CCNE1 and survival of patients with tubo-ovarian high-grade serous carcinoma: an Ovarian Tumor Tissue Analysis consortium study.

Running title: CCNE1 in high-grade serous carcinoma

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Precis (Condensed Abstract) for use in the Table of Contents: Multi-institutional validation of the prognostic value of CCNE1 high-level amplification and overexpression in 3029 tubo-ovarian high-grade serous carcinomas cases supporting its value as prognostic biomarker in this disease.

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Author contributions statement

S. J. Ramus and M. Koebel conceived, designed, and supervised the study. E. Kang and A. Weir contributed equally to the work. E. Kang, K. Farrington, and M. Koebel collected immunohistochemical protein and chromogen in situ hybridization scores. A. Weir performed analyses. E. Kang and M. Koebel drafted the manuscript and revised the manuscript. P.D.P. Pharoah provided statistical advice. All authors contributed through collection, curation and maintenance of respective consortia based, or local institution, collections of patient samples including recruitment and consenting of patients, clinical care, abstraction of clinical data, and updating of outcome and follow up data. All authors revised the manuscript and approved submission of the final version.

Abstract

Background: Cyclin E1 (CCNE1) is a potential predictive marker and therapeutic target in tubo-ovarian high-grade serous carcinoma (HGSC). Smaller studies have revealed unfavorable associations for *CCNE1* amplification and CCNE1 overexpression with survival, but to date no large-scale, histotype-specific validation has been performed. We hypothesized that high-level amplification of *CCNE1* and CCNE1 overexpression, as well as a combination of the two, are linked to shorter overall survival in HGSC.

Methods: Within the Ovarian Tumor Tissue Analysis (OTTA) consortium, we investigated amplification status and protein level in 3029 HGSC cases, and mRNA expression in 2419 samples.

Results: High-level amplification (>8 copies by chromogenic *in situ* hybridization) was found in 8.6% of HGSC and overexpression (>60% with at least 5% demonstrating strong intensity by immunohistochemistry) was found in 22.4%. CCNE1 high-level amplification and overexpression both were linked to shorter overall survival in multivariate survival analysis adjusted for age and stage, with hazard stratification by study (HR=1.26, 95% CI 1.08-1.47, p=0.034, and HR= 1.18, 95% CI 1.05-1.32, p=0.015, respectively). This was also true for cases with combined high-level amplification/ overexpression (HR=1.26, 95% CI 1.09-1.47, p=0.033). *CCNE1* mRNA expression was not associated with overall survival (HR=1.00 per one standard deviation increase, 95% CI 0.94-1.06, p=0.58). We confirmed that *CCNE1* high-level amplification is mutually exclusive with the presence of germline *BRCA1/2* pathogenic variants and shows an inverse association to RB1 loss.

Conclusion: Our study provides large-scale validation that *CCNE1* high-level amplification is associated with a shorter survival, supporting its utility as a prognostic biomarker in HGSC.

Keywords: Ovarian cancer, high-grade serous carcinoma, CCNE1 amplification, prognosis, Cyclin E1 expression

Introduction

Cyclin E1 (CCNE1) is a potential predictive marker and therapeutic target in tubo-ovarian high-grade serous carcinoma (HGSC) [1, 2]. CCNE1 has three main functions in cell cycle progression [3]. First, it is involved in the formation of pre-replication minichromosome maintenance (MCM) protein complexes, which bind origins of DNA replications as cells re-enter G1- from G0-phase of the cell cycle. Second, by forming a complex it activates the cyclin-dependent kinase CDK2 to phosphorylate several targets including RB1, which subsequently abandons its inhibition of E2F transcription factors and initiates the transition from G1- to S-phase [3]. CDK2 inhibition by cyclin-dependent kinase inhibitors 1 (CDKN1a/p21) is dependent on normal TP53 function. Third, the CDK2/CCNE1 complex promotes centrosome duplication[3, 4]. Normal CCNE1 protein levels are tightly regulated peaking in late G1 and decreasing as cells progress through S-phase [5]. In neoplasia, CCNE1 protein overexpression is uncoupled from the cell cycle [6]. Constitutive overexpression of CCNE1, but not of CCND1 or CCNA, induces chromosomal instability and a modest degree of polyploidy [6]. The mechanisms by which CCNE1 causes chromosomal instability are not entirely understood but it has been suggested that cells with deregulated CCNE1 prematurely enter S phase with inadequate nucleotide pools causing replication stress with faulty replication forks engendering DNA double strand breaks [7, 8].

