domains in Parkinson's Disease. *Mov Disord*. 2020 Nov;
 20. Petrou M, Frey K a, Kilbourn MR, Scott PJH, Raffel DM, Bohnen NI, et al. In Vivo Imaging of Human Cholinergic Nerve Terminals with (-)-5-18F-Fluoroethoxybenzovesamicol:

Biodistribution, Dosimetry, and Tracer Kinetic Analyses. *J Nucl Med* [Internet].

2014;55(3):396–404. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24481024>

21. Nejad-Davarani S, Koeppe RA, Albin RL, Frey KA, Müller MLTM, Bohnen NI. Quantification of brain cholinergic denervation in dementia with Lewy bodies using PET imaging with [18F]-FEOBV. *Mol Psychiatry*. 2019;24(3):322–7.
22. van der Zee S, Vallez Garcia D, Elsinga PH, et al. [18F]Fluoroethoxybenzovesamicol in Parkinson's disease patients: Quantification of a novel cholinergic positron emission tomography tracer. *Mov Disord*. 2019 Jun;34(6): 924–926.
23. Boertien JM, van der Zee S, Chrysou A, Gerritsen MJJ, Jansonius NM, Spikman JM, et al. Study protocol of the DUTch PARKinson Cohort (DUPARC): a prospective, observational study of de novo Parkinson's disease patients for the identification and validation of biomarkers for Parkinson's disease subtypes, progression and pathophysiology. *BMC Neurol*. 2020 Jun;20(1):245.
24. Postuma RB, Berg D, Stern M, Poewe W, Olanow CW, Oertel W, et al. MDS clinical diagnostic criteria for Parkinson's disease. *Mov Disord*. 2015 Oct;30(12):1591–601.
25. Schmand B, Bakker D, Saan R, Louman J. The Dutch Reading Test for Adults: a measure of premorbid intelligence level. *Tijdschr Gerontol Geriatr*. 1991 Feb;22(1):15–9.
26. Litvan I, Goldman JG, Tröster AI, Schmand B a., Weintraub D, Petersen RC, et al. Diagnostic criteria for mild cognitive impairment in Parkinson's disease: Movement Disorder Society Task Force guidelines. *Mov Disord*. 2012;27(3):349–56.
27. Goldman JG, Holden S, Ouyang B, Bernard B, Goetz CG, Stebbins GT. Diagnosing PD-MCI by MDS task force criteria: How many and which neuropsychological tests? *Mov Disord*. 2015;30(3):402–6.
28. Spinhoven P, Ormel J, Sloekers PP, Kempen GI, Speckens AE, Van Hemert AM. A validation study of the Hospital Anxiety and Depression Scale (HADS) in different groups of Dutch subjects. *Psychol Med*. 1997;27(2):363–70.

29. Stebbins GT, Goetz CG, Burn DJ, Jankovic J, Khoo TK, Tilley BC. How to identify tremor dominant and postural instability/gait difficulty groups with the movement disorder society unified Parkinson's disease rating scale: comparison with the unified Parkinson's disease rating scale. *Mov Disord*. 2013 May;28(5):668–70.
30. Wellcome Trust Centre for Neuroimaging.
<https://www.fil.ion.ucl.ac.uk/spm/software/spm12/>. University College, London, England.
31. Bohnen NI, Kanel P, Zhou Z, Koeppe RA, Frey KA, Dauer WT, et al. Cholinergic system changes of falls and freezing of gait in Parkinson's disease. *Ann Neurol*. 2019;85(4):538–49.
32. Muller-Gartner HW, Links JM, Prince JL, Bryan RN, McVeigh E, Leal JP, et al. Measurement of radiotracer concentration in brain gray matter using positron emission tomography: MRI-based correction for partial volume effects. *J Cereb Blood Flow Metab*. 1992 Jul;12(4):571–83.
33. Kanel P, Müller MLTM, van der Zee S, Sanchez-Catasus CA, Koeppe RA, Frey KA, et al. Topography of Cholinergic Changes in Dementia With Lewy Bodies and Key Neural Network Hubs. *J Neuropsychiatry Clin Neurosci*. 2020;32(4):370–5.
34. Bohnen NI, Müller MLTM, Kotagal V, Koeppe RA, Kilbourn MR, Gilman S, et al. Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *J Cereb Blood Flow Metab*. 2012;32(8):1609–17.
35. Dautan D, Huerta-Ocampo I, Witten IB, Deisseroth K, Bolam JP, Gerdjikov T, et al. A major external source of cholinergic innervation of the striatum and nucleus accumbens originates in the brainstem. *J Neurosci*. 2014 Mar;34(13):4509–18.
36. Ztaou S, Amalric M. Contribution of cholinergic interneurons to striatal pathophysiology in Parkinson's disease. *Neurochem Int*. 2019 Jun;126:1–10.
37. Ikonomic MD, Abrahamson EE, Isanski BA, Wu J, Mufson EJ, DeKosky ST. Superior frontal cortex cholinergic axon density in mild cognitive impairment and early Alzheimer disease. *Arch Neurol*. 2007 Sep;64(9):1312–7.
38. DeKosky ST, Ikonomic MD, Styren SD, Beckett L, Wisniewski S, Bennett DA, et al.

- Upregulation of choline acetyltransferase activity in hippocampus and frontal cortex of elderly subjects with mild cognitive impairment. *Ann Neurol*. 2002 Feb;51(2):145–55.
39. Kim K, Bohnen NI, Muller MLTM, Lustig C. Compensatory dopaminergic-cholinergic interactions in conflict processing: Evidence from patients with Parkinson’s disease. *Neuroimage*. 2019 Apr;190:94–106.
40. Bohnen NI, Grothe MJ, Ray NJ, Müller MLTM, Teipel SJ. Recent Advances in Cholinergic Imaging and Cognitive Decline—Revisiting the Cholinergic Hypothesis of Dementia. *Curr Geriatr Reports*. 2018;7(1):1–11.
41. Liu S-Y, Wile DJ, Fu JF, Valerio J, Shahinfard E, McCormick S, et al. The effect of LRRK2 mutations on the cholinergic system in manifest and premanifest stages of Parkinson’s disease: a cross-sectional PET study. *Lancet Neurol*. 2018 Apr;17(4):309–16.
42. Bedard M-A, Aghourian M, Legault-Denis C, Postuma RB, Soucy J-P, Gagnon J-F, et al. Brain cholinergic alterations in idiopathic REM sleep behaviour disorder: a PET imaging study with (18)F-FEOBV. *Sleep Med*. 2019 Jun;58:35–41.
43. Postuma RB. Prodromal Parkinson’s disease--using REM sleep behavior disorder as a window. *Parkinsonism Relat Disord*. 2014 Jan;20 Suppl 1:S1-4.
44. Rahayel S, Postuma RB, Montplaisir J, Mišić B, Tremblay C, Vo A, et al. A Prodromal Brain-Clinical Pattern of Cognition in Synucleinopathies. *Ann Neurol*. 2020 Nov;
45. Gilman S, Koeppe RA, Nan B, Wang C-N, Wang X, Junck L, et al. Cerebral cortical and subcortical cholinergic deficits in parkinsonian syndromes. *Neurology*. 2010 May;74(18):1416–23.
46. Wojtala J, Heber IA, Neuser P, Heller J, Kalbe E, Rehberg SP, et al. Cognitive decline in Parkinson’s disease: the impact of the motor phenotype on cognition. *J Neurol Neurosurg Psychiatry*. 2019 Feb;90(2):171–9.
47. Poletti M, Frosini D, Pagni C, Baldacci F, Nicoletti V, Tognoni G, et al. Mild cognitive impairment and cognitive-motor relationships in newly diagnosed drug-naive patients with

Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 2012 Jun;83(6):601–6.

48. Hilker R, Thomas a. V., Klein JC, Weisenbach S, Kalbe E, Burghaus L, et al. Dementia in Parkinson disease: Functional imaging of cholinergic and dopaminergic pathways. *Neurology*. 2005;65(11):1716–22.
49. Müller MLTM, Bohnen NI. Cholinergic dysfunction in parkinson's disease. *Curr Neurol Neurosci Rep*. 2013;13(9).
50. Grace J, Amick MM, Friedman JH. A double-blind comparison of galantamine hydrobromide ER and placebo in Parkinson disease. *J Neurol Neurosurg Psychiatry*. 2009 Jan;80(1):18–23.
51. Mamikonyan E, Xie SX, Melvin E, Weintraub D. Rivastigmine for mild cognitive impairment in Parkinson disease: A placebo-controlled study. *Mov Disord [Internet]*. 2015;00(00):n/a-n/a. Available from: <http://doi.wiley.com/10.1002/mds.26236>

