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**Distribution and phylogenetics of hepatitis E virus genotype 4 in humans and animals**

Short title: Hepatitis E virus genotype 4

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**Author contribution**

YL conceived and designed the study. BL, YS, and XC collected the sequence data and conducted the phylogenetic analysis. BL prepared the manuscript. YL and AW critically revised the manuscript. All authors read the manuscript and approved the final manuscript.

**Key words**

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3 **Impacts**

- 4 ● China and Japan remain endemic for HEV-4, whereas France has documented a high prevalence
- 5 of HEV-4 in Europe.
- 6 ● Globally, prevalent HEV-4 subtypes changed remarkably over the last 18 years.
- 7 ● Increases in affected areas and animal hosts imply consistent cross-border and cross-species
- 8 transmission.

9

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11 Abstract: 250

12 Tables: 4

13 Figures: 4

14 Supplementary tables: 3

15 **Abstract**

16 **Background**

17 Worldwide, hepatitis E virus (HEV) infection is considered a significant public health concern. In  
18 particular, HEV genotype 4 (HEV-4) has spread to more areas and host species. In this study, we  
19 describe the global distribution of HEV-4 and characterize HEV-4 subtypes by host, country, and year  
20 of isolation.

21

22 **Methods**

23 We retrospectively collected HEV-4 sequences available before December 31, 2019, in GenBank.  
24 HEV-4 and its subtypes were determined using phylogenetic comparison with HEV reference  
25 sequences. Information on the isolation of the sequences was extracted from the GenBank or original  
26 publications. Temporal, spatial, and host characteristics of the sequences were summarized and  
27 nucleotide similarity was calculated based on five amplified fragments within HEV genome, stratified  
28 by host, country, and year.

29

30 **Results**

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1 A total of 2295 HEV-4 complete and partial nucleotide sequences were studied. The majority (92.7%)  
2 was isolated in China's mainland, Japan, Hong Kong, and France. A total of 20 animal hosts were  
3 documented, though swine remained predominant (71.7%). Globally, prevalent HEV-4 subtypes  
4 changed remarkably over the last 18 years. Subtype 4a, 4b, 4d, and 4h were most commonly isolated  
5 (80.3%). Subtypes 4c, 4e, 4f, 4g, and 4i remained limited in temporal distribution. High nucleotide  
6 similarities were observed between the sequences amplified in HEV ORF2, in the same and  
7 neighboring countries, and in similar animal hosts.

## 8 **Conclusion**

9 China and Japan are endemic for HEV-4, and have all the subtypes. In Europe, France has a high  
10 prevalence of HEV-4. Increases in affected areas and animal hosts imply consistent cross-border and  
11 cross-species transmission.  
12

## 13 **Key words**

14 Hepatitis E virus; genotype 4; phylogenetic analysis; zoonosis  
15

## 16 **Introduction**

17 Hepatitis E virus (HEV) is a food-borne or waterborne pathogen which can cause liver disease. HEV  
18 has caused numerous outbreaks of hepatitis in multiple low- and middle-income countries, and  
19 subsequently results in a high fatality rate among pregnant women in these geographical areas  
20 (Kmush, Wierzba, Krain, Nelson, & Labrique, 2013). The global burden of HEV was estimated to be  
21 approximately 20 million infections and 3.4 million symptomatic cases every year (Rein, Stevens,  
22 Theaker, Wittenborn, & Wiersma, 2012). However, the real burden may be underestimated, due to  
23 dynamic antibodies response, limited genotypes and sensitivity of assays ("EASL Clinical Practice  
24 Guidelines on hepatitis E virus infection," 2018). Currently, HEV is most prevalent in Eastern Asia  
25 and Southern Asia (WHO, 2019), where the seroprevalence of anti-HEV antibodies have varied from  
26 10% to 50% and 10% to 40%, respectively (WHO, 2010).  
27

28  
29 Hepatitis E virus (HEV) is a non-enveloped, positive-sense single-stranded RNA virus which belongs  
30 to the Genus *Orthohepevirus* within the family of *Hepeviridae* (Purdy et al., 2017; D. B. Smith,

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1 Simmonds, Members Of The International Committee On The Taxonomy Of Viruses Hepeviridae  
2 Study, et al., 2015). HEV genome is approximately 7.2 kb in length, which includes three open  
3 reading frames (ORFs) (Tam et al., 1991; Tsarev et al., 1992). ORF 1 encodes non-structural proteins  
4 consisting of several functional domains such as a methyltransferase domain (Met), Y domain (Y),  
5 papain-like cysteine protease (PCP), hypervariable region (HV), proline-rich domain (Pro), X-domain  
6 (X), helicase (Hel), and RNA-dependent RNA polymerase (RdRp), whereas ORF 2 encodes structural  
7 proteins (Graff, Torian, Nguyen, & Emerson, 2006; Yuchen Nan, Wu, Qin, & Zhou, 2017; Panda &  
8 Varma, 2013). ORF 3, which sometimes overlaps with ORF 1 and ORF 2, encodes a minor structural  
9 protein (Tam et al., 1991).

10  
11 So far, HEV has been genetically classified in four species, Orthohepevirus A to D (Donald B. Smith  
12 & Peter Simmonds, 2018). In the species A, a total of eight genotypes have been identified, with hosts  
13 such as humans (HEV-1, 2, 3, 4, and 7), domestic pigs (HEV-3 and 4), rabbits (HEV-3), wild boars  
14 (*Sus scrofa*) (HEV-3, 4, 5, and 6), and camels (HEV-7 and 8) (Donald B. Smith & Peter Simmonds,  
15 2018). Species B, C, and D have been identified in only non-human animals; however, two cases of  
16 hepatitis E patients infected with rat HEV (species C) were reported in Hong Kong in 2018 (Sridhar et  
17 al., 2018). HEV exhibits high genetic variability by rapid evolution due to its nature of RNA virus and  
18 selection pressure imposed by immune responses of zoonotic hosts (Sanjuán, Nebot, Chirico, Mansky,  
19 & Belshaw, 2010; van Tong et al., 2016), which could lead to increasing genetic diversity and more  
20 emerging genotypes or subtypes. HEV-3 and 4 are the most commonly studied zoonotic genotypes.  
21 HEV-3 is prevalent worldwide (Primadharsini, Nagashima, & Okamoto, 2019), whereas HEV-4 is  
22 mostly limited to Asia, including China, Japan, and countries in Southeastern Asia (Lu, Li, &  
23 Hagedorn, 2006). However, HEV-4 has been recently isolated in Europe (Hakze-van der Honing, van  
24 Coillie, Antonis, & van der Poel, 2011). It remains unexplained why HEV-3 and 4 have different  
25 geographical distributions. HEV-4 was first identified in China in 1999 (Wang et al., 1999) and has  
26 become the predominant genotype in China since 2000 (Zhang et al., 2011). In addition, it is noted  
27 that there is increasing numbers of indigenous cases in Europe (Asma, Stephanie, Eric, Camelia, &  
28 Anne-Marie, 2013; Colson et al., 2012; Fogeda, Avellón, Cilla, & Echevarría, 2009b; Garbuglia et al.,  
29 2013; Midgley, Vestergaard, Dalgaard, Enggaard, & Fischer, 2014; Ole et al., 2008; Tessé et al.,  
30 2012). Currently, a total of nine HEV-4 subtypes, 4a – 4i, have been determined (D. B. Smith et al.,

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1 2020).

