Sequence Comparison of Wild-Type and Cold-Adapted B/Ann Arbor/1/66 Influenza Virus Genes

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Consensus sequences for both wt and ca B/Ann Arbor/1/66 viral PB2, PB1, PA, NP, M, and NS genes were directly determined from vRNA using a combination of chemical and chain-termination sequencing methods. There were 105 sites of difference between the wt and ca sets of these six RNA genes. The differences resulted in 26 amino acid substitutions distributed over the six proteins. The sequence changes were compared to the sequences of other known influenza type B wt viruses to pinpoint those changes that were unique to the ca B/Ann Arbor/1/66 virus. Of the 26 amino acid differences, only 11 were unique to the cold-adapted virus. These unique sites were distributed among five of the six genes. The NS protein had no amino acid substitutions. The sequence changes are discussed in terms of their probable mode of origin and selection, and in terms of their importance to the cold-adapted, temperature-sensitive, and attenuation phenotypes of ca B/AA/1/66 virus. The sequence and organization of the PB2 gene and predicted protein are also given. The PB2 gene was 2396 nucleotides long, and it encoded a predicted protein of 770 amino acids with a molecular weight of 88,035 Da for the wt virus and 88,072 Da for the ca virus. Both proteins were predominantly hydrophilic, and each had an overall charge of +24.5 at pH 7.0. © 1988 Academic Press, Inc.

INTRODUCTION

The cold-adapted (ca) B/Ann Arbor/1/66 (B/AA/1/66) influenza virus line was recently chosen for use in the production of ca reassortant live virus vaccines for testing in man (Maassab et al., 1985). The rationale behind the production of these ca reassortant vaccines is based on the experience amassed from successful trials using influenza type A ca reassortant vaccines. The optimal vaccine reassortant derives the viral genes encoding the two surface antigens, the hemagglutinin (HA) and the neuraminidase (NA), from the wild-type (wt) parent. Its other six genes are derived from the ca donor virus, and it is these genes which carry the characteristic ca, temperature-sensitive (ts), and attenuated (att) phenotypes. Thus, for the purposes of a safe and effective influenza A virus vaccine, all that is required is the assured presence of all six nonsurface genes from the ca A/Ann Arbor/60 donor virus (for review, Maassab et al., 1985). Laboratory studies have confirmed this phenotypic transfer to influenza B reassortant viruses by the six nonsurface genes of ca B/AA/1/66 in primary chick kidney (PCK) tissue culture and in ferrets (Maassab et al., 1986).

Human trials have been done and are being done to extend the validity of this approach for ca influenza B reassortant vaccine viruses (Davenport et al., 1977; Monto et al., 1982; Reeve et al., 1982; Keitel et al., 1986). While the transfer of these phenotypes is guaranteed by the transfer of all six nonsurface genes, the total number of mutations which brings about the ca, ts, and att phenotypes and their locations in the genome are not known. It is also not clear whether these phenotypes are engendered only by the mutations that were selected by the cold-adaptation process, or by other mutations selected in an unknown manner.

Initial studies used various methods to reveal that each of the eight genes of the ca A/Ann Arbor/6/60 (A/AA/6/60) donor virus displayed some difference from the corresponding gene of the wt A/AA/6/60 virus from which it was derived (Cox et al., 1986). Such genomic-wide changes are also probable with the influenza B virus system since just the electrophoretic comparison of the wt and ca B/AA/1/66 viral RNA segments demonstrated that changes occur in at least six of the eight genes (Maassab et al., 1985). Only the NA and NS ca and wt gene pairs did not migrate differently under the various conditions tested. Other studies for both influenza A and B viruses (Snyder et al., 1987; Donabedian et al., 1987) have used a genetic approach in an attempt to analyze the particular gene(s) associated with the three phenotypes. They

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isolated reassortants containing all wt influenza genes except for one gene from the ca parent virus. These "single-gene" reassortants were then analyzed for expression of the ca, ts, and att phenotypes. Single-gene analysis works reasonably well for the ts and att phenotypes, but less well for the ca phenotype since the ability to replicate efficiently at 25°, not a normal wt virus characteristic, can be inhibited by the presence of any wt gene that does not function well at this suboptimal temperature. In addition, some ca viral genes may act only in concert with one or more of the other ca donor virus genes to express these phenotypes, and hence, not be identified by single-gene studies.