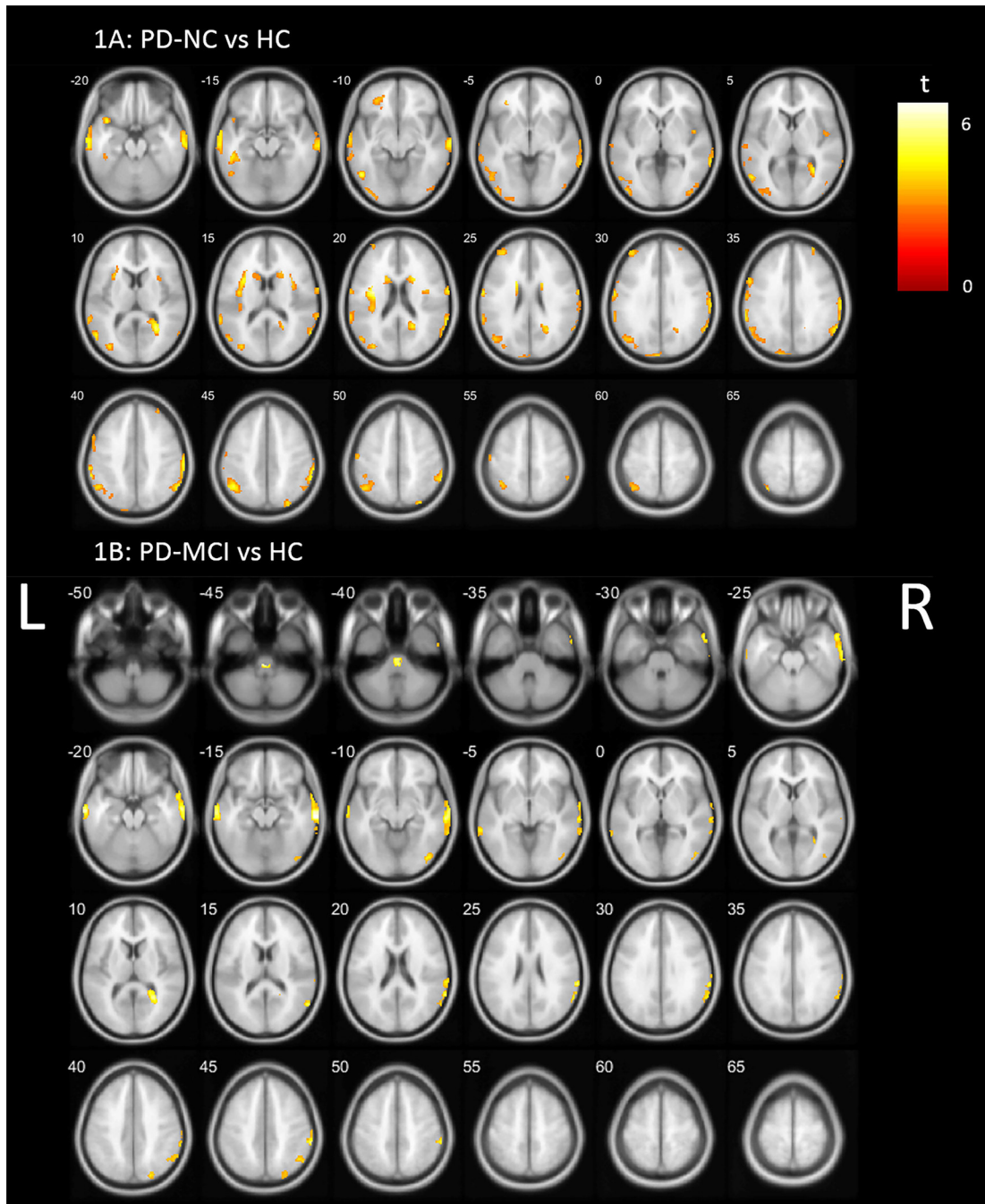


Figure1_vanderZee_denovoPD.jpg

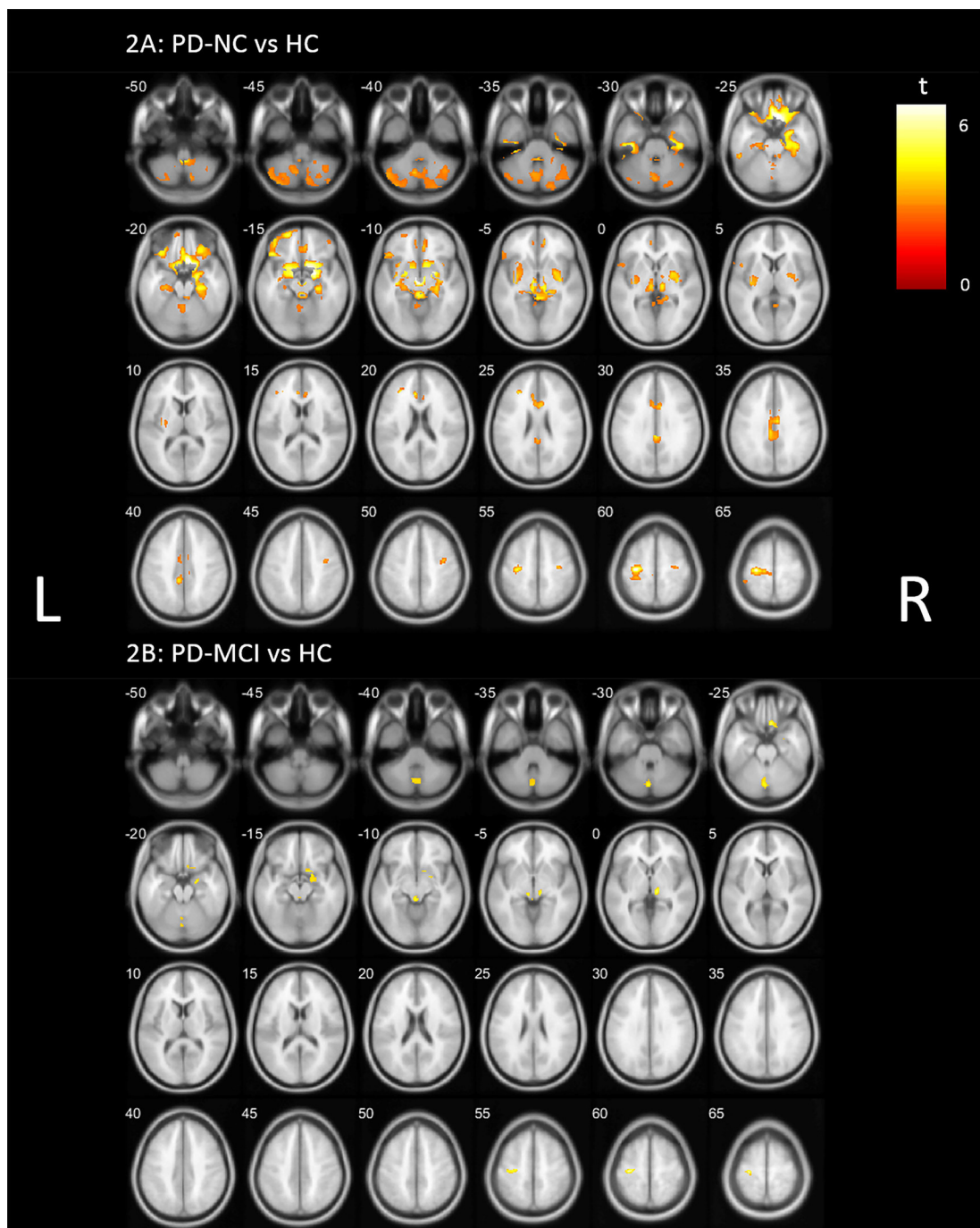


Figure2_vanderZee_denovoPD.jpg

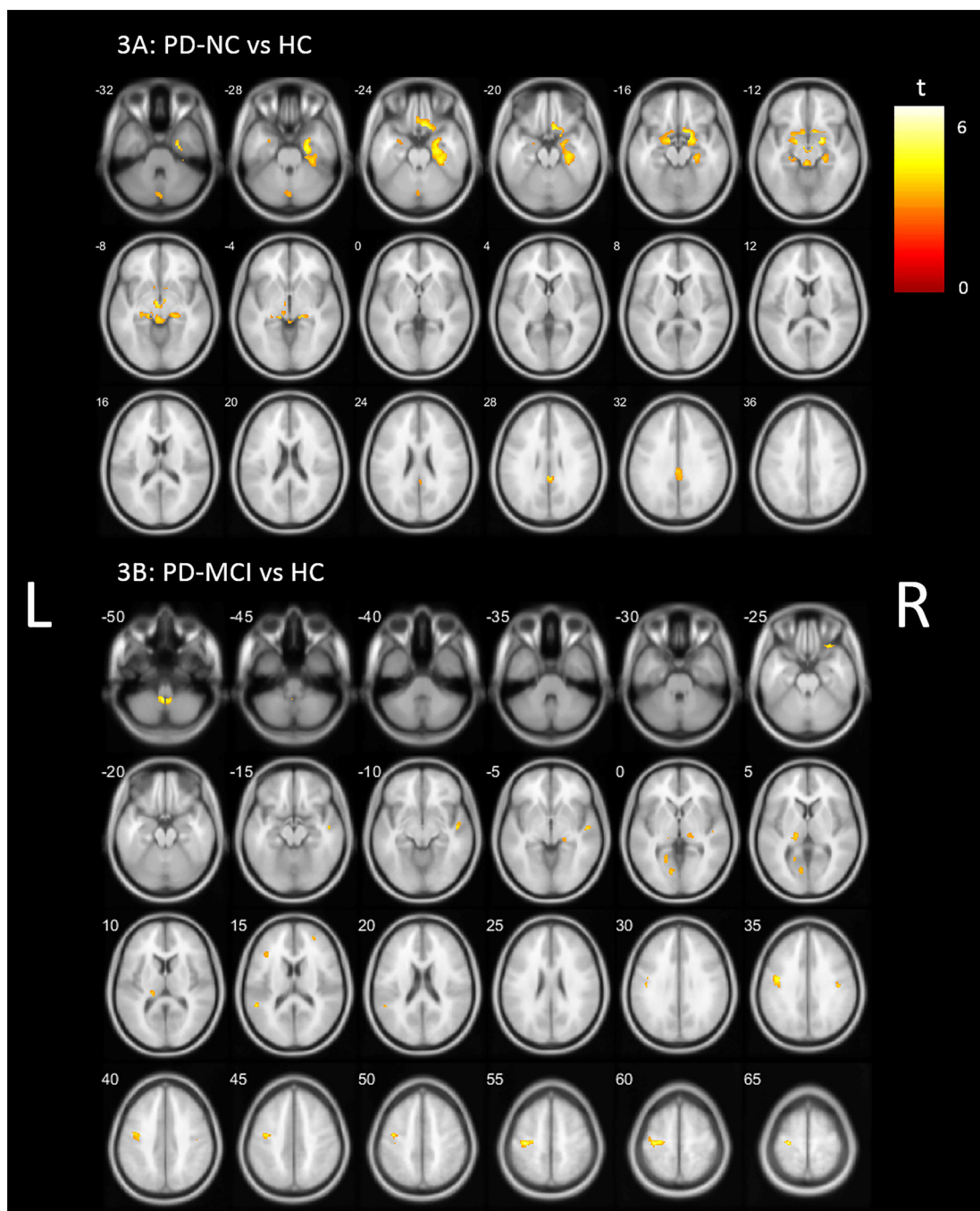


Figure3_vanderZee_denovoPD.jpg

COPYRIGHT TRANSFER AGREEMENT

Date: Dec 15, 2021

Contributor name: Sygrid van der Zee

Contributor address: Hanzeplein 1, 9713 GZ, Groningen, The Netherlands

Manuscript number: MDS-21-1308.R1

Re: Manuscript entitled: Altered cholinergic innervation in de novo Parkinson's disease with and without cognitive impairment (the "Contribution")

for publication in: Movement Disorders (the "Journal")

Published by Wiley on behalf of The International Parkinson and Movement Disorder Society (the "Owner")

Dear Contributor(s):

Thank you for submitting your Contribution for publication. In order to expedite the editing and publishing process and enable the Owner to disseminate your Contribution to the fullest extent, we need to have this Copyright Transfer Agreement executed. If the Contribution is not accepted for publication, or if the Contribution is subsequently rejected, this Agreement shall be null and void. **Publication cannot proceed without a signed copy of this Agreement.**

A. COPYRIGHT

The Contributor assigns to the Owner, during the full term of copyright and any extensions or renewals, all copyright in and to the Contribution, and all rights therein, including but not limited to the right to reproduce, publish, republish, transmit, sell, transfer, distribute, and otherwise use the Contribution in whole or in part in electronic and print editions of the Journal and in derivative works throughout the world, in all languages and in all media of expression now known or later developed, and to license or permit others to do so.