2  
3 In addition, previous studies have provided a large number of nucleotide sequences that facilitate in-  
4 depth study of HEV. In this study, we retrospectively studied all HEV-4 sequences available in the  
5 GenBank to determine the global distribution of HEV-4 and characterized each subtype by host,  
6 country, and year of isolation.

## 7 8 **Materials and methods**

### 9 **HEV-4 sequence retrieval and genotyping**

10 In the GenBank (<https://www.ncbi.nlm.nih.gov/nucleotide>), we defined our search strategies as follows:  
11 ("Hepatitis E virus"[Organism] OR hepatitis e virus [All Fields]) OR ("Hepatitis E virus"[Organism]  
12 OR HEV [All Fields]). Nucleotide sequences with 1 nucleotide (nt) through 10000 nt in length that  
13 were released before December 31, 2019, were retrieved in the GenBank. Those sequences that were  
14 synthesized or patented were excluded from the study. By duplicate alignments with HEV reference  
15 strains (D. B. Smith, Simmonds, Jameel, et al., 2015), we determined if the retrieved sequences were  
16 HEV-4 by constructing a Neighbor-Joining tree and a maximum likelihood tree with bootstrap tests of  
17 1000 replications using the Kimura-2-parameter method in MEGA 7.0 software  
18 ([www.megasoftware.net](http://www.megasoftware.net)). A total of 48 full-length reference sequences representing eight genotypes  
19 of species A, including 11 HEV-4 reference sequences, were utilized for comparison (Supplementary  
20 Table 1) (D. B. Smith et al., 2020). Subtypes of HEV-4 were determined by duplicate comparison  
21 with 48 HEV reference sequences and 11 HEV-4 reference sequences with the Neighbor-Joining  
22 method. If there was conflicting output, the maximum likelihood method was further utilized to  
23 confirm the subtype. HEV genotype and subtype were assigned by Smith *et al* (D. B. Smith *et al.*,  
24 2020). We listed all the uncertain comparison outputs in Supplementary Table 2.

25  
26 Information on the isolation of the retrieved sequences was directly extracted from GenBank,  
27 including species host, country, date of collection, isolate/strain name, ORF, and CDS product. If  
28 there was incomplete information in the GenBank, we further searched the original publications that  
29 had cited the sequences to complete the information. Regarding missing date of collection, we  
30 estimated the year of isolation as one year before the submitted date. If the date was displayed in a

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1 time period, we used the middle year as date of collection. Duplicate sequences that shared identical  
2 nucleotide positions, isolate/strain names, hosts, countries and dates of collection were removed from  
3 the analysis. This search and selection procedure was detailed in Fig 1.

#### 5 **HEV-4 sequence datasets**

6 In this study, we generated six datasets for phylogenetic analysis. In dataset 1, there was a total of  
7 2295 HEV-4 strains with complete information about isolation. Of them, 83 strains were full-length  
8 sequences while the others were partial sequences with different nucleotides in length. Subsequently  
9 we described the global distribution of HEV-4 subtypes by host, country, and year of isolation. Each  
10 strain was determined by excluding duplicate sequences that shared identical isolate/strain name, host,  
11 country and date of collection.

12  
13 In datasets 2-6, the 83 full-length sequences were segmented into the following five partial fragments;  
14 those partial sequences not belonging to the five partial fragments were excluded in the following  
15 analysis. Finally, we included HEV-4 sequences that matched the five partial fragments as follows:  
16 ORF 1 Met (279 nt, n=232, 171-449 nt in subtype 4a reference sequence: AB197673), ORF 1 RdRp  
17 (307 nt, n=446, 4292-4598 nt in AB197673), and ORF 2 (317 nt, n=754, 6026-6342 nt in AB197673;  
18 150 nt, n=368, 6356-6505 nt in AB197673; and 537 nt, n=325, 6449-6985 nt in AB197673).

19 Nucleotide similarities within and between groups stratified by host, country, and year of isolation  
20 were calculated using the Kimura-2-parameter method with bootstrap tests of 1000 replications. HEV-  
21 4 sequences that belonged to the same strain but were in different nucleotide positions within the  
22 HEV genome were kept separately in dataset 2-6.

#### 24 **Ethical Approval statement**

25 This study included the HEV sequences available in the GenBank and involved no ethical issue. Thus,  
26 this study did not require ethical approval.

#### 28 **Results**

##### 29 **Global distribution of HEV-4**

30 HEV-4 was first isolated in a Chinese patient with acute hepatitis in 1992 (Wang et al., 1999) and has

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1 been circulating for decades. A total of 2295 HEV-4 strains were identified in this study (Fig 1).  
2 Information on all sequences was listed in Supplementary Table 3, including year of isolation, host,  
3 country and region, subtype, and GenBank accession number. Temporally, isolation of  
4 HEV-4 strains were very limited before 2000. From 2000 through 2004, the number of isolated strains  
5 increased slightly but remained at a low level. Between 2005 and 2011, the number of isolated strains  
6 remained high every year, with 2007 being the peak. Then the number of strains showed a significant  
7 decline, and began to increase slightly over 100 in 2015 and remained steady until 2018 (Fig 2).

