

# **Variant-to-Gene Mapping and Functional Characterization of Novel Osteoblast Proteins**

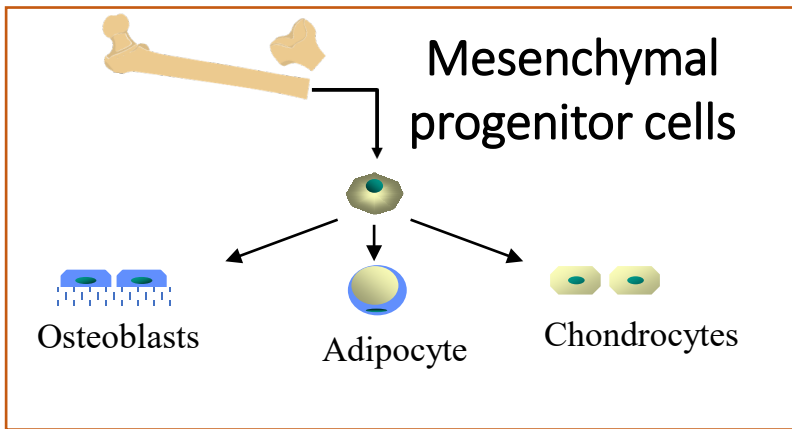
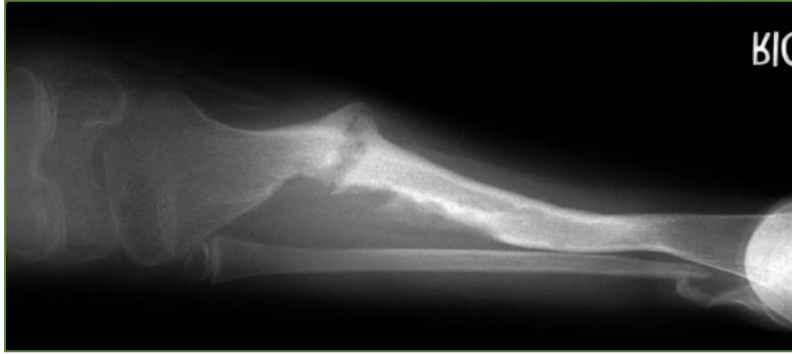
**Yadav Wagley**

**Research Investigator**

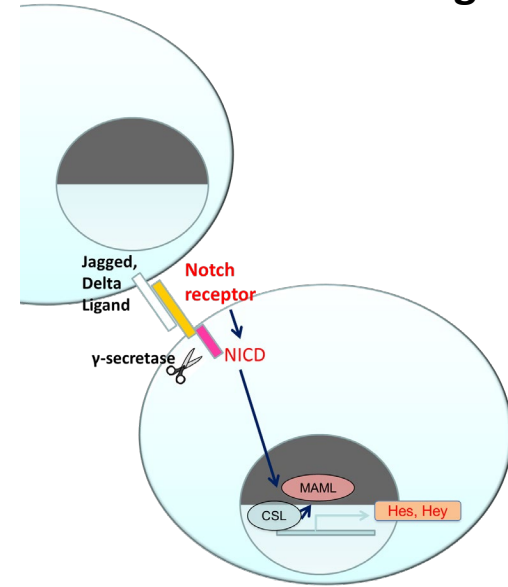
**Orthopaedic Research Laboratories**

**(Hankenson Laboratory)**

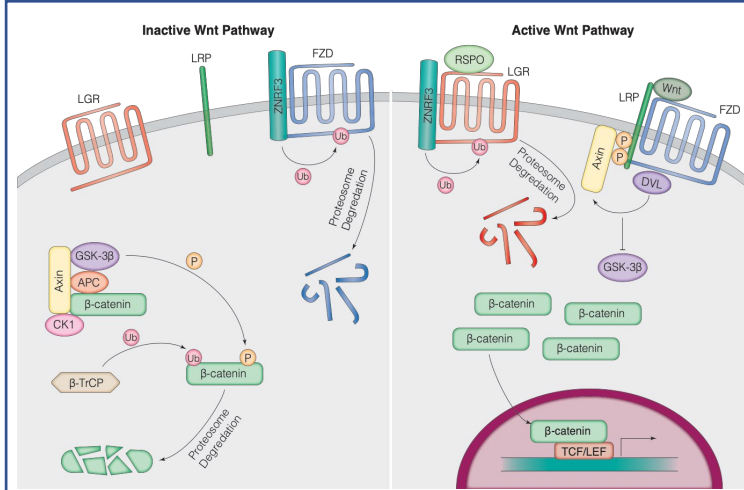
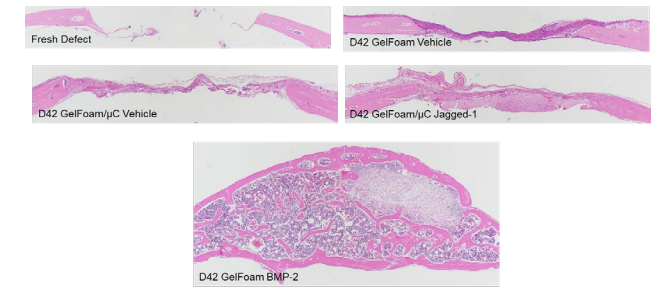
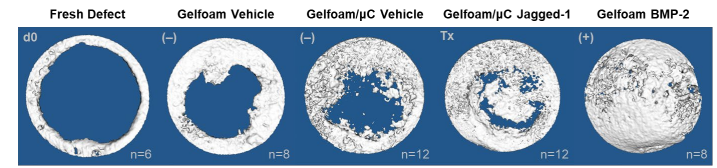
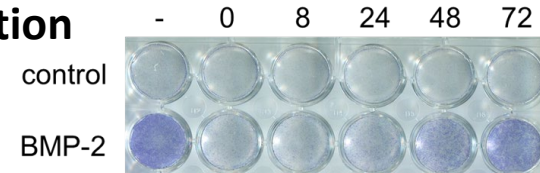
# Hankenson Laboratory— Ortho Res Labs (ORL) – Dept. Ortho. Surgery – Mesenchymal progenitor biology: Exploring extracellular regulators of bone formation and regeneration



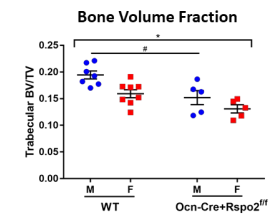
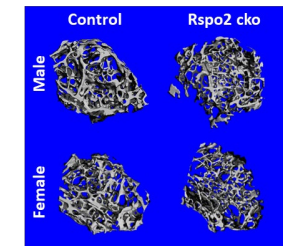
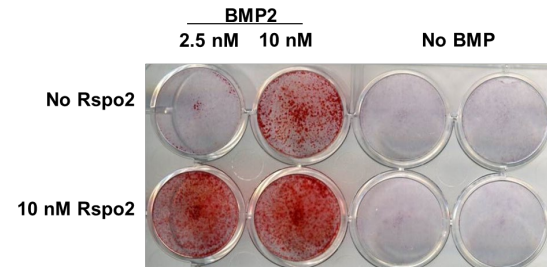
## Notch ligand Jagged-1 regulates BMP2 induced osteoblastogenesis and enhances bone regeneration



Notch signaling inhibitor (DAPT (1 μM) administered after BMP exposure (h)



## R-spondin-2 regulates Wnt signaling, osteoblastogenesis, and bone formation





**MICHIGAN MEDICINE**  
UNIVERSITY OF MICHIGAN



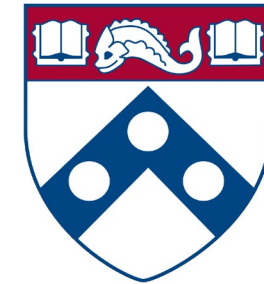
*Orthopaedic Research  
Laboratories*



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James A. Pippin  
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members of **Hankenson Lab** and **Grant Lab**



**Perelman**  
School of Medicine  
UNIVERSITY of PENNSYLVANIA



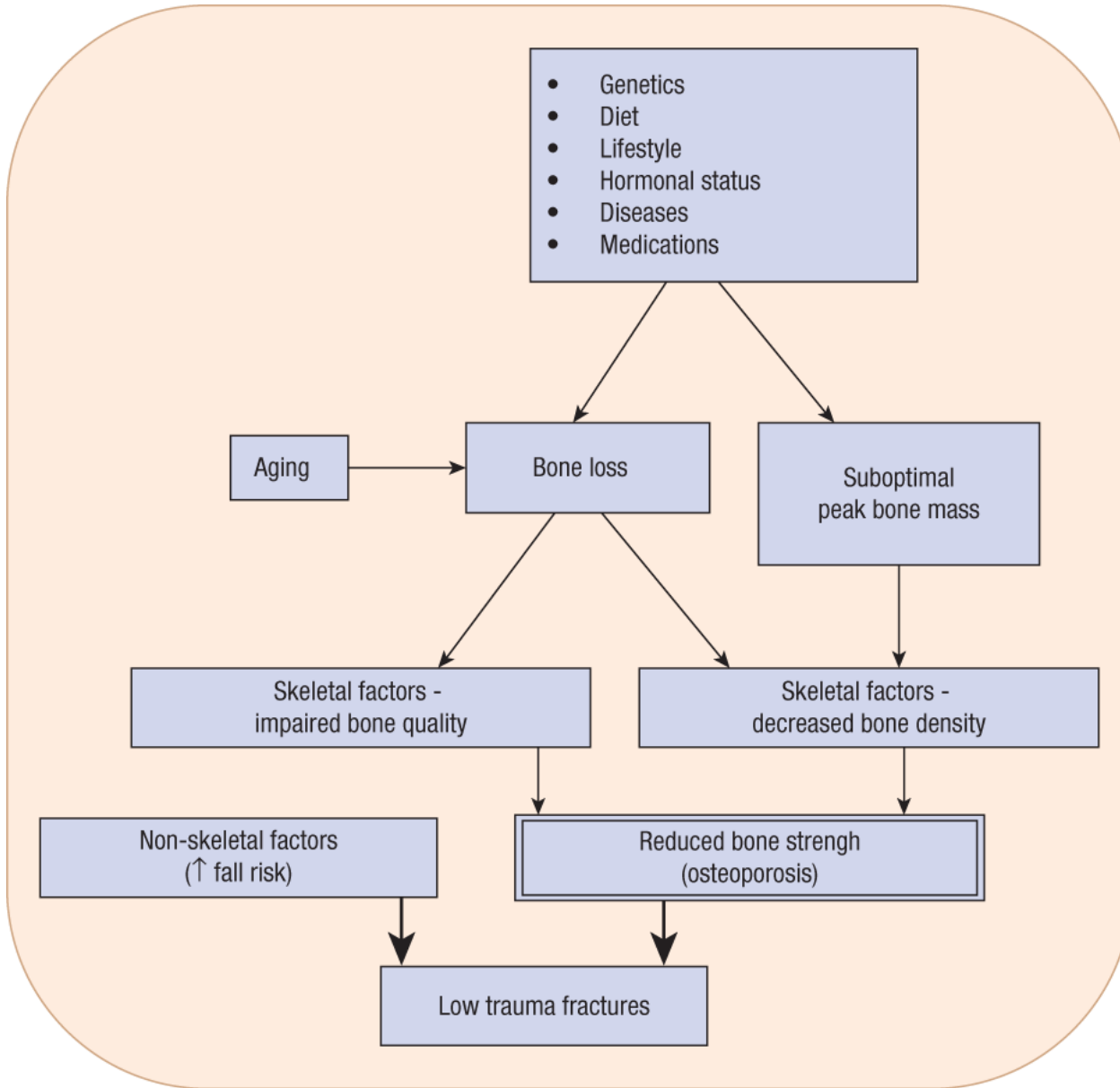
**RO1HG010067**



**RO1AR055607**

**RO1AG072705**

# Osteoporosis: a global issue



- **Over 200 million people worldwide -- 30% of postmenopausal women in USA and in Europe**
- **One in three women and one in five men over the age of 50 will suffer osteoporotic fracture**
- **Significant health problem that will rise by 240% in women and 310% in men over the next 30 years**

# History of Bone Mineral Density (BMD) Genetics

1990s

- Candidate gene studies that described associations between polymorphism in bone-relevant genes (e.g., vitamin D receptor and type I collagen) and BMD

Upto mid 2000s

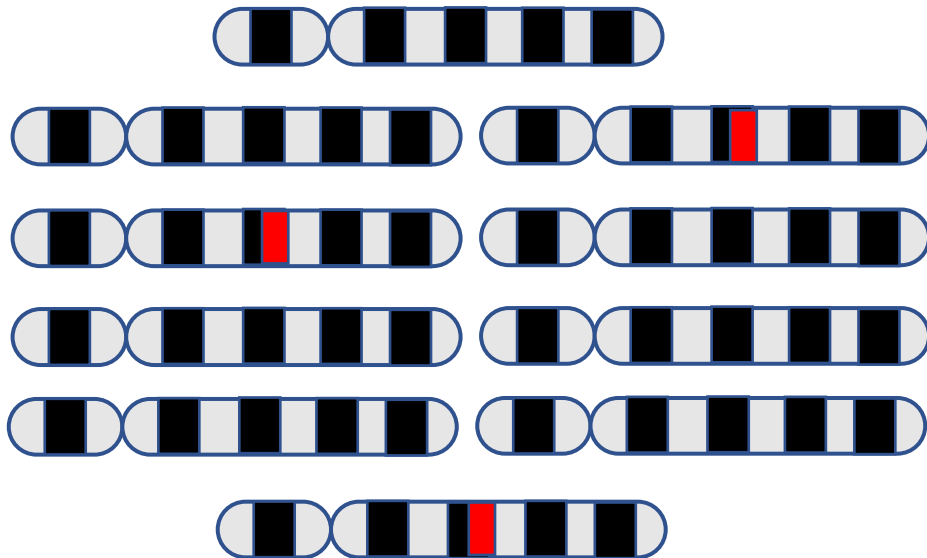
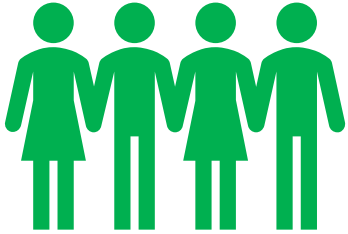
- Candidate gene investigations and linkage scan in families

2007 and onwards

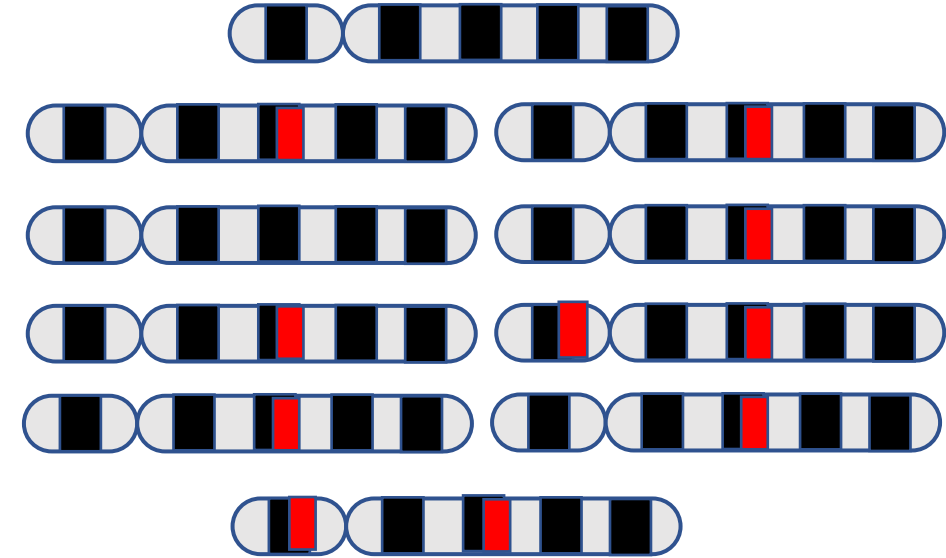
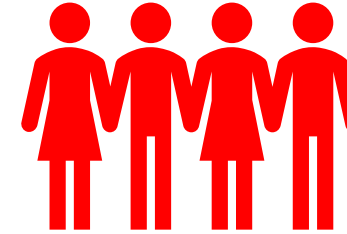
- Genome Wide Association Studies (GWAS)

**BMD GWAS tests the genotype association of millions of single nucleotide polymorphisms (SNPs) across the genome in hundreds of thousands of individuals**

**controls**



**cases**



# Large GWAS analyses for Bone Mineral Density (BMD) illustrates the increase in identified loci as a function of sample size

Study	Phenotype	Sample size	Association count
Morris et al. (2018)	Estimated heel BMD	426824	1103 independent associations (518 loci <sup>a</sup> , 301 novel)
Kemp et al. (2017)	Estimated heel BMD	142487	307 independent associations (203 loci, 153 novel)
Estrada et al. (2012)	Lumbar spine and femoral neck BMD	83894 (32961 discovery, 50933 replication)	64 independent associations (56 loci, 32 novel)
Rivadeneira et al. (2009)	Lumbar spine and femoral neck BMD	19195	20 independent associations (20 loci, 13 novel)

<sup>a</sup>Note that the most recent discovery of 518 loci encompassed nearly all of the previously discovered loci identified in prior studies

Several of the variants identified in individuals with lower BMD are also linked to increased risk of bone fractures

## More than GWAS are required to identify causal genes (or gene-networks)

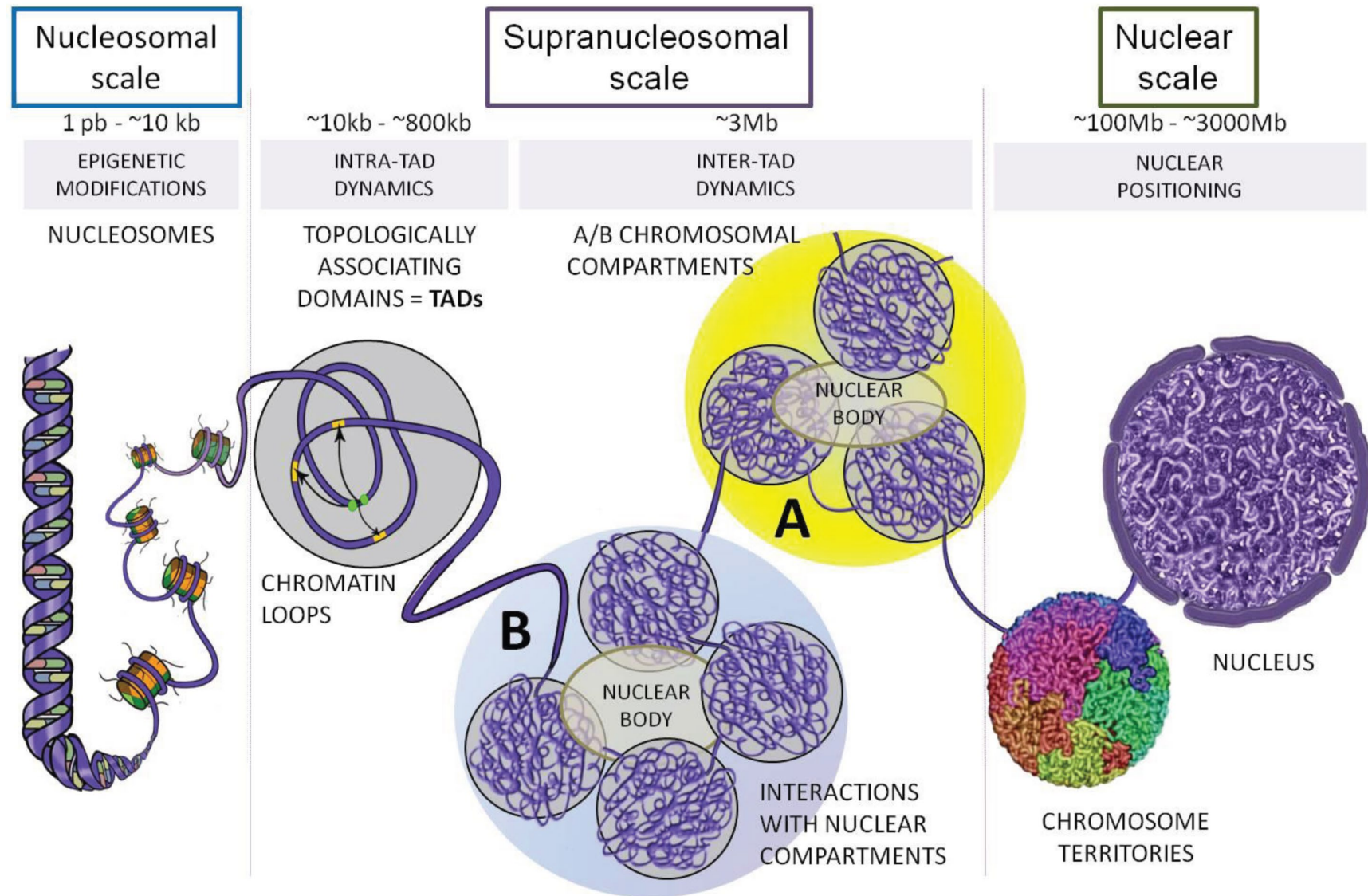
- **GWAS are hypothesis free methods to identify associations between genetic regions (loci) and phenotype**
- **Identifies SNPs across the genome and suggest particular variations that affect a person's response to certain drugs and influence interactions between a person's gene and the environment**
- **CAN NOT tell if the variant is affecting culprit gene or a gene at more distant loci**



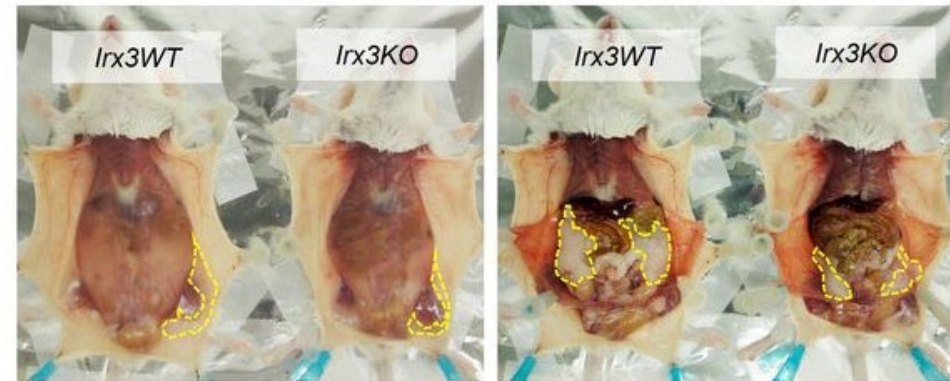
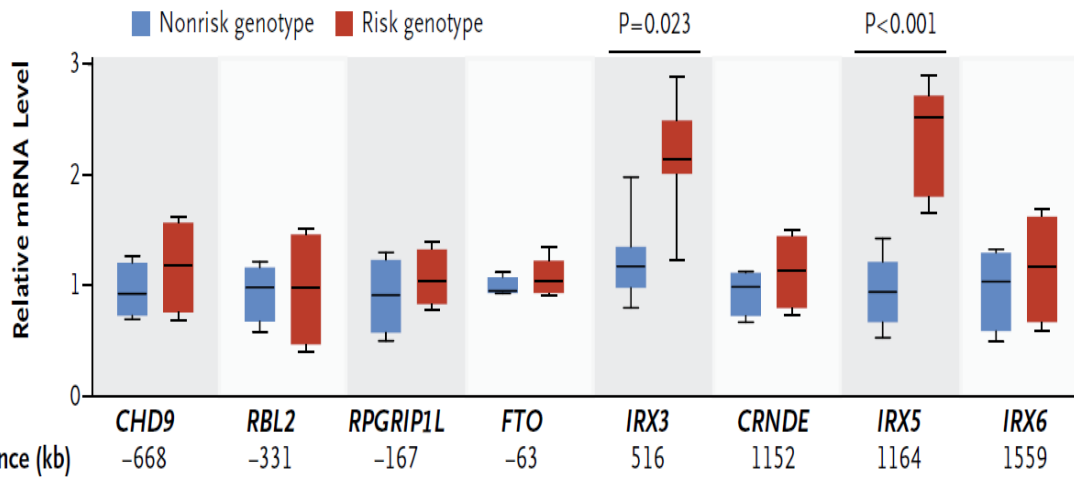
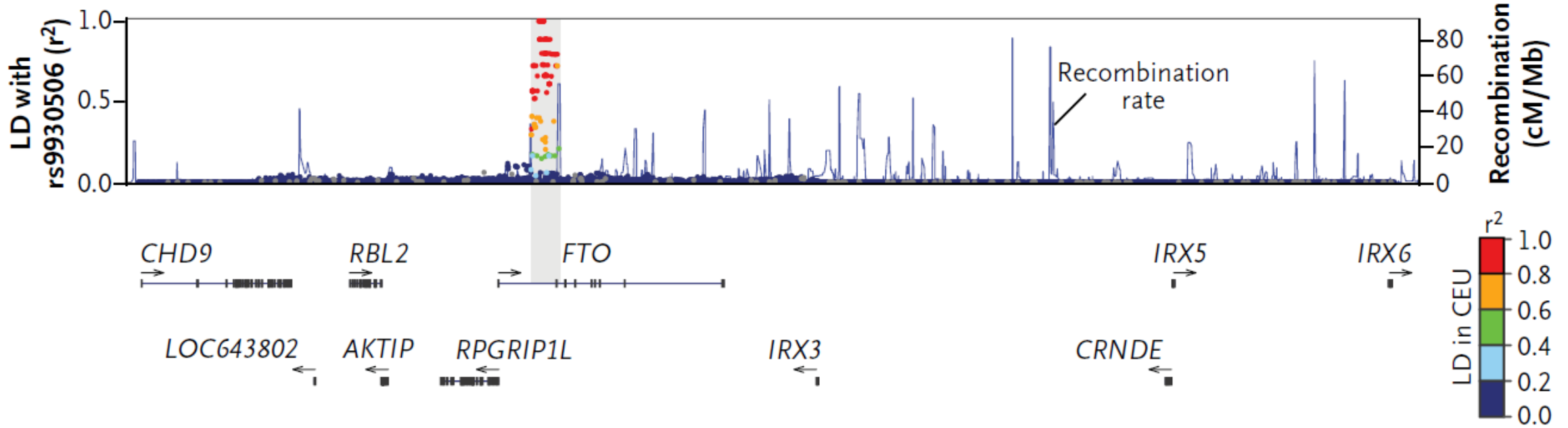
# Variant to gene mapping?