B. RETAINED RIGHTS

Notwithstanding the above, the Contributor or, if applicable, the Contributor's employer, retains all proprietary rights other than copyright, such as patent rights, in any process, procedure or article of manufacture described in the Contribution. This reservation of rights does not affect or limit the rights assigned to Owner in Section A.

C. PERMITTED USES BY CONTRIBUTOR

1. License. The Owner grants to Contributor a non-exclusive, non-transferable and limited license to reproduce and distribute copies of the print or electronic "preprints" of the unpublished Contribution, in the original form submitted to the Journal prior to the peer review process, solely to colleagues within the Contributor's nonprofit organization or educational institution. The Contributor shall make no more than 100 printed copies of the preprints in any calendar year. Such preprints may be posted as electronic files on the Contributor's own personal website, on the Contributor's internal intranet at Contributor's nonprofit organization or educational institution, or on a secure external website at the Contributor's nonprofit organization or educational institution, provided that access is limited to employees and/or students at Contributor's non-profit organization or educational institution. Contributor shall not charge a fee for any

preprints, and Contributor's use under this Section C shall not be for any commercial purpose, or for any systematic external distribution (e.g., posting on a listserve, public website, database connected to a public access server, or automated delivery system). The license grant in this Section does not apply to for-profit corporations, and any proposed use outside of the scope of this Section C must be pre-approved in writing by the Owner. The rights granted to Contributor under this Section C do not include reproduction, distribution or any other use of rating scales, videos or other audiovisual materials associated with the Contribution.

2. Required Citation. Prior to publication, the Contributor must provide full credit and acknowledgement of the Journal in all preprints in the following format: This is a preprint of an article accepted for publication in [Journal Title], Copyright © [year] The International Parkinson and Movement Disorder Society. After publication, the Contributor must provide a citation to the Journal in all preprints in the following format: This is a preprint of an article that was published in [Journal title]: (Title of Article, Contributor, Journal Title and Volume/ Issue, Copyright © [year] The International Parkinson and Movement Disorder Society). An electronic link must be provided to the Journal's website, located at <http://www.interscience.Wiley.com>. The Contributor agrees not to update the preprint or replace it with the published version of the Contribution.

3. Accepted Version. Re-use of the accepted and peer-reviewed (but not the final typeset published) version of the Contribution (the "Accepted Version") is not permitted under this Agreement. There are separate arrangements with certain funding agencies governing reuse of the Accepted Version. Additional terms apply if the Contributor receives or received funding from these agencies. The details of those relationships, and other offerings allowing open web use, are set forth at the following website: <http://www.wiley.com/go/funderstatement>.

4. Additional Terms for Certain Funders. Certain funders, including the NIH, members of the Research Councils UK (RCUK) and Wellcome Trust require deposit of the Accepted Version in a public repository after an embargo period. Details of funding arrangements are set out at the following website: <http://www.wiley.com/go/funderstatement>. Additional terms may be applicable. Please contact the production editor for the journal at MDSprod@wiley.com if you have additional funding requirements.

If any Contributor receiving funds from applicable sources does not choose the Owner's OnlineOpen option, the Contributor will be allowed to self-archive by depositing the Accepted Version in a public repository after the following applicable embargo period has expired, subject to further conditions imposed by the RCUK:

- a. 12 months from first publication online of the final published version of the Contribution for research funded by members of the Research Councils UK (RCUK) other than The Economic and Social Research Council (ESRC) and the Arts and Humanities Research Council (AHRC); or
- b. 24 months from first publication online of the final published version of the Contribution for research funded by ESRC or AHRC.

5. Additional Terms for Certain Institutions. Wiley has arrangements with certain educational institutions to permit the deposit of the Accepted Version in the institutional repository after an embargo period. Details of such arrangements are set out at the following website: <http://olabout.wiley.com/WileyCDA/Section/id-406074.html>. Additional terms may be applicable.

If any Contributor affiliated with these applicable educational institutions does not choose the Owner's OnlineOpen option, the Contributor will be allowed to self-archive by depositing the Accepted Version in the educational institution's repository after the following applicable embargo period has expired. See the following website for details: <http://olabout.wiley.com/WileyCDA/Section/id-817011.html>.

D. CONTRIBUTIONS OWNED BY EMPLOYER

If the Contribution was written by the Contributor in the course of the Contributor's employment (as a "work-made-for-hire" in the course of employment), the Contribution is owned by the company/institution which must execute this Agreement (in addition to the Contributor's signature). In such case, the company/institution hereby assigns to the Owner, during the full term of copyright, all copyright in and to the Contribution for the full term of copyright throughout the world as specified in Section A above.

E. GOVERNMENT CONTRACTS

In the case of a Contribution prepared under U.S. Government contract or grant, the U.S. Government may reproduce, without charge, all or portions of the Contribution and may authorize others to do so, for official U.S. Government purposes only, if the U.S. Government contract or grant so requires. (U.S. Government, U.K. Government, and other government employees: see notes at end.)

F. CONTRIBUTOR'S REPRESENTATIONS

The Contributor represents that the Contribution is the Contributor's original work, all individuals identified as Contributors actually contributed to the Contribution, and all individuals who contributed are included. The Contribution is submitted only to this Journal and has not been published before. (If excerpts from copyrighted works owned by third parties are included, the Contributor will obtain written permission from the copyright owners for all uses as set forth in the Journal's Instructions for Contributors, and show credit to the sources in the Contribution.) The Contributor also warrants that the Contribution contains no libelous or unlawful statements, does not infringe upon the rights (including without limitation the copyright, patent or trademark rights) or the privacy of others, or contain material or instructions that might cause harm or injury. Upon request, Contributor will provide the data or will cooperating fully in obtaining and providing the data on which the Contribution is based for examination by the editors or their assignees.

G. FINANCIAL DISCLOSURES

The Contributor certifies that his/her financial and material support for this research and work, regardless of date, is clearly identified on Exhibit A to this Agreement. The Contributor has also identified on Exhibit A, all other support unrelated to this research, covering the past year from the date of submission (e.g., grants, advisory boards, employment, consultancies, contracts, honoraria, royalties, expert testimony, partnerships, or stock ownership in medically-related fields).

H. VIDEO AND PHOTOGRAPHY CONSENT

In the event that the Contribution includes, discloses or incorporates any content (including, without limitation, any video clip or photograph) which identifies any individual patient(s) ("patient identifiable content"), the Contributor obtained from such patient(s) written consent to such inclusion, disclosure or incorporation and that this consent fully complies with all legal requirements, including without limitation, all of the requirements of the laws of the jurisdiction(s) to which the patient(s) and the patient(s)' physician are subject, including the United States Health Insurance Portability and Accountability Act of 1996 ("HIPAA") if applicable. The Contributor hereby certifies that, if the patient consent form is in a language other than English, such consent form meets all of the requirements set forth in the Instructions to Authors. In addition, the Contributor hereby confirms that he/she obtained from patient(s) written consent to use the patient identifiable content in both print and online (i.e., internet/web-based) publication formats. The Contributor further certifies that the person executing any such patient consent form, to the best of his/her knowledge, had legal capacity under applicable law to execute the form on behalf of the patient.

I. ACKNOWLEDGEMENTS


The Contributor should obtain written permission from all individuals named in the acknowledgement since readers may infer their endorsement of data and conclusions. The Contributor certifies that all individuals named in the acknowledgement section have provided written permission to be named.

J. MISCELLANEOUS

This Agreement may be amended or modified only in a writing executed by both parties. The waiver or failure of any party to exercise any rights under this Agreement shall not be deemed a waiver or other limitation of any other right or any future right. This Agreement shall inure to the benefit of, and shall be binding upon, the parties, their respective successors and permitted assigns. This Agreement may be executed in two (2) or more counterparts, each of which shall be an original and all of which taken together shall constitute one and the same agreement. Executed copies of this Agreement may be delivered by facsimile transmission, pdf/email or other comparable electronic means. If for any reason any provision of this Agreement shall be deemed by a court of competent jurisdiction to be legally invalid or unenforceable, the validity, legality and enforceability of the remainder of this Agreement shall not be affected and such provision shall be deemed modified to the minimum extent necessary to make such provision consistent with applicable law and, in its modified form, such provision shall then be enforceable and enforced. The parties agree to do such further acts and to execute and deliver such additional agreements and instruments from time to time as either may at any time reasonably request in order to assure and confirm unto such requesting party the rights, powers and remedies conferred in the Agreement. This Agreement, including any exhibits attached hereto, contains the entire agreement and understanding of the parties with respect to the subject matter hereof, and supersedes all prior agreements, negotiations, representations and proposals, written and oral, relating thereto.

All Contributors must sign below. Contributors must check one box except that NIH grantees should check both Contributor-owned work and the NIH grantee box. If your Contribution was written during the course of employment, your employer must also sign where indicated.

Please send your original completed and signed forms by fax or email a scanned copy to the Journal production editor. For production editor contact details please visit the Journal’s online author guidelines. Do not send in hard copies of these forms.

<input checked="" type="checkbox"/> Contributor-owned work	 _____ Contributor’s signature	Dec 15, 2021 _____ Date
	Dr. Sygrid van der Zee _____ Type or print name and title	
	_____ Co-Contributor’s signature	_____ Date
	_____ Type or print name and title	

Company/Institution-owned Work (made-for-hire in the Course of employment) _____
Company or Institution (Employer-for-Hire) _____ Date _____

Authorized signature of Employer _____ Date _____

Contributor's signature _____ Date _____

Type or print name and title

ATTACH ADDITIONAL SIGNATURE PAGES AS NECESSARY

U.S. Government work

Note to U.S. Government Employees

A contribution prepared by a U.S. federal government employee as part of the employee's official duties, or which is an official U.S. Government publication, is called a "U.S. Government work", and is in the public domain in the United States. In such case, Paragraph A.1 will not apply but the Contributor must type his/her name (in the Contributor's signature line) above. Contributor acknowledges that the Contribution will be published in the United States and other countries. If the Contribution was not prepared as part of the employee's duties or is not an official U.S. Government publication, it is not a U.S. Government work.