8  
9 Geographically, HEV-4 strains have been isolated in Asia, Europe, and North America involving 22  
10 countries and regions, suggesting extensive distribution across the world. The vast majority of strains  
11 have been isolated in Asia (96.8%, 2223/2295), including China's mainland (n=1745), Hong Kong  
12 (n=92), and Japan (n=243). In addition, sporadic cases have been identified in Europe, and North  
13 America, with the majority isolated in France (n=48) but only one each from the UK and the USA  
14 (Table 1). China and France had the largest number of isolated HEV-4 strains in Asia and Europe,  
15 respectively. Asian countries that have isolated HEV-4, such as South Korea, India, Viet Nam,  
16 Mongolia, and Indonesia, are all neighboring to China. In European countries, such as Denmark, Italy,  
17 Belgium, Spain, and Germany, which are geographically close to France have also reported HEV-4  
18 infection (Fig 3). The findings suggested a geographically central distribution of HEV-4 in China and  
19 France, and circular distribution in the neighboring countries and regions.

20  
21 So far, a total of 20 species have been reported to be the host of HEV-4, including humans (n=1274),  
22 swine (n=732), and other animals (n=289). Generally, the number of human strains was larger than  
23 swine strains in most subtypes; however, swine strains surpassed human ones in subtype 4d (n=210  
24 vs. n=206) and 4e (n=24 vs. n=16). Of the total animal HEV-4 strains, 84.9% (867/1021) were  
25 isolated in China's mainland, covering all the subtypes of animal hosts. In Europe, swine and  
26 cynomolgus monkey (*Macaca fascicularis*) were the only animal hosts. In addition to swine, wild  
27 boar (*Sus scrofa*), deer (*Cervidae*), and other mammals, animal hosts of HEV-4 included birds such as  
28 crowned crane (*Grus japonensis*) (GenBank accession no., EF417589) and silver pheasant (*Lophura*  
29 *nycthemera*) (EF417590), and shellfish such as blood clam (*Scapharca subcrenata*) (KJ816336-  
30 KJ8163340, KJ816346, KJ816347) and Philippine clam (*Ruditapes philippinarum*) (KJ816335,

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1 KJ816342, KJ816343).

2  
3 A total of ten HEV-4 subtypes and clusters were described in this study (Table 1). Subtype 4a (n=642)  
4 was annually isolated with human, swine, and wild boar hosts; a strain was recently isolated in cattle  
5 in South Korea (MK795980). Moreover, 4a was frequently isolated in Asia, except a US strain  
6 (JX426082) in 2010. Similarly, subtype 4c (n=99), 4e (n=40), and 4g (n=48) appeared mainly in Asia  
7 (except a 4e British strain GQ913689 in 2009 and two 4g Swiss strains MK343062 and MK343063 in  
8 2012 and 2015, respectively) and had human, swine, and wild boar hosts. However, 4c, 4e, and 4g  
9 have not been isolated after 2016, 2009, and 2015, respectively. Subtype 4b (n=396) and 4d (n=473)  
10 had a wide range of hosts, including rhesus monkey (*Macaca mulatta*), cynomolgus monkey (*Macaca*  
11 *fascicularis*), cattle (*Bos taurus*), sheep (*Ovis aries*), donkey, wild boar, tiger (Bengal tiger & Siberian  
12 tiger), and shellfish, in addition to human and swine, which were isolated continually in Europe and  
13 Asia. Similarly, subtype 4h (n=331) had a variety of hosts, such as cattle, sheep, goat (*Capra hircus*),  
14 donkey (*Equus asinus*), wild boar, rhesus monkey, tree shrew (*Tupaia belangeri*) and wild yak (*Bos*  
15 *grunniens*). Subtype 4i (n=190) had mammalian hosts (deer, bear (*Ursus thibetanus*), and leopard  
16 (*Panthera pardus*)) and birds; however, it was only isolated in Asia before 2017. Subtype 4f had only  
17 6 strains in humans, and 3 strains in wild boars, in which the latest was isolated in Hong Kong in 2014  
18 (KX752741). Subtype 4 (n=12) was distinct from other subtypes but remained unassigned (Donald B.  
19 Smith, Simmonds, Izopet, Oliveira-Filho, & Purdy, 2016) with human and swine hosts across Asia  
20 and Europe, of which the latest two strains were isolated in France in 2012 (KR027892 and  
21 KR027904). In this study, there was a cluster (n=54) consisted of the HEV-4 sequences being  
22 separated in the phylogenetic tree, which could not be classified into an existing subtype, or having  
23 inconsistent phylogenetic findings using different methods or reference strains (Supplementary Table  
24 2).

25  
26 In China's mainland, the most common subtype was 4a (33.7%), whereas it was 4c (32.1%) in Japan  
27 and 4b (55.4%) in Hong Kong. Subtype 4b (77.5%) was predominant in Europe, particularly in  
28 France (Table 1). Globally, the prevalent subtypes have changed remarkably over the last 18 years. In  
29 total, subtypes 4a, 4b, 4d and 4h were most commonly isolated. In 2005-2010, subtype 4a represented  
30 36.9% of isolated HEV-4 sequences; however, it decreased to 9.1% in 2012-2017. Concurrently, the



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1 proportion of subtype 4h sequences increased from 10.1% to 26.3%, and the proportion of subtype 4d  
2 sequences increased from 16.6% to 33.6%. Subtype 4h was firstly identified in 2000 and then  
3 surpassed 4a in the number of isolation in 2002. In 2018, subtype 4a increased dramatically while  
4 subtype 4h decreased (Fig 4).

#### 6 **Phylogenetic analysis of HEV-4**

7 A phylogenetic analysis was performed based on the Met domain (279 nt) and RdRp domain (307 nt)  
8 in ORF1, and three ORF2 partial fragments (317 nt, 150 nt, and 537 nt). The ORF2 150 nt partial  
9 sequences shared highest nucleotide similarities, whereas the two ORF1 partial sequences  
10 demonstrated lower similarities within and between host groups (human and swine) and country  
11 groups (China's mainland and Japan) (Table 2). Globally, nucleotide similarities increased by year,  
12 particularly between swine sequences on the Met 279 nt, RdRp 307 nt, ORF2 537 nt and human  
13 sequences on the ORF2 150 nt (Table 3). The similarities between human sequences isolated in Japan  
14 were much higher than those in China's mainland (Table 4). Furthermore, HEV-4 strains isolated in  
15 same country shared higher similarities, regardless of host (Table 4).