Although GWAS provides valuable insight on affected loci, the causal variants are often embedded in non-coding regions of the genome which contains important regulatory elements such as enhancers and silencers.

Many such regulatory elements do not control the nearest gene and can reside tens or hundreds of kilobases away



# SNP variant does not always affect implicated gene (Obesity model)



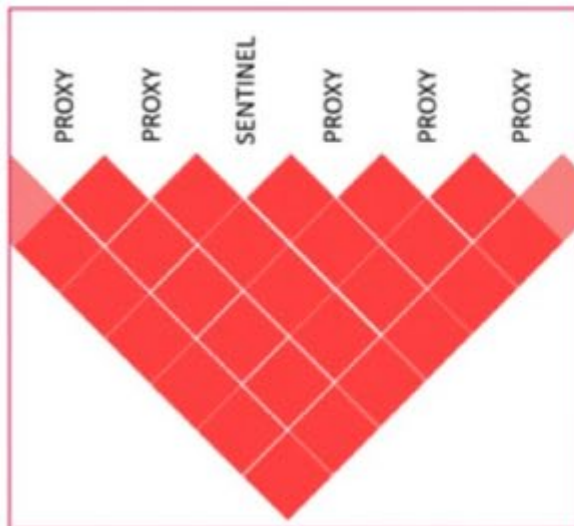
Claussnitzer, M. et al. *N Engl J Med* 373, 895-907 (2015)

Smemo, S. et al. *Nature* 507, 271-275 (2014)

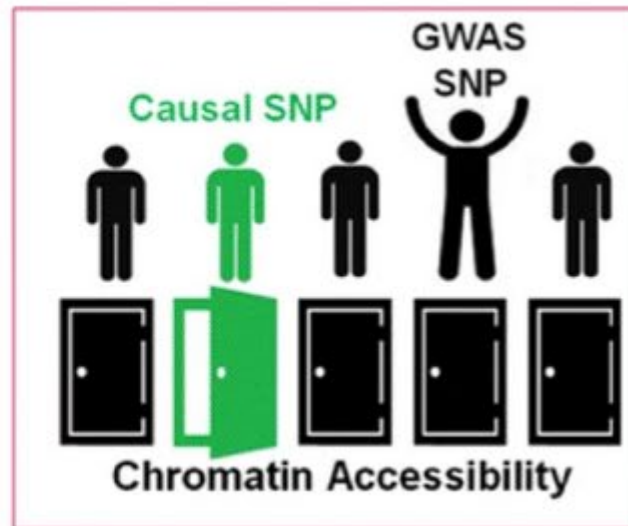
# How to identify chromatin interactions?

- By resolving the topologically associated chromatin domains (closely interacting chromatin loops) by sequencing approach in human MSC derived osteoblasts
- Are those interacting region accessible for epigenomic (or other) regulations?

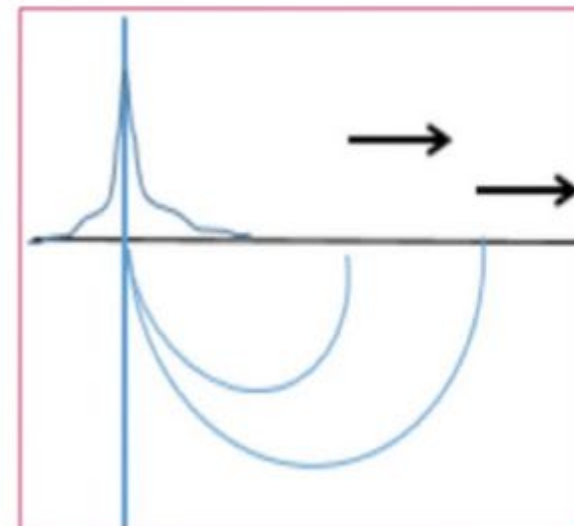
Identify all SNPs in high LD with sentinel associated variant ( $r^2=0.8$ )



Filter by residing in open chromatin as assessed by **ATAC-seq** in hMSC-derived osteoblasts

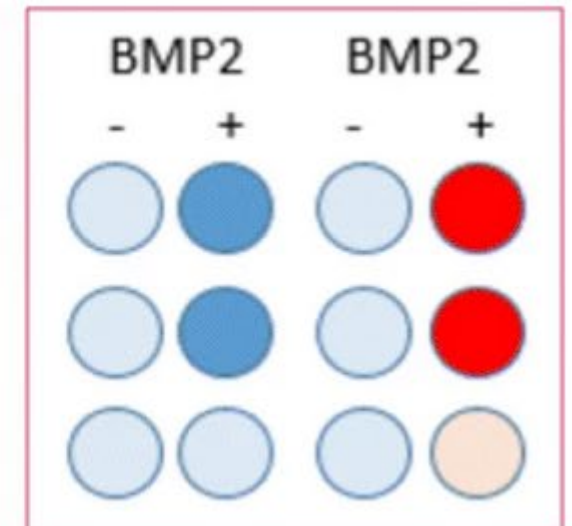


Open chromatin variants are subsequently filtered by being in direct physical contact with gene promoter baits (Promoter-focused **Capture-C**)

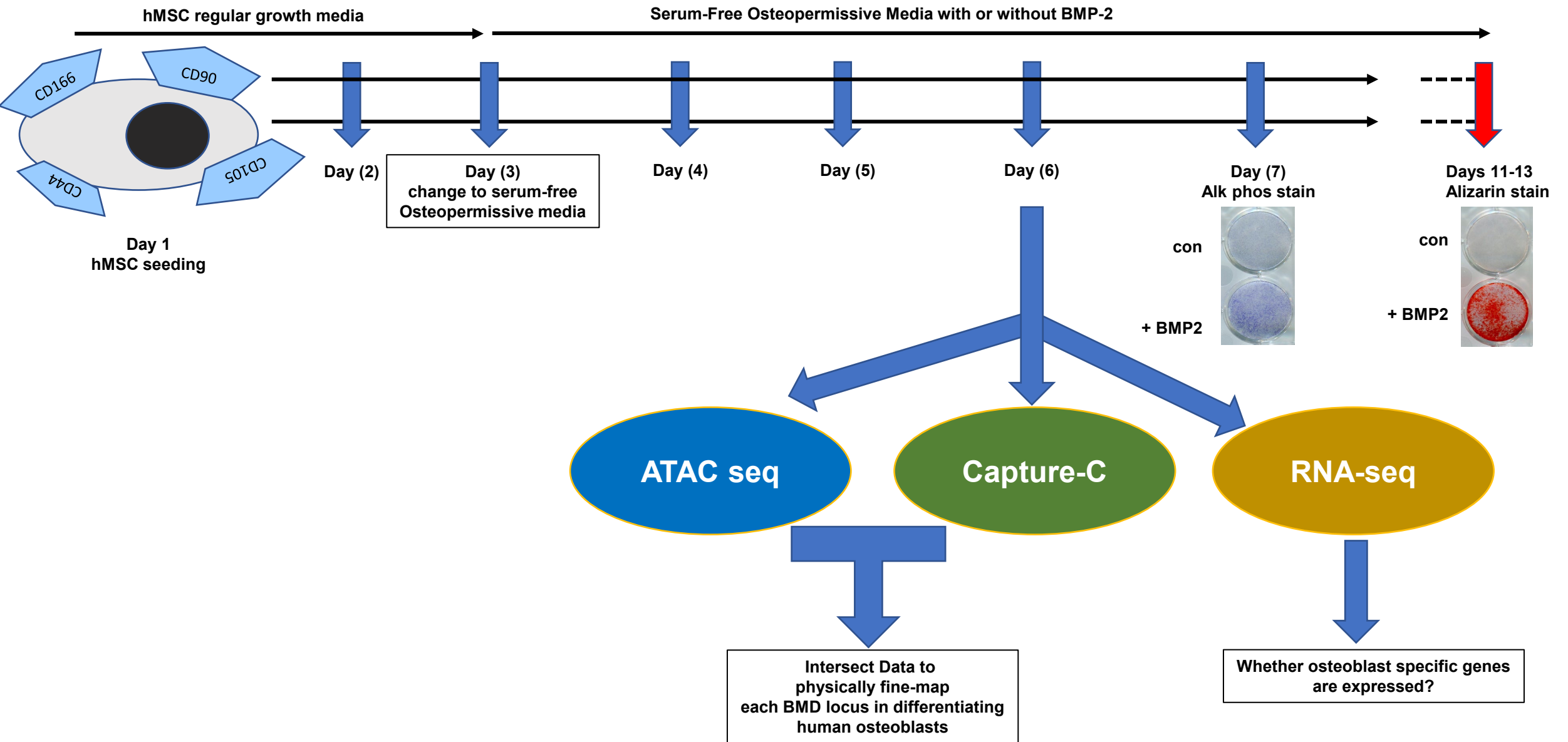


siRNA knockdown experiments are performed for a subset of these contacted genes

**(Functional validation)**



# Summary of Methods



# RNA sequencing on MSC-derived Osteoblasts

1743

2899

1156

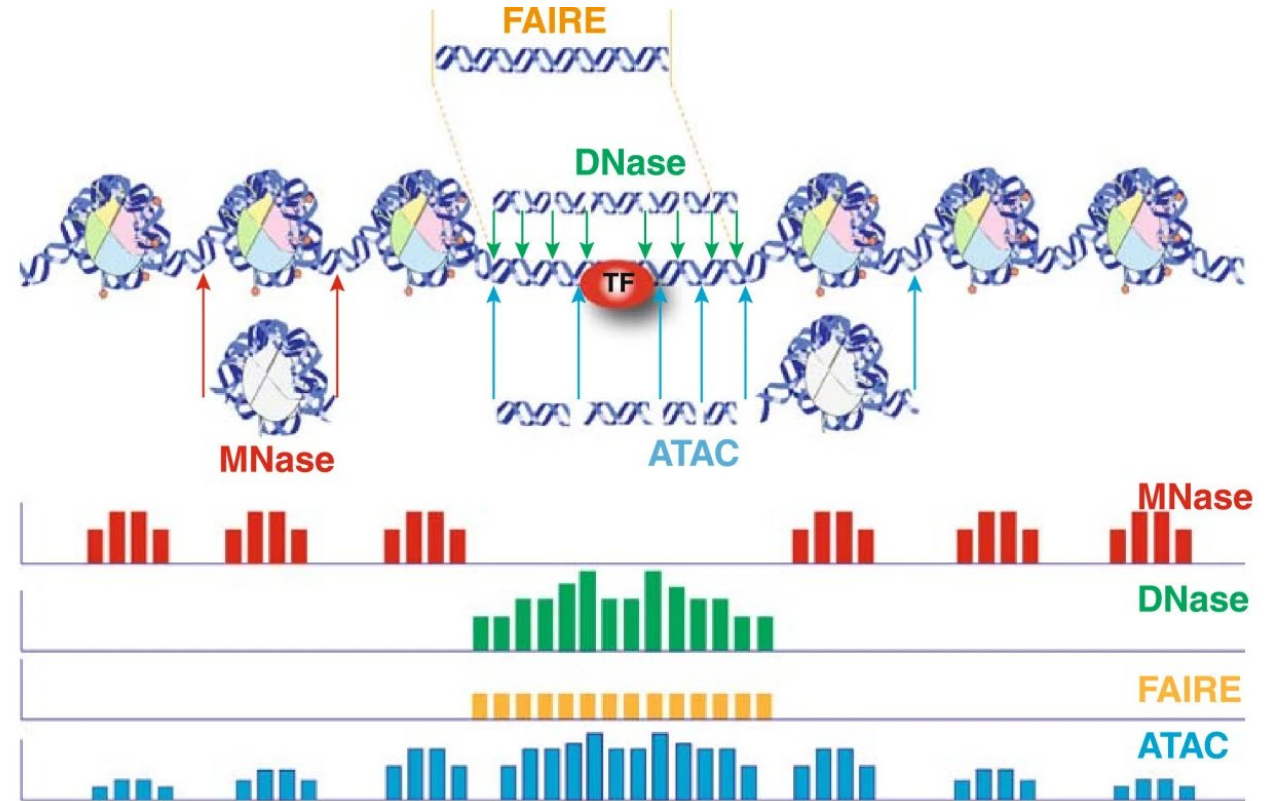
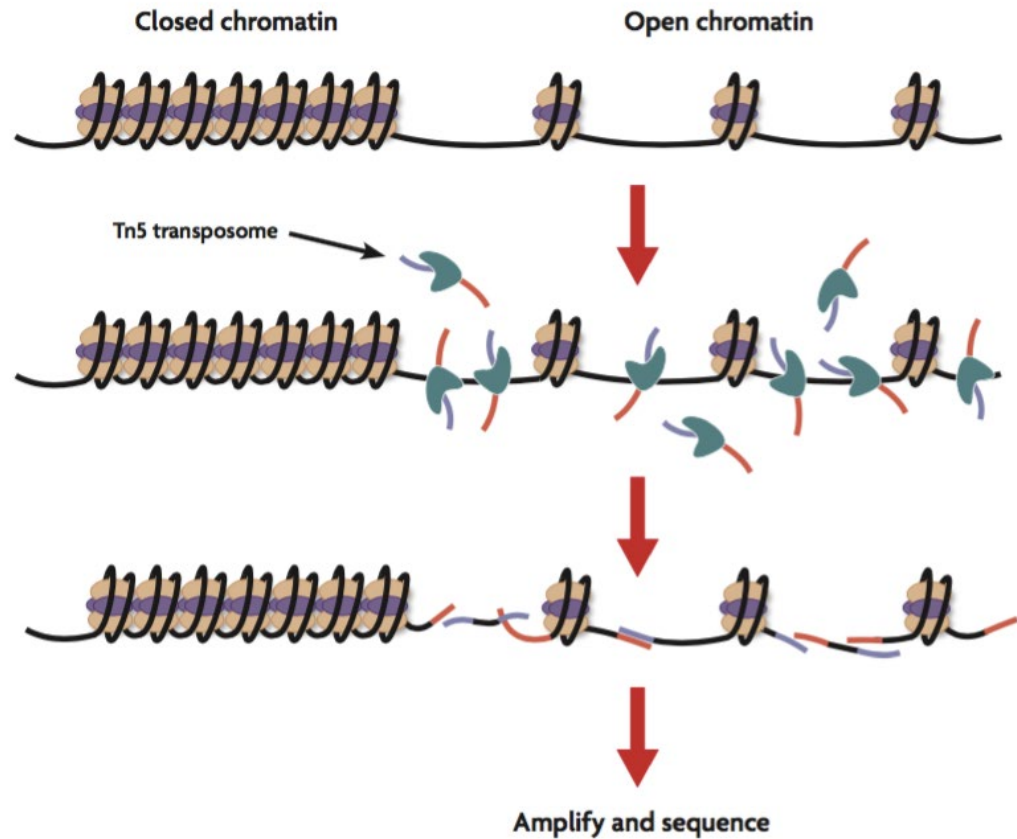
Gene Name	BMP-2 (log2 FC)	Adjusted P. Value
<i>SPP1</i>	3.0496	0.0272
<i>SOST</i>	8.7072	0.0052
<i>DKK1</i>	3.9206	0.0025
<i>SP7</i>	8.3013	0.0004
<i>DMP1</i>	5.5268	0.0229

# Selection of BMD loci

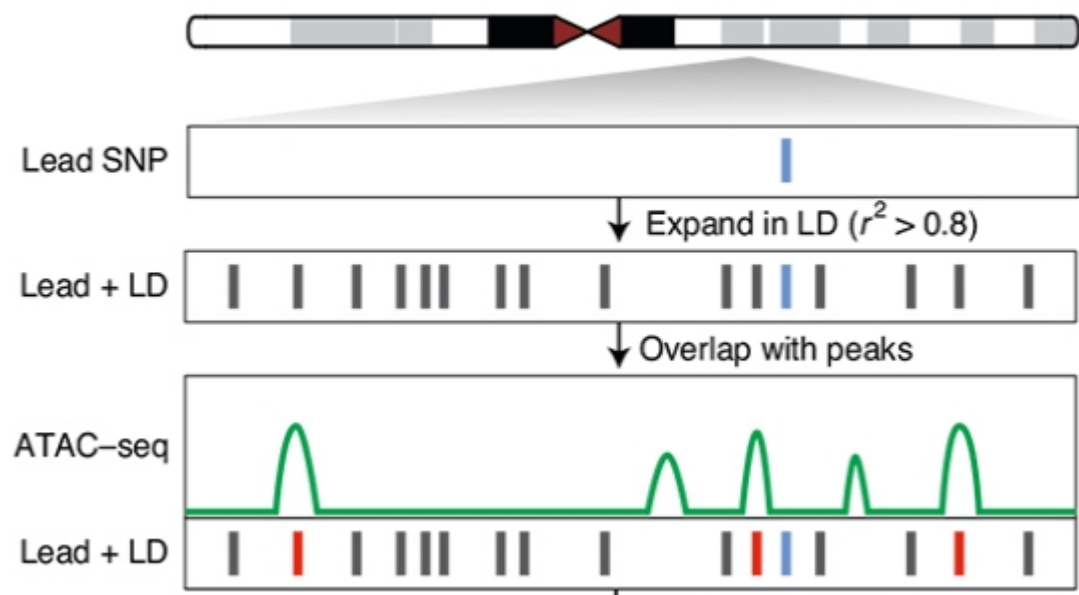
Estrada et al., Nat. Genet. 44, 491-501 (2012)	<b>Genome-wide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture</b>	56 loci (32 new) P<5X10 <sup>-8</sup>
Chesi et al., J Bone Mner Res. 32(6): 1274-1281 (2017)	<b>A Genomewide Association Study Identifies Two Sex-Specific Loci, at SPTB and IZUMO3, Influencing Pediatric Bone Mineral Density at Multiple Skeletal Sites.</b>	5 loci (4 new) 2 were sex specific P<5X10 <sup>-5</sup>
Chesi et al., Hum Mol Genet. 24(17):5053-5059 (2015)	<b>A trans-ethnic genome-wide association study identifies gender-specific loci influencing pediatric aBMD and BMC at the distal radius.</b>	2 loci P<5X10 <sup>-6</sup>
Kemp et al., PloS Genet. 10(6):e1004423 (2014)	<b>Phenotypic dissection of bone mineral density reveals skeletal site specificity and facilitates the identification of novel loci in the genetic regulation of bone mass attainment.</b>	13 loci P<5X10 <sup>-7</sup>
Zheng et al., Nature 526(7571): 112-117 (2015)	<b>Whole-genome sequencing identifies EN1 as a determinant of bone density and fracture.</b>	1 loci P<5X10 <sup>-6</sup>
Medina-Gomez et al., Nat Commun. 8(1):121 (2017)	<b>Bivariate genome-wide association meta-analysis of pediatric musculoskeletal traits reveals pleiotropic effects at the SREBF1/TOM1L2 locus.</b>	8 loci P<5X10 <sup>-8</sup>
Kemp et al., Nat Genet. 49(10):1468-1475 (2017)	<b>Identification of 153 new loci associated with heel bone mineral density and functional involvement of GPC6 in osteoporosis.</b>	203 loci P<6.6X10 <sup>-9</sup>

**107 DEXA BMD loci and 203 loci derived from heel ultrasound BMD**

# Assay for Transposon Accessible Chromatin (ATAC)



# Identifying proxy SNPs in open chromatin and in high LD with GWAS identified sentinel SNP



9 ATAC seq libraries from 4 unique donors

Analyzed using ENCODE ATAC-seq pipeline

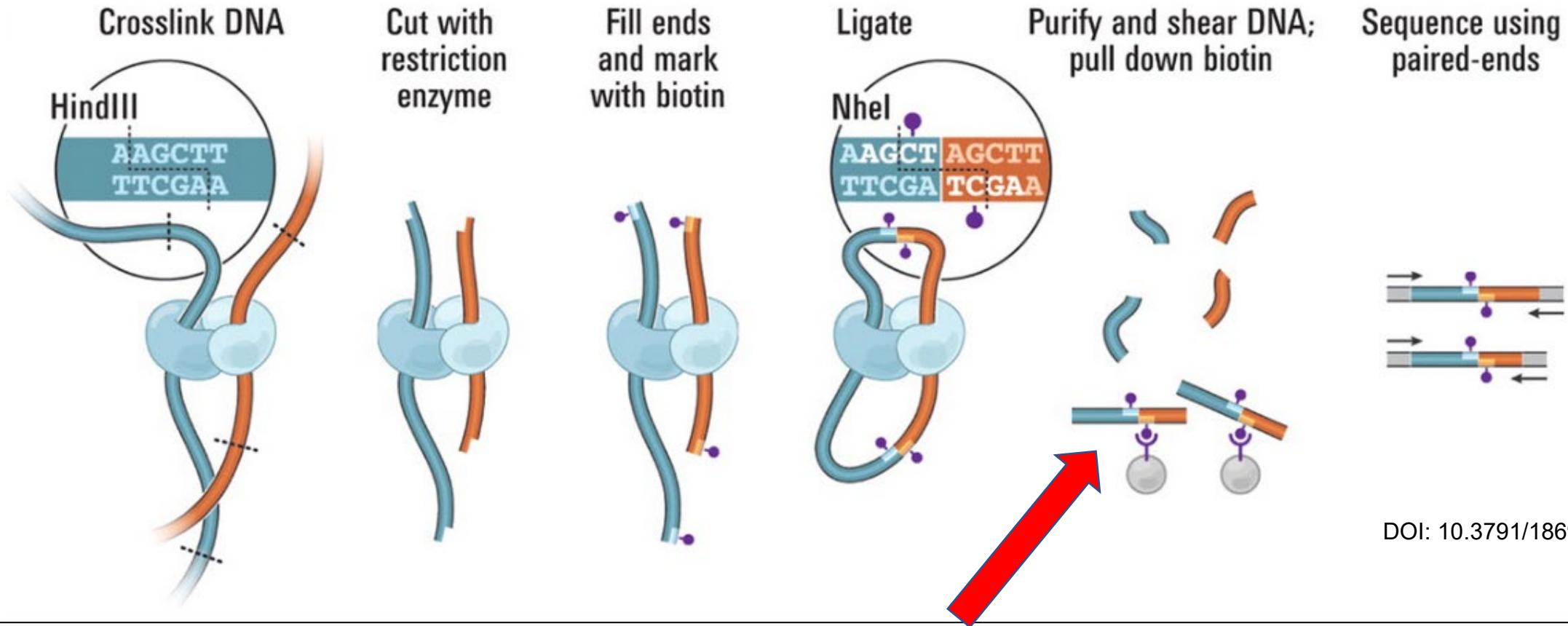
156,406 open chromatin peaks

Determine informative proxy SNP for each of the 110 independent signals at 107 DEXA BMD GWAS loci as well as 307 independent signals at 203 heel ultrasound BMD GWAS loci by overlapping the positions of the open chromatin regions (peaks) with those of the sentinel and proxy SNPs ( $r^2 > 0.8$  to sentinel SNP in Europeans)