In an effort to determine the total extent of the sequence changes that were selected in the six nonsurface genes during the cold-adaptation of B/AA/1/66 wt virus, we sequenced both the wt and ca B/AA/1/66 strains' PB2, PB1, PA, NP, M, and NS genes. These sequence data provide a complete catalog of the nucleotide changes that existed between these two virus lines, and these changed sites were compared to other known wt influenza B virus sequences to pinpoint changes unique to ca B/AA/1/66 viral genes. These data coupled with information derived from the single-gene reassortment studies mentioned above will provide a base of knowledge for future experiments which will attempt to link particular mutations to specific phenotypes. (See following article, Donabedian et al., 1988).

MATERIALS AND METHODS

Chemicals and enzymes

Oligodeoxynucleotide primers were purchased from the University of Michigan DNA Synthesis Facility or were made by one of the authors, C. W. Naeve, with an Applied Biosystems Model 380A DNA Synthesizer. The deoxynucleoside triphosphates, dimethyldichlorosilane, γ -(methacryloxy)propyltrimethoxysilane, diethyl pyrocarbonate (DEP), and sodium borohydride were purchased from Sigma Chemical Co., St. Louis, Missouri. The dideoxynucleoside 5'-triphosphates were obtained from Boehringer-Mannheim, Indianapolis, Indiana. Reverse transcriptase was the standard preparation derived from avian myeloblastosis virus by Life Sciences, Inc., St. Petersburg, Florida. Terminal deoxynucleotidyl transferase (TdT) was either from Life Sciences, Inc., or from Bethesda Research Laboratories, Gaithersburg, Maryland. T4 RNA ligase and T4 polynucleotide kinase were also from Bethesda Research Laboratories. Dimethyl sulfate and aniline were from Aldrich Chemical Co., Milwaukee, Wisconsin. Anhydrous hydrazine was purchased from Fisher Scientific Co., Fair Lawn, New Jersey. $[\alpha^{-32}P]$ deoxyadenosine 5'-triphosphate, tetra-(triethylammonium) salt was purchased from Dupont, NEN Research Products, Boston, Massachusetts, while $[\gamma^{32}P]$ adenosine 5'-triphosphate was from ICN Radiochemicals, Irvine, California.

Viruses and tissue culture

The viruses studied in these experiments were wt and ca B/AA/1/66 strains, and wt B/Melbourne/43 and B/AA/1/86 viruses. The cold-adaptation of the wt B/AA/1/66 virus together with the passage histories for both wt and ca viruses are given in Fig. 1. The original isolate was cultured in primary chick kidney cells from a human nasal wash sample (Maassab *et al.*, 1986). It was not cloned, and therefore the starting virus was a population of individual viruses probably of related but varying individual sequences. The production of viral stocks for the isolation of vRNA for sequencing was performed exactly as presented previously (DeBorde *et al.*, 1986).

Preparation of vRNA

The isolation of vRNA was performed as presented previously (DeBorde et al., 1986), except that the RNA from virus grown in 60–100 embryonated eggs was

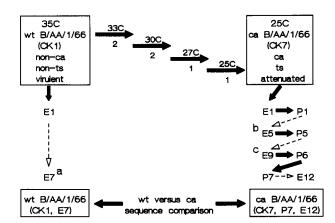


Fig. 1. Cold-adaptation and passage history of wt and ca B/AA/1/66 viruses. The cold-adaptation process is represented by the staggered arrows connecting the top two boxes. The passages were done in PCK cells at the temperatures indicated above each arrow. The number below the arrow represents number of passages at each temperature. The dashed arrows represent a number of replicative steps as follows: ^aConsecutive embryonated egg passages of the wt virus pool at high dilution. ^bConsecutive plaquing and egg amplification of the ca virus pool. ^cConsecutive embryonated egg passages of the ca virus pool at high dilution. The bottom two boxes represent the wt and ca virus pools that were sequenced in this paper.

resuspended in a final volume of 50 μ l, not 50 ml as erroneously stated in that paper.