17 High nucleotide similarities were observed between countries and regions, such as between France  
18 and Denmark (sharing the identity of 96.0%-99.9% on the ORF2 150 nt), South Korea and Mongolia  
19 (87.6%-96.7% on the ORF2 150 nt), Hong Kong and Cambodia (92.9%-96.3% on the ORF2 537 nt,  
20 which was similar to 94.0%-96.7% within Hong Kong). In addition, 4h isolated in rhesus monkey  
21 (n=1), tree shrew (n=1), and wild yak (n=1) shared high similarities based on the two ORF1 regions  
22 (96.1%-99.0% on the Met 279 nt, 96.1%-99.0% on the RdRp 307 nt). Similarly, they shared the  
23 highest nucleotide similarities (98.8%-99.5%) on the ORF2 317 nt (rhesus monkey, n=11; tree shrew,  
24 n=1; wild yak, n=1), whereas 4h sequences in cow (n=52) and goat (n=23) shared high similarities  
25 (98.2%-99.0%) on the ORF2 317 nt.

27 In addition, nucleotide similarities were compared within subtype 4a, 4b, 4d, 4h, and 4i. High  
28 similarities were identified between the sequences: 4a, in human and swine on the ORF2 150 nt  
29 (93.9%-96.3%); 4b, in Viet Nam and Hong Kong on the Met 279 nt (91.8%-95.0%), in France and  
30 Denmark on the ORF2 317 nt (95.5%-97.8%); 4d, in cow and sheep on the ORF2 317 nt (95.6%-

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1 97.8%); 4h, in cow and goat on all the five partial fragments ( $\geq 95.9\%$ ); 4i, in Japan and Malaysia on  
2 the three ORF2 regions (96.1%-98.1% on the ORF2 317 nt, 95.6%-98.5% on the ORF2 150 nt, and  
3 95.3%-97.1% on the ORF2 537 nt).

## 4 5 **Discussion**

6 HEV-4 is mainly isolated in Asia and Europe (Dalton & Izopet, 2018b; M. S. Khuroo, Khuroo, &  
7 Khuroo, 2016). In this study, we found that the majority of HEV-4 sequences were isolated in China's  
8 mainland, Hong Kong, Japan, and France. The geographical distribution was characterized by central  
9 distribution in China and France, and circular distribution in the neighboring countries and regions.  
10 High nucleotide similarities were observed between the HEV-4 sequences in France and Denmark,  
11 Hong Kong and Cambodia, South Korea and Mongolia, suggesting potential cross-border  
12 transmission in neighboring areas. Previous studies have provided possible evidence of cross-border  
13 transmission, such as transmission of HEV-4 from Southeastern Asia to Europe by international  
14 travelers (Fogeda, Avellón, Cilla, & Echevarría, 2009a). In addition, importation pig blood products  
15 from China to Europe as feed growth promoters may have contributed to the sporadic human cases of  
16 HEV-4 infection in Europe; however, it remains lack of direct evidence (Dalton & Izopet, 2018a). On  
17 the other hand, predominant subtypes of HEV-4 varied by area, such as subtype 4a in China's  
18 mainland, 4b in Hong Kong and Europe, and 4c in Japan, suggesting that HEV-4 remains endemic.  
19 Nucleotide similarities remained high within same countries and regions, regardless of host, as  
20 documented elsewhere (Nishizawa et al., 2003). Moreover, the proportion of isolated HEV-4 subtypes  
21 has changed over time. Subtype 4a has decreased, whereas 4h and 4d has increased in 2007-2017. On  
22 the one hand, the changing prevalence and alternating predominant HEV-4 subtypes might be partly  
23 explained by sampling bias, on the other hand, as hypotheses, it may be attributable to increasing  
24 human travel and animal trade regionally and globally (Dalton & Izopet, 2018b; Geng et al., 2010).  
25 The global distribution of HEV-4 is rapidly changing and emerging in different countries, suggesting  
26 the need for more widespread intervention strategies in the future. An HEV vaccine is available (Wu,  
27 Chen, Lin, Hao, & Liang, 2016), although it is currently on the market only in China. However, the  
28 uptake rate of the vaccine remained low and little-known in general population or public health  
29 practitioners (Ren et al., 2019).

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1 HEV-4 subtypes have been increasing in recent years, which are characterized by temporal, spatial,  
2 and host species clustering. Subtype 4a has been isolated annually; however, the years of isolation of  
3 some subtypes (4c, 4e, 4f, 4g and 4i) remain limited, which demonstrates a higher disease burden in  
4 China. So far, swine remains the principal reservoir of HEV-4. Furthermore, a growing number of  
5 mammals and non-mammal animals have been documented to carry HEV-4. In this study, we  
6 summarized the range of hosts, including swine, wild boar, deer, cattle, goat, sheep, donkey, tiger,  
7 leopard, bear, monkey, tree shrew, wild yak, birds, and shellfish. We identified more hosts compared  
8 to a previous review in 2016 (Y. Nan & Zhang, 2016). More important, HEV-4 infection in birds  
9 (Balayart et al., 1983) may suggest an avian-to-mammalian transmission and that in shellfish  
10 (Mohammed Sultan Khuroo, 1980) may suggest a cohabitation in an aquatic environment. It has  
11 raised a new public health concern of multiple cross-species transmission, though there was  
12 possibility that these hosts were merely opportunistic infection with no proliferation of HEV.  
13 Subtypes 4b, 4d, and 4h, which have a variety of animal hosts, were the predominant subtypes in  
14 some Asian and European countries and regions; particularly, subtype 4h had increased in 2012-2017  
15 by almost 3 times, suggesting possibly growing cross-species transmission. In addition, subtype 4h  
16 sequences showed high nucleotide similarities in wild mammals and domestic animals, respectively,  
17 whereas lower similarities between wild mammals and domestic animals, implying continual spread  
18 within similar animal hosts. However, it remained unclear if there is animal-to-animal transmission  
19 and animal-to-human transmission (D. B. Smith & P. Simmonds, 2018), which warrants further study.