U.K. Government work (Crown Copyright)

Note to U.K. Government Employees

The rights in a contribution prepared by an employee of a UK government department, agency or other Crown body as part of his/her official duties, or which is an official government publication, belong to the Crown. Contributors must ensure they comply with departmental regulations and submit the appropriate authorisation to publish. If your status as a government employee legally prevents you from signing this Agreement, please contact the Journal production editor.

Other

Including Other Government work or Non-Governmental Organisation work

Note to Non-U.S., Non-U.K. Government Employees or Non-Governmental Organisation Employees

If your status as a government or non-governmental organisation employee legally prevents you from signing this Agreement, please contact the Journal production editor.

Exhibit A

Financial Disclosure

The Contributor has received financial and material support for this research and work regardless of date from the following sources:

Name: _____

Address: _____

Type of support: _____

This material will be printed with the published article.

In the past year from the date of submission, the Contributor has also received the following support unrelated to this research (e.g., grants, advisory boards, employment, consultancies, contracts, honoraria, royalties, expert testimony, partnerships, or stock ownership in medically-related fields):

Name: _____

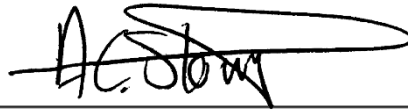
Address: _____

Type of support: _____

This material will be posted on the journal website and may be printed at the Editors' discretion.

ATTACH ADDITIONAL INFORMATION AS NECESSARY

Contributor-owned work



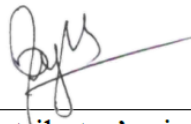
Contributor's signature

Dec. 1st 2021

Date

A.C. Slomp, Msc.

Type or print name and title



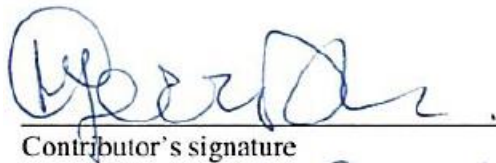
Co-Contributor's signature

12/01/2021

Date

Prabhesh Kanel

Type or print name and title



Contributor's signature

13-12-2021

Date

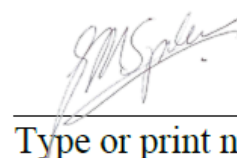
dr. M.Y.Y. Geeritsen

Type or print name and title

Co-Contributor's signature

Date

12-13-2021



Type or print name and title

prof.dr. JM Spikman

Martijn Muller

Contributor's signature


12/8/2021

Date

Martijn Muller, PhD

Type or print name and title

Contributor-owned work



Contributor's signature

12/01/21
Date

Nicolaas Bohnen, MD, PhD

Type or print name and title

U.S. Government work

Note to U.S. Government Employees

A contribution prepared by a U.S. federal government employee as part of the employee's official duties, or which is an official U.S. Government publication, is called a "U.S. Government work", and is in the public domain in the United States. In such case, Paragraph A.1 will not apply but the Contributor must type his/her name (in the Contributor's signature line) above. Contributor acknowledges that the Contribution will be published in the United States and other countries. If the Contribution was not prepared as part of the employee's duties or is not an official U.S. Government publication, it is not a U.S. Government work.

Exhibit A

Financial Disclosure

The Contributor has received financial and material support for this research and work regardless of date from the following sources:

Name: NIH, Bethesda, MD, US

Address: _____

Type of support: _____

This material will be printed with the published article.

In the past year from the date of submission, the Contributor has also received the following support unrelated to this research (e.g., grants, advisory boards, employment, consultancies, contracts, honoraria, royalties, expert testimony, partnerships, or stock ownership in medically-related fields):

Name: VA, Washington, DC, US

Address: _____

Type of support: _____

This material will be posted on the journal website and may be printed at the Editors' discretion.

ATTACH ADDITIONAL INFORMATION AS NECESSARY

Parkinson Foundation, NY, NY, US
Farmer Family Foundation, Cincinnati, OH, US

Contributor-owned work



Contributor's signature

Dec. 3rd 2021

Date

J.M. Boertien, MD

Type or print name and title

Exhibit A

Financial Disclosure

The Contributor has received financial and material support for this research and work regardless of date from the following sources:

Name: _____

Address: _____

Type of support: _____

This material will be printed with the published article.

In the past year from the date of submission, the Contributor has also received the following support unrelated to this research (e.g., grants, advisory boards, employment, consultancies, contracts, honoraria, royalties, expert testimony, partnerships, or stock ownership in medically-related fields):

J.M. Boertien

Name: _____

Tussen beide Markten 33, 9712CC Groningen

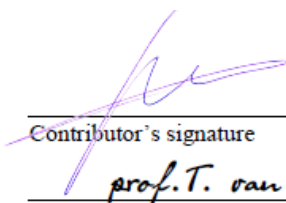
Address: _____

Type of support: Writers fee from Britannia Pharmaceuticals (Kinetic magazine). Owns ETFs that might include stocks from medically related fields.

This material will be posted on the journal website and may be printed at the Editors' discretion.

ATTACH ADDITIONAL INFORMATION AS NECESSARY

[] Contributor-owned work


Contributor's signature

December 6th. 2021
Date

prof. T. van Laar
Type or print name and title

Exhibit A

Financial Disclosure

The Contributor has received financial and material support for this research and work regardless of date from the following sources:

Name: Weston Brain Institute

Address: 22 St. Clair Avenue East, Suite 2001, Toronto, ON
Canada, M4T 2S3

Type of support: Study Grant

This material will be printed with the published article.

In the past year from the date of submission, the Contributor has also received the following support unrelated to this research (e.g., grants, advisory boards, employment, consultancies, contracts, honoraria, royalties, expert testimony, partnerships, or stock ownership in medically-related fields):

Name: prof. T. van Laar

Address: Hanzeplein 1, 9713GZ, Groningen, Netherlands

Type of support: Grant support from: MJFF, Dutch Brain Foundation, UMCG
Consultancy fees from AbbVie, Britannia Pharm. and Neuroderm

This material will be posted on the journal website and may be printed at the Editors' discretion.

Lecture fees from AbbVie, Centrapharm and Britannia Pharm.

ATTACH ADDITIONAL INFORMATION AS NECESSARY

Altered cholinergic innervation in de novo Parkinson's disease with and without cognitive impairment

Running title: Altered cholinergic innervation in de novo Parkinson

Sygrid van der Zee, PhD,^{1,2}, Prabesh Kanel, PhD,^{3,4}, Marleen J.J. Gerritsen, PhD,^{1,2}, Jeffrey M. Boertien, MD,¹, Anne C. Slomp, MSc,^{1,2}, Martijn L.T.M. Müller, PhD,^{3,4}, Nicolaas I. Bohnen, MD, PhD,^{3,4,5,6,7}, Jacoba M. Spikman, PhD,^{1,2}, Teus van Laar, MD, PhD,¹

¹ Department of Neurology, University of Groningen, University Medical Center Groningen, Groningen, the Netherlands

² Department of Neurology, Division of Clinical Neuropsychology, University of Groningen, University Medical Center, The Netherlands.

³ Department of Radiology, University of Michigan, Ann Arbor, MI, United States.

⁴ Morris K. Udall Center of Excellence for Parkinson's Disease Research, University of Michigan, Ann Arbor, MI, United States.

⁵ Department of Neurology, University of Michigan, Ann Arbor, MI, United States.

⁶ Neurology Service and GRECC, Veterans Administration Ann Arbor Healthcare System, Ann Arbor, MI, United States

⁷ University of Michigan Parkinson's Foundation Center of Excellence, Ann Arbor, MI, United States

Correspondence: dr. S. van der Zee Department of Neurology, University Medical Center Groningen, Hanzeplein 1, 9713 GZ Groningen, the Netherlands. +31 50361 2401. Email: s.van.der.zee01@umcg.nl

Wordcount: 3665 words

Funding sources for the study: Weston Brain institute, 22 St. Clair Avenue East, Suite 2001, Toronto, ON Canada M4T 2S3, Email: info@westonbrain.org, Telephone: +1-416-967-7979

Trial registration: NCT04180865

No conflicts of interest related to the manuscript

Abstract

Background: Altered cholinergic innervation plays a putative role in cognitive impairment in Parkinson's disease (PD), at least in advanced stages. Identification of the relationship between cognitive impairment and cholinergic innervation early in the disease will provide better insight in disease prognosis and possible early intervention.

Objective: To assess regional cholinergic innervation status in de novo patients with PD, with and without cognitive impairment.

Methods: 57 newly diagnosed, treatment-naïve, PD patients (32 males, mean age 64.6 ± 8.2 years) and 10 healthy control subjects (5 males, mean age of 54.6 ± 6.0 years) were included. All participants underwent cholinergic [^{18}F]Fluoroethoxybenzovesamicol PET, and detailed neuropsychological assessment. PD patients were classified as either cognitively normal (PD-NC) or mild cognitive impairment (PD-MCI). Whole brain voxel-based group comparisons were performed.

Results: Results show bidirectional cholinergic innervation changes in PD. Both PD-NC and PD-MCI groups showed significant cortical cholinergic denervation compared to controls ($p < 0.05$, FDR corrected), primarily in the posterior cortical regions. Higher than normal binding was most prominently present in PD-NC in both cortical and sub-cortical regions, including the cerebellum, cingulate cortex, putamen, gyrus rectus, hippocampus and amygdala.

Conclusion: Altered cholinergic innervation is already present in de novo patients with PD. Posterior cortical cholinergic losses were present in all patients independent of cognitive status. Higher than normal binding in cerebellar, frontal and subcortical regions in cognitively intact patients may reflect compensatory cholinergic upregulation in early-stage PD. Limited or failing cholinergic upregulation may play an important role in early, clinically evident cognitive impairment in PD.

Abbreviations:

[¹⁸F]FEOBV: [¹⁸F]Fluoroethoxybenzovesamicol

FDR: False Discovery Rate

HC: healthy controls

LED: Levodopa Equivalent Dose

MDS-UPDRS-III: Movement Disorder Society - Revised Unified Parkinson's Disease Rating Scale part III

PD: Parkinson's disease

PD-MCI: Parkinson's disease Mild Cognitive Impairment

PC-NC: Parkinson's disease Normal Cognition

PET: Positron Emission Tomography

VACHT: Vesicular Acetylcholine Transporter

Introduction

Cognitive impairment is common in Parkinson's disease (PD) and a major source of disability and lower quality of life (1,2). Mild cognitive impairment (PD-MCI) is already present in 25-30% of newly diagnosed patients and is a major risk factor for the development of PD dementia (PDD) (3–5). Cognitive impairment in PD is heterogeneous with multiple domains affected and great variability in onset and progression (6).

