DIFFERING CALCIUM REQUIREMENTS FOR REGULATORY EFFECTS OF ATP, ATP $_{\gamma}$ S AND ADENOSINE ON 0 $_{\dot{7}}$  RESPONSES OF HUMAN NEUTROPHILS

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In formyl peptide stimulated human neutrophils the enhancement of  $0\frac{1}{2}$  responses by ATP and ATPyS requires extracellular calcium. In contrast, the inhibitory effects of adenosine are independent of a calcium requirement. Rates of  $0\frac{1}{2}$  generation are not affected by these adenine compounds. Rather, ATP and ATPyS cause a sustained period of generation whereas adenosine causes an abrupt early termination of the  $0\frac{1}{2}$  response. The differing calcium requirements for regulatory effects of adenine compounds on  $0\frac{1}{2}$  responses of stimulated neutrophils suggests that ATP (or ATPyS) and adenosine may exert their effects at different points in the pathway of signal transduction events. • 1988 Academic Press, Inc.

Superoxide anion  $(0\frac{1}{2})$  responses of human neutrophils stimulated by the chemotactic peptide, N-formyl-Met-Leu-Phe (fMLP) are enhanced by the presence of ATP or its analogues such as ATP $\gamma$ S (1,2) and inhibited by the presence of adenosine (3,4). The current studies were designed to assess if extracellular calcium is required for the regulatory effects of ATP, ATP $\gamma$ S and adenosine on  $0\frac{1}{2}$  responses of fMLP stimulated human neutrophils.

# MATERIALS AND METHODS

 $\frac{Reagents}{Sigma}$ . Unless otherwise indicated, all chemicals were purchased from  $\frac{Reagents}{Sigma}$ .

<u>Preparation of Neutrophils.</u> Venous blood was obtained from healthy adult humans, using a 1:6 ratio of acid-citrate dextrose and blood, yielding a

<u>Abbreviations</u> used are: HBSS, Hanks balanced salt solution; fMLP, N-formyl-Met-Leu-Phe; ATP $_{\gamma}$ S, adenosine 5',-0-(3-thiotriphosphate).

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final citrate concentration of 2.5 mM. Neutrophils were fractionated by Ficoll-Hypaque techniques as described in detail elsewhere (1). Generation of  $0\frac{1}{2}$  was determined by superoxide dismutase inhibitable reduction of ferricytochrome c (1).

Reactions Conditions. As indicated, neutrophils (5 x  $10^6/\text{ml}$ ) were incubated with  $25\mu\text{M}$  ATP, ATP $\gamma\text{S}$ , pr adenosine for 5 min at  $37^\circ\text{C}$  in HBSS containing Mg and, as indicated, Ca . Control groups were similarlly treated but in the absence of adenine compound. The cells were then washed once, resuspended in HBSS lacking calcium, and then placed into the reaction mixtures, to which 1.8mM calcium was added where indicated. The mixtures were warmed for 5 min at  $37^\circ\text{C}$  and then stimulated with fMLP and the  $0\frac{1}{2}$  responses measured. MgSO<sub>4</sub> (0.4mM) was constantly present in all salt solutions, unless otherwise indicated.

<u>Statistical Analysis</u>. Data were expressed as  $\bar{x}$  ± standard error of the mean (S.E.M.). A paired t-test was used to compare the response between two treatments. Statistical significance was defined as p<0.05. Specifics regarding the number of experiments, etc., are contained in the text or within each Table.

#### **RESULTS**

Contrasting Requirements of Calcium for Regulatory Effects of ATP, ATPyS and Adenosine on 05 Responses of Stimulated Neutrophils.

In order to determine if calcium is required for the enhancing effects of ATP and ATP $_{\rm Y}$ S on  $0\frac{1}{2}$  generation in fMLP stimulated neutrophils, experiments were carried out using 25 $\mu$ M ATP or ATP $_{\rm Y}$ S. The requirement for extracellular calcium was assessed employing Hanks buffered salt solution (containing 0.4mM Mg $^{++}$ ) in the presence or absence of 1.8mM calcium. When calcium was present both during exposure of neutrophils to ATP or ATP $_{\rm Y}$ S ("prewash" phase) as well as during exposure to fMLP ("postwash" phase), there was a statistically significant rise (25.8% and 38.1%) in the  $0\frac{1}{2}$  response (Table I).