20  
21 Our study has limitations. Firstly, our study was conducted based on the sequences available in the  
22 GenBank, which may represent a sample of HEV-4 strains circulating across the world. Sequencing is  
23 associated with multiple factors, such as awareness, research priority, and economic development in  
24 every single country and region. In low- and middle-income countries and regions, intention to detect  
25 and sequence HEV may be limited, compared to other infectious diseases that may be more important,  
26 resulting in sampling and sequencing bias. However, the GenBank is the principal platform for  
27 microbial sequences, in which HEV-4 full-length and partial sequences available may be appropriate  
28 for understanding the real epidemic scenario of HEV-4, as we could not obtain a more representative  
29 sample. Moreover, this study provides a preliminary and quantitative evidence of changing trends in  
30 HEV-4 isolation worldwide. Second, that the phylogenetic analysis was performed based on the five

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1 partial fragments within HEV ORF1 and ORF2. Due to all the fragments not parallel available for the  
2 same HEV-4 strains and their inconsistent number of isolations by year/country/host, we calculated  
3 the nucleotide similarities based on a pooled database. Our analysis with a global perspective may  
4 bridge the gap in the absence of each fragment. Third, we identified an uncertain cluster within HEV-  
5 4. Some of the sequences were similar to that of subtype 4; however, the sequences could not be  
6 currently classified into existing HEV-4 subtypes. It may suggest a new subtype, which should be  
7 confirmed with further evidence. At the same time, it showed a limitation in the identification of  
8 HEV-4 subtypes based on a short partial fragment (62.9% of the sequences have less than 300 nt in  
9 length), as suggested by inconsistent findings in the phylogenetic analysis. Thus, further amplification  
10 of longer partial fragment is warranted to determine their subtypes.

11  
12 In conclusion, China and Japan remain endemic for HEV-4. The number of strains isolated in France  
13 may indicate endemicity and highlights the spread of HEV-4 in Europe. HEV-4 subtypes are  
14 characterized by temporal, spatial, and host species clustering. Increases in affected areas and animal  
15 hosts imply consistent cross-border and cross-species transmission.

#### 17 **Conflict of Interest Statement**

18 The authors declare no conflict of interest.

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#### 25 **References**

- 26 Asma, J., Stephanie, H. B., Eric, M., Camelia, M., & Anne-Marie, R.-A. (2013). Genotype 4  
27 Hepatitis E Virus in France: An Autochthonous Infection With a More Severe  
28 Presentation. *Clinical Infectious Diseases An Official Publication of the*  
29 *Infectious Diseases Society of America*, 57(4), 122-126. doi:10.1093/cid/cit291  
30 Balayart, M. S., Andjaparidze, A. G., Savinskaya, S. S., Ketiladze, E. S., Braginsky, D.

- 
- 1 M., Savinov, A. P., & Poleschuk, V. F. (1983). Evidence for a Virus in Non-A, Non-B  
2 Hepatitis Transmitted via the Fecal-Oral Route. *Intervirology*, *20*(1), 23-31.  
3 doi:10.1159/000149370
- 4 Colson, P., Romanet, P., Moal, V., Borentain, P., Purgus, R., Benezech, A., . . .  
5 Gérolami, R. (2012). Autochthonous Infections with Hepatitis E Virus Genotype 4,  
6 France. *Emerging infectious diseases*, *18*(8), 1361-1364. doi:10.3201/eid1808.111827
- 7 Dalton, H. R., & Izopet, J. (2018a). Transmission and Epidemiology of Hepatitis E Virus  
8 Genotype 3 and 4 Infections. *Cold Spring Harb Perspect Med*, *8*(11).  
9 doi:10.1101/cshperspect.a032144
- 10 Dalton, H. R., & Izopet, J. (2018b). Transmission and Epidemiology of Hepatitis E Virus  
11 Genotype 3 and 4 Infections. *Cold Spring Harbor Perspectives in Medicine*, *8*(11),  
12 20. doi:10.1101/cshperspect.a032144
- 13 EASL Clinical Practice Guidelines on hepatitis E virus infection. (2018). *J Hepatol*,  
14 *68*(6), 1256-1271. doi:10.1016/j.jhep.2018.03.005
- 15 Fogeda, M., Avellón, A., Cilla, C. G., & Echevarría, J. M. (2009a). Imported and  
16 autochthonous hepatitis E virus strains in Spain. *J Med Virol*, *81*(10), 1743-1749.  
17 doi:10.1002/jmv.21564
- 18 Fogeda, M., Avellón, A., Cilla, C. G., & Echevarría, J. M. (2009b). Imported and  
19 autochthonous hepatitis E virus strains in Spain. *Journal of Medical Virology*,  
20 *81*(10), 1743 - 1749. doi:10.1002/jmv.21564
- 21 Garbuglia, A. R., Scognamiglio, P., Petrosillo, N., Mastroianni, C. M., Sordillo, P.,  
22 Gentile, D., . . . Capobianchi, M. R. (2013). Hepatitis E Virus Genotype 4  
23 Outbreak, Italy, 2011. *Emerging infectious diseases*, *19*(1), 110-114.  
24 doi:10.3201/eid1901.120983
- 25 Geng, Y., Wang, C., Zhao, C., Yu, X., Harrison, T. J., Tian, K., & Wang, Y. (2010).  
26 Serological prevalence of hepatitis E virus in domestic animals and diversity of  
27 genotype 4 hepatitis E virus in China. *Vector borne and zoonotic diseases*, *10*(8),  
28 765-770. doi:10.1089/vbz.2009.0168
- 29 Graff, J., Torian, U., Nguyen, H., & Emerson, S. U. (2006). A Bicistronic Subgenomic mRNA  
30 Encodes both the ORF2 and ORF3 Proteins of Hepatitis E Virus. *Journal of Virology*,