215 informative proxy SNPs corresponding to 58 DEXA loci and 282 proxy SNPs corresponding to 112 heel loci in high LD with the sentinel SNP of the BMD loci investigated

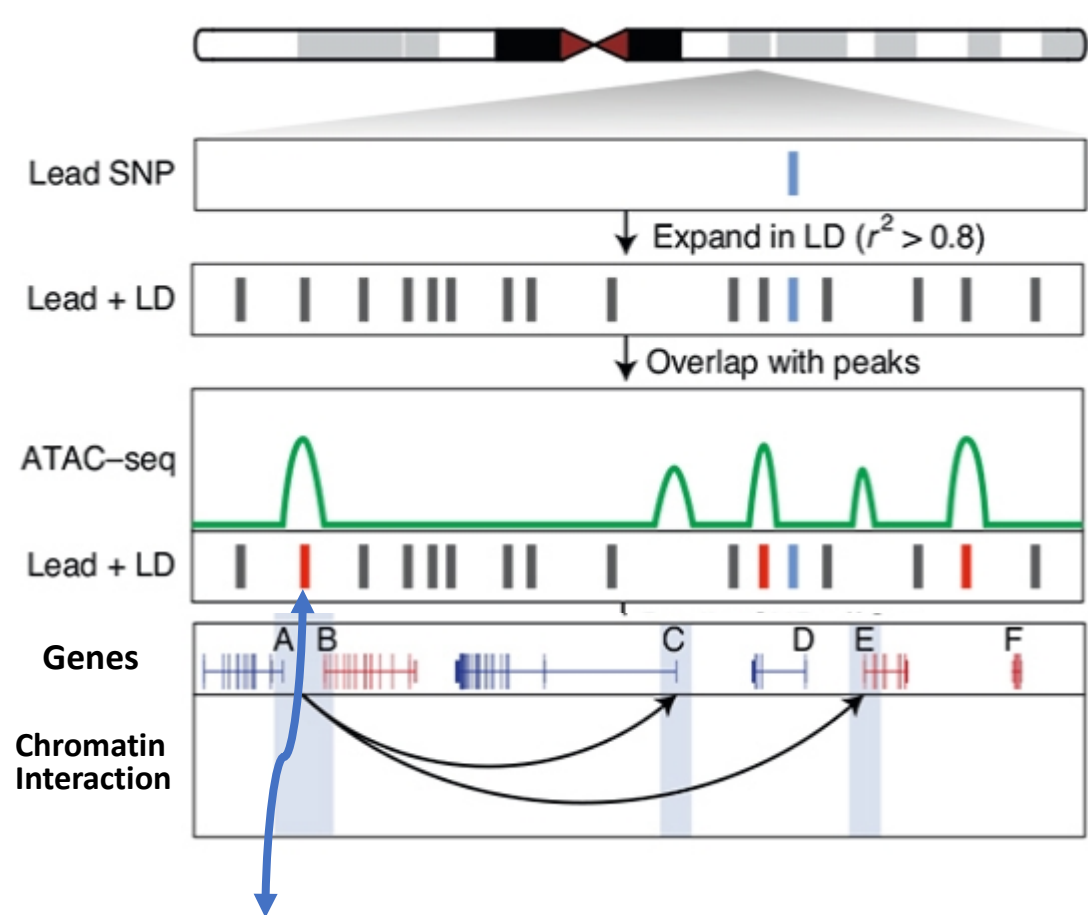


# Chromatin Conformation Capture



ligated library was hybridized with custom-designed Capture-C library consisting of both ends of DpnII restriction fragments encompassing promoters of all human coding genes and non-coding RNAs to enrich promoter interacting region

**One of the most comprehensive 3D promoter interactome analyzed !**



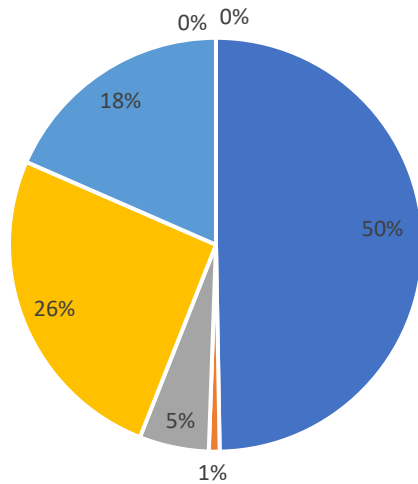
Open chromatin variants are subsequently filtered by being in direct physical contact with gene promoter baits (Promoter-focused Capture-C)

Would this proxy SNP affect gene C or gene E?

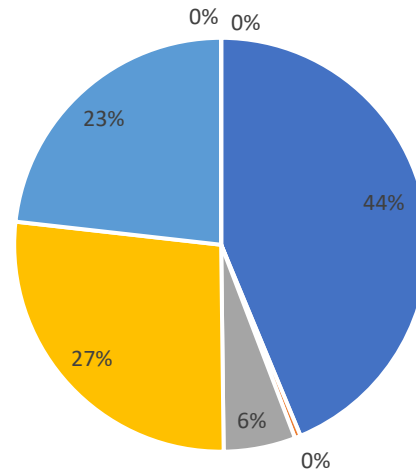
# Hi-C results for each library

~1.6 billion reads per sample; >40% valid read pairs and >75% capture efficiency

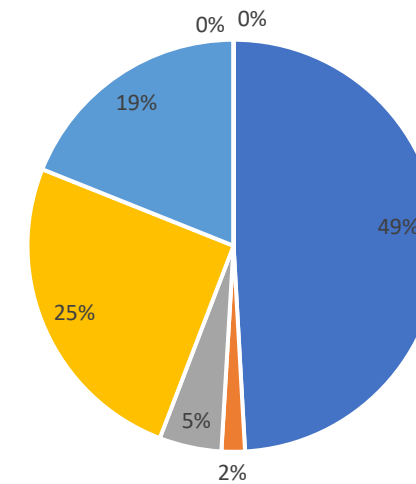
Filtering of Pairs Aligned



Filtering of Pairs Aligned



Filtering of Pairs Aligned



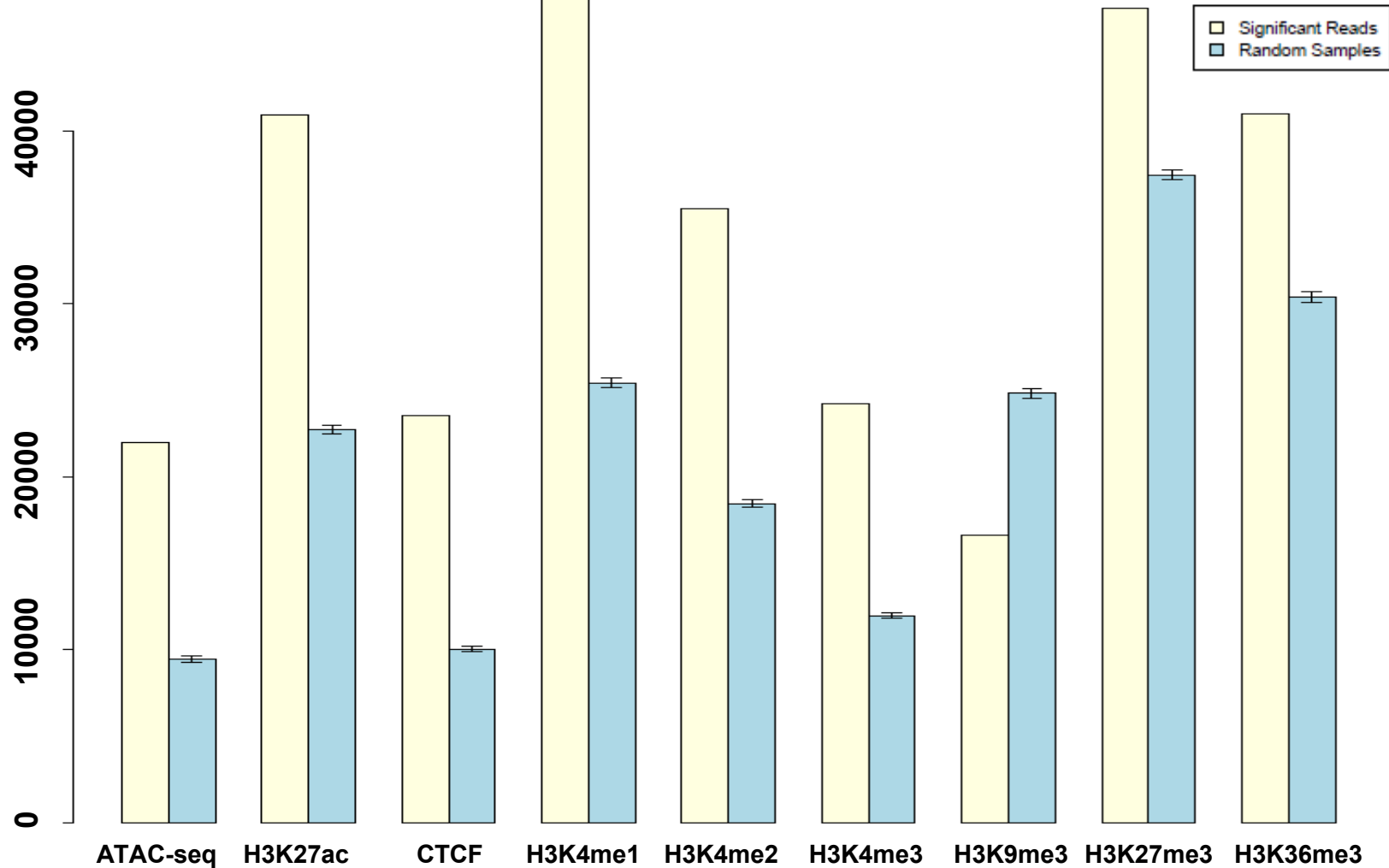
- Valid Pairs
- Same Circularised
- Same Dangling Ends
- Same Internal
- Re-ligation
- Contiguous Sequence
- Wrong Size

For further analysis: we combined 1 fragment resolution reads and 4 fragments resolution reads to achieve reads for short interactions as well as increased sensitivity

**295,422 interactions (~14% were bait to bait; median distance for cis interactions 50.5kb, trans-interactions 1%)**

**Most of the non-bait promoter-interacting regions (PIRs) had contacts with a single baited region (84%), while only 1% contacted more than 4**

# Identified Promoter interacting regions (PIRs) were significantly enriched for histone marks associated with active chromatin regions in primary human osteoblasts from the ENCODE project



# Combining the open chromatin landscape (ATAC-seq data) with chromatin interaction data (Capture-C) is an efficient approach to link biologically significant genes with their active regulatory elements

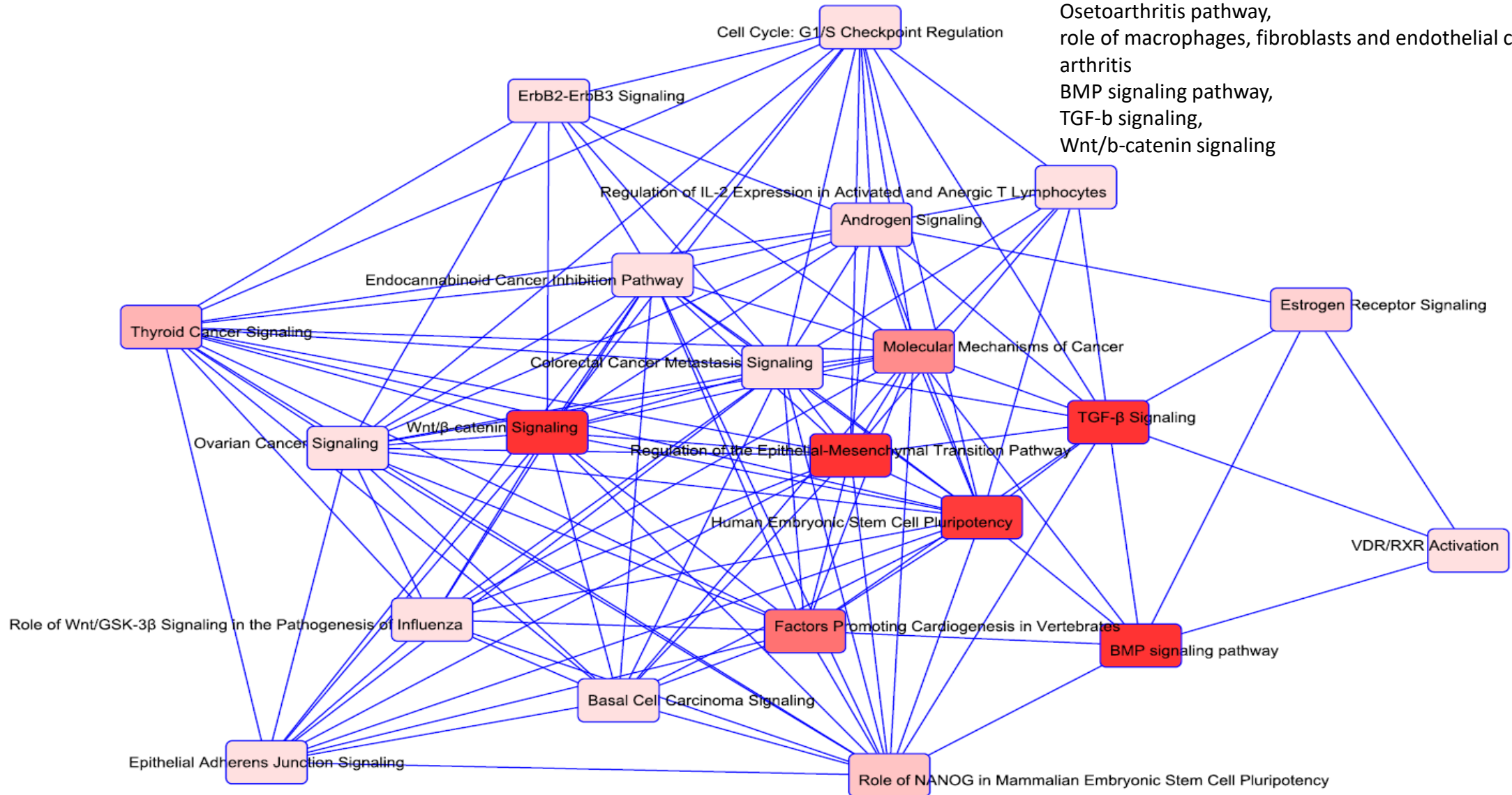
Of the BMD GWAS loci in MSC-derived osteoblasts, 46 GWAS loci revealed at least one or more BMD proxy SNPs in open chromatin (and not residing in a baited promoter region) interacting with an open gene promoter

A total of 77 open baited regions corresponding to 81 gene promoters were linked to 84 open chromatin regions harboring one or more BMD proxy SNPs through 104 distinct chromatin looping interactions

Among the implicated genes in those loci, several have a known role in osteogenesis such as *SMAD3*, *SMAD9*, *SPP1*, *WLS*, *FRZB*, *NOG* and *MIR31HG*; while several are completely novel

# Ingenuity Pathway Analysis of the implicated genes reveal pathways relevant to osteoblastic differentiation

role of osteoblasts, osteoclasts and chondrocytes in rheumatoid arthritis  
Osetoarthritis pathway,  
role of macrophages, fibroblasts and endothelial cells in rheumatoid arthritis  
BMP signaling pathway,  
TGF-b signaling,  
Wnt/b-catenin signaling



## **SNP-promoter interactions shows both close and distant genes**

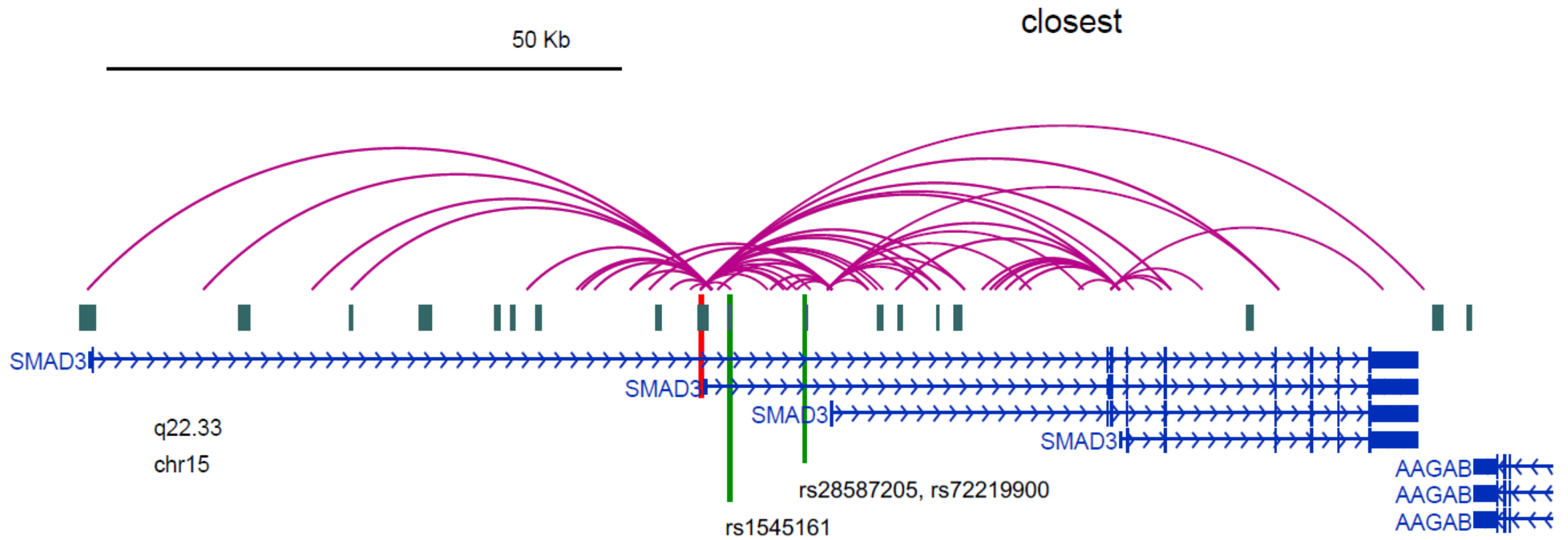
**SNP-promoter interactions (for SNPs not residing in baits) fell in to three types of observations:**

- 1. to nearest gene only (30%)**
- 2. to both nearest and more distant genes (13%) and**
- 3. only to distant genes (57%)**



# Example of SNP promoter interaction: Nearest

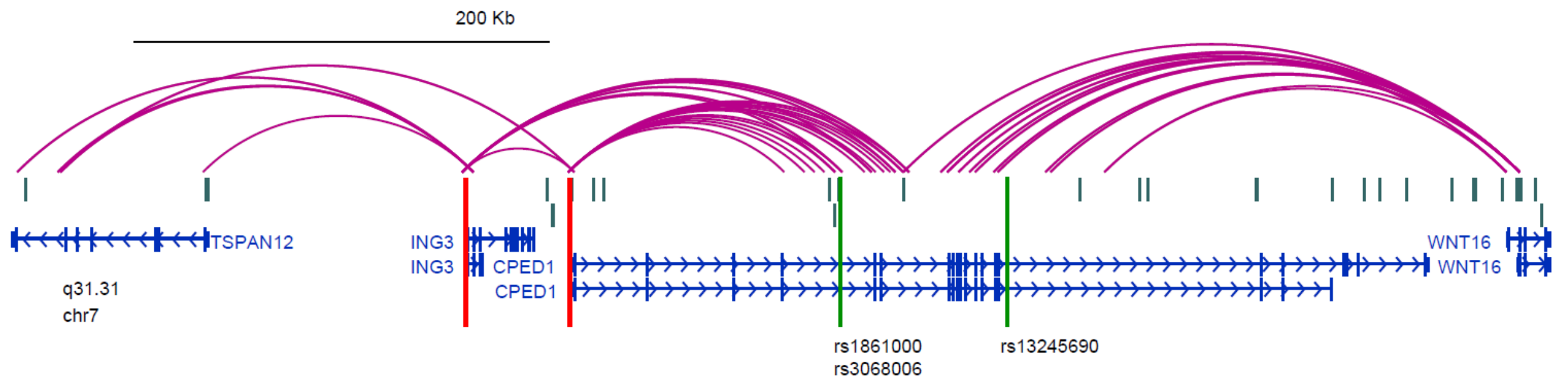
## SMAD3 at 'SMAD3' (sentinel rs1545161)



# Example of SNP promoter interaction: Nearest and Distant

ING3 and CPED1 at 'CPED1' (sentinel rs13245690)

distant and closest

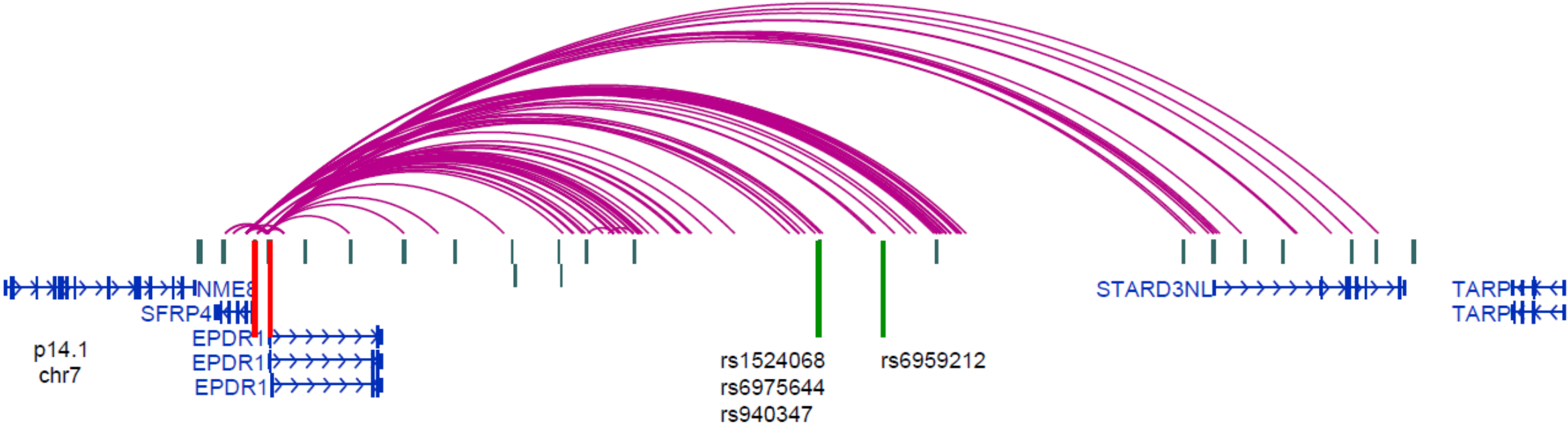


# Example of SNP promoter interaction: Distant

EPDR1 and SFRP4 at 'STARD3NL' (sentinel rs6959212)