If calcium was omitted during "prewash" cell exposure to ATP or ATP $_{Y}$ S but added in the subsequent "postwash" exposure of neutrophils to fMLP, there was no increase in the  $0\frac{1}{2}$  response. In contrast, if calcium was present when cells were exposed to ATP or ATP $_{Y}$ S but absent in the cell suspension when fMLP was added, there was a statistically significant enhancement (40.3% and 42.5%) of the  $0\frac{1}{2}$  response. Not surprisingly, in the continuous absence of calcium, not only was the  $0\frac{1}{2}$  response to fMLP blunted, no increment occurred in cells that had also been exposed to ATP or ATP $_{Y}$ S. These data indicate that the ability of ATP and ATP $_{Y}$ S to bring about enhanced  $0\frac{1}{2}$  responses in fMLP stimulated neutrophils requires the presence of extracellular calcium at the time of neutrophil contact with nucleotide.

Using a similar protocol, a companion series of studies was carried out to determine if the inhibitory effects of adenosine on  $0\frac{1}{2}$  responses in fMLP stimulated neutrophils require the presence of extracellular calcium. As shown in Table II, it is evident that the inhibitory effects of adenosine were independent of a requirement for extracellular calcium. In the con-

				Table I					
Requirement	of	Calcium	for	Enhancing	Effects	of	ATP	and	ATPγS
		on S	uper	oxide Resp	onses*				

material added during prewash**	presence of prewash	f calcium*** postwash	$0\frac{1}{2}$ production (nmol/30 min)	change (percent)
Exp. I				
HBSS	+	+	$21.3 \pm 4.7$	
ATP	+	+	26.8 ± 4.3	+25.8 (p<.004)****
HBSS	-	+	$21.1 \pm 4.1$	
ATP	-	+	$20.8 \pm 4.4$	-1.4 (p,N.S.)
HBSS	+	-	16.6 ± 3.4	
ATP	+	=	$22.3 \pm 4.1$	+40.3 (p<.03)
HBSS	-	-	$15.5 \pm 3.0$	
ATP	-	-	$15.2 \pm 2.6$	-1.9 (p,N.S.)
Exp. II				
HBSS	+	+	14.7 ± 1.70	
ATPγS	+	+	$20.3 \pm 2.62$	+38.1 (p<.008)
HBSS	-	+	15.9 ± 1.70	-
ATPyS	_	+	15.6 ± 1.53	-1.8 (p,N.S.)
HBSS	+	_	12.0 ± 1.49	***
ATPyS	+	-	17.1 ± 210	+42.5 (p<.003)
HBSS	-	-	$12.3 \pm 1.70$	
ATPyS	-	-	$12.1 \pm 1.63$	-1.6 (p,N.S.)

<sup>\*</sup>Conditions employed: 5 x  $10^5$  human blood neutrophils, 5 x  $10^{-7}$ M fMLP, 25  $\mu$ M ATP $\gamma$ S or ATP. In Experiment I, n=4; in Experiment II, n=8.  $0\frac{1}{2}$  data are expressed as  $\bar{x}$   $\pm$  S.E.M.

tinuing presence of calcium, adenosine caused a 33% inhibition in the  $0\frac{1}{2}$  responses. Omission of calcium in the salt solution when cells were exposed to adenosine ("prewash" phase) followed by addition of calcium when fMLP was

Table II

Lack of Requirement of Calcium for
Inhibitory Effects of Adenosine on Superoxide Responses\*

material added during prewash		of calcium postwash	05 production (nmol/30 min)	change (percent)
HBSS	+	+	10.0 ± 1.56	
adenosine	+	+	6.67 ± 1.34	-33.3 (p<.025)**
HBSS	_	+	$8.80 \pm 1.84$	-,
adenosine	-	+	5.14 ± 1.77	-41.5 (p<.05)
HBSS	+	_	$7.23 \pm 1.46$	-,
adenosine	+	-	5.09 ± 1.39	-29.5 (p<.035)
HBSS	-	-	$7.61 \pm 1.13$	•
adenosine	-	-	4.18 ± 0.68	-45.0 (p<.02)

<sup>\*</sup>Conditions employed:  $5\times10^5$  human blood neutrophils,  $5\times10^{-7}$ M fMLP,  $25~\mu$ M adenosine. The number of experiments was 6. Other experimental details are as described in Table I.

<sup>\*\*\*</sup>HBSS, Hanks balanced salt solution, Tris buffered, pH 7.4.

<sup>\*\*\*</sup>When present, the concentration of calcium was 1.8mM. "Prewash" refers to the first treatment of neutrophils (presence or absence of ATP or ATPyS) while "postwash" refers to time of addition of fMLP (in the absence of ATP or ATPyS).

\*\*\*\*For statistical comparisons, 0; values obtained by cell exposure to ATP or ATPyS were compared to the HBSS values in each of the pairs within the experimental groupings. N.S., not significant.

<sup>\*\*</sup>Paired t-test, comparing 0; values of cells subjected to the presence or absence of adenosine in each of the four pairs of data.