- 
- 1           80(12), 5919–5926. doi:10.1128/JVI.00046–06
- 2 Hakze–van der Honing, R. W., van Coillie, E., Antonis, A. F., & van der Poel, W. H.
- 3           (2011). First isolation of hepatitis E virus genotype 4 in Europe through swine
- 4           surveillance in the Netherlands and Belgium. *PLoS One*, *6*(8), e22673.
- 5           doi:10.1371/journal.pone.0022673
- 6 Khuroo, M. S. (1980). Study of an epidemic of non-A, non-B hepatitis. Possibility of
- 7           Another Human Hepatitis Virus Distinct From Post-Transfusion non-A, non-B Type.
- 8           *American Journal of Medicine*, *68*(6), 818–824. doi:10.1016/0002-9343(80)90200-4
- 9 Khuroo, M. S., Khuroo, M. S., & Khuroo, N. S. (2016). Hepatitis E: Discovery, global
- 10          impact, control and cure. *World J Gastroenterol*, *22*(31), 7030–7045.
- 11          doi:10.3748/wjg.v22.i31.7030
- 12 Kmush, B., Wierzba, T., Krain, L., Nelson, K., & Labrique, A. B. (2013). Epidemiology of
- 13          hepatitis E in low- and middle-income countries of Asia and Africa. *Semin Liver*
- 14          *Dis*, *33*(1), 15–29. doi:10.1055/s-0033-1338111
- 15 Lu, L., Li, C., & Hagedorn, C. H. (2006). Phylogenetic analysis of global hepatitis E
- 16          virus sequences: genetic diversity, subtypes and zoonosis. *Rev Med Virol*, *16*(1), 5–
- 17          36. doi:10.1002/rmv.482
- 18 Midgley, S., Vestergaard, H. T., Dalgaard, C., Enggaard, L., & Fischer, T. K. (2014).
- 19          Hepatitis E virus genotype 4, Denmark, 2012. *Emerg Infect Dis*, *20*(1), 156–157.
- 20          doi:10.3201/eid2001.130600
- 21 Nan, Y., Wu, C., Qin, Z., & Zhou, E. M. (2017). Zoonotic Hepatitis E Virus: An Ignored
- 22          Risk for Public Health. *Frontiers in Microbiology*, *8*, 2396.
- 23          doi:10.3389/fmicb.2017.02396
- 24 Nan, Y., & Zhang, Y. J. (2016). Molecular Biology and Infection of Hepatitis E Virus.
- 25          *Frontiers in Microbiology*, *7*, 1419. doi:10.3389/fmicb.2016.01419
- 26 Nishizawa, T., Takahashi, M., Mizuo, H., Miyajima, H., Gotanda, Y., & Okamoto, H. (2003).
- 27          Characterization of Japanese swine and human hepatitis E virus isolates of genotype
- 28          IV with 99 % identity over the entire genome. *The Journal of general virology*,
- 29          *84*(Pt 5), 1245–1251. doi:10.1099/vir.0.19052–0
- 30 Ole, W., Sven, S., Judith, K., Martin, K., Camilla, R., Annelie, P., . . . Klaus, S.

- 
- 1 (2008). Phylogenetic and Case-Control Study on Hepatitis E Virus Infection in  
2 Germany. *Journal of Infectious Diseases*, *198*, 1732 - 1741. doi:10.1086/593211
- 3 Panda, S. K., & Varma, S. P. K. (2013). Hepatitis E: Molecular Virology and Pathogenesis.  
4 *Journal of Clinical & Experimental Hepatology*, *3*(2), 114-124.  
5 doi:10.1016/j.jceh.2013.05.001
- 6 Primadharsini, P. P., Nagashima, S., & Okamoto, H. (2019). Genetic Variability and  
7 Evolution of Hepatitis E Virus. *Viruses*, *11*(5). doi:10.3390/v11050456
- 8 Purdy, M. A., Harrison, T. J., Jameel, S., Meng, X. J., Okamoto, H., Van der Poel, W. H.  
9 M., . . . Ictv Report, C. (2017). ICTV Virus Taxonomy Profile: Hepeviridae. *J Gen*  
10 *ViroL*, *98*(11), 2645-2646. doi:10.1099/jgv.0.000940
- 11 Rein, D. B., Stevens, G. A., Theaker, J., Wittenborn, J. S., & Wiersma, S. T. (2012). The  
12 global burden of hepatitis E virus genotypes 1 and 2 in 2005. *Hepatology*, *55*(4),  
13 988-997. doi:10.1002/hep.25505
- 14 Ren, H., Wagner, A. L., Xie, J. Y., Chen, K. Y., Lu, Y. H., Zheng, X. B., . . . Chen, X.  
15 X. (2019). How Do Experts and Nonexperts Want to Promote Vaccines? Hepatitis E  
16 Vaccine as Example. *Health Serv Insights*, *12*, 1178632919897276.  
17 doi:10.1177/1178632919897276
- 18 Sanjuán, R., Nebot, M. R., Chirico, N., Mansky, L. M., & Belshaw, R. (2010). Viral  
19 mutation rates. *J ViroL*, *84*(19), 9733-9748. doi:10.1128/jvi.00694-10
- 20 Smith, D. B., Izopet, J., Nicot, F., Simmonds, P., Jameel, S., Meng, X. J., . . . Purdy,  
21 M. A. (2020). Update: proposed reference sequences for subtypes of hepatitis E  
22 virus (species Orthohepevirus A). *J Gen ViroL*, *101*(7), 692-698.  
23 doi:10.1099/jgv.0.001435
- 24 Smith, D. B., & Simmonds, P. (2018). Classification and Genomic Diversity of Enterically  
25 Transmitted Hepatitis Viruses. *Cold Spring Harb Perspect Med*, *8*, a031880.  
26 doi:10.1101/cshperspect.a031880
- 27 Smith, D. B., & Simmonds, P. (2018). Classification and Genomic Diversity of Enterically  
28 Transmitted Hepatitis Viruses. *Cold Spring Harb Perspect Med*, *8*(9).  
29 doi:10.1101/cshperspect.a031880
- 30 Smith, D. B., Simmonds, P., Izopet, J., Oliveira-Filho, E. F., & Purdy, M. A. (2016).

- 
- 1 Proposed reference sequences for Hepatitis E virus subtypes. *Journal of General*  
2 *Virology*, 97(3), 537. doi:10.1099/jgv.0.000393
- 3 Smith, D. B., Simmonds, P., Jameel, S., Emerson, S. U., Harrison, T. J., Meng, X.  
4 J., . . . Purdy, M. A. (2015). Consensus proposals for classification of the family  
5 Hepeviridae. *Journal of General Virology*, 96(Pt\_10), 2223-2232.  
6 doi:10.1099/vir.0.068429-0
- 7 Smith, D. B., Simmonds, P., Members Of The International Committee On The Taxonomy Of  
8 Viruses Hepeviridae Study, G., Jameel, S., Emerson, S. U., Harrison, T. J., . . .  
9 Purdy, M. A. (2015). Consensus proposals for classification of the family  
10 Hepeviridae. *J Gen Virol*, 96(Pt 5), 1191-1192. doi:10.1099/vir.0.000115
- 11 Sridhar, S., Yip, C., Wu, S., Cai, J., Zhang, J., Leung, K., . . . Teng, L. (2018). Rat  
12 Hepatitis E Virus as Cause of Persistent Hepatitis after Liver Transplant. *Emerging*  
13 *infectious diseases*, 24(12), 2241-2250. doi:10.3201/eid2412.180937
- 14 Tam, A. W., Smith, M. M., Guerra, M. E., Huang, C.-C., Bradley, D. W., Fry, K. E., &  
15 Reyes, G. R. (1991). Hepatitis E virus (HEV): Molecular cloning and sequencing of  
16 the full-length viral genome. *Transboundary and emerging diseases*, 185(1), 120-131.  
17 doi:10.1016/0042-6822(91)90760-9
- 18 Tessé, S., Lioure, B., Fornecker, L., Wendling, M.-J., Stoll-Keller, F. o., Bigaillon, C.,  
19 & Nicand, E. (2012). Circulation of genotype 4 hepatitis E virus in Europe: First  
20 autochthonous hepatitis E infection in France. *Journal of clinical virology : the*  
21 *official publication of the Pan American Society for Clinical Virology*, 54(2), 197-  
22 200. doi:10.1016/j.jcv.2012.02.007
- 23 Tsarev, S. A., Emerson, S. U., Reyes, G. R., Tsareva, T. S., Letgers, L. J., Malik, I.  
24 A., . . . Purcell, R. H. (1992). Characterization of a Prototype Strain of  
25 Hepatitis E Virus. *Proceedings of the National Academy of Sciences of the United*  
26 *States of America*, 89(2), 559-563. doi:10.1073/pnas.89.2.559
- 27 van Tong, H., Hoan, N. X., Wang, B., Wedemeyer, H., Bock, C. T., & Velavan, T. P. (2016).  
28 Hepatitis E Virus Mutations: Functional and Clinical Relevance. *EBioMedicine*, 11,  
29 31-42. doi:10.1016/j.ebiom.2016.07.039
- 30 Wang, Y., Ling, R., Erker, J. C., Zhang, H., Li, H., Desai, S., . . . Harrison, T. J.