150 Kb

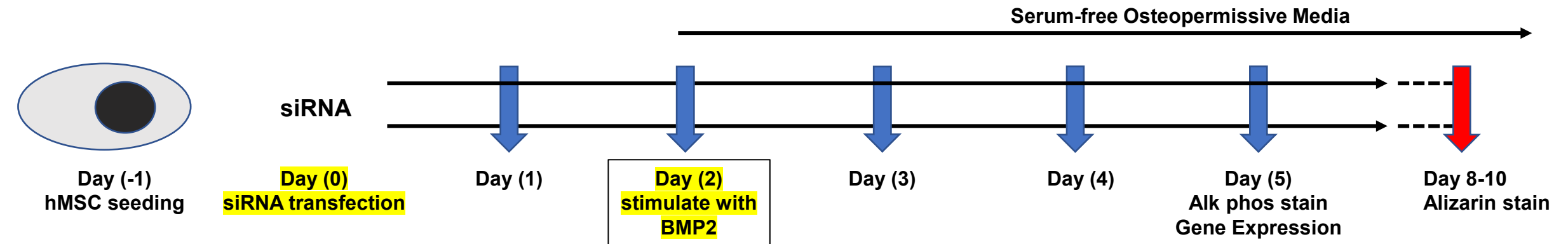
distant



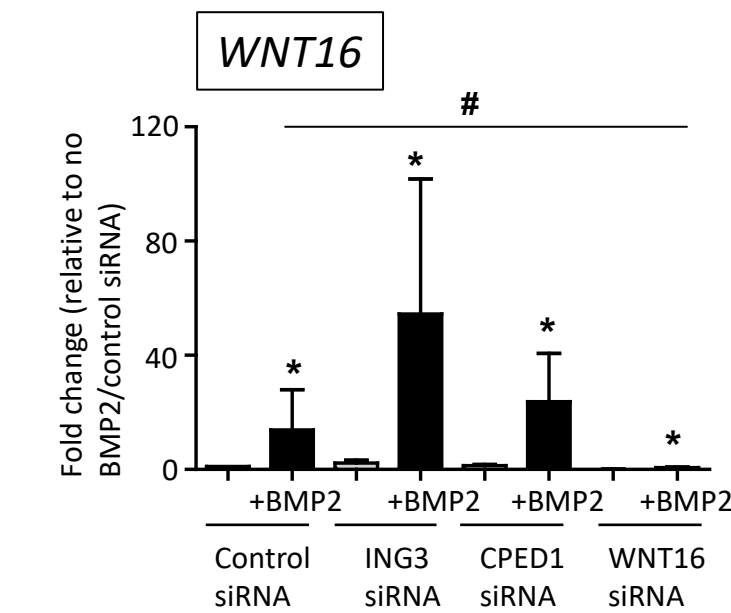
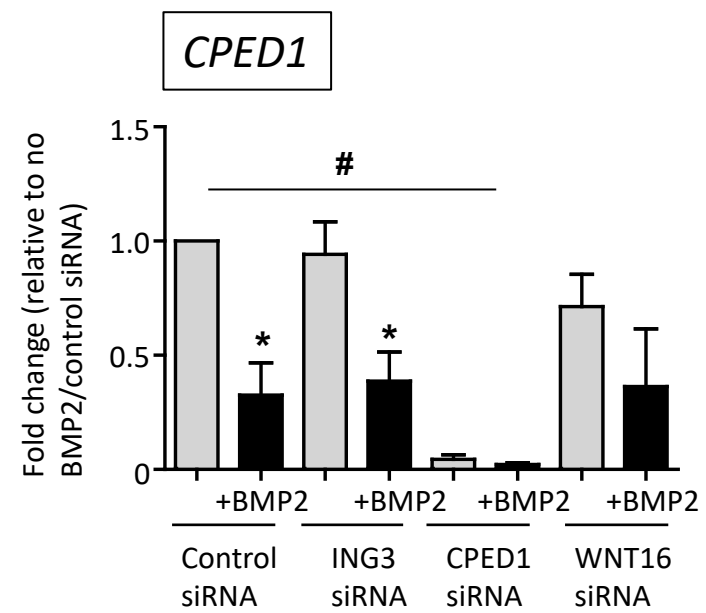
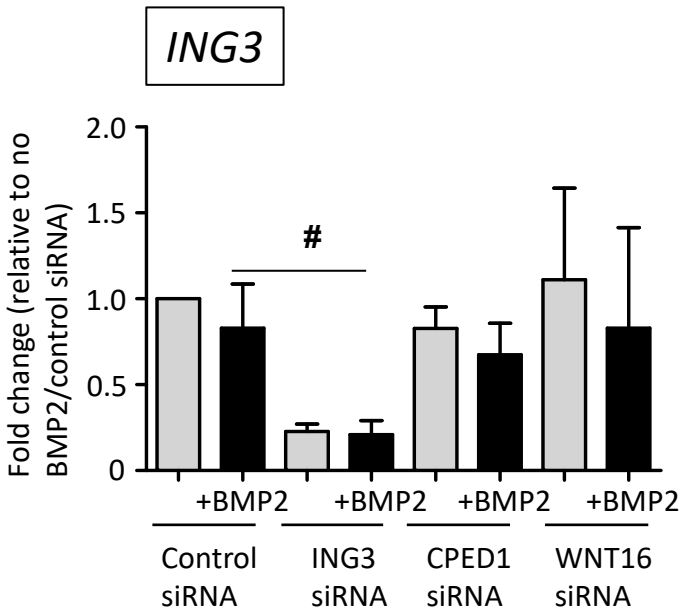
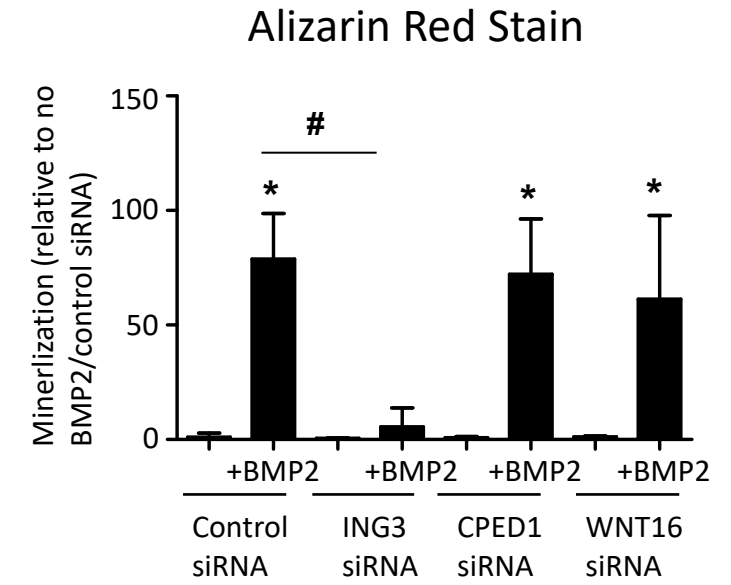
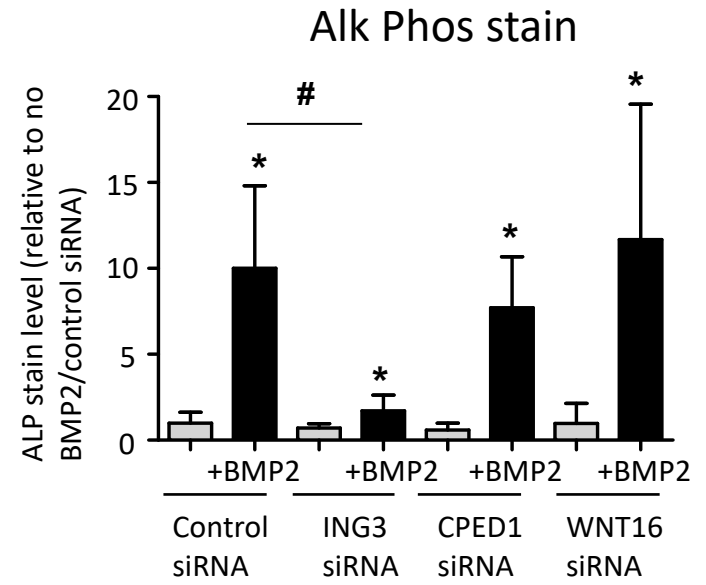
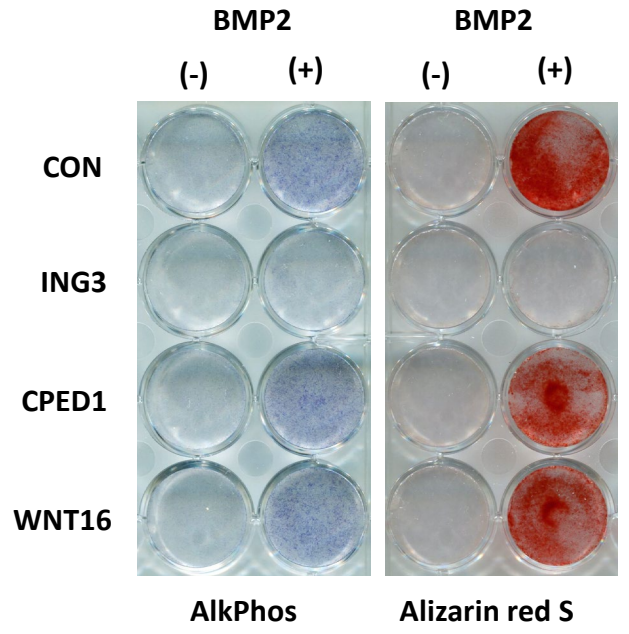
# Functional Experiments

• *CPED1* locus ➔ *ING3*, *CPED1* and *WNT16*

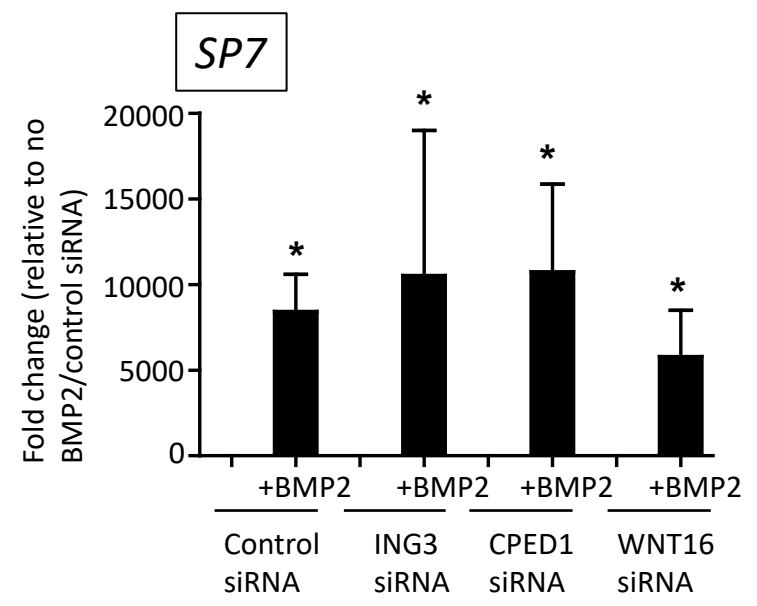
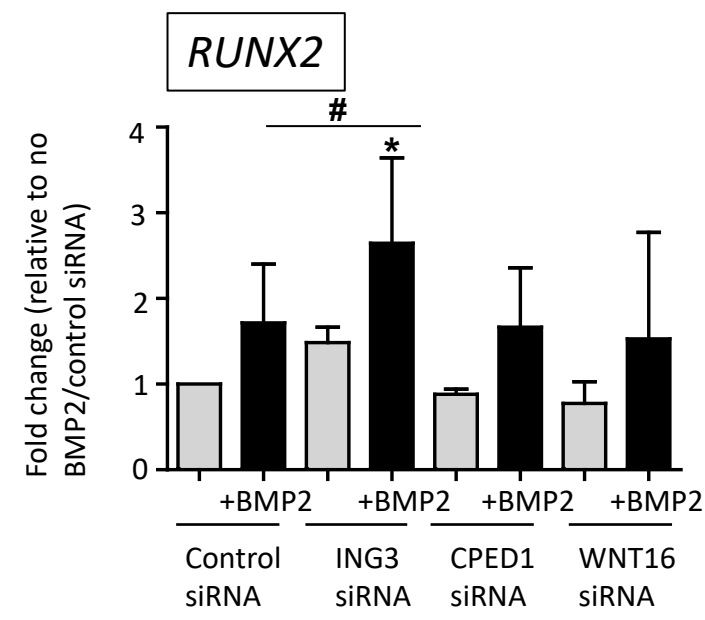
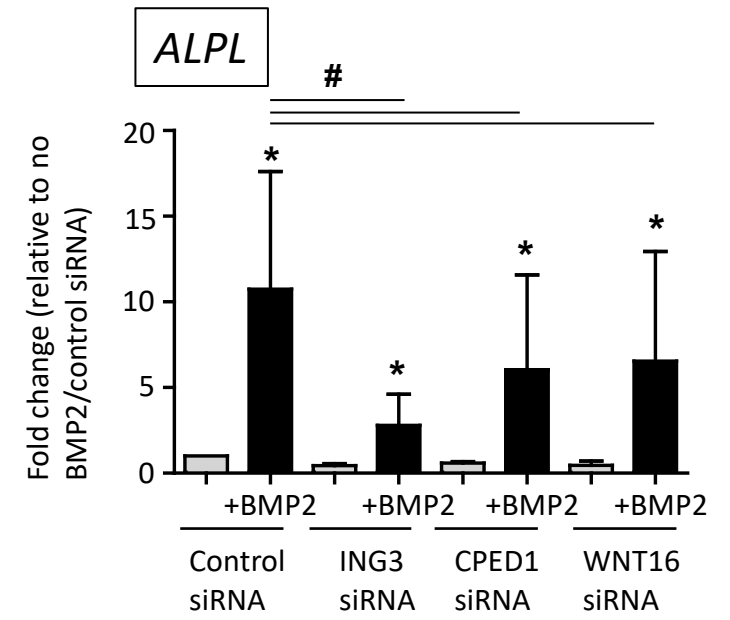
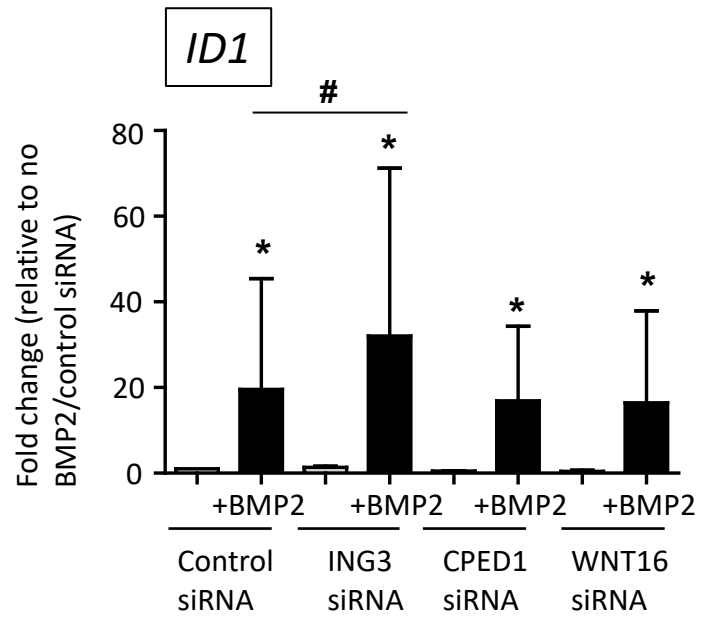
• *EPDR1* locus ➔ *SFRP4* and *EPDR1*



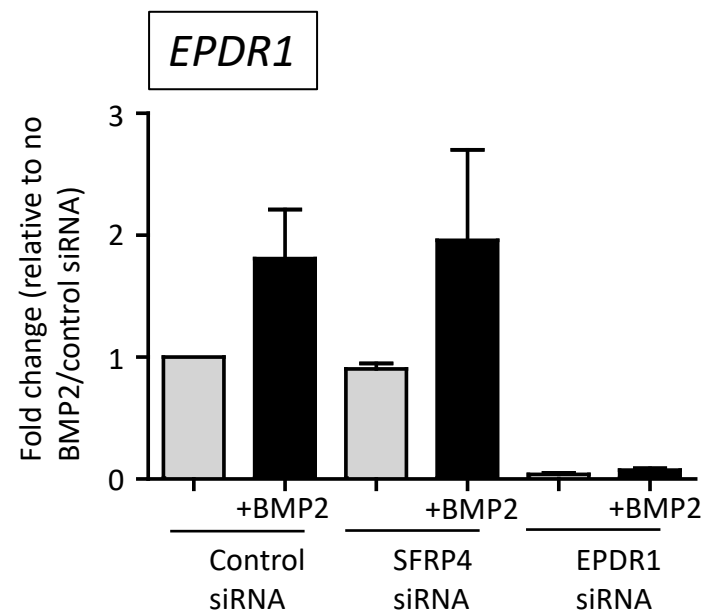
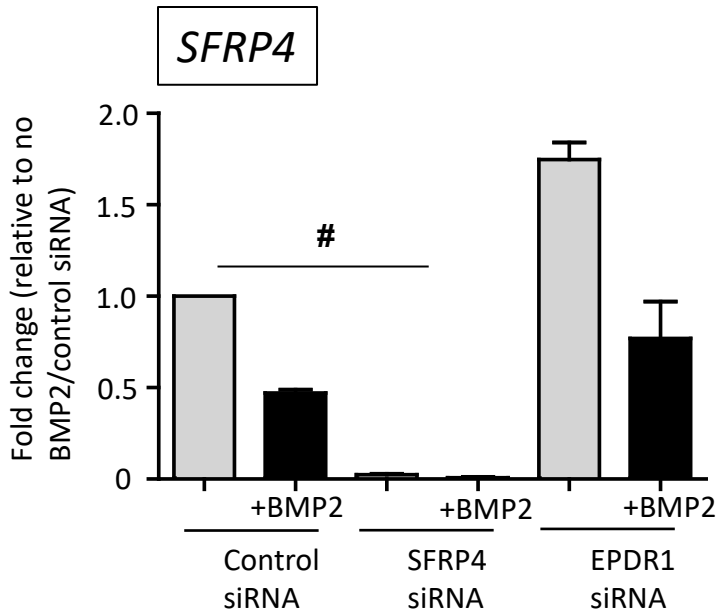
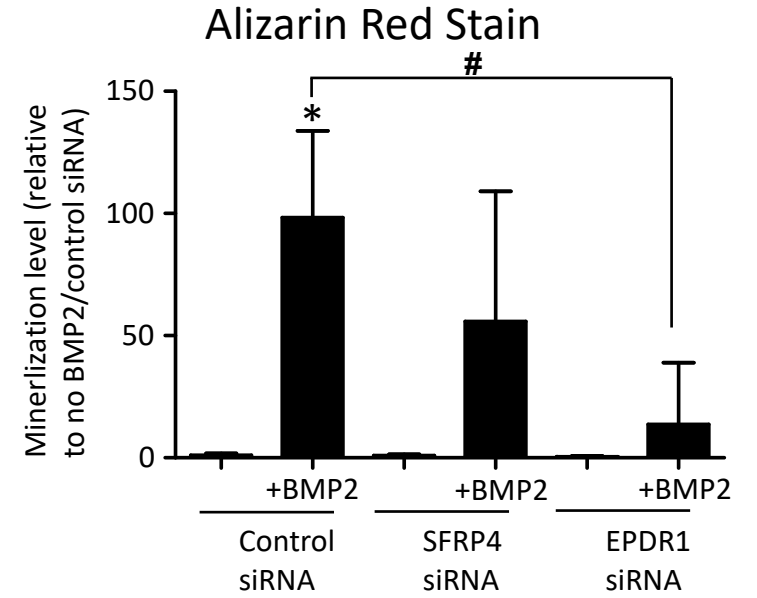
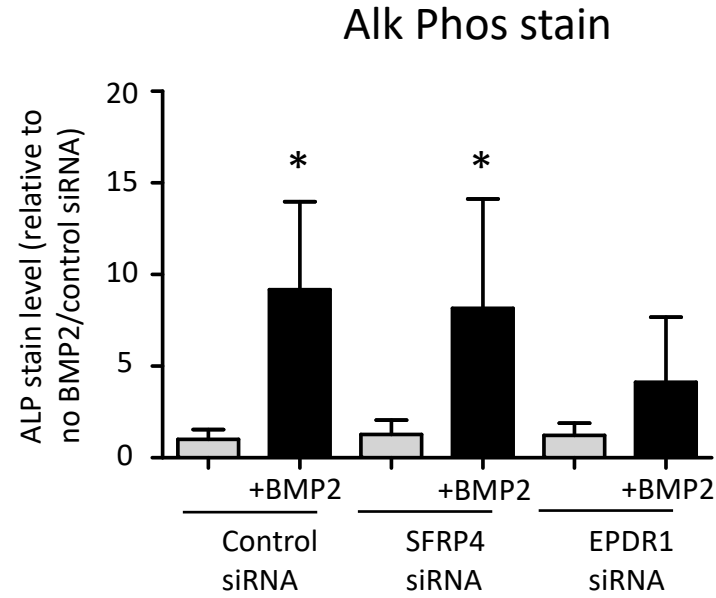
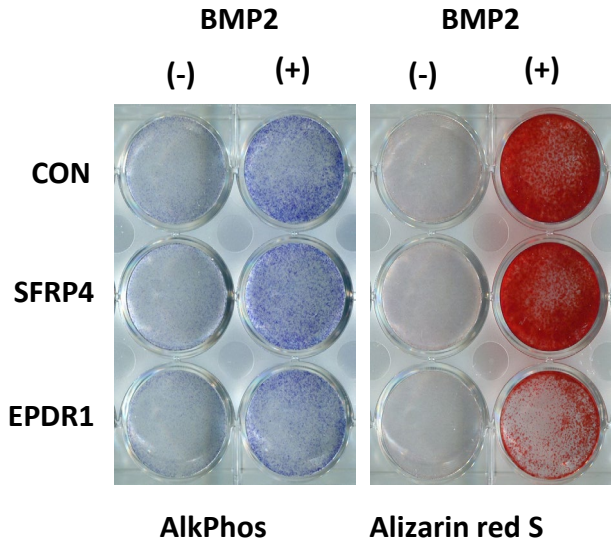
# ING3 silencing reduces human osteoblast differentiation



