A Tail Withdrawal Procedure for Assessing Analgesic Activity in Rhesus Monkeys

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Rhesus monkeys were restrained in chairs from which their tails hung free so that their tails could be immersed into a thermos of water. Monkeys consistently kept their tails in 38–40°C water for at least 20 sec, but withdrew them from 55°C water in 1–4 sec. Tail withdrawal latencies from 55°C water remained consistent over a period of 3 hr. Morphine produced dose-dependent increases in tail withdrawal latencies from 55°C water, whereas pentobarbital, haloperidol, and phencyclidine did not increase tail withdrawal latencies except at doses that produced marked sedation.

Key Words: Tail withdrawal; Analgesia; Morphine; Rhesus monkeys

INTRODUCTION

The tail withdrawal procedure that was originally developed by Janssen et al. (1963) has been used extensively to examine the effects of morphine-like compounds in rats. A number of studies (Van Bever et al., 1976; Sewell and Spencer, 1976; Tyers, 1980; Upton et al., 1982) have shown that morphine-like compounds produce dose-dependent increases in the time it takes rats to withdraw their tails from 55°C water. Similar increases in tail withdrawal latencies have been reported following morphine administration in squirrel monkeys (Genovese and Dykstra, 1985).

In most rodent tail withdrawal procedures, withdrawal latencies are only determined once or twice in each rat, thereby limiting the contribution of conditioning factors to the withdrawal response. Since it is not practical to use large numbers of monkeys in a procedure such as this, additional training is necessary to assure that the monkeys’ tail withdrawal response is dependent on the temperature of the water. It is also important to determine if the tail withdrawal response can be measured repeatedly in the same monkey, both within and between experimental sessions.

METHODS

Animals

Eight male or female rhesus monkeys (Macaca mulatta) weighing between 4.1 and 7.6 kg were used. All monkeys had been previously adapted to restraint chairs.
Monkeys' tails were shaved, as needed, to keep them free of hair. Monkeys were allowed free access to food and water in their home cages. In addition, their diet was supplemented with fresh fruit.

Apparatus

Five primate restraint chairs were used. These loosely restrained the monkeys at the neck and positioned them in a seated position from which their tails hung free, as shown in Figure 1. The monkeys' arms were also restrained. The restraint chairs were lined up against a wall, with the monkeys' backs and tails facing away from the wall. Adjacent to the restraint chairs was a controlled-temperature water bath. A wide-mouth, one-pint thermos was used to carry water from the water bath to the monkeys, and a digital timer was used to measure the reaction time.

Procedure

Monkeys were first placed into restraint chairs where they remained throughout the 3-hr experimental session. Daily experimental sessions were divided into several trials spaced 30 min apart. On the first trial of each experimental session, the lower 10–12 cm of each monkey's tail was immersed into a thermos of tap water (38–42°C). Only monkeys that kept their tails in tap water for at least 20 sec continued with the experimental protocol. On subsequent trials, the monkeys' tails were immersed
into water obtained from a water bath maintained at 55°C. Tail withdrawal latencies were determined on each trial. If a monkey did not remove his tail from the water within 20 sec, the tail was removed by the experimenter and the latency was assigned a value of 20 sec (20 sec cutoff time). Throughout the experiment, control probes with tap water were randomly interspersed between trials with 55°C water. If a monkey did not keep his tail in tap water for 20 sec during these probes, he was eliminated from the experiment for that day. Experimental sessions were run approximately twice a week, between 9 and 12 a.m. On days that experimental sessions were not run, monkeys were placed in the restraint chairs for at least 3 hr, and control probes with tap water were routinely administered. Of the eight monkeys used, two were only used in the initial control experiments and the remaining six were used for dose–effect determinations with each drug examined in at least four monkeys.

**Dose–Effect Determinations**

Dose–effect curves were obtained by administering cumulative doses of morphine (0.3–10.0 mg/kg), pentobarbital (3.0–30 mg/kg), haloperidol (0.03–0.56 mg/kg), or phencyclidine (0.1–1.0 mg/kg). In the cumulative dosing procedure, monkeys received a drug injection after each trial (i.e., once every 30 min). The amount of drug administered after each trial increased the cumulative dose by either ¼ or ½ log unit. In this way an entire dose–effect curve was generated for each drug in one day. All injections were subcutaneous in the back and delivered in a volume of 0.1 ml/kg.

**RESULTS**

**Control Performance**

In order to determine if tail withdrawal latencies were temperature dependent, monkeys’ tails were immersed into water maintained at different temperatures, and tail withdrawal latencies were determined. Figure 1 shows temperature–effect curves obtained from three different monkeys. It can be seen that the tail withdrawal latencies from 38–40°C water were at least 20 sec; withdrawal latencies decreased as the temperature of the water increased.

In order to determine if tail withdrawal latencies were consistent over time, monkeys’ tails were immersed in 55°C water once every 30 min over a period of 3 hr. Figure 2 shows time–effect curves obtained for three monkeys. Tail withdrawal latencies did not change as a function of time.