1 (1999). A divergent genotype of hepatitis E virus in Chinese patients with acute  
2 hepatitis. *The Journal of general virology*, *80*, 169–177. doi:10.1099/0022-1317-80-  
3 1-169

4 WHO. (2010). Global prevalence of hepatitis E: a systematic review. Available at:  
5 [http://whqlibdoc.who.int/hq/2010/WHO\\_IVB\\_10.14\\_eng.pdf](http://whqlibdoc.who.int/hq/2010/WHO_IVB_10.14_eng.pdf).

6 WHO. (2019). Hepatitis E Available at: [https://www.who.int/news-room/fact-](https://www.who.int/news-room/fact-sheets/detail/hepatitis-e)  
7 [sheets/detail/hepatitis-e](https://www.who.int/news-room/fact-sheets/detail/hepatitis-e).

8 Wu, X., Chen, P., Lin, H., Hao, X., & Liang, Z. (2016). Hepatitis E virus: Current  
9 epidemiology and vaccine. *Hum Vaccin Immunother*, *12*(10), 2603–2610.  
10 doi:10.1080/21645515.2016.1184806

11 Zhang, S., Wang, J., Yuan, Q., Ge, S., Zhang, J., Xia, N., & Tian, D. (2011). Clinical  
12 characteristics and risk factors of sporadic Hepatitis E in central China. *Virology*  
13 *Journal*, *8*(1), 152. doi:10.1186/1743-422X-8-152

14  
15  
16 **Table 1** Geographical distribution of HEV-4 strains (n=2295)

Countries and regions	Number of sequences	Predominant subtype/cluster	Other existing subtypes/clusters
<b>Asia</b>			
China's mainland	1745	4a (33.7%)	4, 4b, 4c, 4d, 4f, 4g, 4h, 4i, uncertain
Japan	243	4c (32.1%)	4, 4a, 4b, 4d, 4e, 4f, 4g, 4h, 4i, uncertain
Hong Kong, China	92	4b (55.4%)	4, 4a, 4d, 4f, 4h, 4i, uncertain
South Korea	37	4a (70.3%)	4c,4d, uncertain
India	27	4e (88.9%)	4b
Taiwan, China	21	4c (57.1%)	4, 4a, 4b, uncertain
Laos	13	4g (38.5%)	4, 4a, 4f, uncertain
Viet Nam	15	4b (73.3%)	4a, 4c, 4h
Mongolia	12	4a (100%)	-

Indonesia	9	4b (77.8%)	4i, uncertain
Malaysia	8	4i (75.0%)	4a, 4g
Cambodia	1	4b	-
Subtotal	2223		
<b>Europe</b>			
France	48	4b (91.7%)	4, 4d, 4h
Russia	9	4b (88.9%)	4
Denmark	3	4b (66.7%)	4d
Italy	3	4d (100%)	-
Belgium	3	uncertain (100%)	-
Switzerland	2	4g	-
Spain	1	4b	-
Germany	1	4	-
United Kingdom	1	4e	-
Subtotal	71		
<b>America</b>			
USA	1	4a	-

1 \* Cluster uncertain consisted of the HEV-4 sequences being separated in the phylogenetic tree, which  
2 could not be classified into an existing subtype, or was composed of the HEV-4 sequences that had  
3 inconsistent phylogenetic findings using neighbor-joining and maximum methods or 48 full-length  
4 HEV reference strains and 10 HEV-4 reference

5

6 **Table 2** Nucleotide identities on the five partial fragments stratified by principal hosts and countries  
7 (SE)

	Met 279 nt	RdRp 307 nt	ORF2 317 nt	ORF2 150 nt	ORF2 537 nt
<b>Hosts</b>					
within human	85.9 (1.6)	83.7 (1.6)	86.4 (1.3)	90.5 (1.5)	86.3 (1.1)

	n=177	n=289	n=304	n=208	n=276
within swine	86.5 (1.6)	85.3 (1.5)	86.2 (1.4)	88.7 (1.8)	85.6 (1.1)
	n=32	n=141	n=243	n=126	n=33
between human & swine	84.6 (1.7)	84.0 (1.6)	85.0 (1.5)	89.0 (1.8)	85.8 (1.1)
<b>Countries</b>					
within China's mainland	86.5 (1.5)	84.7 (1.5)	86.1 (1.4)	89.5 (1.7)	86.7 (1.1)
	n=55	n=375	n=482	n=311	n=266
within Japan	88.9 (1.3)	87.8 (1.3)	89.9 (1.1)	92.3 (1.5)	90.0 (0.9)
	n=111	n=56	n=181	n=393	n=38
between China's mainland & Japan	83.6 (1.8)	81.5 (1.9)	84.95 (1.5)	88.6 (1.9)	84.4 (1.2)

1

2 **Table 3** Nucleotide identities on the five partial fragments stratified by principal hosts and years (SE)