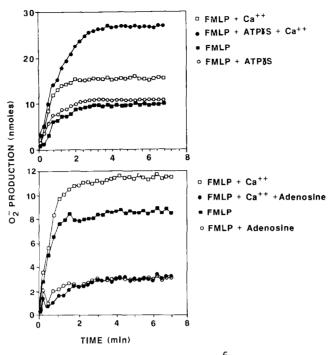


Figure 1: Upper frame, generation of  $0\frac{1}{2}$  by  $10^6$  human neutrophils stimulated at time 0 with 5 x  $10^7$  M fMLP in the presence or absence of 10  $\mu$ M ATP $_{Y}$ S, as a function of the presence or absence of 1.8mM extracellular calcium. Lower frame, similar studies employing 10  $\mu$ M adenosine in the presence or absence of calcium.

subsequently added ("postwash" phase) resulted in a 41.5% reduction of the  $0\frac{1}{2}$  response. The presence of calcium during cell exposure to adenosine but omission of calcium when fMLP was added to the cells resulted in a 29.5% reduction in the  $0\frac{1}{2}$  response. Finally, even in the continuing absence of calcium, adenosine was able to inhibit (by 45.0%) the  $0\frac{1}{2}$  response. Thus, the inhibitory effects of adenosine on  $0\frac{1}{2}$  responses of fMLP stimulated human neutrophils are independent of a requirement for extracellular calcium, in striking contrast to the calcium requirement for the enhancing effects of ATP and ATP $\gamma$ S.

Changes in Rates of  $0\frac{1}{2}$  Generation. Experiments were carried out to determine if ATP, ATP $_{Y}$ S and adenosine affect the rates of  $0\frac{1}{2}$  generation in the presence and absence of calcium. Representative data from four separate experiments are shown in Figure 1. In fMLP stimulated neutrophils, there was a prompt  $0\frac{1}{2}$  response which reached a plateau within 1.5 min. (Figure 1, upper frame). Using similar conditions in the presence of 10  $_{\mu}$ M ATP $_{Y}$ S, the initial rates of  $0\frac{1}{2}$  production were unaffected as defined by statistical analysis (data not shown), but the presence of ATP $_{Y}$ S caused a sustained production of  $0\frac{1}{2}$ , the plateau not being reached until approximately 3 min. When calcium was omitted from the salt solution, the total  $0\frac{1}{2}$  response in fMLP stimulated

cells was, as expected, greatly reduced, the plateau occurred earlier, and the presence of ATPyS was virtually without effect. Thus, the ability of ATP $\gamma$ S to enhance  $0\frac{1}{2}$  responses of fMLP stimulated neutrophils requires extracellular calcium and is related to sustained periods of production of  $0\frac{1}{2}$ . A similar pattern was found with ATP (data not shown).

Companion experiments were carried out with 10 µM adenosine, the results of which are shown in Figure 1, lower frame. As expected, calcium was required for the maintenance of the  $0\frac{1}{2}$  response in fMLP stimulated neutrophils. The addition of adenosine, whether in the presence or absence of calcium, led to an abrupt cessation of the 02 response. Although the initial rate of  $0\frac{1}{2}$  production was similar to that found with stimulated cells in the absence of adenosine (data not shown), the most remarkable effect of adenosine was to cause very early termination (in <1min) of the  $0\frac{1}{2}$  response, extracellular calcium not being necessary for this outcome.

## **DISCUSSION**

ATP and ATPyS may "prime" neutrophils for enhanced superoxide responses to fMLP in a manner that is analogous to the effect of bacterial endotoxin (5) or very low levels of either fMLP (6) or platelet activating factor (7). However, while ATP, per se, generates calcium transients (1,8) other agents such as bacterial endotoxin and phorbol ester prime neutrophils but do not usually produce measurable calcium transients. In one sense, the ability of ATP or ATP $\gamma$ S to prime neutrophils for enhanced  $0\frac{1}{2}$  responses might be analogous to the ability of calcium ionophore A23187 to alter neutrophils for enhanced  $0\frac{1}{2}$  responses when subsequently stimulated with phorbol ester (9). Nucleotides could alter fMLP receptors, the g-binding proteins, protein kinase C or the process of its translocation, or they might conceivably directly affect NADPH oxidase once the neutrophil has been activated. Alternatively, ATP and ATPYS could cause fusion of secondary granules to the cell membrane, resulting in an increase in numbers of fMLP receptors on the cell membrane. Since this fusion process is known to require the presence of extracellular calcium (10), the possibility that fMLP receptor adduction to the cell membrane is the explanation for the observed effects of the ATP compounds seems possible and is a current topic of study.

#### **ACKNOWLEDGEMENT**

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