In ovarian carcinoma, *CCNE1* amplification has been associated with resistance to platinum-based chemotherapy and shorter overall survival [9, 10]. However, the cut-off for amplification varies between studies and larger studies such as The Cancer Genome Atlas (TCGA) project reported only a suggestive trend towards shorter overall survival (p=0.0718) and another study of 179 HGSC showed evidence for a significant association only with progression-free survival [11, 12]. Amplification of the chromosomal region 19q12 containing the *CCNE1* gene is common (~20%) in HGSC, which across all tumor sites ranks third in

frequency after endometrial carcinosarcoma and urothelial carcinoma [13]. *CCNE1* amplification is inversely associated with germline pathogenic *BRCA1/2* variants, which becomes mutually exclusive for high-level amplifications (defined by > 8 copies) [14, 15]. *CCNE1* high-level amplified HGSC require proficient homologous recombination (HR), including BRCA1/2 function to maintain cell viability [14, 15]. *CCNE1* high-level amplification is the lead alteration for both the copy number signature 6 and the fold back inversion mutation signature, which characterize to HR-proficient HGSC [16, 17]. HGSC patients with HR-proficient tumors do not respond well to chemotherapy or poly (ADP-ribose) polymerase (PARP) inhibitors. For example, PARP maintenance therapy for patients with HR-proficient HGSC with partial chemotherapy response resulted in a median progression-free survival of 8.3 months compared with 21.9 months for patients with HR-deficient HGSC [18].

While no association of *CCNE1* mRNA expression with survival in HGSC has been observed [15, 19], CCNE1 protein overexpression has been associated with unfavorable outcomes in ovarian carcinomas, but only in studies conducted before the era of histotype-specific analysis [20-22]. Two recent studies suggested that combining *CCNE1* amplifications and CCNE1 overexpression is associated with shorter survival [15, 23]. We recently validated a CCNE1 chromogenic *in situ* hybridization (CISH) assay orthogonally against other copy number assays to be applicable on tissue microarrays and refined the cut-off for immunohistochemistry to detect *CCNE1* high-level gene amplifications[15].

Here our objectives were to validate previously reported associations of CCNE1 alterations with overall survival, assess correlations between *CCNE1* high-level gene amplifications, *CCNE1* mRNA, and CCNE1 protein expression, and explore associations with selected biomarkers in a large cohort of HGSC samples from the international Ovarian Tumor Tissue Analysis (OTTA) consortium.

Methods

Study cohort

Twenty studies from the Ovarian Tumor Tissue Analysis (OTTA) consortium participated in the current study [24]. Each study enrolling patients received local ethics review board approval (Table S1). Tissue microarrays were constructed from formalin-fixed paraffin-embedded (FFPE) tumor specimens obtained from debulking surgery representing each tumor with 1-3 cores. 0.6 to 1.mm in size. For both CISH and immunohistochemistry (IHC), data were successfully obtained in 3029 individual HGSC patient samples. Clinical covariates, time to follow up, and status were centrally standardized. Cases were collected during the pre-PARP inhibitor era. Platinum-based chemotherapy was given in the majority as adjuvant after primary debulking surgery (PDS) or as neoadjuvant chemotherapy (NACT). Information on specific drugs was not collected. Previously generated IHC data within the OTTA consortium for TP53, CDKN2A and RB1 were used [25-27].

CCNE1 DNA CISH

A previously published in-house CISH protocol using a commercial Digoxigenin (DIG)-labeled *CCNE1* DNA probe (Empire Genomics, Buffalo, NY, USA) was utilized [15]. De-paraffinized 4 µm tissue microarray sections were pretreated with proteinase K (3 min) and citrate-based antigen retrieval buffer at 80°C (1 hr) followed by pepsin (45 sec), then dehydrated and air-dried. Hybridization with the DIG-labelled *CCNE1* probe was carried out at 37°C for 16-18 hours in HybEZ™ II (Advanced Cell Diagnostics, Minneapolis, MN, USA). A levamisole solution was used (15 min) to remove endogenous alkaline phosphatase activity, followed by a blocking solution (30 min) of 10% normal sheep serum, 2% bovine

serum albumin, and 0.05% Tween-20. An Alkaline phosphatase (AP)-conjugated sheep anti-DIG antibody (dilution 1:800; Roche, Basel, Switzerland) was incubated for 2 hours. AP substrate was applied, and the reaction was stopped with 50mM Tris, 150 mM NaCl, 10 mM KCl buffer when slides reached the desired intensity of staining. Counterstaining was performed with hematoxylin.

CCNE1 immunohistochemistry

Four µm sections from tissue microarrays were de-paraffinized, rehydrated, and subjected to heat-induced epitope retrieval on the DAKO Omnis platform (Agilent Technologies, Santa Clara, CA, USA), followed by incubation with the CCNE1 antibody (1:600, clone EP126, Cell Marque, Rocklin, CA, USA, 30-10R-30) at room temperature and in the EnVision FLEX (Agilent Technologies). The reaction was visualized using 3,3-diaminobenzidine tetrahydrochloride (DAB) for 10 minutes and counterstained with hematoxylin.