It has been increasingly recognized that comorbid cholinergic dysfunction is a major contributor to the pathophysiology of cognitive impairment in PD (7,8). The four major human brain cholinergic systems are the basal forebrain corticopetal (BF) projection system, cholinergic efferents of the peduncopontine-laterodorsal tegmental complex (PPN/LDTC), medial vestibular nucleus (MVN) cholinergic neurons projecting to the cerebellum, and striatal cholinergic interneurons (9–12).

Previous *in vivo* neuroimaging assessment of cholinergic innervation, using acetylcholinesterase PET, has demonstrated predominantly posterior cholinergic denervation in PD, with more severe cholinergic degeneration in PDD (13–16). There is additional evidence that the cholinergic system is a major driver of cognitive impairment in PD even in the absence of dementia (17). We previously showed that deficits in attention, executive functioning and memory correlated with loss of cholinergic activity on both a global cortical level (16,18) as well as on a regional (sub)cortical level (19). Although cholinergic denervation has been shown in cognitively impaired patients with PD, less is known about cholinergic innervation changes in early-stage disease and before the onset of cognitive changes. Elucidation of the relationship between cognitive impairment and cholinergic integrity in a very early stage of the disease may provide new clues that may inform novel therapeutic strategies.

The goal of this study was to characterize cholinergic innervation status in newly diagnosed, treatment-naïve PD patients with and without cognitive deficits. We compared vesicular acetylcholine transporter (VAChT) [¹⁸F]Fluoroethoxybenzovesamicol ([¹⁸F]FEOBV) PET imaging between *de novo* patients (with and without mild cognitive impairment) and healthy control (HC) subjects. Unlike previously used acetylcholinesterase PET ligands, [¹⁸F]FEOBV PET allows for detailed assessment of not only low level cortical but also high binding subcortical structures, such as the basal ganglia and the cerebellum (20–22).

Methods

Participants

57 Newly diagnosed patients with PD and 10 healthy control (HC) subjects were included in this cross-sectional study. Patients were enrolled in the Dutch Parkinson Cohort (DUPARC) study between 2017 and 2019 (For details: Boertien et al., 2020) (23). Inclusion criteria for patients consisted of PD diagnosis by a movement disorders specialist according to Movement Disorder Society (MDS) Clinical Diagnosis Criteria for PD (24) and with a confirmed dopaminergic striatal deficit on 3,4-dihydroxy-6-18F-fluoro-1-phenylalanine (¹⁸F-FDOPA) PET. HC had a normal neurological examination and did not have a history of neurological or psychiatric disorders. Exclusion criteria for both PD and HC subjects included the inability to provide written informed consent, the use of dopaminergic and (anti-)cholinergic medication and an estimated low premorbid intelligence level (estimated IQ <70, on the Dutch Adult Reading test (25)). All subjects gave written informed consent and the study was approved by the local ethics committee.

Clinical examination

All patients underwent comprehensive neuropsychological assessment covering all cognitive domains (23). A selection of outcome measures of tests and subtests of the cognitive test battery was made a priori, meeting level II criteria for PD-MCI (26,27), listed in table 1. Subject scores for each of the cognitive tests were compared to established test-specific normative data generated by age, gender and education. A performance of >1.5 standard deviation (SD) below normative values was considered abnormal. Patients were categorized as either PD with normal cognition (PD-NC) or PD-MCI. PD-MCI was based on level II criteria for PD-MCI and required below-threshold performance on at least two neuropsychological tests (26). Any patient or clinical characteristics possibly influencing performance on the neuropsychological assessments and MCI grouping, including visual difficulties, color blindness, speech problems and significant mood disorders were taken into account at the time of assessment and prior to data analysis, and if necessary excluded. HC subjects underwent cognitive testing using the Montreal Cognitive Assessment (MoCA) test.

Additional clinical assessment included the subjective duration of motor complaints before PD diagnosis and the Hospital Anxiety and Depression scale (HADS) (28). All PD subjects underwent motor examination using the MDS-revised Unified Parkinson's Disease Rating Scale part III (MDS-UPDRS-III). In addition, specific items from the MDS-UPDRS parts II and III were used for the classification of motor phenotypetype, using criteria previously formulated by Stebbins et al. (29). Patients were classified into tremor dominant (TD), postural instability and gait difficulty (PIGD), and indeterminate motor phenotypes

Image acquisition

All subjects underwent brain MRI and VAcH PET imaging, using (^{18}F)FEObv. MR imaging of PD subjects was acquired using Siemens Magnetom Prisma 3-Tesla magnetic resonance imaging scanners (Best, Netherlands) equipped with SENSE-8 channel head coils. For each subject, anatomical T1-weighted images were obtained using a sagittal 3-dimensional gradient-echo T1-weighted sequence with 0.9 x 0.9 x 0.9 mm acquisition. HC subjects underwent a T1-weighted MRI scan (3T Inera, Philips, The Netherlands) with 1.0 x 1.0 x 1.0 mm acquisition. (^{18}F)FEObv imaging was performed on the same day as MR imaging. After a low-dose CT for attenuation and scatter correction, participants were scanned using either a Biograph 40-mCT or 64-mCT (Siemens Healthcare, USA). Both these systems are EARL certified and identical in software version, acquisition- and reconstruction-protocols and PET detectors, and only differ in the number of CT slices. Patient and control subjects were randomly divided over the two PET scanners. (^{18}F)FEObv was injected using an intravenous bolus and delayed imaging was performed over 30 minutes (in six 5-minute frames) starting 210 minutes after injection.

Image processing was performed using Statistical Parametric Mapping (SPM) software (30). (^{18}F)FEObv PET imaging frames were spatially coregistered within subjects with a rigid-body transformation. The cropped T1-weighted MR scan was coregistered with the subject PET image. Freesurfer software package (Laboratory for Computational Neuroimaging, Athinoula A. Martinos Center for Biomedical Imaging, Boston, MA, USA) was used to segment MRI into cortical and subcortical brain regions. We calculated the distributed volume ratio (DVR) of each grey matter target region using the summed six delayed imaging frames and the white matter reference region as previously described (21,31). A parametric image for individual subjects was created by using the average of six delayed imaging frames divided by the mean of the white matter reference region. Partial volume correction on our parametric images was done using the Muller-Gartner method (32).

Imaging analysis

Voxel-based analysis was performed as previously described (33). Parametric PET images were spatially normalized to the Montreal Neurological Institute stereotactic template and smoothed with a Gaussian kernel of 8 mm full width half maximum. The relevant brain areas were displayed in Montreal Neurological Institute atlas coordinates (in millimeters) in the stereotactic space using the automated anatomical labeling toolbox.

Statistical analyses

To evaluate brain cholinergic innervation in *de novo* patients with PD, we first compared HC, PD-NC and PD-MCI groups on baseline demographic and clinical characteristics. Comparisons between all

three groups were performed using ANOVA for parametric variables and χ^2 testing for dichotomous variables. Comparisons between the two PD groups were performed using an independent samples t-test or a Mann-Whitney U test, for either normal and skewed distributed data, respectively. Statistical analysis was considered significant for $\alpha < 0.05$. Statistical analyses were performed using SPSS Statistics for Windows, Version 24.0 (IBM Statistics, USA) and SPM software.

Whole brain voxel-based analyses were performed to assess difference in cholinergic innervation between groups. Parametric [^{18}F]FEOBV DVR images were used for a two-sample voxel-wise statistical comparison between groups, in both positive and negative directions, controlling for age. The minimum cluster size was set to 50. The false discovery rate (FDR) approach was used for correction for multiple testing effects in the voxel-based analyses. Additional post-hoc Freesurfer-based volume-of-interest (VOI) analysis was performed on regions selected based on the voxel-based analysis. The percentage change of these regions was calculated by dividing the difference between the mean PD and HC group by the mean HC group.

Results

Demographic and clinical characteristics

The 57 *de novo* PD patients (32 males) had a mean (SD) age of 64.6 (8.2) years and a mean (SD) MDS-UPDRS-III score of 30.8 (11.4). 17 (29.8%) patients were classified as PD-MCI. Demographic and clinical characteristics of the different groups are listed in table 2. HC subjects had a significantly lower age and higher MoCA score compared to both PD groups ($p < 0.001$). The PD-MCI group had higher MDS-UPDRS-III scores ($p < 0.01$) and Hoehn and Yahr stage ($p = 0.05$) compared to the PD-NC group, reflecting more severe motor impairment in the PD-MCI group. No significant difference was found between the PD groups on the motor phenotype (TD, PIGD or indeterminate). Information on duration of motor complaints was missing for 11 patients. Available data showed no significant difference in the duration of motor symptom complaints prior to PD diagnosis.

Cognitive functioning

Overall, 15 out of 17 patients classified as PD-MCI showed multi-domain cognitive impairment, reflecting impaired scores in two or more cognitive domains. The most frequently affected domains were memory and executive functions, with 12 and 10 patients presenting impairments in these domains, respectively. The attention and visuospatial domains were affected in 7 out of 17 PD-MCI patients. Language was the least affected domain with 4 PD-MCI patients showing impaired scores. Detailed cognitive performance on the neuropsychological tests for both PD groups is listed in the supplementary materials.

Cholinergic innervation compared to controls.

Whole brain voxel-based analyses were performed to compare regional brain VAcHt binding between the PD groups (PD-MCI and PD-NC) and the HC subjects in positive and negative directions, while controlling for age. The PD groups showed evidence of bidirectional changes in different topographic patterns compared to the control subjects.

Significantly lower VAcHt binding was found in both PD groups, primarily in the temporal and posterior cortical regions, compared to HC (Figure 1; $p < 0.05$, FDR corrected). The topographic profiles for PD-NC and PD-MCI were comparable, with main affected regions including the parietal, parieto-occipital junction and lateral temporal cortices. PD-NC presented with additional lower binding in the frontal cortex, insula and caudate (tail), and PD-MCI with additional lower binding in the pons-medulla regions. A detailed overview of significant clusters is provided in the supplemental data.