End-labeling of vRNA

Purified vRNA (30 to 50 μg) and 100 μCi of [32P]cytidine bis-phosphate (pCp) were dried down together in a vacuum concentrator and resuspended in 5 μ l of 2× RNA ligase buffer, (100 mM HEPES, pH 7.5, 20 μg/ml BSA, 6.6 μM ATP, 30 mM MgCl₂), 1 μ l dimethylsulfoxide, and 1 μ I 0.5 mM ATP. Thirty to thirty-five pCp ligation units of T4 RNA ligase was added, and the total volume was adjusted to 10 µl with H2O if necessary. The reaction was incubated at 4° overnight. Two hundred microliters of 0.3 M sodium acetate was added to the ligation mixture, followed by 600 μ l of 95% ethanol, and the RNA was precipitated at -20° for at least 4 hr. The RNA was pelleted for 5 min in a microfuge, washed once with 70% ice-cold ethanol, pelleted once more, and dried in a vacuum concentrator. The RNA was resuspended in H₂O and mixed with an equal volume of loading buffer (0.2× TBE, 0.2% SDS, 20% sucrose, and 0.01% bromphenol blue).

Isolation of individual vRNA segments

For the chemical sequencing reactions, the purified and [32P]pCP 3' end-labeled vRNA was separated into its eight segments on a 3% polyacrylamide gel in 1X TBE, (0.09 M Trizma base, 0.09 M boric acid, and 2 mM Na₂EDTA) (Peacock and Dingman, 1968) with 0.01% sodium dodecyl sulfate (SDS). This level of SDS still protected the RNA from any contaminating RNases during the long electrophoretic run, but did not preclude visualization of the RNA with ethidium bromide. These conditions separated all segments except RNAs 4 and 5 (NP and HA). To separate these two RNAs, 6 M urea was added to the above gel. The RNA was electrophoresed at 37° for 16 hr at 200 V, constant voltage for the nondenaturing gel, and at 37° for 21 hr at 240 V, constant voltage for the 6 M urea gel. After electrophoresis the vRNA segments were visualized by immersing the gel in 1 liter of 1X TBE buffer containing 0.01% SDS and 1 μ g/ml ethidium bromide for 25 min at room temperature. The gel was then removed from the ethidium bromide solution, rinsed with 1× TBE buffer, and placed on a sheet of plastic wrap on a uv transilluminator. The desired RNA was excised from the gel with little or no excess gel material. The end-labeled vRNA segments were then eluted from the gel slices using an International Biotechnologies, Inc. (New Haven, CT), unidirectional electroeluter. The running buffer was 0.5× TBE buffer plus 0.01% sarcosine. The high salt retention buffer was 3.0~M sodium acetate with 0.03% sarcosine and 0.01% bromphenol blue. Elution proceeded at 100~V, constant voltage, for l-hr intervals. The smaller segments, NS and M, were completely eluted in 1 hr; NP, HA, and NA segments took 2 to 3 hr; and the three polymerase segments, PB2, PB1 and PA took 3 to 4 hr. Completion of elution was monitored by Cerenkov counting of the original gel slice and hourly samples. Recovery was 70 to 90% of the original sample. The isolated vRNA segments were precipitated at -20° overnight with 3 vol of 95% ethanol.

Chain-termination sequencing reactions and gels

These procedures, including the use of TdT to eliminate ambiguities, were performed exactly as described previously (DeBorde *et al.*, 1986). Only one compression area was encountered. It was in the PB2 gene and was resolved by running the standard sequence reaction products on a gel that was a 7 *M* urea, 8% polyacrylamide gel modified by the addition of 40% deionized formamide (Martin, 1987). To determine the wt and ca sequences, the sequencing reaction products were run side by side on the same electrophoretic gel. Several independent runs, with and without TdT enzyme treatment, were performed for each primer.

Chemical sequencing reactions and gels

These procedures were taken from Peattie (1979) and followed exactly, including the electrophoresis and autoradiography procedures. Recommendations given in Peattie (1979) for the safe use and disposal of sodium borohydride, aniline, DEP, and hydrazine were followed.

Sequence analysis programs

All sequence analyses, comparisons, manipulations, and calculations were performed using the programs developed by Queen and Korn (1984) and distributed by Beckman Instruments, Inc., Palo Alto, Calfornia, as the Microgenie Sequence Software package.