CCNE1 CISH and IHC scoring

The *CCNE1* CISH assay was previously orthogonally validated to detect *CCNE1* high-level amplifications (presence of clusters > 8 copies) against digital PCR and nCounter Cancer CN Assay [15]. CCNE1 protein expression showed a wide and relatively even distribution from 5-90% of positive tumor cells, but previous receiver operating characteristic curve (ROC) analysis established an optimal cut-off for IHC to detect high-level amplification at > 60% overall staining cells with at least 5% showing strong intensity [15]. The interpretation of tissue microarrays was performed by three pathologists. Training was provided on a set of 90 HGSC cases guided by illustrated examples. Subsequently, interobserver reproducibility was tested on 415 cases. The interobserver observer reproducibility for paired observers achieved a Cohen's

kappa of 0.48, 0.55, and 0.77 for CISH and 0.65, 0.75, and 0.85 for IHC using a binary categorization. Subsequently, examples of discordant cases were discussed at a multiheaded microscope to further align interpretational thresholds and equivocal categories were allowed both for CISH and IHC [15]. Each observer subsequently scored approximately one-third of the cases by using the following criteria for CISH: score 0 – no clusters = negative for high-level amplification ($CCNE1^{nonamp}$), score 1 – equivocal favor negative, score 2 - equivocal favor high, score 3 – nuclear clusters of CISH signal = high-level amplification ($CCNE1^{amp}$), and for IHC: score 0 – < 60% positive tumor cells ($CCNE1^{lo}$), score 1 – equivocal favor low , score 2 – equivocal favor high, score 3 – ≥ 60% positive tumor cells with at least 5% strongly staining cells ($CCNE1^{hi}$).

CCNE1 mRNA expression by NanoString

FFPE tumor specimens (n=2419) with partial overlap to above (1612/3029) were obtained from additional cores or sections [28]. RNA extraction methods, assay run parameters, data processing, and control/reference samples were previously described [29]. *CCNE1* mRNA expression was assessed using the NanoString nCounter technology; the *CCNE1* target sequence was CCTCCAGACACCAGTGCGTGCTCCCGATGCTGCTATGGAAGGTGCTACTTGACCTAAGGGACTCCCACAACAACAA AAGCTTGAAGCTGTGGAGGGCCAC, and *CCNE1* mRNA data were normalized against housekeeping genes [29]. Quality assurance of the assay was previously performed with high duplicate sample correlation [19, 30].

Statistical analyses

Correlations between *CCNE1* mRNA, gene amplification (ISH), and protein (IHC) overexpression were measured using Pearson correlation coefficients. Chi-square proportions testing was undertaken to

evaluate clinical and molecular variables across CCNE1 combinations. Univariate and multivariate survival analyses of *CCNE1* profiles were performed. Overall survival (death from any cause) was the primary endpoint. Potential survival bias introduced by the time between diagnosis and study enrolment was moderated by left-truncation. Deaths potentially unrelated to HGSC were right-censored at 10 years from diagnosis. The Kaplan-Meier method, alongside log-rank testing, was used to assess overall survival by *CCNE1* profile. Multivariate Cox proportional hazards regression modelling, stratified by OTTA study, complemented this analysis through estimation of hazard ratios (HRs) with 95% confidence intervals (CIs). The covariates, age, stage, completeness of surgical cytoreduction (residual disease vs. no residual disease (sensitivity analysis), and *CCNE1* profiles were adjusted for, and different baseline hazards of OTTA studies were stratified, in multivariate models. Scaled Schoenfeld residuals assessed the assumption of proportional hazards. All statistical analyses were carried out using RStudio v1.1.463 or GraphPad Prism v7.02. Statistical significance was defined by $p<0.05$.

Results

Prevalence of CCNE1 high-level amplification and association with overall survival

CCNE1 CISH showed high-level amplification (score 3) in 259/3029 (8.6%) cases and 2426/3029 (80.2%) demonstrated no evidence of amplification (score 0). The remainder were equivocal with 67/3029 (2.2%) favored high-level amplification (score 2), and 277/3029 (9.1%) not favored (score 1). Kaplan-Meier survival analysis showed a significantly different overall survival between the groups (log-rank p=0.00016, Figure 1A). In multivariate analysis adjusted for age and stage and stratified for OTTA study, *CCNE1* high-level amplified HGSC showed a hazard ratio of 1.26 (95% CI 1.08-1.47) compared with the reference group with no evidence of amplification (Table 1). Data on the completeness of surgical cytoreduction was available for a subset of cases (66.9%) and within this group, a sensitivity analysis adjusted for age, stage, completeness of surgical cytoreduction, and stratified for OTTA study, resulted in the same hazard ratio of 1.27 (95% CI 1.06-1.52, Table S2).

Prevalence of CCNE1 protein overexpression and association with overall survival

CCNE1 IHC showed overexpression (score 3) in 678/3029 (22.4%) cases and 1824/3029 (60.2%) had low CCNE1 protein levels (score 0) (Table 1). The remainder were equivocal, with 233/3029 (7.7%) favored to express high protein levels (score 2) and 301/3029 (9.9%) favored to express low levels (score 1). Kaplan-Meier survival analysis showed a significantly different survival between the groups (log-rank p=0.021, Figure 1B). In multivariate analysis adjusted for age and stage, and stratified for OTTA study, CCNE1 high protein level HGSC showed a hazard ratio of 1.18 (95% CI 1.05-1.32) compared with the group with low CCNE1 protein levels (Table 1). In a sensitivity multivariate analysis adjusted for age, stage, completeness of surgical cytoreduction, and stratified for OTTA study, a similar hazard ratio of 1.20 (95% CI 1.05-1.37) was obtained (Table S2).