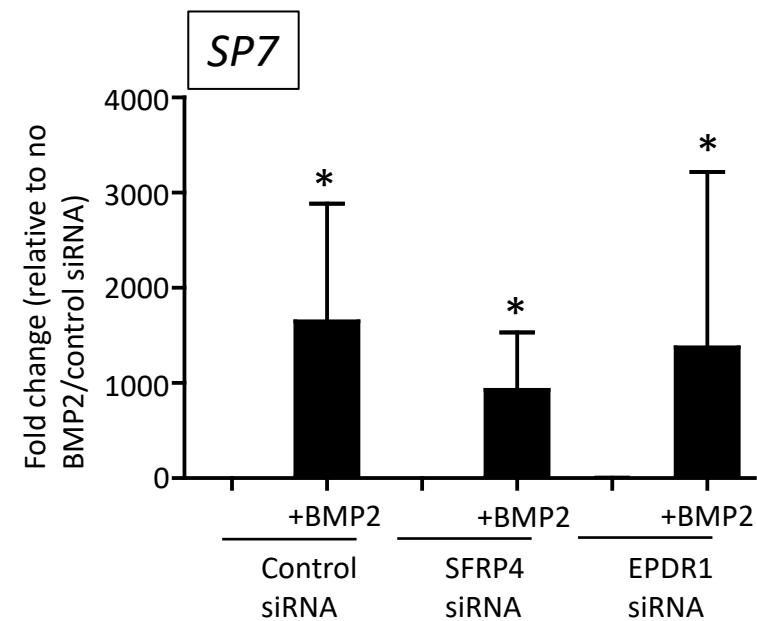
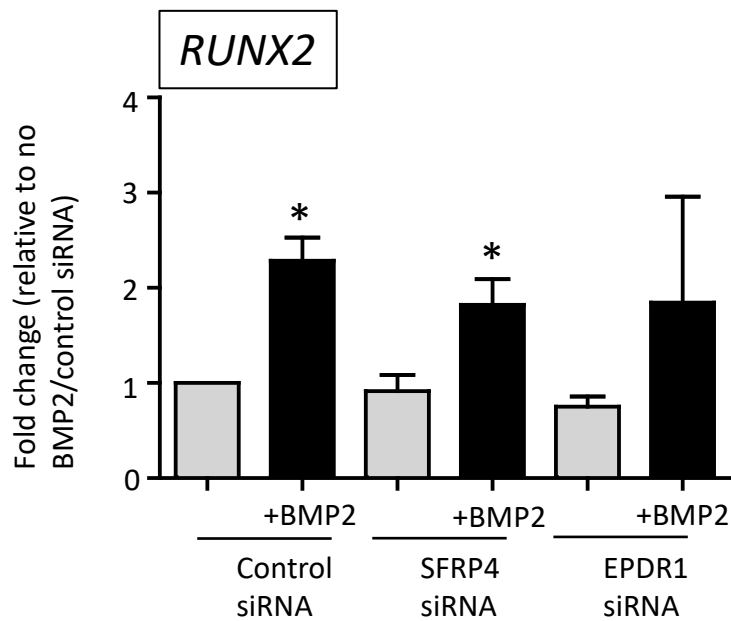
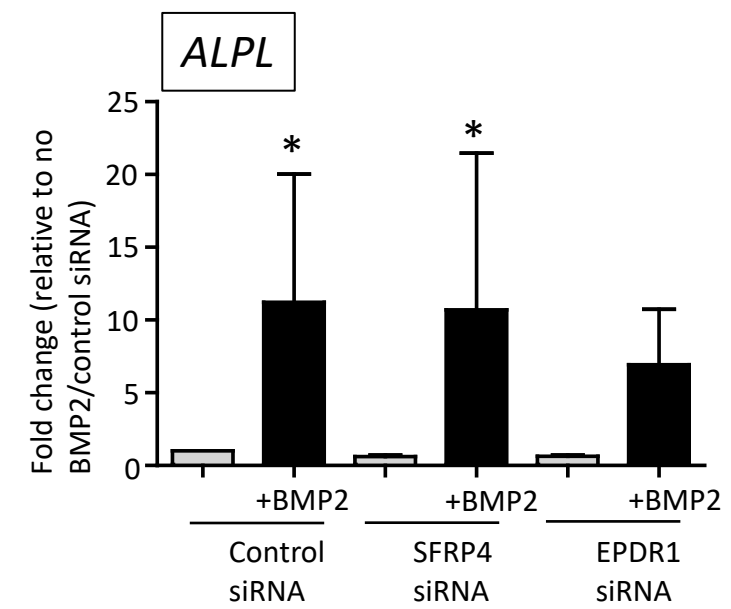
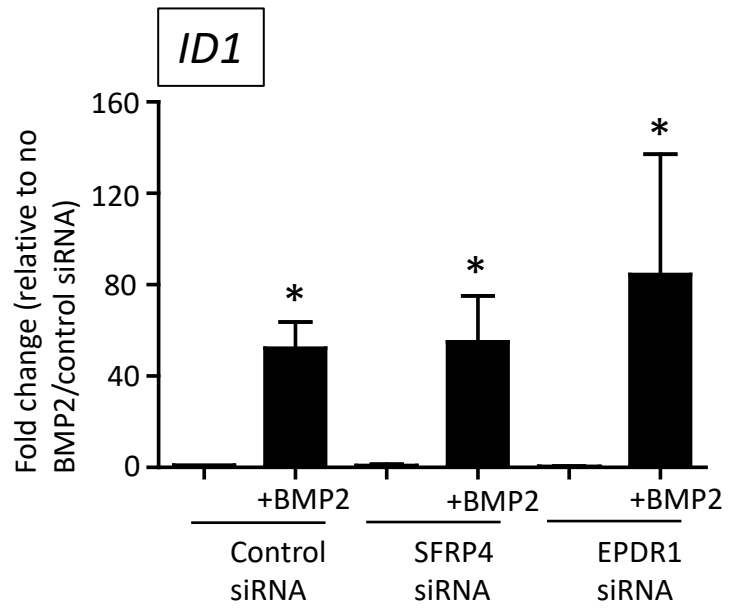
# ING3 silencing does not decrease RUNX2 and SP7



# EPDR1 silencing reduces human osteoblast differentiation

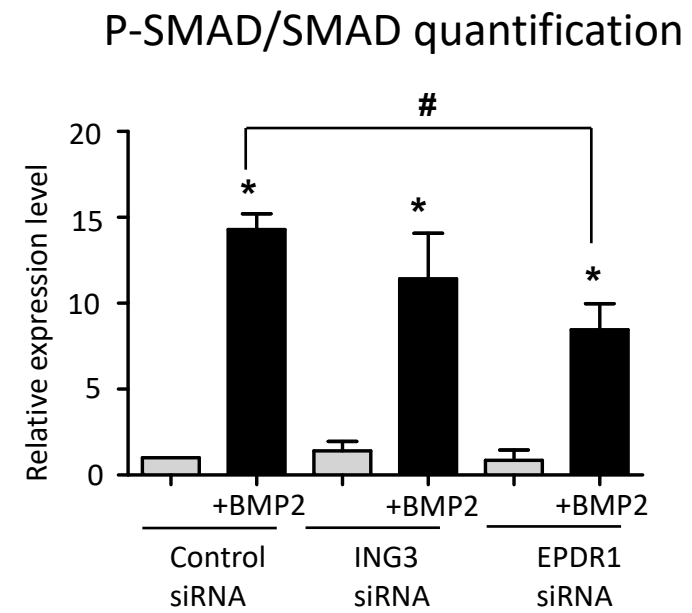
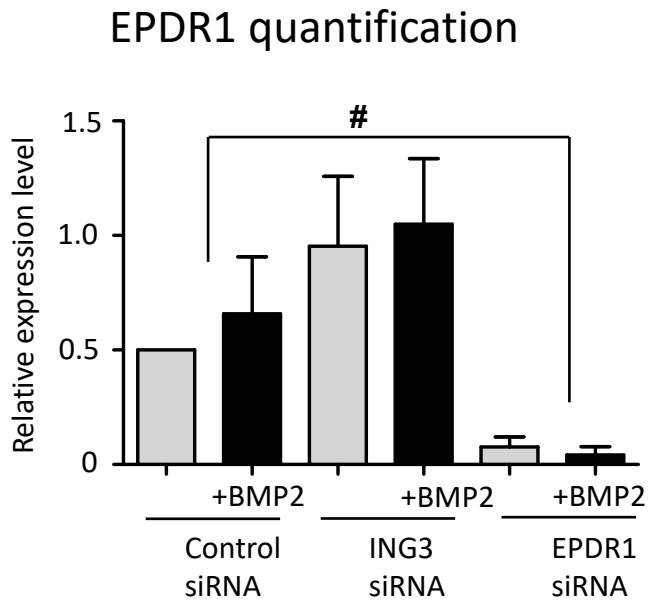
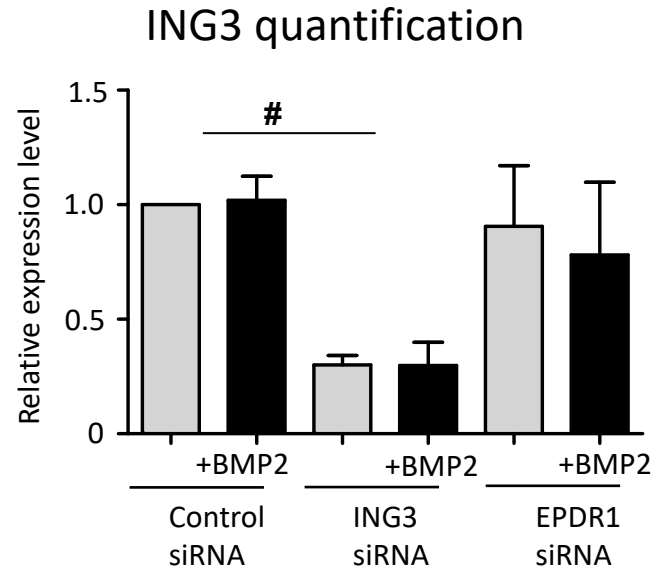
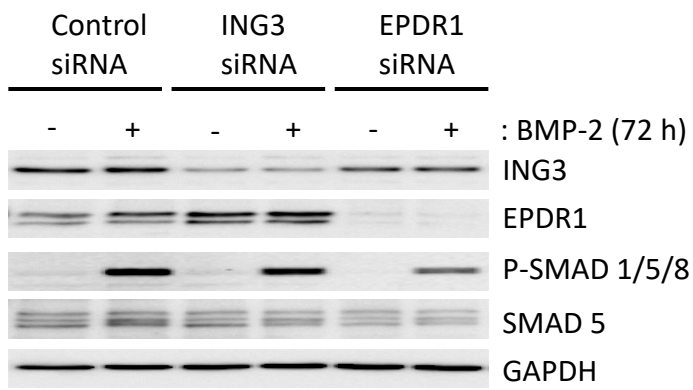


# EPDR1 silencing reduces human osteoblast differentiation





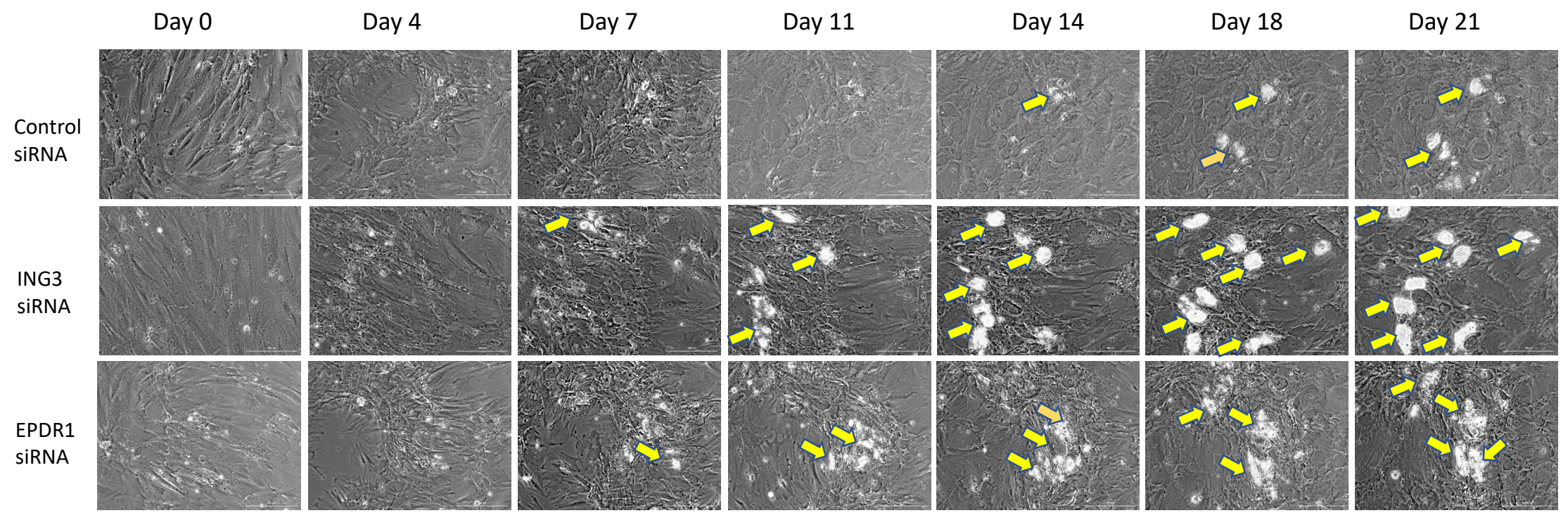
# ING3 and EPDR1 silencing demonstrated at protein levels



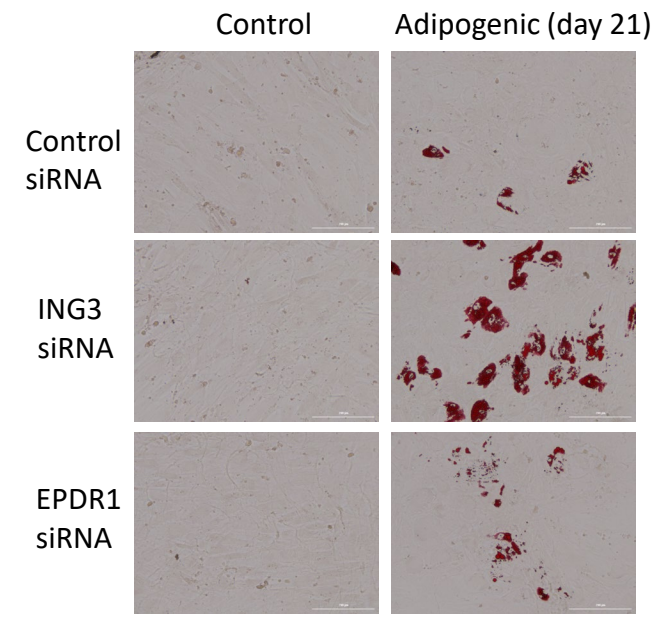
**Are *ING3* and *EPDR1* fundamental genes required for general cell metabolism and whether these genes affect MSC differentiation into adipocytes?**

# *ING3* and *EPDR1* silencing increases adipogenesis

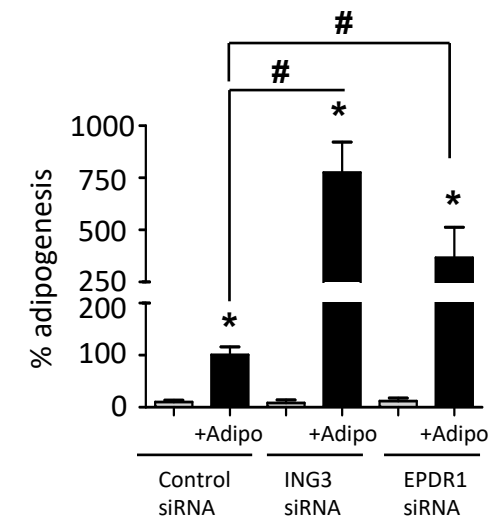
Adipogenic differentiation 



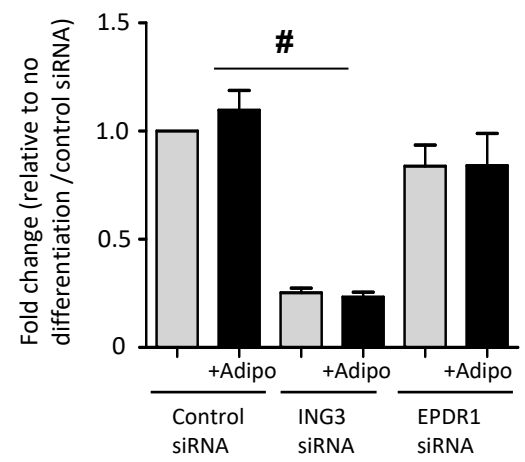
# ING3 and EPDR1 silencing increases adipogenesis



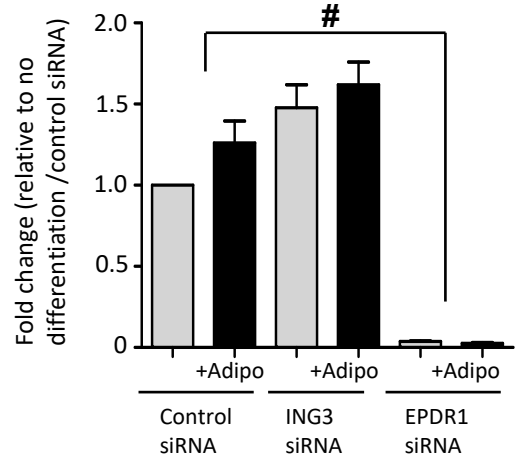
Lipid-droplet accumulation



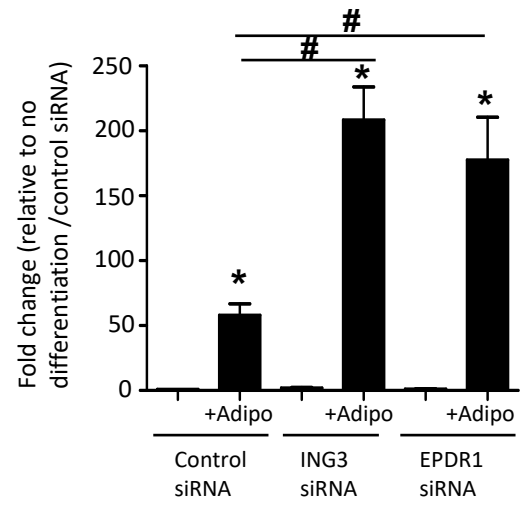
ING3



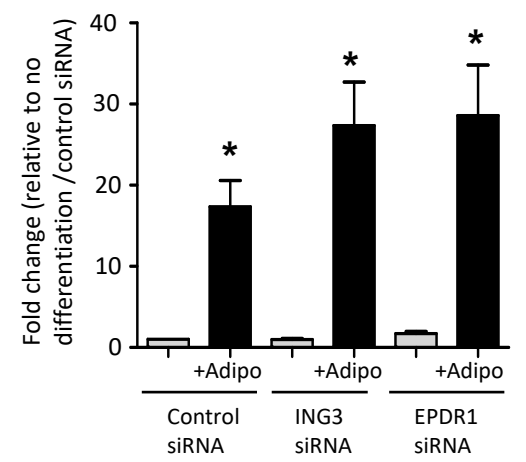
EPDR1



C/EBP alpha



PPAR gamma



Article | [Open Access](#) | [Published: 19 March 2019](#)

## Genome-scale Capture C promoter interactions implicate effector genes at GWAS loci for bone mineral density

[Alessandra Chesj](#), [Yadav Wagley](#), [Matthew E. Johnson](#), [Elisabetta Manduchi](#), [Chun Su](#), [Sumei Lu](#), [Michelle E. Leonard](#), [Kenya M. Hodge](#), [James A. Pippin](#), [Kurt D. Hankenson](#), [Andrew D. Wells](#) & [Struan F. A. Grant](#) 

*Nature Communications* **10**, Article number: 1260 (2019) | [Cite this article](#)

8491 Accesses | 32 Citations | 79 Altmetric | [Metrics](#)



**46 GWAS loci revealed at least one or more BMD proxy SNPs in open chromatin (and not residing in a baited promoter region) interacting with an open gene promoter**

Research | [Open Access](#) | [Published: 04 January 2021](#)

## Genome-wide association study implicates novel loci and reveals candidate effector genes for longitudinal pediatric bone accrual

[Diana L. Cousminer](#) , [Yadav Wagley](#), [...] [Struan F. A. Grant](#) 

*Genome Biology* **22**, Article number: 1 (2021) | [Cite this article](#)

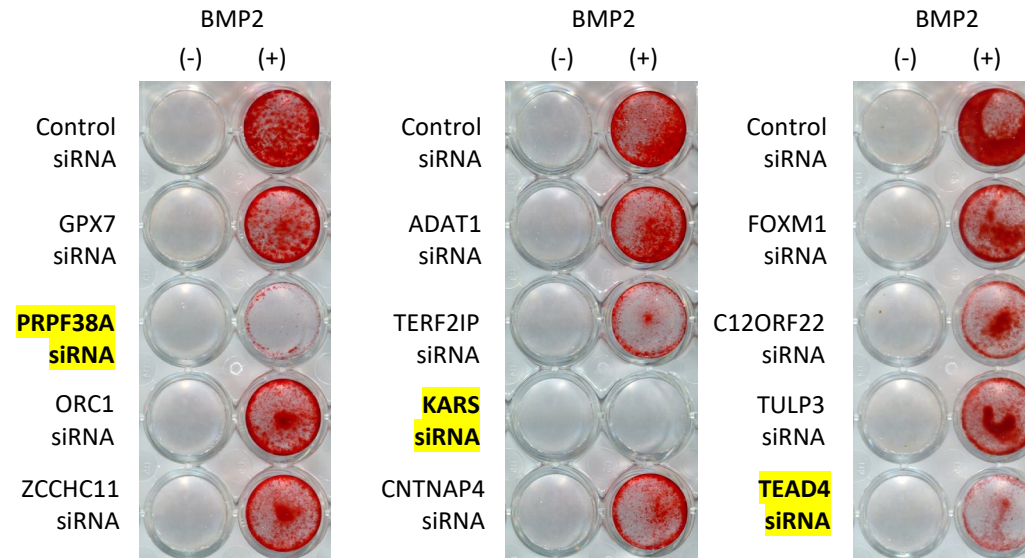
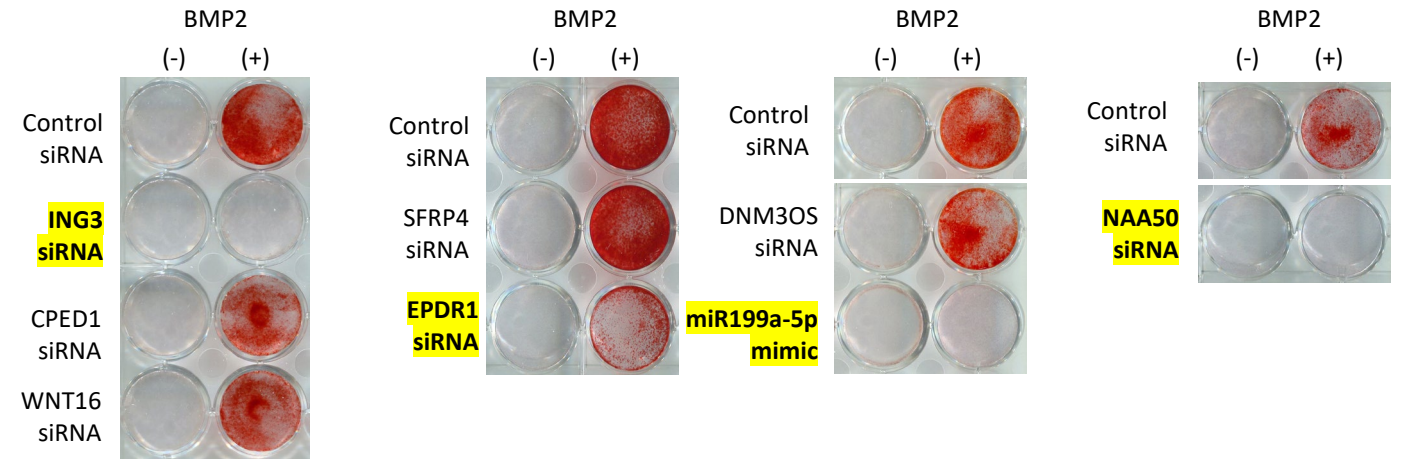
2290 Accesses | 4 Citations | 81 Altmetric | [Metrics](#)



**40 prioritized loci using multiple inclusion criteria (traditional GWAS threshold, suggestive significance level) of which 35 signals are novel**

# For 7 loci, causal effector genes have been established

Locus	Implicated genes
<i>CPED1</i>	<i>ING3, CPED1, WNT16</i>
<i>STARD3NL</i>	<i>EPDR1, SFRP4</i>
<i>DNM3</i>	<i>DNM3OS, MIR199A2</i>
<i>KIAA2018</i>	<i>NAA50</i>
<i>CC2D1B</i>	<i>GPX7, PRPF38A, ORC1, ZCCHC11</i>
<i>TERF2IP</i>	<i>ADAT1, TERF2IP, KARS, CNTNAP4</i>
<i>TSPAN9</i>	<i>FOXM1, C12ORF22, TULP3, TEAD4</i>



Functional follow up studies with EPDR1 and miR199a-5p are ongoing

High-throughput CRISPRi assay to identify effector genes at remaining loci is being developed