Reverse direction voxel-based comparison between HC and both PD groups showed significantly higher VAcHt binding in PD subjects compared to HC ($p < 0.05$, FDR corrected). This higher cholinergic innervation was most prominently apparent in the PD-NC group (Figure 2A), including the cerebellum, the cingulate cortex, the gyrus rectus, the anteroventral striatum, putamen, the dorsal tegmentum, thalamus, metathalamus (lateral geniculate nucleus (LGN) and medial geniculate nucleus (MGN)), pulvinar, the amygdala, the hippocampus and parahippocampal regions. The PD-MCI groups showed a more limited topography of higher VAcHt binding, including the cerebellar vermis, the right gyrus rectus, amygdala, hippocampus and metathalamus, dorsal tegmentum, right inferior thalamus/pulvinarthalamus, and the left precentral gyrus (figure 2B). A detailed overview of significant clusters is provided in the supplemental data.

Most prominent representative examples of the magnitude of absolute binding changes with HC based on VOI comparison include reduced binding in the lateral occipital gyri (-13.3%), the pericalcarine (-19.0%), the transverse temporal gyri (-23.1%), and increases in the cerebellar cortex (+14.2%), the medial and lateral orbitofrontal region (resp +8.8% and +6.5%), frontal pole (+6.1%), the bilateral amygdala (+4.8%), and anterior cingulate (+3.8%) in the PD-NC group. For the PD-MCI group, representative examples include the lateral occipital gyri (-15.7%), the cuneus (-19.6%), pericalcarine (-23.9%), the middle temporal gyri (-14.2%) and the inferior parietal gyri (-15.4%) with lower VAcHt binding in PD-MCI, and increases in the cerebellar cortex (+13.2%), the right medial and lateral orbitofrontal region (resp +2.6% and +3.9%), and the right amygdala (+2.9%).

PD-MCI vs PD-NC group comparison

Lower VAcHt binding was found in the PD-MCI group in the fusiform and hippocampal region, the anterior insula and the left prefrontal cortex (uncorrected, $p < 0.001$). No significant differences were found after correction for multiple comparisons.

Post hoc: controlling for motor impairment

Clinical characteristics comparing both PD groups demonstrated a significant higher MDS-UPDRS-III score in patients with PD-MCI compared to PD-NC (table 2), indicating more severe motor impairment in the PD-MCI group. Bidirectional whole brain voxel-based analysis comparing both PD groups with controls was therefore repeated, controlling for both age and MDS-UPDRS-III.

Limited lower VAcHt binding was found in PD-NC and PD-MCI compared to HC ($p < 0.001$, uncorrected), including small regional differences in occipital and temporal regions.

Reverse direction analysis showed higher VAcHt binding in both PD-NC and PD-MCI compared to controls (figure 3; $p < 0.001$, uncorrected). PD-NC showed most extensive higher binding when compared to controls, with a cholinergic topography including the brainstem, gyrus rectus, posterior cingulate cortex, amygdala, and hippocampus and right parahippocampal region. The PD-MCI group demonstrated less widespread higher VAcHt binding, limited to the right superior temporal region, the left pulvinar, left optic radiation/occipital region and precentral gyrus.

Discussion

We investigated the topography of regional cholinergic innervation status in newly diagnosed, treatment-naïve patients with PD, with and without cognitive impairment, compared to healthy control subjects. We found altered cholinergic innervation, demonstrated by both higher and lower binding cholinergic brain regions. First, we demonstrate that cortical cholinergic denervation is already present at the start of the disease in both cognitively impaired and cognitively unaffected patients with PD. Second, we also found evidence of increased cholinergic binding in the PD groups. Cognitively unimpaired PD patients showed most robust evidence of higher-than-normal VAcHT binding compared to the PD-MCI group.

Previous studies have established the importance of the cholinergic system in cognitive impairment in PD, even in the absence of dementia (13,34). However, the majority of previous in vivo imaging studies are based on PET imaging using acetylcholinesterase tracers, which limit accurate estimates of high binding brain areas. The use of a VAcHT PET tracer, like [¹⁸F]FEOBV, allows for more reliable and regionally specific cholinergic assessment, especially in high binding areas such as the striatum and cerebellum (20–22). To our knowledge, this is the first assessment of regional cholinergic innervation in de novo patients with PD, with and without cognitive impairment, on a detailed cortical and subcortical level.

29.8% of patients in our study met PD-MCI criteria, which is in line with previous research on the cognitive impairment in early PD (3). In addition, the PD-MCI group presented with more severe motor impairment. However, there was no difference between PD-NC and PD-MCI in terms of motor phenotype (TD, PIGD or indeterminate) or duration of subjective motor complaints prior to PD diagnosis.

A striking finding in our study was the higher-than-normal cholinergic binding in de novo patients with PD compared to controls, most prominent in the PD-NC group. The brain topography represented by higher VAcHT binding included the cerebellum, parts of the thalamic complex, putamen, hippocampus and parahippocampal region, amygdala, gyrus rectus and cingulate cortex. Higher VAcHT binding in the cerebellum and thalamus indicate a substantial role of the cholinergic projections from the medial vestibular nucleus and PPN/LDTC, respectively (10,11). In addition, the involvement of the hippocampus, parahippocampal region, amygdala, gyrus rectus and cingulate cortex suggests involvement of projections originating from the basal forebrain, including Ch1-2 and Ch4 (Nucleus basalis of Meynert) groups (12). From the Ch4 cell group two major pathways can be identified; the medial pathway joining the white matter of the gyrus rectus and projecting to the

cingulum, and the lateral pathway of which the capsular division projects to the amygdala and temporal lobe (11). Finally, the altered [^{18}F]FEOBV uptake in the putamen and caudate nucleus indicate a role of intrinsic cholinergic striatal interneurons or PPN/LDTC projections (35). A possible role of the dopaminergic system should also be considered, as previous studies have shown an intricate interaction between the dopaminergic and cholinergic system; it is the multiplicative and interacting effects of the two systems that lead to cognitive deficits (17,36).

The higher cholinergic innervation in de novo PD suggests a compensatory cholinergic upregulation in the early phase of the disease. A compensatory role of the cholinergic system has previously been suggested in Alzheimer's disease (AD) where cholinergic upregulation was found in prodromal patients with MCI (37,38). Bohnen et al. (2015) and Kim et al. (2019) expanded on these findings by showing both independent and interactive roles of the cholinergic and dopaminergic system in cognitive functioning in PD (17,39). In the context of dopaminergic losses, preservation or even upregulation of cholinergic innervation may help preserve cognitive functioning (40). In addition, Liu et al (2018) reported increased cortical acetylcholinesterase activity related to LRRK2 gene mutation in (premanifest) PD (41). Bedard et al. (2019) recently showed significantly higher [^{18}F]FEOBV uptake in patients with idiopathic REM sleep behavior disorder (RBD) (42). RBD is considered an important marker of prodromal PD (43) and associated with cognitive impairment conversion (44). Our study is the first to demonstrate evidence of higher-than-normal cholinergic innervation in PD patients, substantiating the hypothesis of a compensatory cholinergic upregulation in very early PD.

Interestingly, the higher cholinergic VAcHT binding was less profound in the PD-MCI subgroup than in the cognitively unimpaired PD group, even though both groups consisted of newly diagnosed, treatment-naïve patients with PD with a similar duration of motor complaints prior to PD diagnosis. These findings suggest that a possible lack of compensatory cholinergic upregulation may be related to clinically evident cognitive impairment in early PD patients. In addition, the demonstrated denervation pattern in both PD-NC and PD-MCI groups provides further support for this hypothesis, as the relative extent of the denervation changes in the two PD subgroups had remarkable overlap and PD-MCI did not show more profound cholinergic denervation than PD-NC when compared to controls. In contrast, the PD-NC group showed slightly more extent denervation changes, possibly related to the higher number of subjects in this group providing more statistical power.

However, it should be noted that an alternative explanation for the difference in higher-than-normal VAcHT binding between PD-NC and PD-MCI may be related to the severity of motor impairment. The PD-MCI group presented with a higher MDS-UPDRS-III score than PD-NC, indicating more severe motor impairment. Therefore, a post-hoc analysis was performed, controlling for motor

impairment. Our data showed that the higher cholinergic binding was still present in PD-NC, but less widespread and robust. The upregulation in the cerebellum and thalamic complex was less profound, as well as the striatal regions, suggesting an important role of increased PPN cholinergic innervation and cholinergic striatal interneurons in motor performance in early PD. This is in line with previous research showing cholinergic PPN projections and striatal interneurons strongly contributing to motor symptoms in PD, especially in the motor subtype presenting with postural instability and gait disorder (31,45). In contrast, regions with basal forebrain cholinergic projections, including the posterior cingulate cortex, gyrus rectus, amygdala, hippocampus and parahippocampal region remained present after correction for this confounder variable. Interestingly, these regions show partial overlap with the regional cholinergic topography we previously found to be related to cognitive functioning in PD on the level of multiple cognitive domains (19), substantiating the cholinergic role of these regions in cognitive functioning in PD. On the other hand, controlling for MDS-UPDRS-III scores might cause overcorrection, as more severe motor and cognitive impairment often coincide (46,47) and MDS-UPDRS-III scores are significantly correlated with the majority of included cognitive tests (supplementary). The post-hoc analysis controlling for motor performance may therefore give an underestimation of the regional topography of higher cholinergic binding related to cognitive status.

In contrast to earlier findings (14,15,48), we found only limited cortical cholinergic denervation in patients with PD-MCI, without a substantial VAcH binding differences versus PD-NC. A possible explanation for this lack of difference may be the heterogeneity of cognitive performance in patients with PD. In our study, the majority of patients with PD-MCI showed multidomain cognitive impairment, with a variety of domains affected. In addition, not only heterogeneity in cognitive functioning, but also in other PD symptoms, including motor profile and non-motor symptoms, may result in cholinergic heterogeneity (49). The finding of a limited cholinergic denervation pattern may also contribute to the understanding why previous studies have found limited effectiveness of cholinesterase inhibitors in PD-MCI (50,51). We suggest that early cognitive decline is the result of failing cholinergic compensation, rather than cholinergic denervation per se. A better stratification, including the specific profile of dopaminergic and cholinergic innervation, in addition to clinical cognitive performance, might therefore improve the effectiveness of cholinergic treatment in PD.

Limitations

One limitation of this study is the relatively small sample size of the PD subgroups. In addition, the PD-MCI group presented with a heterogeneous cognitive profile, affecting multiple domains. The clinical heterogeneity of this group and the relatively small sample size of the subgroups may have

contributed to the limited difference found between PD-MCI and PD-NC. Future studies with a larger sample size can allow for more detailed stratification and improve our understanding on possible compensatory mechanisms specific to cognitive functioning across different domains. Second, even though MDS guidelines were followed, the grouping of patients into PD-NC and PD-MCI is an arbitrary process, as cognitive changes occur along a continuous spectrum. This might add to the heterogeneity. Furthermore, the cross-sectional design of our study does not allow assessment of temporal changes. More detailed data on cognitive performance over time and the progression to PD-MCI and PDD will enhance our understanding of the cholinergic role in the progression of the disease and the suggested compensatory mechanism. Finally, the HC group had a significantly lower age than both PD groups. We previously demonstrated an important role of age in the relationship between cognition and cholinergic innervation (19). Although the analyses we corrected for age, a possible role of the age difference can not be ruled out.