Associations of combined CCNE1 high-level amplification and protein level with overall survival

After binarization of CISH and IHC scores into scores 0/1 versus scores 2/3, 265/326 (81.3%) of high-level amplified cases showed high CCNE1 protein levels, and conversely, 2064/2703 (76.4%) of non-high-level amplified cases showed low CCNE1 protein levels. We then combined CCNE1 CISH and IHC into four groups (Figure 2, Table S3): first, negative for *CCNE1* high-level amplification with low CCNE1 protein expression ($CCNE1^{nonamp\_lo}$) comprising 68.1% (2064/3029) of the cases; second, negative for *CCNE1* high-level amplification but with CCNE1 protein overexpression ($CCNE1^{nonamp\_hi}$) comprising 21.1% (639/3029); third, *CCNE1* high-level amplification but low CCNE1 protein expression ($CCNE1^{amp\_lo}$) comprising 2.0% (61/3029); and fourth, *CCNE1* high-level amplification with CCNE1 protein overexpression ($CCNE1^{amp\_hi}$) comprising 8.8% (265/3029) of cases (Table 1). Kaplan-Meier survival analysis showed a significantly different overall survival between the groups (log-rank $p<0.0001$, Figure 2). Patients in the $CCNE1^{amp\_hi}$ group had a 5-year survival rate of 28.3% compared with 41.9% in the $CCNE1^{nonamp\_lo}$ group (Table 1). This difference remained significant in multivariate modeling, following adjustment for age and stage and stratified for OTTA study. The $CCNE1^{amp\_high}$ group had a higher risk of death (hazard ratio 1.26, 95% CI 1.09-1.47) compared with the reference $CCNE1^{nonamp\_lo}$ group (Table 1). In a sensitivity analysis adjusted for age, stage, completeness of surgical cytoreduction, and stratified for OTTA study, the hazard ratio for the $CCNE1^{amp\_high}$ group compared with the reference $CCNE1^{nonamp\_lo}$ group was 1.20 (95% CI 1.00-1.43; Table S2).

Associations of combined CCNE1 high-level amplification and protein expression with clinical parameters and biomarkers in HGSC

The univariate associations of the combined groups with clinicopathological parameters are shown in Table 2. Patients diagnosed with *CCNE1* high-level amplified HGSC were older, with a trend towards a higher likelihood of residual disease after debulking surgery. No associations were observed for stage (FIGO I, II (local/regional) compared with FIGO III/IV (distant)) or the timing of the primary chemotherapy regimen (adjuvant versus neoadjuvant, Table S4). For subsets with available data, the four groups showed significant associations with TP53 IHC (available data for 65.9% of cases), *BRCA1/2* germline variant (available data for 31.5% of cases), CDKN2A IHC (available data for 64.2 of cases), and RB1 IHC status (available data for 71.1% of cases, Table 3). Normal TP53 IHC was most prevalent in the CCNE1^{nonamp_lo} group. However, the abnormal TP53 IHC patterns, which are surrogates for the functional groups of *TP53* mutation [31], were not different. Germline *BRCA1/2* mutations were rarely present in *CCNE1* high-level amplified HGSC. Only two HGSC cases had protein-truncating deleterious *BRCA2* variants and both cases had a *CCNE1* CISH score of 2 (equivocal favor high). The CCNE1^{amp_high} group had the highest frequency of CDKN2A block expression, a surrogate for RB pathway activation, but there were no cases with complete absence of CDKN2A expression, a surrogate for a deleterious deletion of *CDKN2A*. *CCNE1* high-level amplification was inversely associated with loss of RB1.

CCNE1 mRNA expression by NanoString in HGSC

For 1612/3029 overlapping cases with *CCNE1* mRNA expression, there was moderate correlation between *CCNE1* mRNA expression and CISH scores (r=0.476) and CCNE1 IHC scores (r=0.545, Figure 3). *CCNE1* mRNA expression was also different across the four combined groups, with the highest level observed in CCNE1^{amp_high}, followed by CCNE1^{amp_lo} and CCNE1^{nonamp_hi} (Figure 3). Lastly, we evaluated the associations of *CCNE1* mRNA expression with overall survival in 2419 HGSC cases. The clinicopathological characteristics of these cases are shown in Table S5. When considering one standard

deviation increase in *CCNE1* mRNA expression score, there was no association with overall survival (HR=1.00, 95% CI 0.94 - 1.06, p=0.96, Table S6). This also was the case when using a cut-off at the top 10% versus the remainder ((HR=1.06, 95% CI 0.88 - 1.27, p=0.53,Table S6).