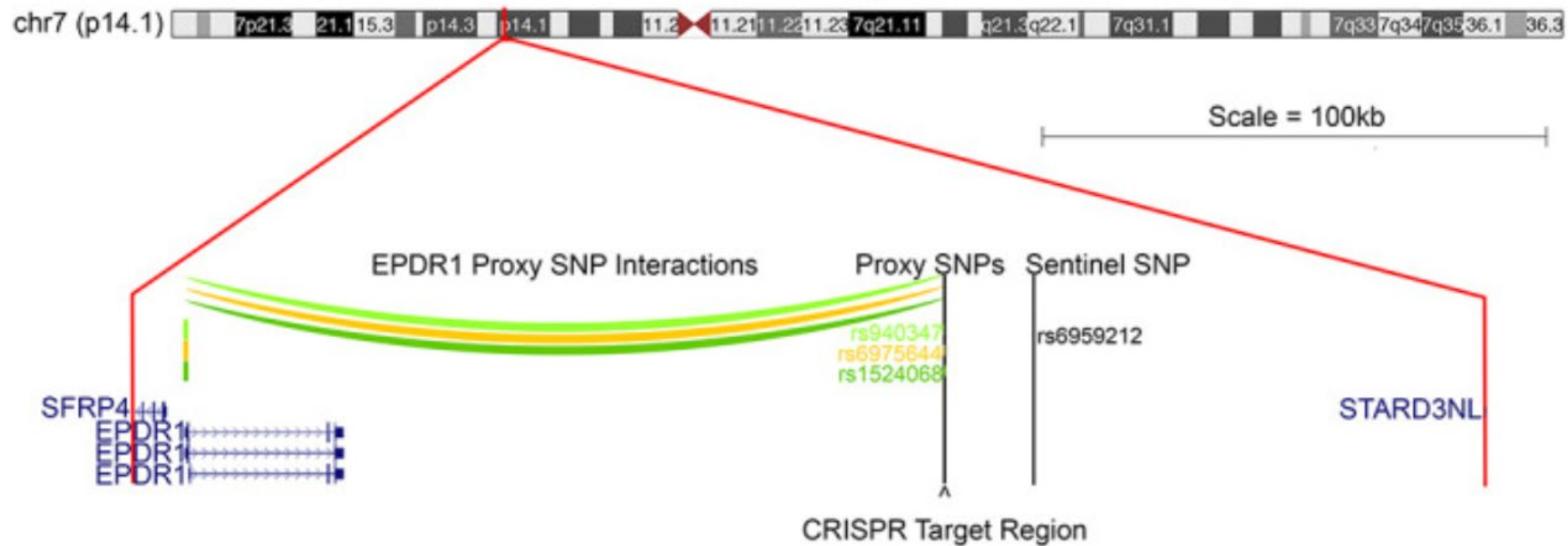
# Summary

- **ATAC-seq coupled with chromatin confirmation capture can be used for implicating gene variants associated with a particular phenotype**
- ***ING3* at the 'WNT16-CPED' locus and *EPDR1* at the 'STARD3NL' locus play a role in human osteoblast differentiation.**

**The open chromatin region spanning three proxy SNPs at the 'STARD3NL' locus constitutes EPDR1 enhancer and is functional during osteoblast differentiation**

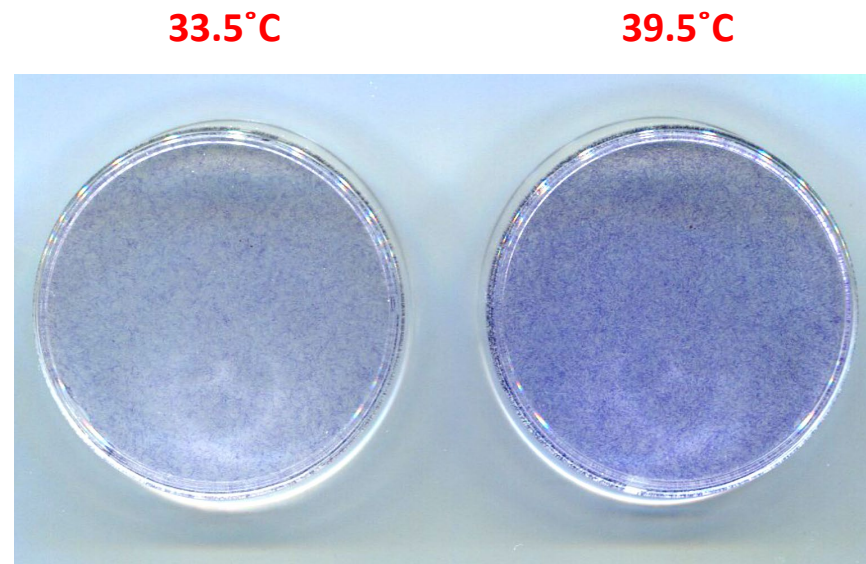


## EPDR1 at 'STARD3NL' Locus (Sentinel rs6959212)



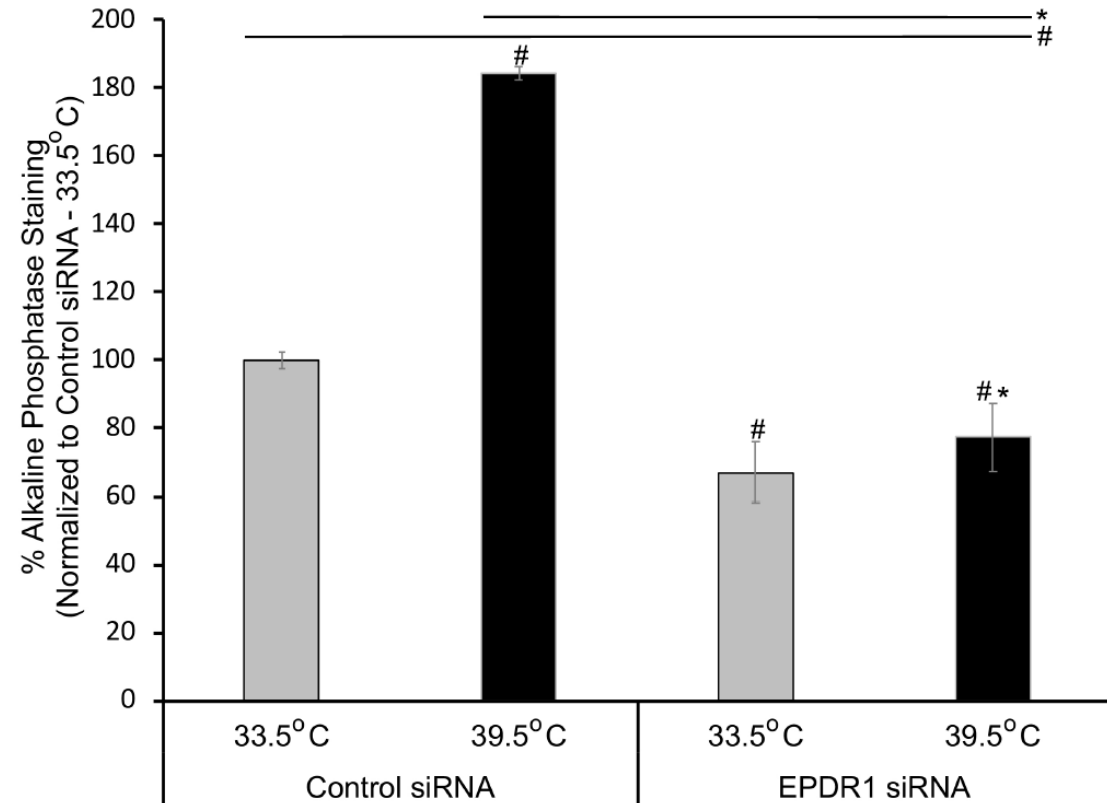
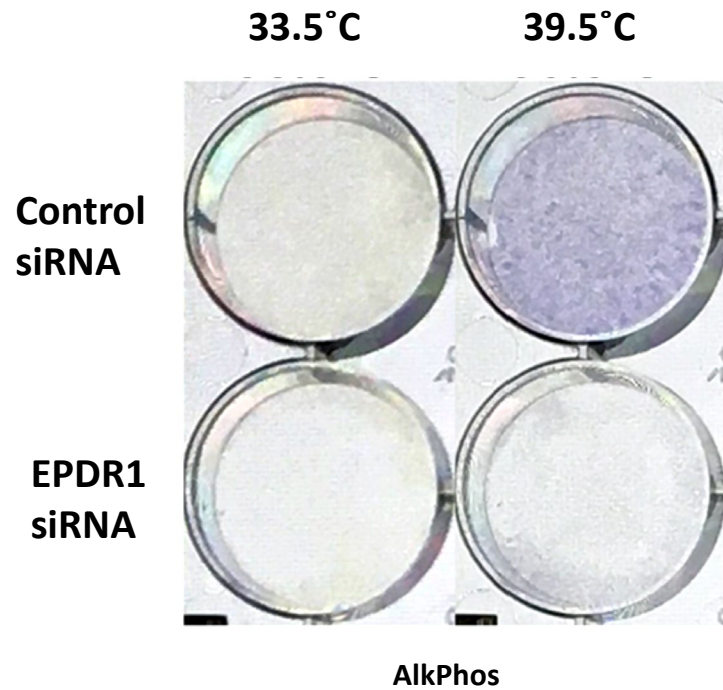
# hFOB1.19 cells as an alternate model to study proxy SNP regulation of EPDR1

- Immortalized human fetal osteoblastic cell line—easy expanding and passaging
- Contains temperature sensitive mutant, tsA58, of the SV40 large T antigen that allows for the genome-edited cells to proliferate at 33.5°C and to differentiate at 39.5°C



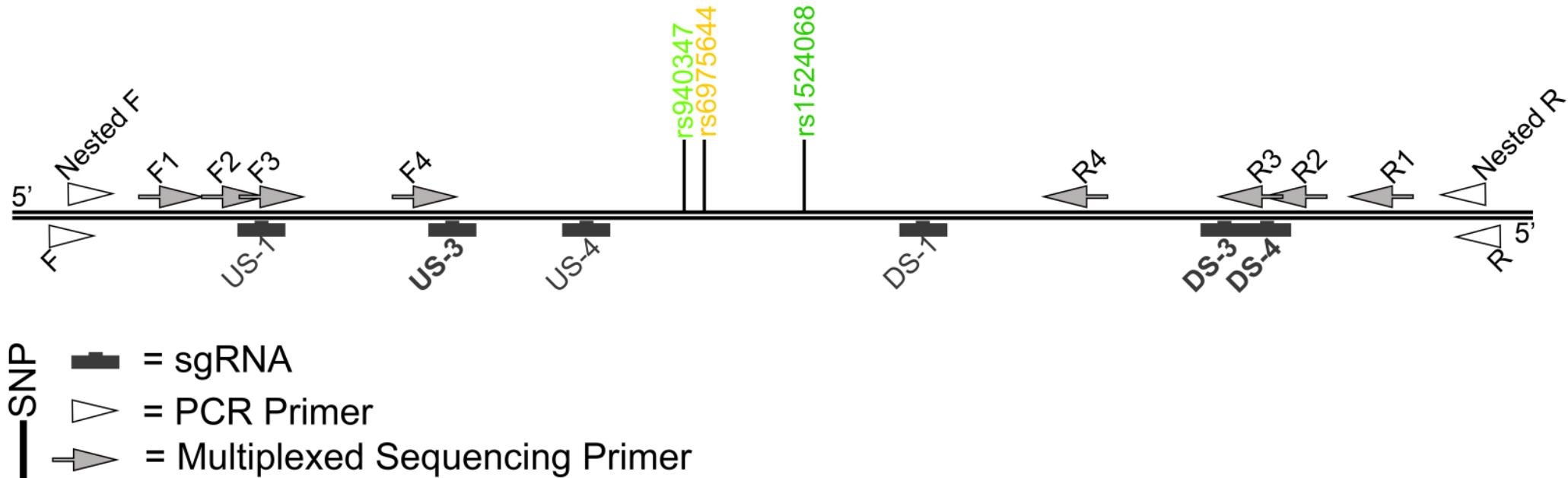
AlkPhos

- **EPDR1 knockout in hFOB1.19 cells shows reduced Alk Phos expression and therefore is a suitable cell model to study CRISPR-Cas9 genome editing of proxy SNPs**

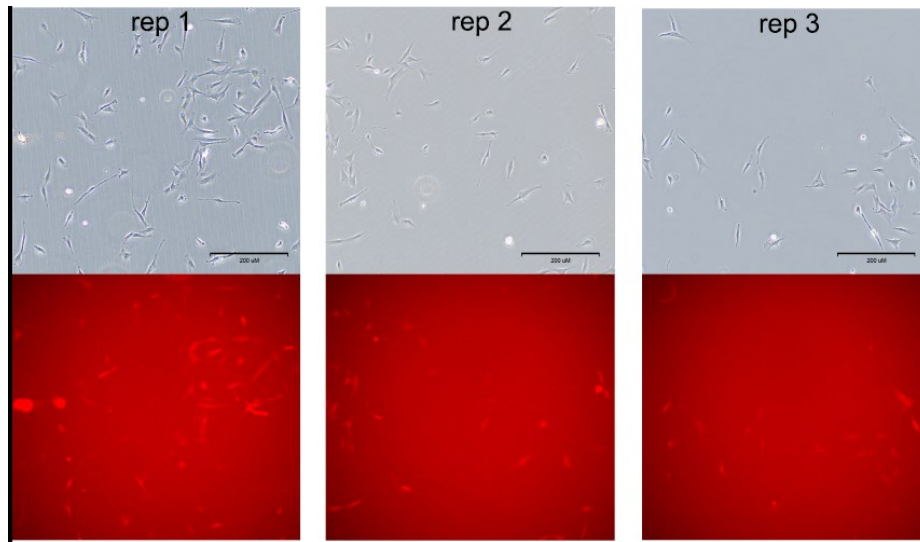


**Pooled lentivirus was prepared using three sets of guides on each upstream and downstream site of the proxy SNPs**

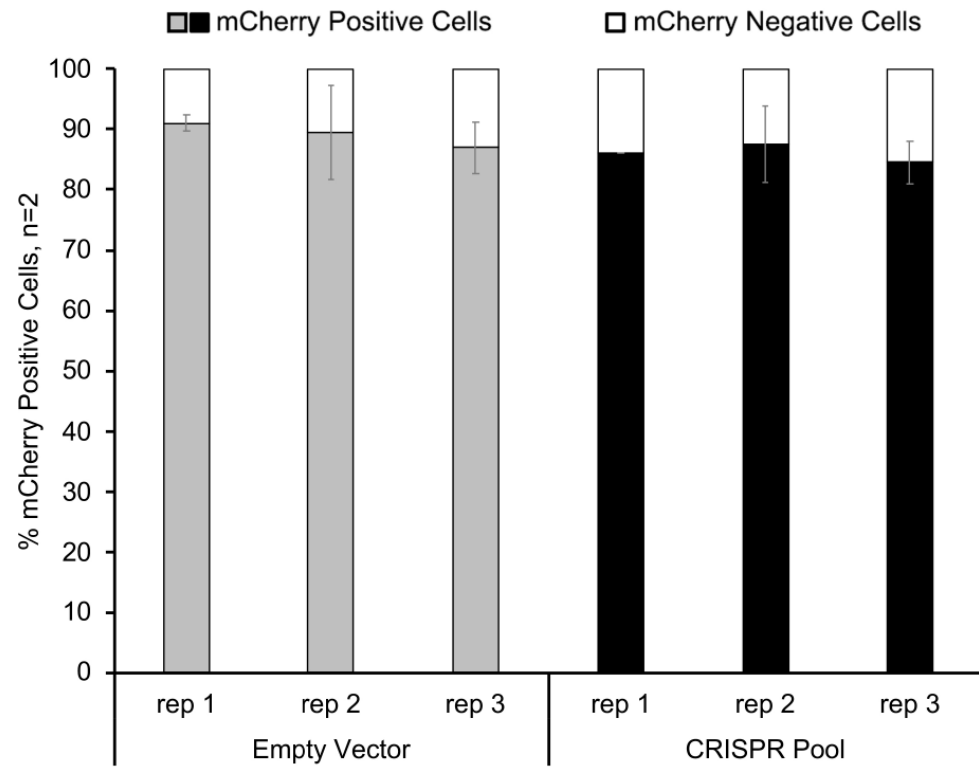
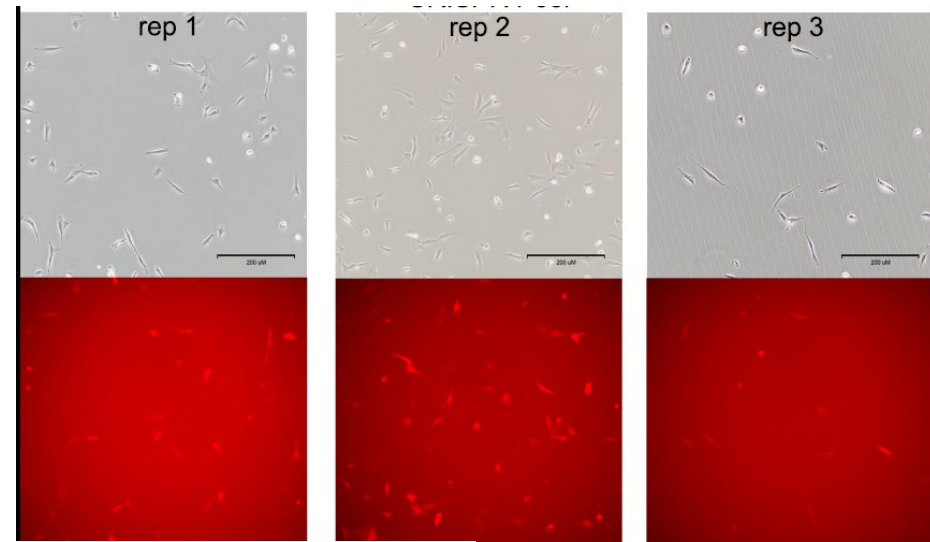
### CRISPR Design at '*STARD3NL*' Locus (Sentinel rs6959212) Proxy SNPs



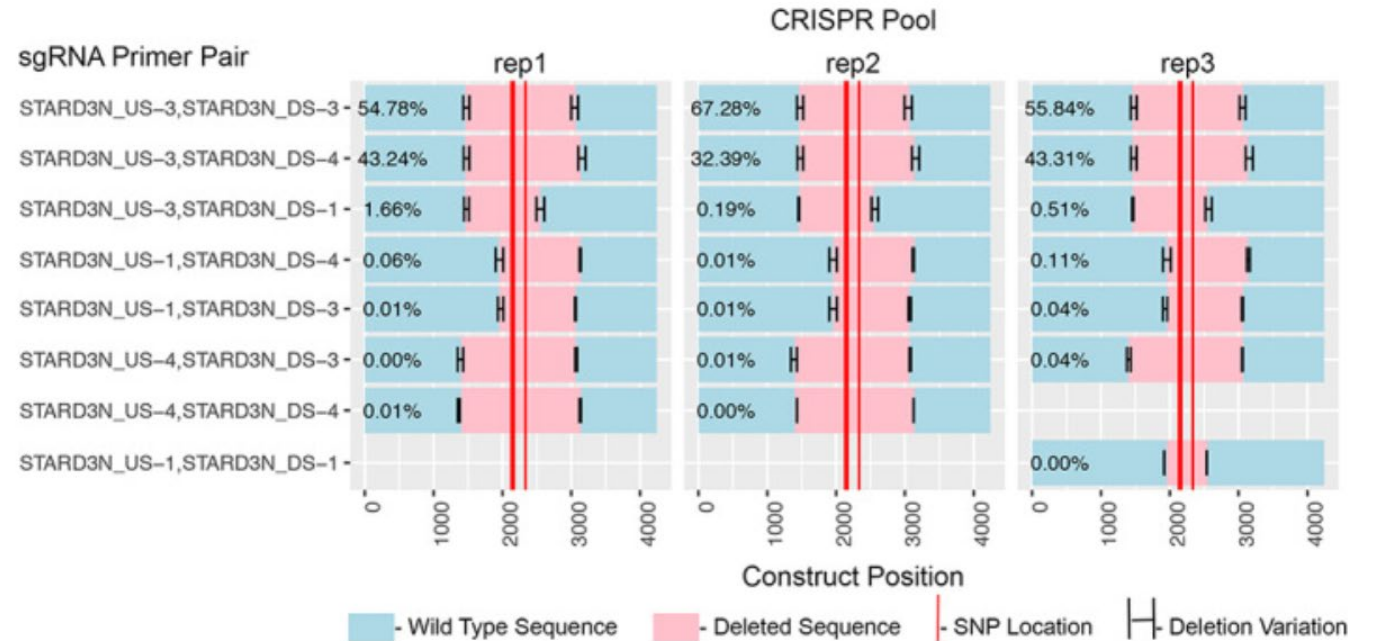
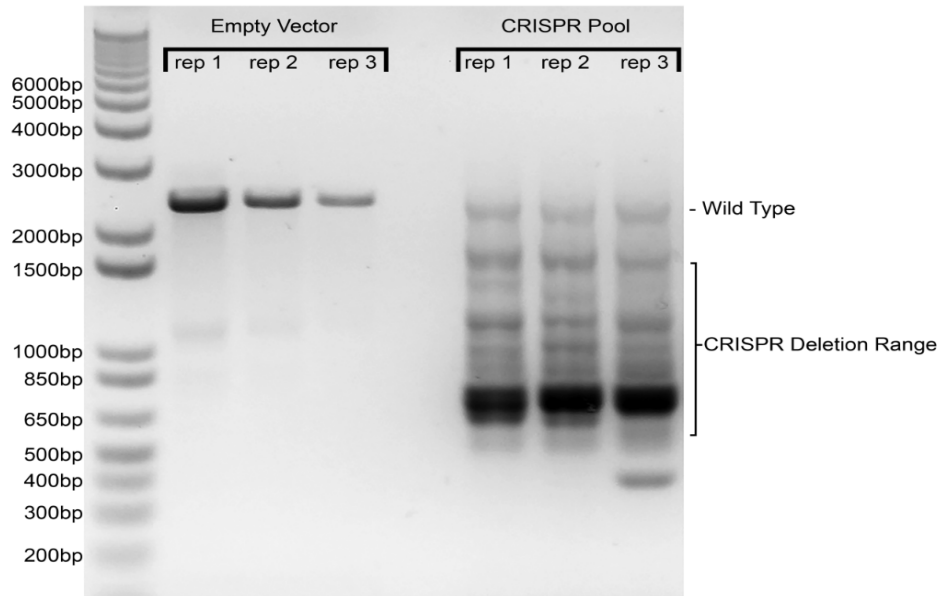
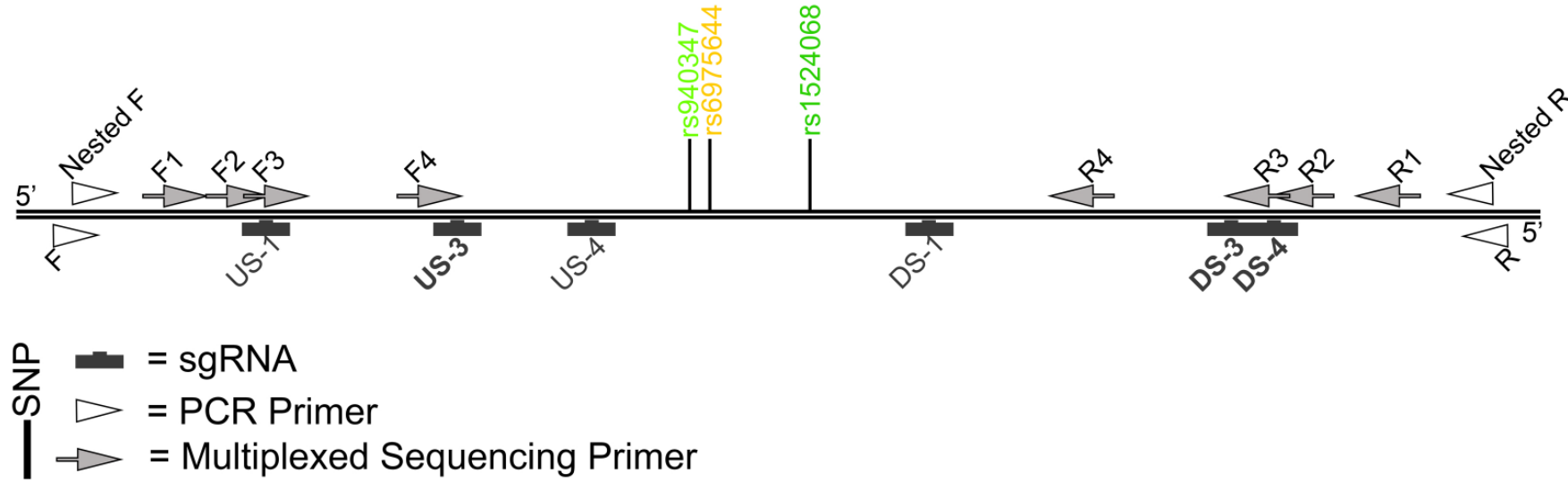
### Empty Vector