Conclusion

This study demonstrates evidence of bidirectional changes in cholinergic innervation in de novo patients with PD, with and without cognitive impairment, compared to healthy controls. Increased cholinergic binding in early PD, especially in the cognitively intact patients, suggests a compensatory cholinergic upregulation in this group. Taken together, we postulate that in early, treatment-naïve patients with PD, the clinical syndrome of PD-MCI may be related to limited or failing cholinergic upregulation, instead of a more progressed (posterior cortical) cholinergic denervation.

Author contributions

Study concept and design: van der Zee, Boertien, Gerritsen, Spikman, van Laar

Acquisition, analysis or interpretation of the data: van der Zee, Kanel, Slomp, Boertien, van Laar

Drafting of the manuscript: van der Zee

Critical revision of the manuscript: all authors

Statistical analysis: van der Zee, Kanel

Obtained funding: van der Zee, van Laar

Conflict of Interest Disclosures

Dr. van der Zee, dr. Kanel, ms Slomp, dr. Gerritsen and dr. Spikman have nothing to disclose. JM Boertien has received a writer's fee from Britannia Pharmaceuticals (Kinetic Magazine) and owns exchange traded funds (ETFs) that might include stocks from medically related fields. Dr. Müller has received research funding from the NIH, Department of Veteran Affairs and the Michael J. Fox Foundation. Dr. Bohnen has received research funding from the NIH, Department of Veterans Affairs, Parkinson's Foundation, Farmer Family Foundation, the Michael J. Fox Foundation, Eisai, and EIP Pharma. He has participated in an Eisai advisory board and received in kind research support from Scion Neurostim, Expansion Therapeutics and Innovative Health Solutions. Dr. van Laar has received research funding from Weston Brain Institute, Lysosomal Therapeutics, the Michael J Fox Foundation, the Dutch Brain Institute and the UMCG. He received lecture- and consultancy fees from AbbVie, Britannia and PureIMS.

Additional contributors: The authors thank all patients, caregivers, health care professionals and students who have contributed and collaborated in this project. The inclusion of patients was established with the help of the collaborative Parkinson Platform Northern Netherlands (PPNN).

References

1. Post B, Muslimovic D, van Geloven N, Speelman JD, Schmand B, de Haan RJ, et al. Progression and prognostic factors of motor impairment, disability and quality of life in newly diagnosed Parkinson's disease. *Mov Disord*. 2011 Feb 15;26(3):449–56.
2. Lawson RA, Yarnall AJ, Duncan GW, Khoo TK, Breen DP, Barker RA, et al. Severity of mild cognitive impairment in early Parkinson's disease contributes to poorer quality of life. *Parkinsonism Relat Disord*. 2014;20(10):1071–5.
3. Aarsland D, Bronnick K, Williams-Gray C, Weintraub D, Marder K, Kulisevsky J, et al. Mild cognitive impairment in Parkinson disease: a multicenter pooled analysis. *Neurology*. 2010 Sep 21;75(12):1062–9.
4. Domellöf ME, Ekman U, Forsgren L, Elgh E. Cognitive function in the early phase of Parkinson's disease, a five-year follow-up. *Acta Neurol Scand*. 2015;132(2):79–88.
5. Aarsland D, Andersen K, Larsen JP, Lolk A, Kragh-Sorensen P. Prevalence and characteristics of dementia in Parkinson disease: an 8-year prospective study. *Arch Neurol*. 2003;60(3):387–92.
6. Palavra NC, Naismith SL, Lewis SJG. Mild cognitive impairment in Parkinson's disease: a review of current concepts. *Neurol Res Int*. 2013;2013:576091.
7. Halliday GM, Leverenz JB, Schneider JS, Adler CH. The neurobiological basis of cognitive impairment in Parkinson's disease. *Mov Disord*. 2014 Apr 15;29(5):634–50.
8. Bohnen NI, Albin RL. The cholinergic system and Parkinson disease. *Behav Brain Res*. 2011 Aug 10;221(2):564–73.
9. Ballinger EC, Ananth M, Talmage DA, Role LW. Basal Forebrain Cholinergic Circuits and Signaling in Cognition and Cognitive Decline. *Neuron* [Internet]. 2016;91(6):1199–218. Available from: <http://dx.doi.org/10.1016/j.neuron.2016.09.006>
10. Zhang C, Zhou P, Yuan T. The cholinergic system in the cerebellum: from structure to function. *Rev Neurosci*. 2016 Dec;27(8):769–76.
11. Selden NR, Gitelman DR, Salamon-Murayama N, Parrish TB, Mesulam MM. Trajectories of

- cholinergic pathways within the cerebral hemispheres of the human brain. *Brain* [Internet]. 1998;121 (Pt 1:2249–57. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/9874478>
12. Mesulam MM. The systems-level organization of cholinergic innervation in the human cerebral cortex and its alterations in Alzheimer's disease. *Prog Brain Res*. 1996;109:285–97.
 13. Hilker R, Thomas A V., Klein JC, Weisenbach S, Kalbe E, Burghaus L, et al. Dementia in Parkinson disease: Functional imaging of cholinergic and dopaminergic pathways. *Neurology*. 2005;65(11):1716–22.
 14. Klein JC, Eggers C, Kalbe E, Weisenbach S, Hohmann C, Vollmar S, et al. Neurotransmitter changes in dementia with Lewy bodies and Parkinson disease dementia in vivo. *Neurology*. 2010;74(11):885–92.
 15. Shimada H, Hirano S, Shinotoh H, Aotsuka a., Sato K, Tanaka N, et al. Mapping of brain acetylcholinesterase alterations in Lewy body disease by PET. *Neurology*. 2009;73(4):273–8.
 16. Bohnen NI, Müller MLTM, Kotagal V, Koeppe RA, Kilbourn MR, Gilman S, et al. Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *J Cereb Blood Flow Metab*. 2012;32(8):1609–17.
 17. Bohnen NI, Albin RL, Müller MLTM, Petrou M, Kotagal V, Koeppe R a, et al. Frequency of Cholinergic and Caudate Nucleus Dopaminergic Deficits Across the Predemented Cognitive Spectrum of Parkinson Disease and Evidence of Interaction Effects. *JAMA Neurol* [Internet]. 2015;72(2):194–200. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/25506674>
 18. Bohnen NI, Kaufer DI, Hendrickson R, Ivanko LS, Lopresti BJ, Constantine GM, et al. Cognitive correlates of cortical cholinergic denervation in Parkinson's disease and parkinsonian dementia. *J Neurol*. 2006;253(2):242–7.
 19. van der Zee S, Müller MLTM, Kanel P, van Laar T, Bohnen NI. Cholinergic Denervation Patterns Across Cognitive Domains in Parkinson's Disease. *Mov Disord*. 2020 Nov;
 20. Petrou M, Frey K a, Kilbourn MR, Scott PJH, Raffel DM, Bohnen NI, et al. In Vivo Imaging of Human Cholinergic Nerve Terminals with (-)-5-18F-Fluoroethoxybenzovesamicol:

Biodistribution, Dosimetry, and Tracer Kinetic Analyses. *J Nucl Med* [Internet].

2014;55(3):396–404. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24481024>

21. Nejad-Davarani S, Koeppe RA, Albin RL, Frey KA, Müller MLTM, Bohnen NI. Quantification of brain cholinergic denervation in dementia with Lewy bodies using PET imaging with [18F]-FEOBV. *Mol Psychiatry*. 2019;24(3):322–7.
22. van der Zee S, Vallez Garcia D, Elsinga PH, et al. [18F]Fluoroethoxybenzovesamicol in Parkinson's disease patients: Quantification of a novel cholinergic positron emission tomography tracer. *Mov Disord*. 2019 Jun;34(6): 924–926.
23. Boertien JM, van der Zee S, Chrysou A, Gerritsen MJJ, Jansonius NM, Spikman JM, et al. Study protocol of the DUTch PARKinson Cohort (DUPARC): a prospective, observational study of de novo Parkinson's disease patients for the identification and validation of biomarkers for Parkinson's disease subtypes, progression and pathophysiology. *BMC Neurol*. 2020 Jun;20(1):245.
24. Postuma RB, Berg D, Stern M, Poewe W, Olanow CW, Oertel W, et al. MDS clinical diagnostic criteria for Parkinson's disease. *Mov Disord*. 2015 Oct;30(12):1591–601.
25. Schmand B, Bakker D, Saan R, Louman J. The Dutch Reading Test for Adults: a measure of premorbid intelligence level. *Tijdschr Gerontol Geriatr*. 1991 Feb;22(1):15–9.
26. Litvan I, Goldman JG, Tröster AI, Schmand B a., Weintraub D, Petersen RC, et al. Diagnostic criteria for mild cognitive impairment in Parkinson's disease: Movement Disorder Society Task Force guidelines. *Mov Disord*. 2012;27(3):349–56.
27. Goldman JG, Holden S, Ouyang B, Bernard B, Goetz CG, Stebbins GT. Diagnosing PD-MCI by MDS task force criteria: How many and which neuropsychological tests? *Mov Disord*. 2015;30(3):402–6.
28. Spinhoven P, Ormel J, Sloekers PP, Kempen GI, Speckens AE, Van Hemert AM. A validation study of the Hospital Anxiety and Depression Scale (HADS) in different groups of Dutch subjects. *Psychol Med*. 1997;27(2):363–70.