Discussion

In this study, we validate the association of combined *CCNE1* high-level gene amplification and CCNE1 protein overexpression with overall survival in a large cohort of patients with HGSC from the OTTA consortium. Our results demonstrate that assessing *CCNE1* at a DNA copy number level and protein level is a more robust determinant of prognosis than mRNA expression. We also confirm that *CCNE1* high-level amplification is essentially mutually exclusive with pathogenic *BRCA1/2* germline alterations and associated with biomarker changes in the RB1 pathway.

For the association of CCNE1 protein expression with survival, the genomic context seems to matter. The fairly large group of CCNE1^{nonamp_hi} shows a similar survival compared with the CCNE1^{nonamp_lo} reference but longer survival relative to the CCNE1^{amp_hi} group. Both the CCNE1^{nonamp_hi} and CCNE1^{amp_hi} group express similarly high protein levels but CCNE1^{amp_hi} express higher mRNA levels than CCNE1^{nonamp_hi} suggesting that amplification-driven CCNE1 overexpression is due to higher transcriptional activity, while CCNE1 overexpression in CCNE1^{nonamp_hi} cases may be more dependent on protein stabilization/lack of degradation [32]. We, however, demonstrate that differences in *CCNE1* mRNA expression were not associated with overall survival in HGSC. We obtained consistent hazard ratios close to 1.0 by studying 2419 samples in the current study, confirming the results from a previous OTTA study [19]. Despite the strong correlation of mRNA with amplification status and protein levels, the lack of survival association may be caused by dilution of the mRNA signal in tumor bulk analysis with varying tumor cellularity compared to the spatially controlled CISH and IHC assays. The survival differences of the two groups with high CCNE1 protein levels still creates a conceptual dilemma since it is the protein that is exerting the function, and the mechanism of protein accumulation should not matter, unless there is a difference in the timing of expression in relation to the cell cycle or the functional quality of

CCNE1 protein. In high CCNE1-expressing breast cancer, CCNE1 can be proteolytically cleaved into low-molecular weight derivatives [33]. An alternative explanation might be that in the CCNE1^{amp_hi} group other oncogenes co-amplified with CCNE1 on 19q12, such as *URI* contribute to survival [34].

While we confirm that the group of CCNE1^{amp_hi} is associated with the shortest overall survival, we also show that this association is mainly driven by DNA copy number status, achieving a better stratification than protein level. However, protein level seems to provide additional information by singling out the small group of CCNE1^{amp_lo}, which in the main analysis had a similar HR compared with the reference CCNE1^{nonamp_lo}. In a sensitivity analysis including residual tumor, the HR was more similar to the high-risk group CCNE1^{amp_hi}. Albeit, this was not statistically significant with small case numbers in the CCNE1^{nonamp_lo} subgroup and this sensitivity analysis may have introduced bias for the small subgroups that are not comparable to the overall cohort. This raises a question about the importance of the level of CCNE1 protein expression in the context of *CCNE1* high-level amplification. Both CCNE1^{amp_hi} and CCNE1^{amp_lo} groups are similar regarding clinical parameters (i.e., age, residual disease) and rarely harbored *BRCA1/2* germline alterations and loss of RB1 was uncommon. While this suggests no difference and assessment of the DNA copy number status would be sufficient, both groups differed in regard to the abnormal block CDKN2A expression status, which was highest in CCNE1^{amp_hi}, indicating a higher RB1 pathway dependent on the CCNE1 protein level. Based on out observed differences in survival and *CCNE1* mRNA expression, together with previous study findings [15, 23], we interpret that CCNE1^{amp_hi} is different from CCNE1^{amp_lo}. By focusing on the CCNE1^{amp_hi} group, IHC can be used to screen HGSC samples for CCNE1 overexpression followed by copy number assessment for clinical trial inclusion, which would pragmatically circumvent the limited sensitivity of CCNE1 IHC. However, the clinical significance of this relatively small group remains uncertain. We cannot entirely exclude a misclassification based on the CISH or the IHC assay. Future studies should use full section IHC to exclude potential intratumoral heterogeneity of the protein expression and alternative copy number

assay for the small group of CCNE1^{amp_lo} tumors. But some *CCNE1* high-level amplified tumors may not express high protein levels. TCGA reported that low *CDKN2A* mRNA expression is mutually exclusive with *CCNE1* amplification [11]. We observed a small number of cases with loss of CDKN2A protein (a surrogate for *CDKN2A* deep deletions) in the CCNE1^{amp_lo} but not in the CCNE1^{amp_hi}, suggesting that another concurrent RB1 pathway alteration could prevent CCNE1 protein overexpression in the context of *CCNE1* high-level DNA amplifications.