### CRISPR pool

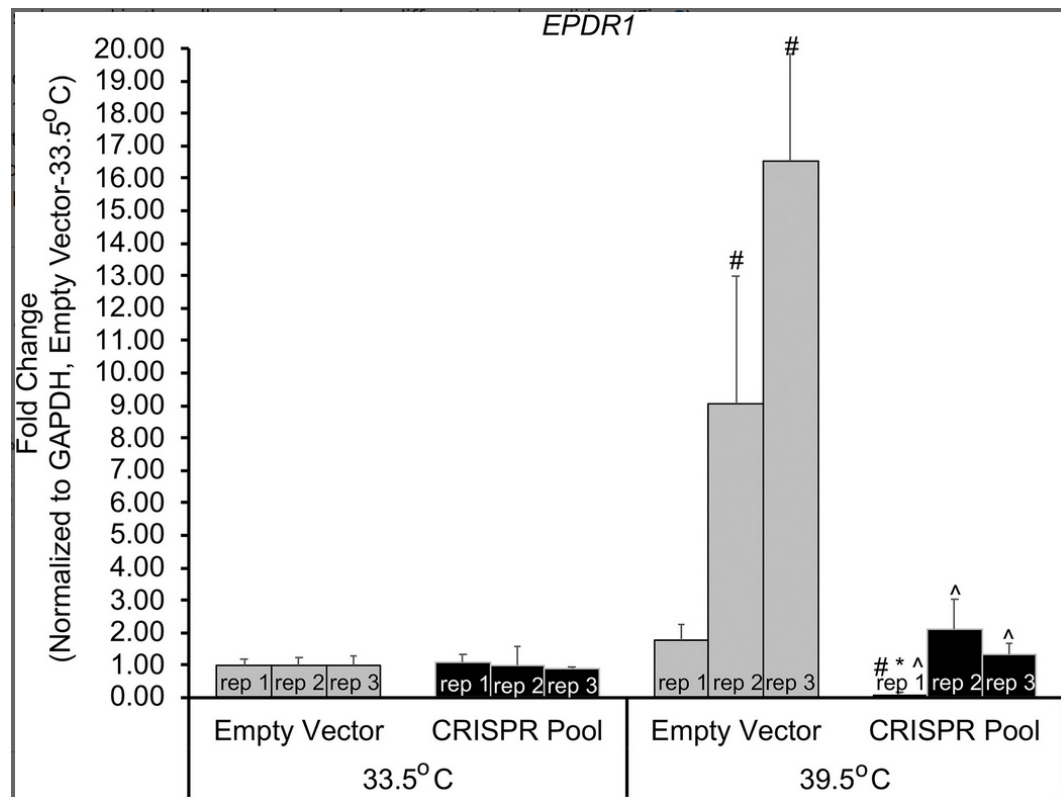


# CRISPR Design at 'STARD3NL' Locus (Sentinel rs6959212) Proxy SNPs

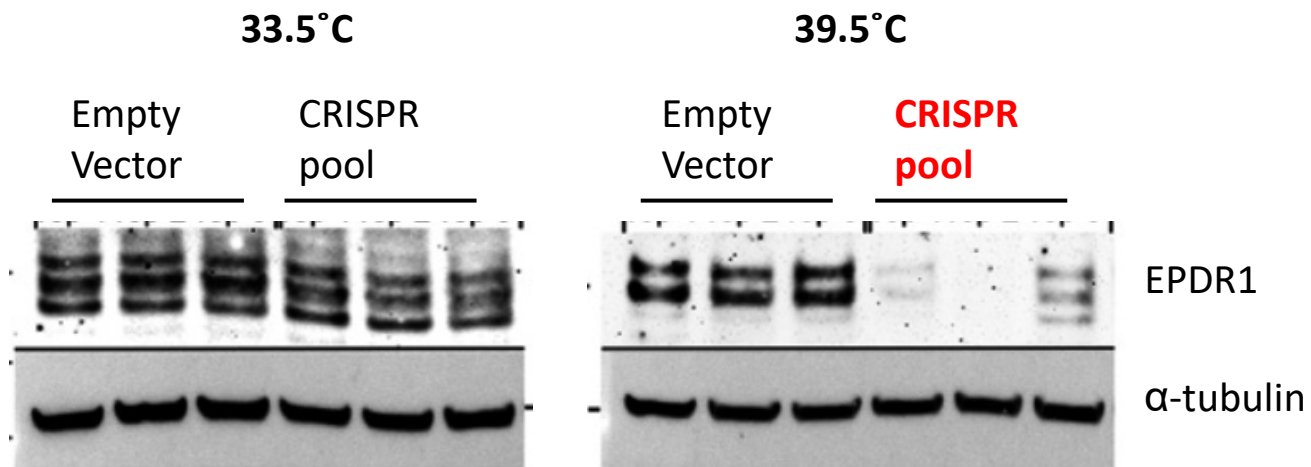


# Deletion of proxy SNPs reduces *EPDR1* expression in cells undergoing differentiation

## RT-qPCR



## Western Blot



33.5°C

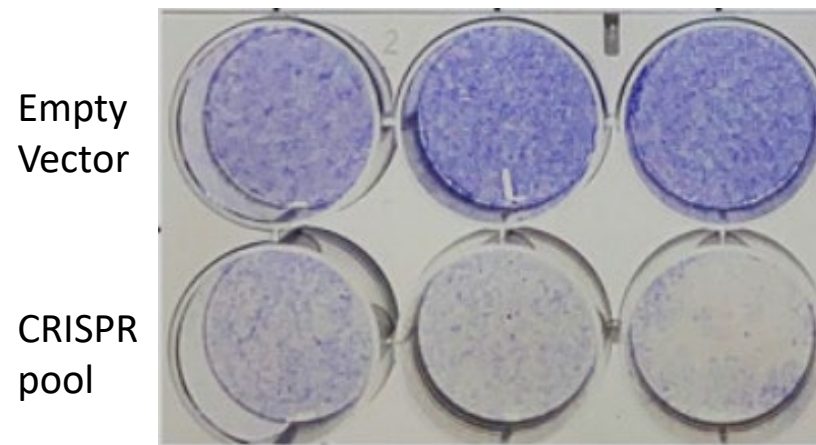


Empty Vector

CRISPR pool

AlkPhos

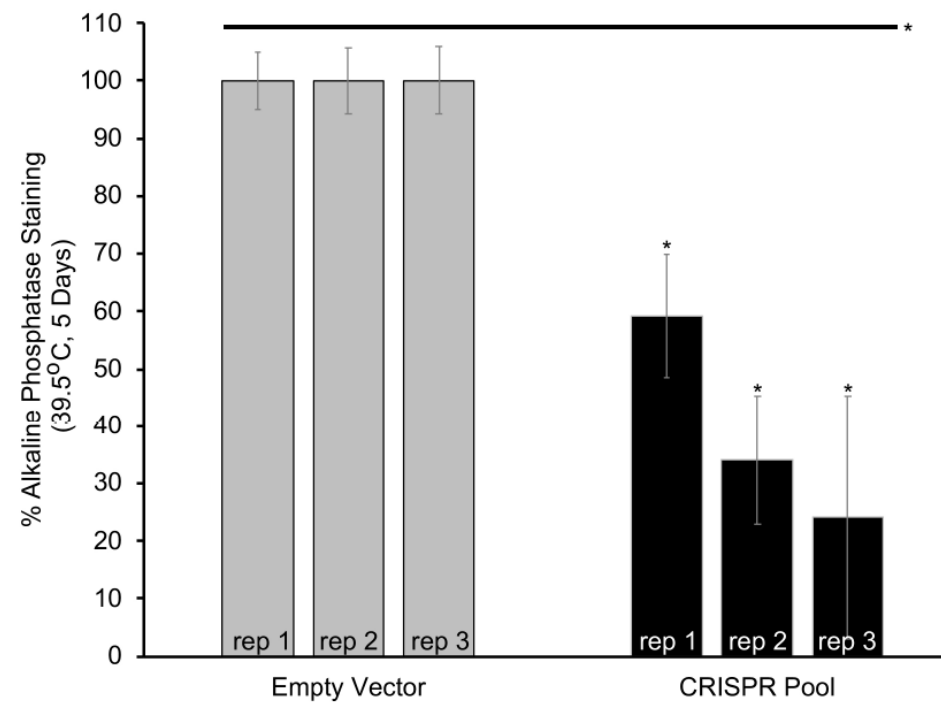
39.5°C



Empty Vector

CRISPR pool

AlkPhos

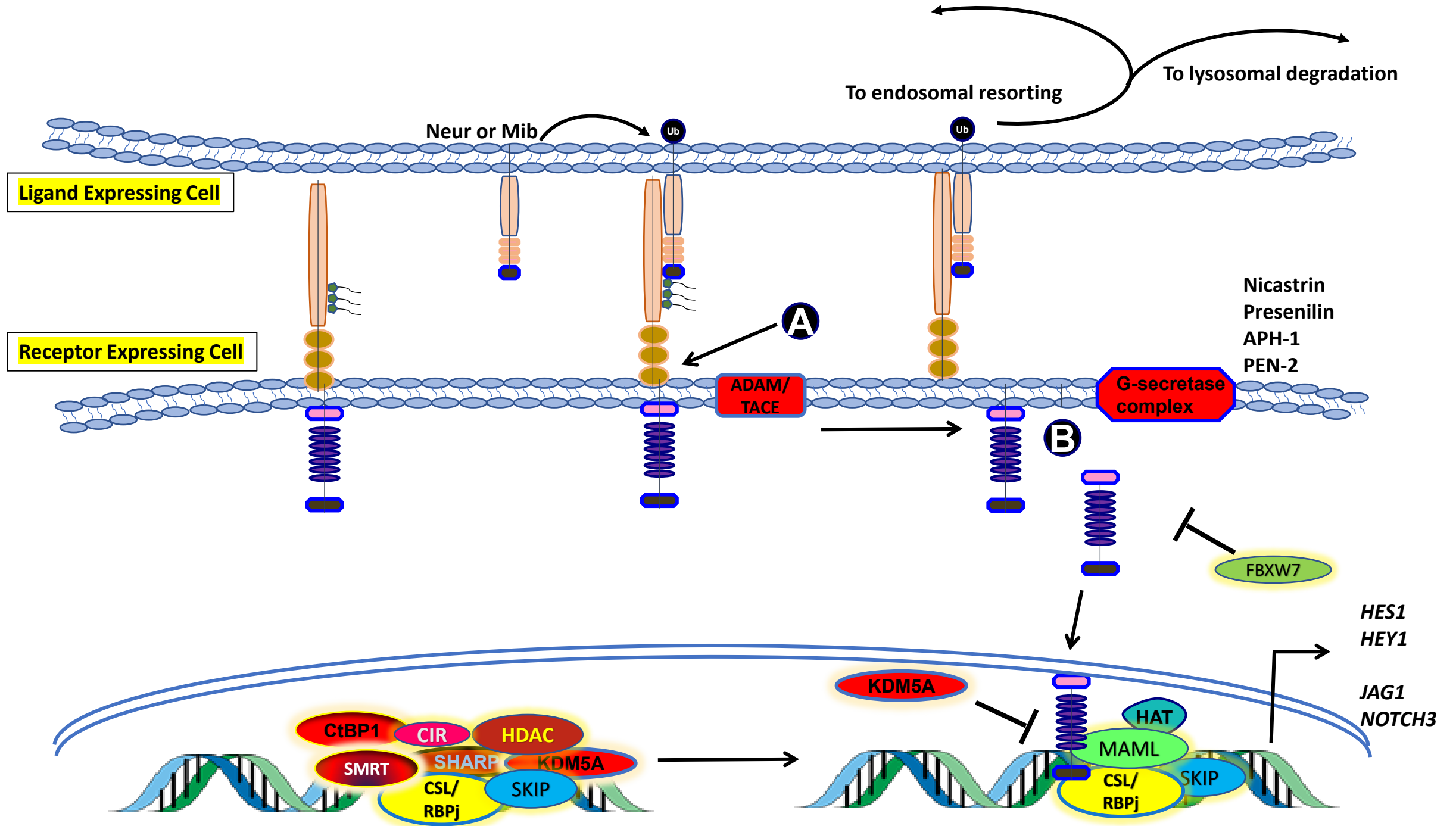




## Summary

- **EPDR1 plays a regulatory role during osteoblast differentiation of hFOB1.19 cells**
- **CRISPR-Cas9 deletion of three proxy SNPs at the 'STARD3NL' locus shows reduced EPDR1 expression during hFOB1.19 osteoblast differentiation**

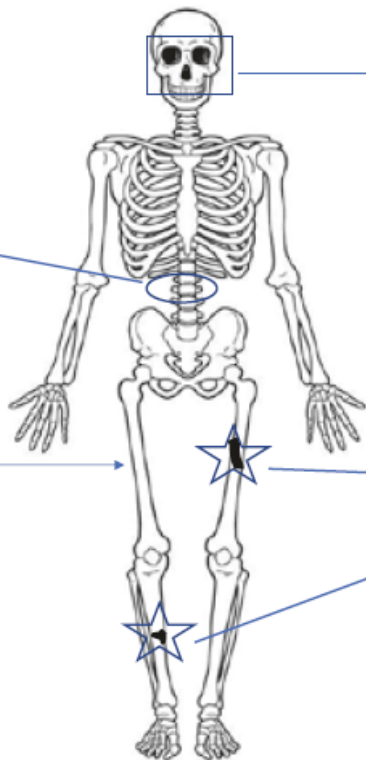
**EPDR1 is required for Jagged1 mediated human osteoblast differentiation**





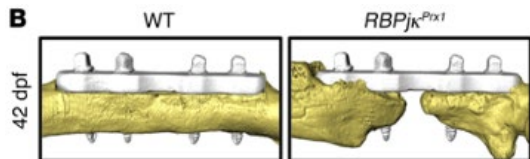
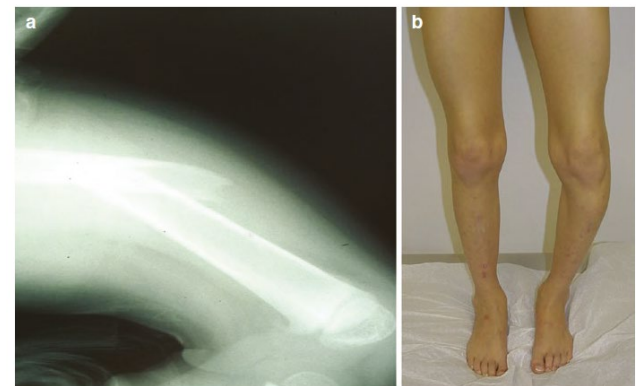
Butterfly vertebrae result from abnormal somite development

- ALGS patients have reduced bone density.
- Mice with genetic alterations in Notch signaling show highly variable phenotypes.
- Mice with *Jag1* disruption in bone progenitors show reduced bone regeneration.

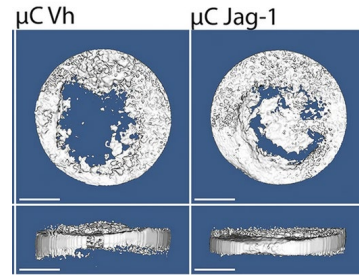
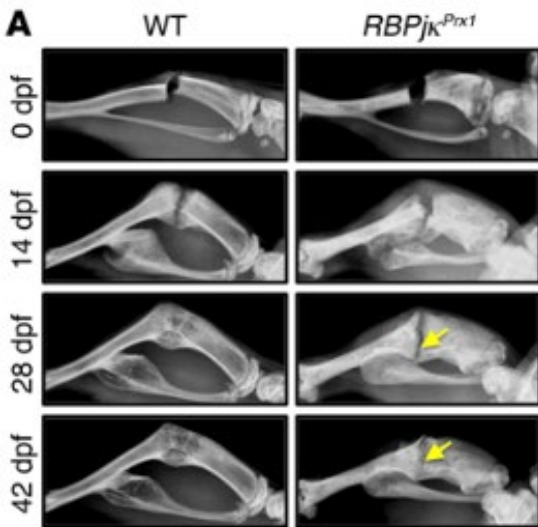


- Characteristic facial features
- Prominent forehead w/pointed chin
  - Deeply set eyes
  - Straight nose with bulbous tip

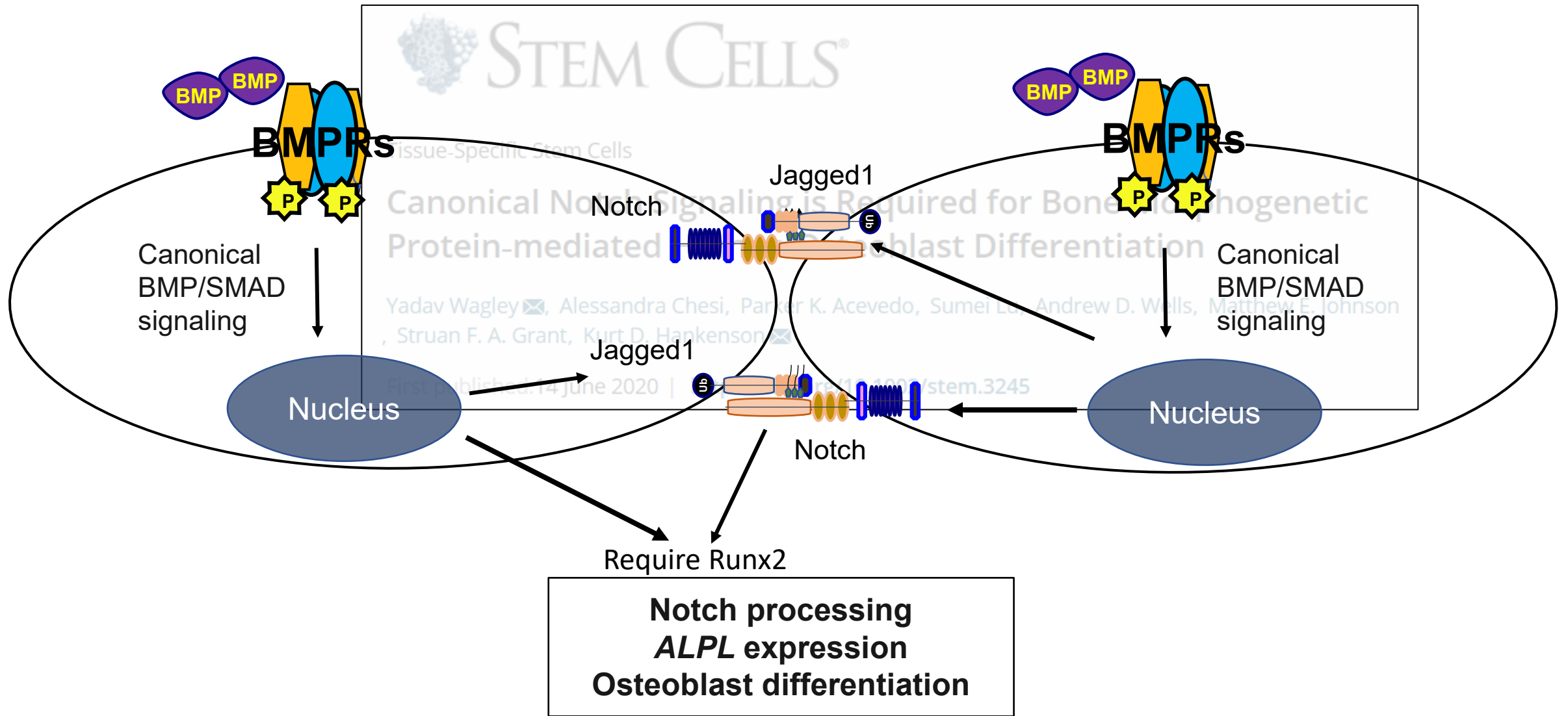
- ALGS patients have increased fracture risk and poor fracture healing.
- Mice with canonical Notch signaling disrupted do not heal bone defects.
- Jagged-1 delivery to bone injuries promotes fracture healing



Wang et al., (2016) JCI



Youngstrom et al. (2017), NPJ Regen Med

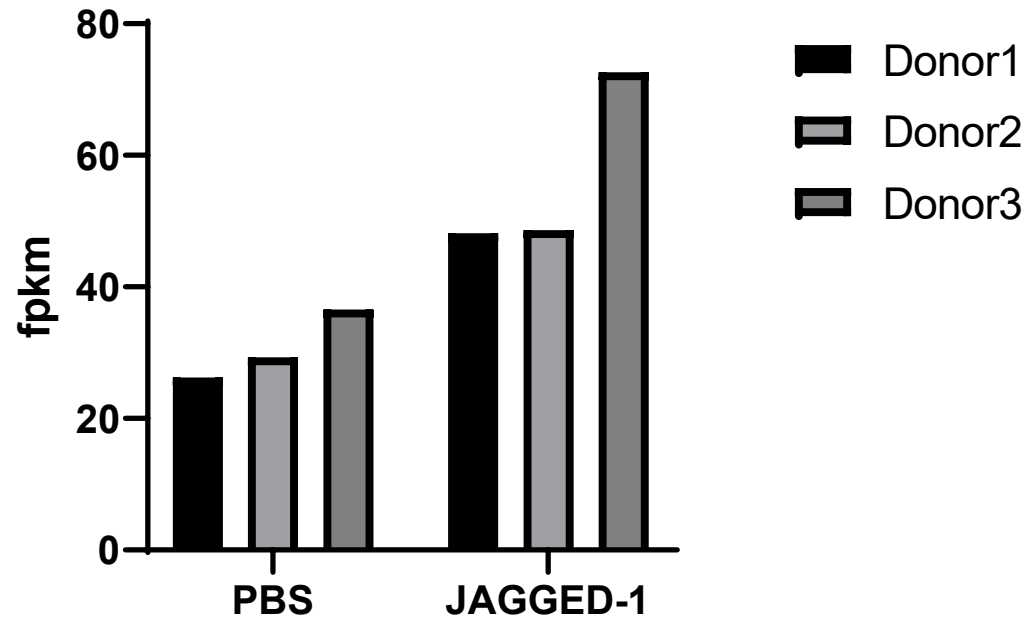


BMP stimulation of bone-marrow derived human mesenchymal progenitor cells (hMSCs) increases Notch proteins via increased Notch ligand Jagged1 expression. Canonical Notch signaling is required for BMP-induced *ALPL* expression and osteoblastic commitment of hMSCs. Both BMP-induced osteoblastogenesis and Notch-induced osteoblastogenesis require Runx2 and RBPj

# hMSC upregulates *EPDR1* upon Jagged1 stimulation

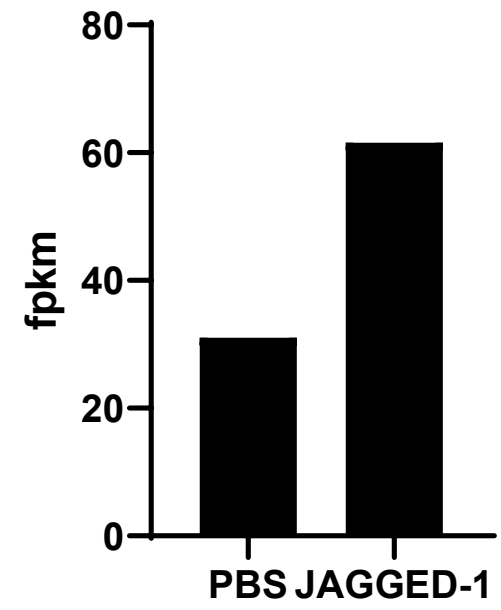
ENSEMBLE_GENE_ID	GENE	FPKM.JAG11	FPKM.JAG12	FPKM.JAG13	FPKM.Contr1	FPKM.Contr2	FPKM.Contr3	FC	FDR
ENSG00000086289.7	EPDR1	48.1494242955653	48.61702414	72.63829631	26.24704744	29.27442245	36.59837206	1.85085841	0.129099636