	Met 279 nt	RdRp 307 nt	ORF2 317 nt	ORF2 150 nt	ORF2 537 nt
Before 2000, Human	88.1 (1.5) n=20	87.8 (1.4) n=19	87.8 (1.3) n=29	92.9 (1.4) n=14	90.7 (0.8) n=14
2001-2005, Swine	84.8 (2.6) n=2	NA	87.2 (1.5) n=35	92.1 (1.5) n=17	NA
2001-2005, Human	87.3 (1.5) n=68	84.8 (1.6) n=46	89.0 (1.2) n=75	91.8 (1.4) n=63	86.7 (1.1) n=44
2006-2010, Swine	85.7 (1.7) n=14	84.5 (1.6) n=87	87.9 (1.3) n=79	89.6 (1.6) n=67	85.1 (1.2) n=14
2006-2010, Human	87.3 (1.5) n=55	84.5 (1.6) n=185	90.2 (0.9) n=56	91.0 (1.4) n=95	86.9 (1.0) n=174
2011-2015, Swine	88.0 (1.6) n=7	87.3 (1.3) n=44	89.3 (1.1) n=81	93.2 (1.3) n=20	86.4 (1.1) n=9
2011-2015,	86.8 (1.6)	83.5 (1.7)	85.7 (1.4)	89.6 (1.8)	85.5 (1.1)

Human	n=31	n=26	n=90	n=21	n=41
2016-2019,	93.7 (0.9)	91.5 (1.1)	87.1 (1.4)	91.6 (1.7)	92.2 (0.7)
Swine	n=9	n=9	n=48	n=22	n=9
2016-2019,	88.2 (2.0)	86.2 (1.6)	89.9 (1.1)	94.1 (1.3)	89.3 (0.7)
Human	n=3	n=13	n=54	n=15	n=3

1 \* NA: there was a single sequence.

2 **Table 4** Nucleotide identity on the five partial fragments stratified by combined principal hosts and  
3 countries (SE)

	Met 279 nt	RdRp 307 nt	ORF2 317 nt	ORF2 150 nt	ORF2 537 nt
<b>Within the group</b>					
Japan, human	89.7 (1.2)	87.95 (1.4)	91.3 (1.0)	92.4 (1.5)	90.1 (0.9)
	n=100	n=54	n=141	n=37	n=36
Japan, swine	84.8 (2.5)	84.1 (2.5)	92.6 (1.1)	87.2 (3.3)	87.8 (1.6)
	n=2	n=2	n=16	n=2	n=2
China's mainland, human	86.1 (1.6)	84.7 (1.6)	86.4 (1.4)	91.2 (1.5)	87.1 (1.0)
	n=14	n=223	n=106	n=156	n=221
China's mainland, swine	86.7 (1.6)	85.5 (1.5)	86.5 (1.4)	89.0 (1.8)	86.0 (1.1)
	n=27	n=137	n=218	n=122	n=30
<b>Between the groups</b>					
Japan, human & swine	89.0 (1.4)	85.8 (1.7)	89.4 (1.4)	91.1 (1.9)	89.3 (1.0)
China's mainland, human & swine	85.3 (1.6)	84.8 (1.5)	84.9 (1.5)	89.3 (1.7)	86.4 (1.1)
Human, Japan & China's mainland	83.6 (1.8)	81.4 (1.9)	84.9 (1.6)	89.2 (1.9)	84.4 (1.3)
Swine, Japan & China's mainland	84.1 (1.9)	82.3 (1.9)	84.5 (1.7)	88.3 (2.0)	83.4 (1.4)

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Japan, swine & China's mainland, human	84.6 (1.8)	83.3 (1.8)	85.0 (1.6)	89.6 (1.9)	83.6 (1.4)
Japan, human & China's mainland, swine	83.3 (1.9)	81.1 (2.0)	84.7 (1.6)	88.3 (1.9)	84.3 (1.3)

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1 \* NA: there was a single sequence.

2 **Figure legends**

3 **Fig 1** The roadmap of the inclusion of hepatitis E virus genotype 4 (HEV-4) sequences and  
4 phylogenetic analysis in the study.

5

6 **Fig 2** Temporal distribution of isolated hepatitis E virus genotype 4 (HEV-4) strains in 1992-2018.

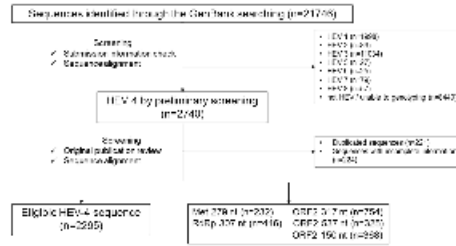
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8 **Fig 3** Geographical distribution of isolated hepatitis E virus genotype 4 (HEV-4) strains.

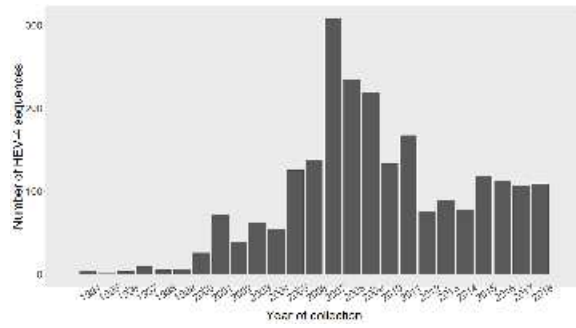
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10 **Fig 4** Isolation of hepatitis E virus genotype 4 (HEV-4) subtypes in 1992-2018.

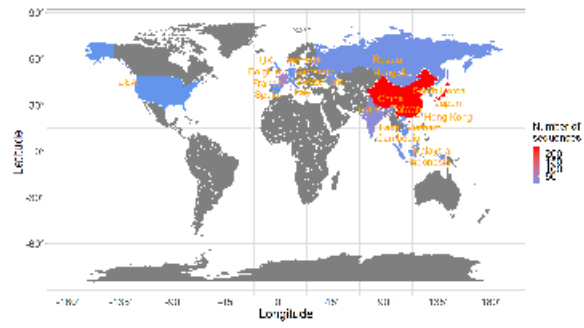
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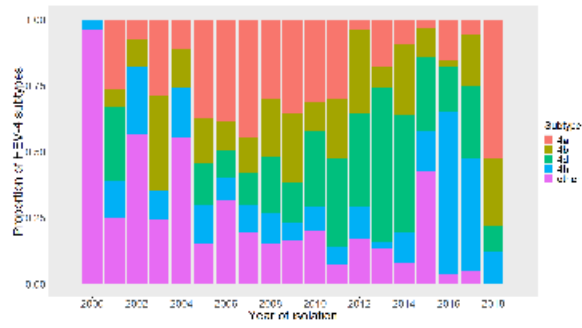


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