Our results confirm that CCNE1 high-level DNA amplifications are essentially mutually exclusive with pathogenic *BRCA1/2* germline alterations. The two exceptional cases with pathogenic *BRCA2* germline variants that were grouped as CCNE1^{amp_hi} were scored as equivocal favor high by CISH. These rare cases of "double classifiers" may require additional assays such as validated HRD assays or copy number signatures to assign as HR-deficient or HR-proficient. From a treatment perspective, the CCNE1^{amp_hi} group had a shorter survival likely in part due to lower response to platinum-based chemotherapy, which correlates with insensitivity to PARP inhibitors. Therefore, the CCNE1^{amp_hi} group may not respond to PARP inhibitors, making CCNE1^{amp_hi} a candidate biomarker that could be used as a negative predictive test for PARP inhibitors. This hypothesis could be tested in secondary analyses of clinical trials that include unselected HGSC patients treated with PARP inhibitors [18].

Novel treatment approaches are required for women diagnosed with CCNE1^{amp_hi} HGSC [35]. Bowtell and colleagues observed decreased tumorigenesis in *CDK2*-knockout HGSC cell lines with *CCNE1* amplifications. However, the CDK2 inhibitor, dinaciclib, did not suppress tumorigenesis, probably because it is not entirely specific for CDK2 [36]. Perhaps, a more specific CDK2 inhibitor could be tested on HGSC patients with CCNE1^{amp_hi} HGSC. It remains to be seen whether redundancies in the CDK2/CCNE1

pathway (CDK1 for CDK2, CCNE2 for CCNE1) observed in normal cells pose another challenge of targeting this pathway in cancers [3,37]. In post-hoc analysis of a clinical trial investigating the Wee1 inhibitor adavosertib in combination with gemcitabine, *CCNE1* amplified tumors were more likely to respond [38]. Through phosphorylation of the CDK1/CCNB complex, Wee1 kinase is an inhibitor of the G2/M transition, which is more critical for HGSC with deficient G1/S transition. Notably, in a recent phase II trial adavosertib has also shown promising response rates in CCNE1 overexpressing recurrent HGSC irrespective of amplification status [39]. Alternatively, using a CRISPR–Cas9-screen *PKMYT1*, a protein kinase also involved in G2/M transition, was identified as synthetic lethal target for CCNE1 high expressing cells, which were sensitive to inhibition by a selective *PKMYT1* inhibitor [40]. This suggests that perhaps both CCNE1 expression and amplification status should be assessed when testing CCNE1 as predictive marker for new molecular therapy.

While the main CCNE1 function is in cell cycle progression, it is important to note that the main oncogenic effect may be independent from proliferation. High proliferating HGSC are associated with longer survival, likely due to better response to standard chemotherapy [27, 28], whereas CCNE1 alterations are associated with shorter survival and poor response to chemotherapy. CCNE1 protein expression is only weakly correlated with proliferation markers (Ki67, MCM3)[28]. While uncontrolled cell cycle entry remains the main known function of CCNE1, overall, these data suggest that much of CCNE1's oncogenic function is related to a premature S-phase entry resulting in chromosomal instability rather than increased proliferation [7, 8].

The main limitation of our study was the assay resolution. We did not count the DNA copy number ratio by using a CEP19 control probe but focused on the presence of *CCNE1* clusters as a surrogate for high-

level amplification defined by > 8 copy numbers, which was previously orthogonally validated using the NanoString CNV assay and digital PCR [15]. Not using ratios prevented us from assessing low level gains. The prevalence of *CCNE1* high-level amplifications is about half compared to previous studies reporting *CCNE1* amplification (frequency of ~20%) which is due to the higher cut-off we used [11, 12]. Our present study utilized CISH analysis, which a well-established and clinically adopted technique to interrogate genetic amplification such as evaluation of *ERBB2* amplification in breast and gastric cancer. However, Next Generation Sequencing (NGS)-based assays such as whole genome/exome sequencing or targeted panel sequencing are being increasingly utilized in the clinical setting to provide more comprehensive molecular characterization of tumor, including copy number alterations. In contrast to CISH (or FISH) assays that provide spatially focused analysis that evaluates signals only from carcinoma cells, the NGS-based assays typically use bulk tumor samples in which tumor content can vary and it may have lower sensitivity compared to spatially controlled assays such as ISH, particularly from samples with low tumor content in the settings of core needle biopsies or post-treatment (neoadjuvant chemotherapy) samples. Another consideration is that CISH analysis generally requires less amount of tumor tissue than NGS-based analysis, which may be relevant in cases where a limited amount of diagnostic tissue is available. Future studies are required to determine the clinical utility and limitations of NGS-based assay for *CCNE1* copy number evaluation. There were limited data annotations for some analyses due to missing data for residual disease and germline *BRCA1/2* status.

In conclusion, our large-scale validation with survival data supports the notion that CCNE1 is the most promising biomarker to define the largest subgroup of HR-proficient HGSC. CCNE1 high-level amplifications should be studied as negative predictive markers for current standard therapies (chemotherapy, PARP inhibitors) and should be evaluated in clinical trials assessing novel treatment approaches. We propose to focus initially on the $CCNE1^{amp\_hi}$ group; CCNE1 IHC could be used as a screening tool, followed by an assessment of DNA copy number status.