**Effects of Morphine**

Figure 3 shows the effects of increasing doses of morphine on tail withdrawal latencies from 55°C water. Mean control latencies in 55°C water were 2.3 (+0.21) sec. Morphine produced dose-dependent increases in tail withdrawal latencies. After tail withdrawal latencies were determined at the highest dose of morphine, 1.0 mg/kg of naloxone was administered. Thirty minutes after naloxone, tail withdrawal latencies were redetermined and found to be at control levels.
Effects of Phencyclidine, Pentobarbital, and Haloperidol

Pentobarbital (3.0–30 mg/kg) and haloperidol (0.03–0.56 mg/kg) generally did not increase tail withdrawal latencies. At the highest dose of pentobarbital (30 mg/kg), all monkeys appeared to be asleep; however, they awoke when their tails were immersed in water and withdrew them within 2–4 sec. Similarly, low doses of phencyclidine (0.1–0.3 mg/kg) did not increase tail withdrawal latencies; however, at 1.0 mg/kg of phencyclidine, tail withdrawal latencies exceeded the cutoff point of 20 sec, and the monkeys showed signs of marked sedation.

<table>
<thead>
<tr>
<th>TEMPERATURE (°C)</th>
<th>TAIL WITHDRAWAL LATENCY (SEC)</th>
<th>10 MG/KG MOR</th>
<th>1.0 MG/KG PCP</th>
</tr>
</thead>
<tbody>
<tr>
<td>55</td>
<td>&gt;20</td>
<td></td>
<td>20</td>
</tr>
<tr>
<td>58</td>
<td>14.0 (4.21)†</td>
<td>20</td>
<td>20</td>
</tr>
<tr>
<td>60</td>
<td>8.1 (4.74)</td>
<td>&gt;20</td>
<td>20</td>
</tr>
<tr>
<td>62</td>
<td>6.3 (3.61)</td>
<td>&gt;20</td>
<td>20</td>
</tr>
</tbody>
</table>

† Standard deviations are shown in parentheses; n = 4.
FIGURE 3. Tail withdrawal latencies in seconds from 55°C water following administration of increasing doses of morphine (mg/kg), administered cumulatively. The point at C represents the mean latency obtained prior to drug administration. Each point is the mean of one observation in at least four monkeys. Brackets indicate one standard error. The point at 1.0 Nx shows the effect of 10 mg/kg morphine in combination with 1.0 mg/kg naloxone.

Effects of Morphine and Phencyclidine at Different Temperatures

Table 1 shows the effects of 10 mg/kg of morphine and 1.0 mg/kg of phencyclidine on tail withdrawal latencies from water maintained at 55, 58, 60, and 62°C. In this part of the study, trials were 10 min rather than 30 min apart. The 10-mg/kg dose of morphine increased tail withdrawal latencies from 55°C water to 20 sec or more; however, this effect was reduced as the water temperature was increased. In contrast, the effects of 1.0 mg/kg phencyclidine were not attenuated at higher water temperatures.

DISCUSSION

The present report shows that the tail withdrawal procedure can be adapted for use with rhesus monkeys and that the same monkey can be used repeatedly within and between experimental sessions. This procedure requires very little training except for intermittent control probes with tap water to ensure that the tail with-
The tail withdrawal procedure is under control of the water temperature. Therefore, the tail withdrawal procedure provides a measure of analgesia that can be coordinated with other behavioral observations in the same monkey or used to monitor the development of drug-induced changes in analgesia that might occur as the result of tolerance or dependence.

Tail withdrawal latencies were shown to be temperature dependent and to remain consistent over periods up to 3 hr. Moreover, morphine produced dose-dependent increases in the time it took rhesus monkeys to withdraw their tails from 55°C water, and this effect was antagonized by naloxone. In contrast to the effects observed with morphine, neither pentobarbital nor haloperidol increased tail withdrawal latencies, and phencyclidine only increased them at doses that produced marked sedation. It is important to note that the doses of haloperidol, pentobarbital, and phencyclidine examined here were large enough to disrupt responding in other situations. For example, rates of food maintained responding in rhesus monkeys are either eliminated or markedly decreased following 0.01 mg/kg haloperidol (Woods et al., 1976), 0.32 mg/kg phencyclidine (Soloman et al., 1982), and 17.8 mg/kg pentobarbital (Herling et al., 1979).

The effects of morphine and phencyclidine could be further differentiated by examining their effects at different water temperatures. Although 10 mg/kg of morphine increased tail withdrawal latencies from 55°C water to 20 sec or more, this effect was reduced at higher temperatures. In contrast, the effects of 1.0 mg/kg of phencyclidine were not temperature dependent, suggesting that the monkey was not able to remove his tail from the water.

The data presented here are similar to those reported previously under the rat tail withdrawal procedure (Janssen et al., 1963). That is, the tail withdrawal procedure in rats is selective for morphine-like compounds with no increases in latency seen following pentobarbital, phencyclidine, and haloperidol, as well as a number of phenothiazines, anticholinergics, and psychomotor stimulants. This suggests that the tail withdrawal procedure can be used successfully to examine the analgesic effects of opioid-like compounds in monkeys.

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**REFERENCES**