29. Stebbins GT, Goetz CG, Burn DJ, Jankovic J, Khoo TK, Tilley BC. How to identify tremor dominant and postural instability/gait difficulty groups with the movement disorder society unified Parkinson's disease rating scale: comparison with the unified Parkinson's disease rating scale. *Mov Disord*. 2013 May;28(5):668–70.
30. Wellcome Trust Centre for Neuroimaging.
<https://www.fil.ion.ucl.ac.uk/spm/software/spm12/>. University College, London, England.
31. Bohnen NI, Kanel P, Zhou Z, Koeppe RA, Frey KA, Dauer WT, et al. Cholinergic system changes of falls and freezing of gait in Parkinson's disease. *Ann Neurol*. 2019;85(4):538–49.
32. Muller-Gartner HW, Links JM, Prince JL, Bryan RN, McVeigh E, Leal JP, et al. Measurement of radiotracer concentration in brain gray matter using positron emission tomography: MRI-based correction for partial volume effects. *J Cereb Blood Flow Metab*. 1992 Jul;12(4):571–83.
33. Kanel P, Müller MLTM, van der Zee S, Sanchez-Catasus CA, Koeppe RA, Frey KA, et al. Topography of Cholinergic Changes in Dementia With Lewy Bodies and Key Neural Network Hubs. *J Neuropsychiatry Clin Neurosci*. 2020;32(4):370–5.
34. Bohnen NI, Müller MLTM, Kotagal V, Koeppe RA, Kilbourn MR, Gilman S, et al. Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *J Cereb Blood Flow Metab*. 2012;32(8):1609–17.
35. Dautan D, Huerta-Ocampo I, Witten IB, Deisseroth K, Bolam JP, Gerdjikov T, et al. A major external source of cholinergic innervation of the striatum and nucleus accumbens originates in the brainstem. *J Neurosci*. 2014 Mar;34(13):4509–18.
36. Ztaou S, Amalric M. Contribution of cholinergic interneurons to striatal pathophysiology in Parkinson's disease. *Neurochem Int*. 2019 Jun;126:1–10.
37. Ikonomic MD, Abrahamson EE, Isanski BA, Wu J, Mufson EJ, DeKosky ST. Superior frontal cortex cholinergic axon density in mild cognitive impairment and early Alzheimer disease. *Arch Neurol*. 2007 Sep;64(9):1312–7.
38. DeKosky ST, Ikonomic MD, Styren SD, Beckett L, Wisniewski S, Bennett DA, et al.

- Upregulation of choline acetyltransferase activity in hippocampus and frontal cortex of elderly subjects with mild cognitive impairment. *Ann Neurol*. 2002 Feb;51(2):145–55.
39. Kim K, Bohnen NI, Muller MLTM, Lustig C. Compensatory dopaminergic-cholinergic interactions in conflict processing: Evidence from patients with Parkinson’s disease. *Neuroimage*. 2019 Apr;190:94–106.
40. Bohnen NI, Grothe MJ, Ray NJ, Müller MLTM, Teipel SJ. Recent Advances in Cholinergic Imaging and Cognitive Decline—Revisiting the Cholinergic Hypothesis of Dementia. *Curr Geriatr Reports*. 2018;7(1):1–11.
41. Liu S-Y, Wile DJ, Fu JF, Valerio J, Shahinfard E, McCormick S, et al. The effect of LRRK2 mutations on the cholinergic system in manifest and premanifest stages of Parkinson’s disease: a cross-sectional PET study. *Lancet Neurol*. 2018 Apr;17(4):309–16.
42. Bedard M-A, Aghourian M, Legault-Denis C, Postuma RB, Soucy J-P, Gagnon J-F, et al. Brain cholinergic alterations in idiopathic REM sleep behaviour disorder: a PET imaging study with (18)F-FEOBV. *Sleep Med*. 2019 Jun;58:35–41.
43. Postuma RB. Prodromal Parkinson’s disease--using REM sleep behavior disorder as a window. *Parkinsonism Relat Disord*. 2014 Jan;20 Suppl 1:S1-4.
44. Rahayel S, Postuma RB, Montplaisir J, Mišić B, Tremblay C, Vo A, et al. A Prodromal Brain-Clinical Pattern of Cognition in Synucleinopathies. *Ann Neurol*. 2020 Nov;
45. Gilman S, Koeppe RA, Nan B, Wang C-N, Wang X, Junck L, et al. Cerebral cortical and subcortical cholinergic deficits in parkinsonian syndromes. *Neurology*. 2010 May;74(18):1416–23.
46. Wojtala J, Heber IA, Neuser P, Heller J, Kalbe E, Rehberg SP, et al. Cognitive decline in Parkinson’s disease: the impact of the motor phenotype on cognition. *J Neurol Neurosurg Psychiatry*. 2019 Feb;90(2):171–9.
47. Poletti M, Frosini D, Pagni C, Baldacci F, Nicoletti V, Tognoni G, et al. Mild cognitive impairment and cognitive-motor relationships in newly diagnosed drug-naive patients with

Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 2012 Jun;83(6):601–6.

48. Hilker R, Thomas a. V., Klein JC, Weisenbach S, Kalbe E, Burghaus L, et al. Dementia in Parkinson disease: Functional imaging of cholinergic and dopaminergic pathways. *Neurology*. 2005;65(11):1716–22.
49. Müller MLTM, Bohnen NI. Cholinergic dysfunction in parkinson's disease. *Curr Neurol Neurosci Rep*. 2013;13(9).
50. Grace J, Amick MM, Friedman JH. A double-blind comparison of galantamine hydrobromide ER and placebo in Parkinson disease. *J Neurol Neurosurg Psychiatry*. 2009 Jan;80(1):18–23.
51. Mamikonyan E, Xie SX, Melvin E, Weintraub D. Rivastigmine for mild cognitive impairment in Parkinson disease: A placebo-controlled study. *Mov Disord [Internet]*. 2015;00(00):n/a-n/a. Available from: <http://doi.wiley.com/10.1002/mds.26236>

Table 1: Selection of cognitive tasks used for the assessment of PD-MCI

Domain	Function	(Sub)test
Learning and memory	Verbal learning	RAVLT immediate recall ^a
	Verbal recall	RAVLT delayed recall ^a
	Visual learning	LLT(52) immediate recall ^b
	Visual recall	LLT delayed recall ^b
Attention and processing speed	Basic processing speed	Vienna test system Reaction time test(53): S1 decision time ^c
	Complex information processing speed	Vienna test system Reaction time test: S3 decision time ^c
	Selective attention	Stroop III: color-word
	Attention and memory span	Digit Span
Executive function	Set Shifting	WCST perseverative errors ^d
	Problemsolving	WCST trials to complete first category ^d
	Flexibility	Trail Making Test part B
Visual perception	Visuospatial perception	Judgment of Line orientation
	Visual search and attention	TEA Map search
Language	Verbal fluency	Category fluency: animal naming
	Naming ability	Boston Naming test

^{a, b, c, d} – concerns two subtasks within one cognitive test. If both scores are considered impaired, it will be counted as one impaired test.

PD-MCI: Parkinson's disease Mild Cognitive Impairment; RAVLT: Rey Auditory Verbal Learning Test; LLT: Location Learning Test; WCST: Wisconsin Card Sorting Test; TEA: Test of everyday attention.

Author Manuscript

Table 2. Demographic and Clinical Characteristics of the study participants^a

	HC (n=10)	PD-NC (n=40)	PD-MCI (n=17)	P Value
Age, y	54.6 (6.0)	63.4 (7.4)	67.5 (9.6)	<0.001^d
Gender, male n (% male)	5 (50%)	19 (47.5%)	13 (76.5%)	0.124
Educational level ^b	5.0 (1.0)	5.0 (2.0)	6.0 (1.0)	0.704
Motor symptom duration, months		20.2 (11.8)	25.1 (17.2)	0.273
MDS-UPDRS-III, total score		28.2 (9.1)	36.8 (14.1)	0.008
Motor phenotype, n TD/PIGD/Indeterminate		15/18/7	4/12/1	0.189
Hoehn and Yahr stage ^c		2.0 (1.0); 26.7	2.0 (1.0); 34.5	0.050
MoCA, total score	28.4 (0.8)	26.1 (2.6)	23.7 (3.2)	< 0.001^e
HADS anxiety, total score		4.5 (2.6)	4.7 (2.4)	0.778
HADS depression, total score		3.4 (2.4)	4.6 (2.6)	0.085

Abbreviations: HC, healthy controls; PD-NC, Parkinson's Disease Normal Cognition; PD-MCI, Parkinson's Disease Mild Cognitive Impairment; MDS-UPDRS-III, Movement Disorder Society Unified Parkinson's Disease Rating Scale part III; TD, Tremor Dominant; PIGD, Postural Instability and Gait Difficulty; MoCA, Montreal Cognitive Assessment; HADS, Hospital Anxiety and Depression Scale;

^a Data are presented as mean (standard deviation) unless otherwise indicated.

^b Educational level according to the Dutch Verhage scale (Verhage, 1964), listed as median (interquartile range)

^c Hoehn and Yahr stage is listed as median (interquartile range); mean rank

^d Post-hoc analysis: HC<PD-NC; HC<PD-MCI

^e Post-hoc analysis: HC>PD-NC>PD-MCI

Table 2. Demographic and Clinical Characteristics of the study participants^a

	HC (n=10)	PD-NC (n=40)	PD-MCI (n=17)	P Value
Age, y	54.6 (6.0)	63.4 (7.4)	67.5 (9.6)	<0.001^d
Gender, male n (% male)	5 (50%)	19 (47.5%)	13 (76.5%)	0.124
Educational level ^b	5.0 (1.0)	5.0 (2.0)	6.0 (1.0)	0.704
Motor symptom duration, months		20.2 (11.8)	25.1 (17.2)	0.273
MDS-UPDRS-III, total score		28.2 (9.1)	36.8 (14.1)	0.008
Motor phenotype, n TD/PIGD/Indeterminate		15/18/7	4/12/1	0.189
Hoehn and Yahr stage ^c		2.0 (1.0); 26.7	2.0 (1.0); 34.5	0.050
MoCA, total score	28.4 (0.8)	26.1 (2.6)	23.7 (3.2)	< 0.001^e
HADS anxiety, total score		4.5 (2.6)	4.7 (2.4)	0.778
HADS depression, total score		3.4 (2.4)	4.6 (2.6)	0.085

Abbreviations: HC, healthy controls; PD-NC, Parkinson's Disease Normal Cognition; PD-MCI, Parkinson's Disease Mild Cognitive Impairment; MDS-UPDRS-III, Movement Disorder Society Unified Parkinson's Disease Rating Scale part III; TD, Tremor Dominant; PIGD, Postural Instability and Gait Difficulty; MoCA, Montreal Cognitive Assessment; HADS, Hospital Anxiety and Depression Scale;

^a Data are presented as mean (standard deviation) unless otherwise indicated.

^b Educational level according to the Dutch Verhage scale (Verhage, 1964), listed as median (interquartile range)

^c Hoehn and Yahr stage is listed as median (interquartile range); mean rank

^d Post-hoc analysis: HC<PD-NC; HC<PD-MCI

^e Post-hoc analysis: HC>PD-NC>PD-MCI