Figure legends

Figure 1. Kaplan Meier overall survival analysis for (A) CISH score levels and (B) IHC score levels.

Figure 2. A. *CCNE1* DNA CISH and IHC combinations resulting in 4 groups: CCNE1^{nonamp_lo} CISH showing no high-level amplification and IHC < 60% positive tumor cell nuclei, CCNE1^{nonammp_hi} CISH showing no high-level amplification and IHC > 60% positive and >5% strongly staining tumor cell nuclei, CCNE1^{amp_lo} CISH showing high-level amplification and IHC < 60% positive tumor cell nuclei, CCNE1^{amp_hi} CISH showing high-level amplification and IHC > 60% positive and >5% strongly staining tumor cell nuclei. B. Kaplan Meier overall survival analysis for four combined CISH/IHC groups. C. Risk table indicating the number of patients within the cohort that are at risk of death, observed at a yearly.

Figure 3. A. Correlation of CCNE1 DNA CISH score with normalized mRNA expression, B. Correlation of CCNE1 protein IHC score with normalized RNA expression. C. Association of four combined CISH/IHC groups with normalized RNA expression. Pearson's correlation analysis given by r. *p<0.05

Supplemental material

Table S1 OTTA study cohorts

Table S2 Sensitivity analyses: Multivariable association between the expression and amplification of CCNE1 and HGSC OS (n=2026) – sensitivity analysis including residual tumor

Table S3 Binarized CCNE1 amplification and protein level combination

Table S4 Univariate associations of the combined CCNE groups with type of chemotherapy regimen

Table S5 Clinicopathological parameters of mRNA expression/NanoString 2 cohort (n=2419

Table S6 Multivariable survival analysis of CCNE1 mRNA expression

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Table 1 Multivariate association between the expression and amplification of CCNE1 and overall survival in high-grade serous ovarian carcinoma (n=3029)

CCNE1 profile	**n** [a]	**5-yr survival (% ± SE)**	**Hazard ratio (95% CI)** [b]	**p-value**
CCNE1 CISH score 0	2426	41.9 ± 1.1	ref	**0.034***
CCNE1 CISH score 1	277	40.2 ± 3.1	0.98 (0.84-1.14)	
CCNE1 CISH score 2	67	32.1 ± 6.2	0.97 (0.72-1.31)	
CCNE1 CISH score 3	259	29.5 ± 3.0	**1.26 (1.08-1.47)***	
CCNE1 IHC score 0	1824	41.6 ± 1.2	ref	**0.015***
CCNE1 IHC score 1	301	43.0 ± 3.0	0.99 (0.85-1.16)	
CCNE1 IHC score 2	233	42.8 ± 3.4	0.92 (0.78-1.10)	
CCNE1 IHC score 3	678	35.4 ± 2.0	**1.18 (1.05-1.32)***	
$CCNE1^{nonamp\_lo}$	2064	41.9 ± 1.2	ref	**0.033***
$CCNE1^{nonamp\_hi}$	639	41.0 ± 2.1	1.04 (0.93-1.16)	
$CCNE1^{amp\_lo}$	61	37.8 ± 6.6	0.97 (0.71-1.34)	
$CCNE1^{amp\_high}$	265	28.3 ± 3.0	**1.26 (1.09-1.47)***	

[a] The same cohort was assessed in univariate survival analysis

[b] Hazard ratio adjusted for patient age and stage, with stratification by OTTA study; Cox proportional regression modelling was used to calculate p-values and define significance. Statistically significant values shown in bold; * $p<0.05$

Cyclin E1 (CCNE1); High-grade serous ovarian carcinoma (HGSC); Overall survival (OS); Standard error (SE); Hazard ratio (HR); Confidence interval (CI); Chromogenic in situ hybridization (CISH); Immunohistochemistry (IHC); Negative for CCNE1 high-level amplification ($CCNE1^{nonamp}$); CCNE1 high-level amplification ($CCNE1^{amp}$); Negative for CCNE1 protein overexpression by immunohistochemistry ($CCNE1^{lo}$); CCNE1 protein overexpression by immunohistochemistry ($CCNE1^{hi}$)

Table 2 Clinicopathological parameters by combined CCNE1 protein and amplification status (n=3029)

Clinicopathological variable	CCNE1 profile				p-value [a]	Total
	CCNE1^{nonamp_lo}	CCNE1^{nonamp_hi}	CCNE1^{amp_lo}	CCNE1^{amp_hi}		
Number of cases, n (%) [b]	2064 (68.1)	639 (21.1)	61 (2.0)	265 (8.8)		3029 (100.0)
Age at diagnosis, years						
Mean ± SD	60.9 ± 11.4	61.7 ± 10.9	65.0 ± 9.11	65.0 ± 9.8		61.5 ± 11.2
Median	61	62	66	65		62
Range	21-93	30-92	40-84	38-91		21-93
Stage, n (%) [c]					0.3848	
FIGO I, II (local/regional)	350 (17.0)	124 (19.4)	9 (14.8)	41 (15.5)		525 (17.3)
FIGO III, IV (distant)	1714 (83.0)	515 (80.6)	52 (85.2)	224 (84.5)		2527 (82.7)
Completeness of survival cytoreduction					0.0563 [d]	
No residual disease, n (%) [c]	555 (40.7)	200 (44.5)	10 (34.5)	61 (33.2)		826 (40.8)
Residual disease present, n (%) [c]	809 (59.3)	249 (55.5)	19 (65.5)	123 (66.9)		1200 (59.2)
Unknown, n [c]	700	190	32	81		1003