RESULTS

Figures 2-7 present the cRNA (plus strand) sequences and the predicted polypeptide sequences for the PB2, PB1, PA, NP, M, and NS genes. Each gene sequence was determined by a combination of chemical and chain-termination sequencing procedures (see Materials and Methods for details and/or

B/AA/1/66 ca PB2 cRNA (+) B/AA/1/66 wt PB2 cRNA (+) B/AA/1/66 wt PB2 Protein B/AA/1/66 ca PB2 Protein	5'
	AÁC ÀUÀ AUÀ AGA ÁAÁ UÚC ÁÁU ÁCÁ ÚCÁ ÁGÁ ÁUÚ GÁÁ ÁGG ÁÚC ÚÁU ÚÁÁ ÚÚÚÁ ÁGG ÁÚG ÁGG ÁÚG ÁGG GCC ÁUG ÚGÚ ÚCÚ ÁÁÚ ÚÚÚ CÉC ÚÚG GCÚ Asn Ile Ile Arg Lys Phe Asn Thr Ser Arg Ile Glu Arg Asn Pro Ser Leu Arg Met Lys Trp Ala Met Cys Ser Asn Phe Pro Leu Ala (60)
CUG ACC AAG GGU GAU AUG GCA Leu Thr Lys Gly Asp Met Ala	A (256). A du da
Val Thr Trp Trp Asn Thr Tyr	A (341) U (359). GGA CCC AUA GGA GAU ACU GAA AGG GUC GAA AAG GUC UAC GAA AGC UUU UUU CUC AGA AAG AUG AGA CUU GAC AAU GCC ACU UGG GGC CGA (425) Gly Pro Ile Gly Asp Thr Glu Gly Phe Glu Lys Val Tyr Glu Ser Phe Phe Leu Arg Lys Met Arg Leu Asp Asn Ala Thr Trp Gly Arg (134)
AUA ACU UUU GGC CCA GUU GAA Ile Thr Phe Gly Pro Val Glu	AGA GUG AGA AAA AGG GUA CUG CUA AAC CCC CUC ACC AAG GAA AUG CCU CCA GAU GAA GCG AGC AAU GUG AUA AUG GAA AUA UUG UUC CCU (536) Arg Val Arg Lys Arg Val Leu Leu Asn Pro Leu Thr Lys Glu Met Pro Pro Asp Glu Ala Ser Asn Val Ile Met Glu Ile Leu Phe Pro (171)
Lys Glu Ala Gly Ile Pro Arg	.A (572). GAA UCU ACU UGG AUG CAU AGG GAA CUG AUA AAA GAA AAA AGA GAA AAA UUG AAA GGA ACG AUG AUA ACC CCC AUU GUA CUG GCA UAC AUG (647) GTU Ser Thr Trp Met His Arg Glu Leu Ile Lys Glu Lys Arg Glu Lys Leu Lys Gly Thr Met Ile Thr Pro Ile Val Leu Ala Tyr Met (208)
CUU GAG AGA GAA CUG GUU GCC Leu Glu Arg Glu Leu Val Ala	CGA ÁGA ÁGG UÚC CUG CCA GUG GCA GGA GCA ÁCA ÚCA GCC GAG UÚC AUA GAA AUG CUA CAU ÚGC UÚA CAA GGÚ GAA ÁAÚ ÚGG ÁGA CÁA ÁUA (758) Árg Árg Árg Phe Leu Pro Val Ala Gly Ala Thr Ser Ala Glu Phe Ile Glu Met Leu His Cys Leu Gln Gly Glu Asn Trp Arg Gln Ile (245)
UAU CAC CCA GGA GGG AAU AAA Tyr His Pro Gly Gly Asn Lys	CUÁ ÁCU GÁA UCU ÁGG UCU CÁA UCA ÁUG ÁUU GUÁ GCU UGU ÁGA ÁAA ÁUA GUC ÁGA ÁGA ÚCA ÁUA GUC ÁGA ÚCA ÁAC CCA CUÁ GÁG CÚÁ GCU Leu Thr Glu Ser Arg Ser Gln Ser Met Ile Val Ala Cys Arg Lys Ile Val Arg Arg Ser Ile Val Ala Ser Asn Pro Leu Glu Leu Ála (282)