*EPDR1*

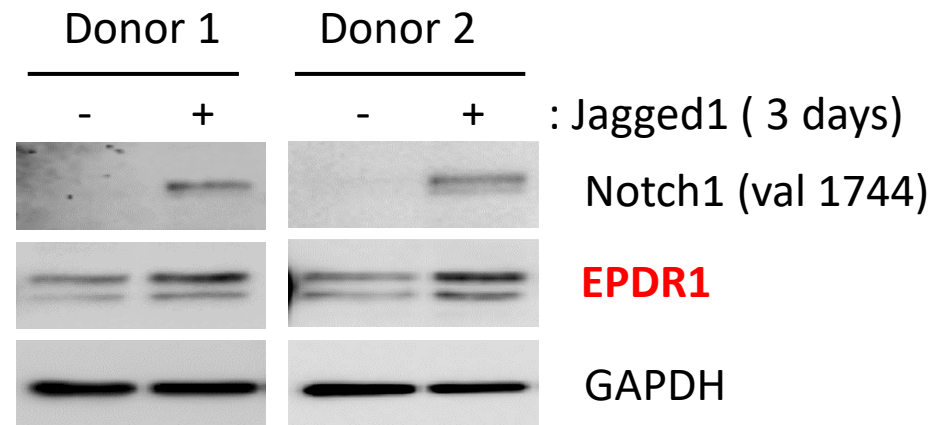
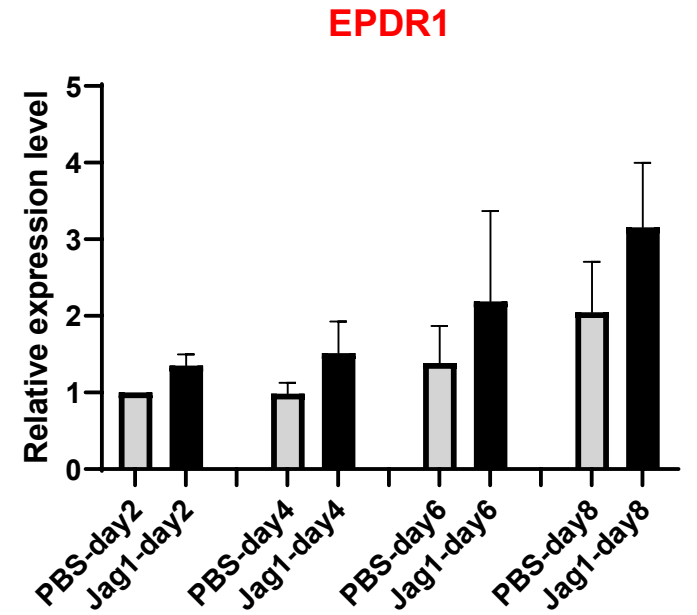
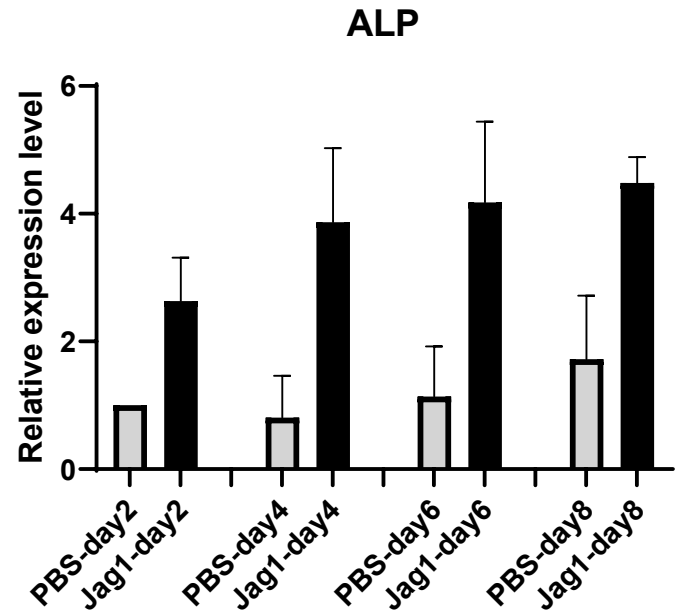
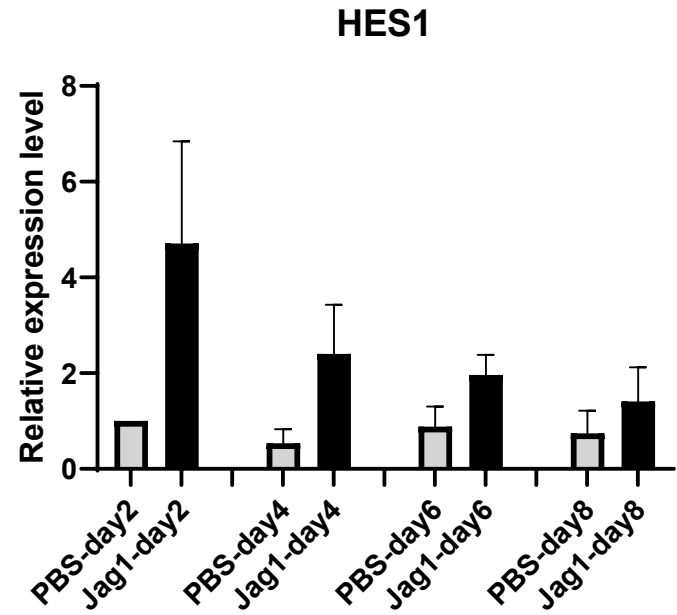


Separate sequencing library for Donor3

*EPDR1*

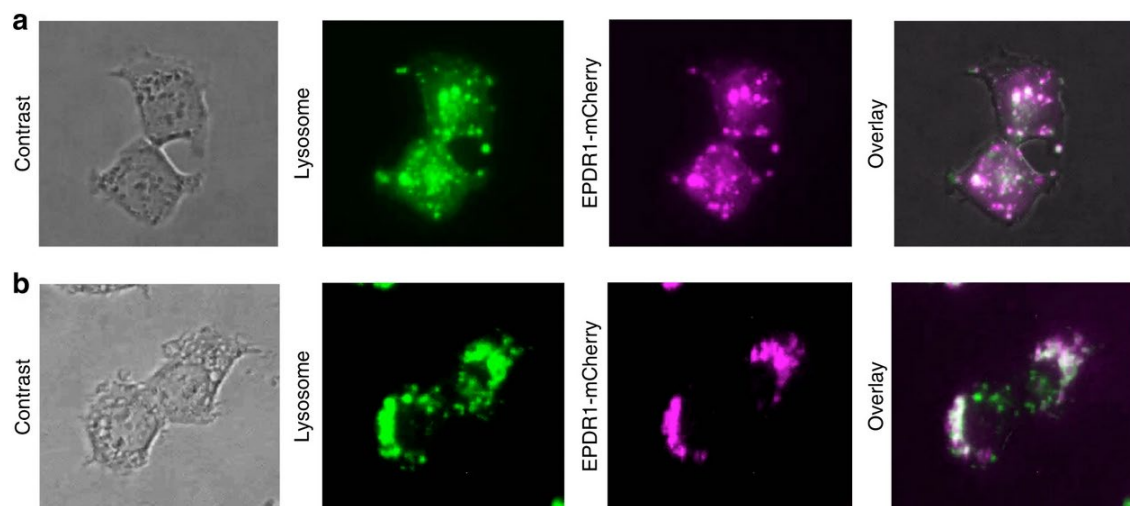


# hMSC upregulates *EPDR1* upon Jagged1 stimulation

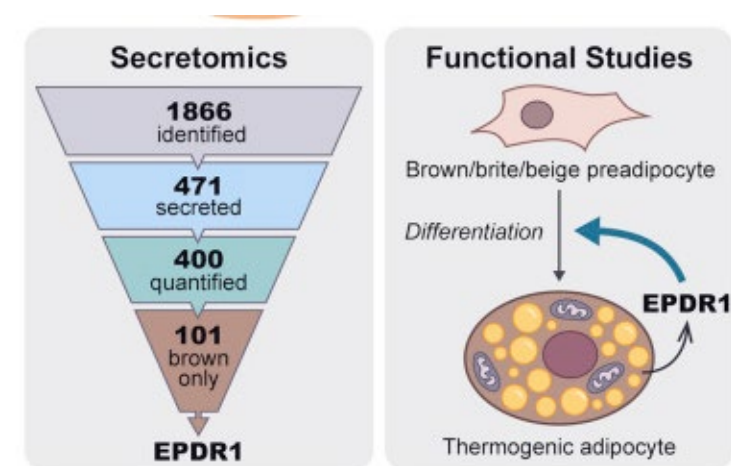


[Entrez Gene Summary for EPDR1 Gene](#)

The protein encoded by this gene is a type II transmembrane protein that is similar to two families of cell adhesion molecules, the protocadherins and ependymins. This protein may play a role in calcium-dependent cell adhesion. This protein is glycosylated, and the orthologous mouse protein is localized to the lysosome. Alternative splicing results in multiple transcript variants. A related pseudogene has been identified on chromosome 8. [provided by RefSeq, Aug 2011]



Wei et al, Communications Biology, 2(52): 2019

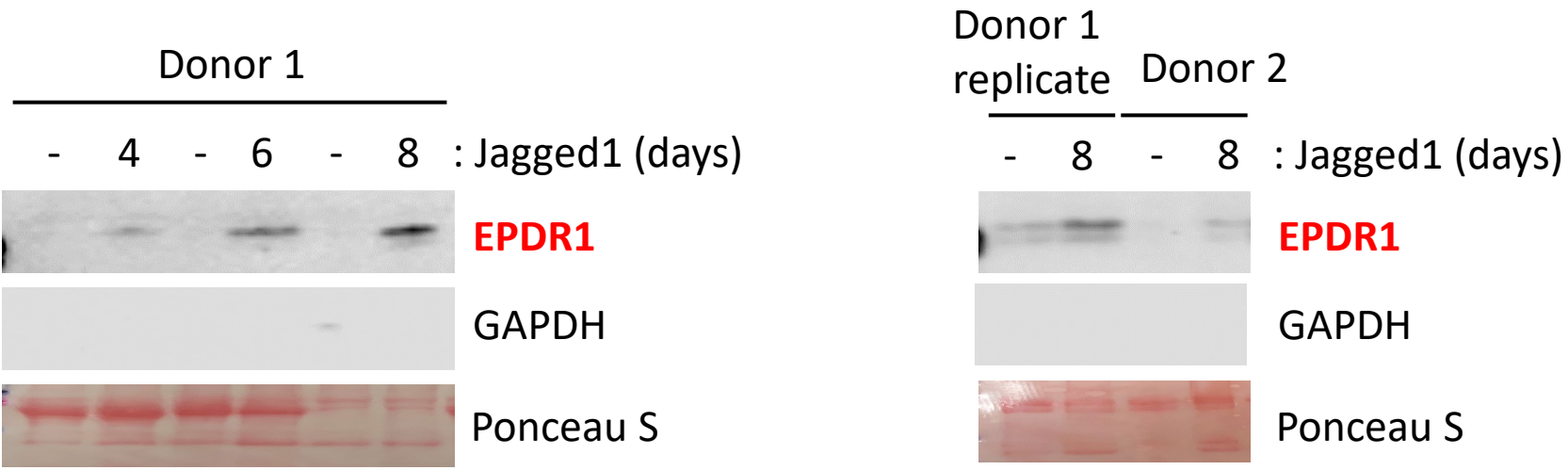


Deshmukh et al., Cell metabolism, 30(5): 963-975.e7, 2019)

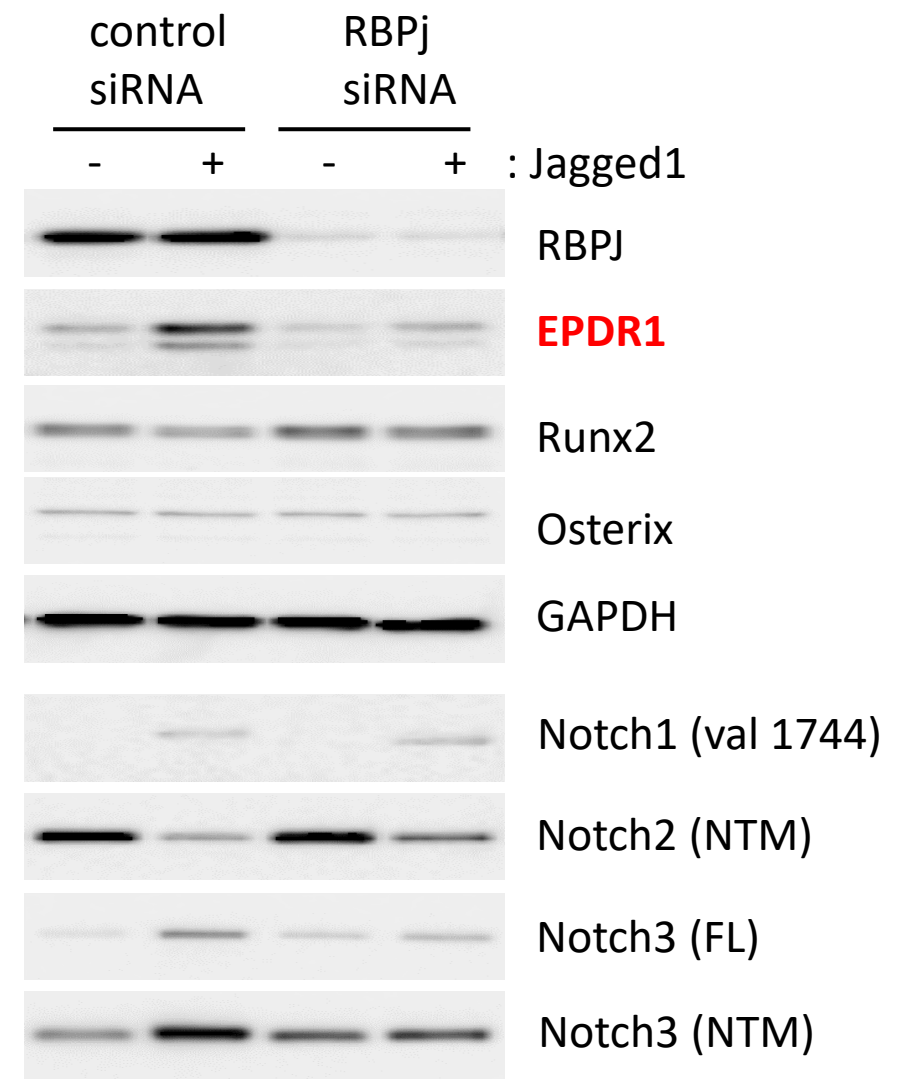
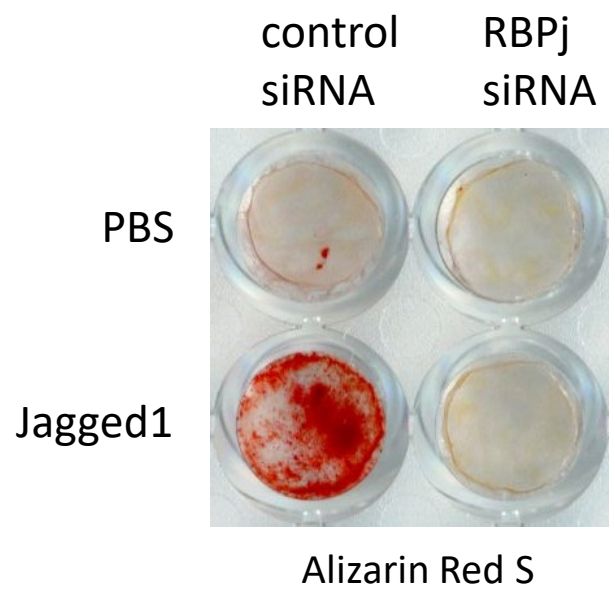
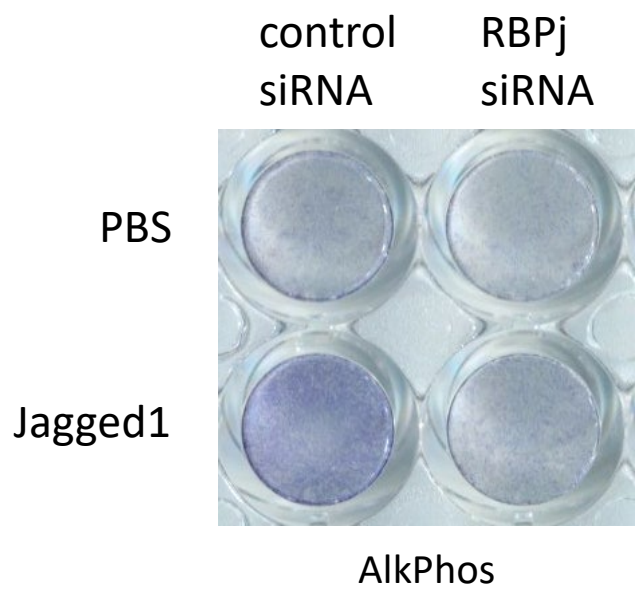




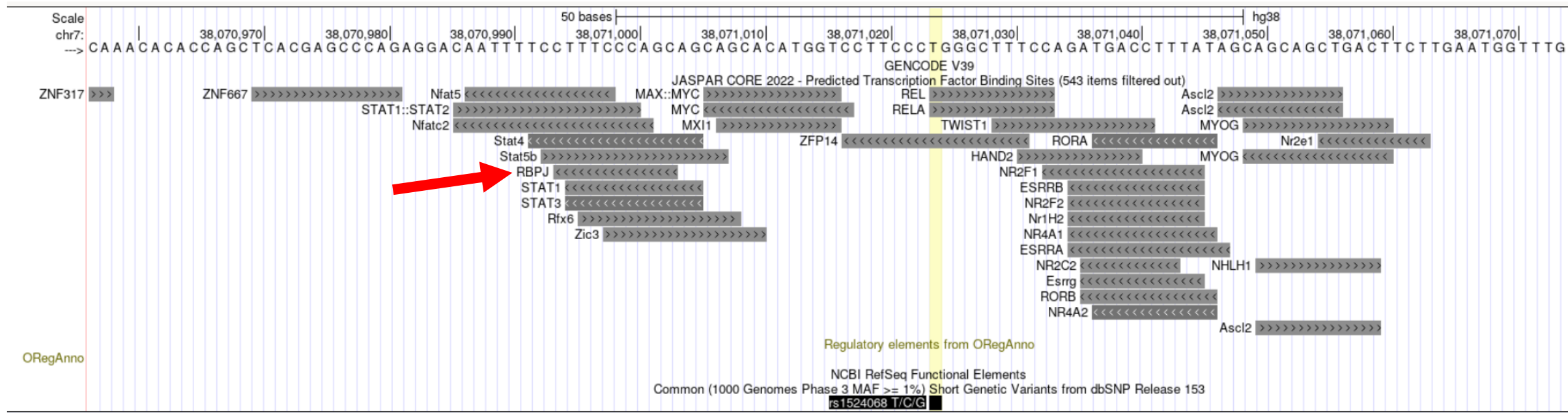
# EPDR1 protein secretion is increased in Jagged1 stimulated cells



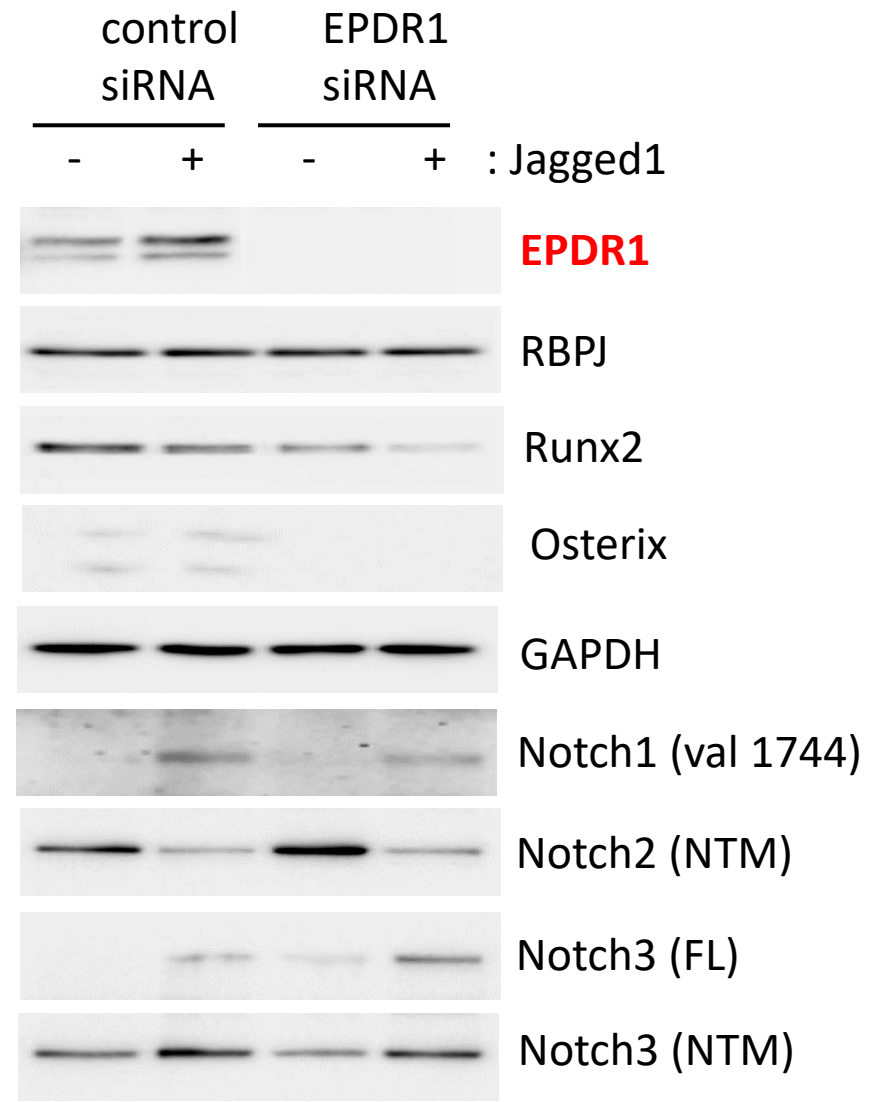
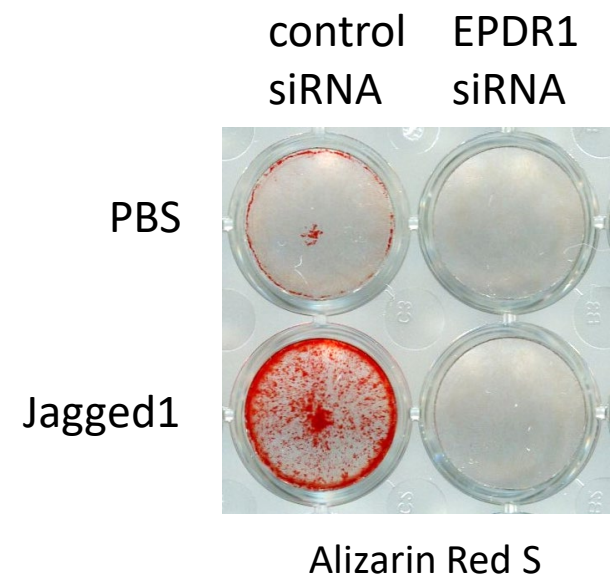
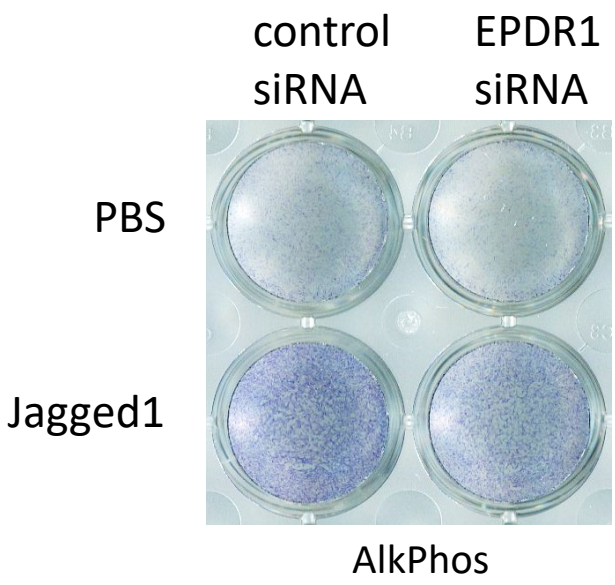
# RBPJ silencing using siRNA demonstrates reduction of EPDR1 protein expression



# RBPJ binding site is predicted upstream of rs1524069



# EPDR1 silencing using siRNA impairs Jagged1 mediated osteoblastogenesis



## Summary and future directions

- **EPDR1 is a secreted protein that plays a key role during human osteoblast differentiation**
- **Notch signaling enhances EPDR1 transcriptional activation**
  
- **Confirm the binding of RBPJ upstream of proxy SNP rs1524069**
- **Assess overexpression of EPDR1 during MSC differentiation.**
- **Evaluate the signaling potential of soluble EPDR1 protein during MSC differentiation.**

**Thank you**

**QUESTIONS?**