[a] Chi-squared testing was used to calculate p-values. Statistically significant values shown in bold; *p<0.05.
[b] The proportion of cases in each score stratum is given as a percentage of the total patients examined
[c] The proportion of cases is given as a percentage of the total cases within each score stratum
[d] Chi-squared testing to compare the proportions of cases with absent vs present residual disease status. This does not include cases where residual disease status was unknown.

Cyclin E1 (CCNE1); Standard error (SE); Negative for CCNE1 high-level amplification (CCNE1^{nonamp}); CCNE1 high-level amplification (CCNE1^{amp}); Negative for CCNE1 protein overexpression by immunohistochemistry (CCNE1^{lo}); CCNE1 protein overexpression by immunohistochemistry (CCNE1^{hi}); Standard deviation (SD); International Federation of Gynecology and Obstetrics (FIGO)

Table 3 Univariable associations with selected biomarkers by combined CCNE1 protein and amplification status (n=3029)

Molecular marker [b]	Status	CCNE1 profile [a]				Total [c]	p-value [d]
		CCNE1^{nonamp_lo}	CCNE1^{nonamp_hi}	CCNE1^{amp_lo}	CCNE1^{amp_hi}		
TP53	**Abnormal**	1202 (89.9)	413 (94.3)	42 (100.0)	172 (95.6)	1829 (91.6)	**0.0008**
	Normal	135 (10.1)	25 (5.7)	0 (0.0)	8 (4.4)	168 (8.4)	
	Unknown	727	201	19	85	1032	
Abnormal TP53 IHC patterns [e]	**Abnormal OE**	830 (69.1)	296 (71.7)	31 (73.8)	120 (69.8)	1277 (69.8)	0.8706
	Abnormal CA	311 (25.9)	98 (23.7)	10 (23.8)	46 (26.7)	465 (25.4)	
	Abnormal CY	61 (5.1)	19 (4.6)	1 (2.4)	6 (3.5)	87 (4.8)	
***BRCA1/2* germline pathogenic variant**	**Present**	111 (16.9)	33 (16.5)	0 (0.0)	2 (2.9)	146 (15.3)	**0.0020**
	Absent	546 (83.1)	167 (83.5)	28 (100.0)	67 (97.1)	808 (84.7)	
	Unknown	1407	439	33	196	2075	
CDKN2A	**Normal**	630 (48.2)	125 (29.7)	12 (30.8)	33 (18.6)	800 (41.2)	**<0.0001**
	Abnormal block positive	591 (45.2)	288 (68.4)	24 (61.5)	144 (81.4)	1047 (53.9)	
	Abnormal complete absence	86 (6.6)	8 (1.9)	3 (7.7)	0 (0.0)	97 (5.0)	
	Unknown	757	218	22	88	1085	
RB1	**Normal (retained)**	1153 (81.1)	402 (83.6)	44 (97.8)	187 (91.2)	1786 (83.0)	**0.0001**
	Abnormal (loss)	269 (18.9)	79 (16.4)	1 (2.2)	18 (8.8)	367 (17.0)	
	Unknown	642	158	16	60	876	
Total [c]		2064 (68.1)	639 (21.1)	61 (2.0)	265 (8.8)	3029 (100.0)	

[a] CCNE1 profile amplification is defined by chromogenic *in situ* hybridization and, and protein expression is defined by immunohistochemistry.
[b] The proportion of cases with a particular molecular marker status is given as a percentage of the total patients examined in each CCNE1 profile. This does not include cases where mutational status was unknown.
[c] The proportion of cases in each CCNE1 profile is given as a percentage of the total patients examined. This does not include cases where mutational status was unknown.
[d] Chi-squared testing was used to calculate p-values. Statistically significant values, where $p<0.05$, have been shown in bold. This does not include cases where mutational status was unknown.
[e] TP53 type of abnormal mutation-type immunohistochemical pattern: OE – overexpression, CA - complete absence, CY - cytoplasmic.

Cyclin E1 (CCNE1); Negative for CCNE1 high-level amplification (CCNE1^{nonamp}); CCNE1 high-level amplification (CCNE1^{amp}); Negative for CCNE1 protein overexpression by immunohistochemistry (CCNE1^{lo}); CCNE1 protein overexpression by immunohistochemistry (CCNE1^{hi}); Immunohistochemistry (IHC); Overexpression (OE); Complete absence (CA); Cytoplasmic (CY)