Val Glu Ile Ala Asn Lys Thr	U (899). GUG AUA GAC ACU GAA CCU UUA AAA UCA UGU CUG GCA GCC AUA GAC GGA GGU GAU GUA GCC UGU GAC AUA AUA AGA GCU GCA UUA GGA CUA (980) Val Ile Asp Thr Glu Pro Leu Lys Ser Cys Leu Ala Ala Ile Asp Gly Gly Asp Val Ala Cys Asp Ile Ile Arg Ala Ala Leu Gly Leu (319)
G (983) AAA AUC AGA CAA AGA Lys Ile Arg Gln Arg Gln Arg	
GGA AUA UGG GAC GGA GAA GAG Gly lle Trp Asp Gly Glu Glu	GAG UUC CAU GUA AGA UGU GGU GAA UGC AGG GGA AUA UUA AAA AAG AGU AAA AUG AGA AUG GAA AAA CUG CUG AUA AAU UCA GCC AAA AAG (1202) Glu Phe His Val Arg Cys Gly Glu Cys Arg Gly Ile Leu Lys Lys Ser Lys Met Arg Met Glu Lys Leu Leu Ile Asn Ser Ala Lys Lys (393)
GAG GAC AUG AAA GAU UUA AUA Glu Asp Met Lys Asp Leu Ile	ÂUC DUG DEC ÂUE GUA DUU DEU CAA GAC ÂCU ÂGE ÂUG DUC CAA GGA GUG ÁGA GGA GAA ÁUA ÁAU DU <u>U COU ÁAU CGA GCA GG</u> C CAA COU DUA (1313) Ile Leu Cys Met Val Phe Ser Gln Asp Thr Arg Met Phe Gln Gly Val Arg Gly Glu Ile Asn Phe Leu Asn Ârg Âla Gly Gln Leu Leu (430)
UCU CCA AUG UAC CAA CUC CAA	(1334). CGA UAU UUU UUG AAU AGG AGC AAC GAC CUU UUU GAU CAA UGG GGG UAU GAG GAA CCA CCC AAA GCA AGU GAA CUA CAU GGG AUA AAU GAA (1424) Arg Tyr Phe Leu Asn Arg Ser Asn Asp Leu Phe Asp Gin Trp Gly Tyr Glu Glu Pro Pro Lys Ala Ser Glu Leu His Gly Ile Asn Glu (467)
UUA AUG AAU GCA UCU GAC UAU Leu Met Asn Ala Ser Asp Tyr	ÁCG ƯƯG AÀA GGG GƯƯ GUÁ GUÁ ÁCA AÀA AÀU GUG ÁUU GAU GAU GÁC ƯƯƯ ÁGU UCU ÁCU GÁA ÁCA GÁA AÀA GUÁ ÚCU ÁUÁ ÁCÁ ÁÀÁ ÁÀU CỦÁ ÁGU (1535) Thr Leu Lys Gly Val Val Val Val Thr Lys Asn Val lle Asp Asp Phe Ser Ser Thr Glu Thr Glu Lys Val Ser Ile Thr Lys Asn Leu Ser (504)
	GUC AUA AUG GGG GCU AAU GAC GUA AGU GAA UUA GAA UCA CAA GCA CAG CUA AUG AUA ACA UAU GAU ACA CCU AAG AUG UGG GAG AUG GGA (1646) Val 11e Met Gly Ala Asn Asp Val Ser Glu Leu Glu Ser Gln Ala Gln Leu Met 11e Thr Tyr Asp Thr Pro Lys Met Trp Glu Met Gly (541)
Thr Thr Lys Glu Leu Val Gln	ÁAC ÁCC UÁC CÁA UGG GUG CỦA ÁAA ÁAU DUG GUA ÁCA CUG ÁÁG GCU CÁG DUU CUU CUG GGÁ ÁÁA GÁÁ GÁC ÁUG DÚC CÁÁ DGG GÁU GCÁ DÚÚ (1757) Asn Thr Tyr Gin Trp Val Leu Lys Asn Leu Val Thr Leu Lys Ala Gin Phe Leu Leu Gly Lys Glu Asp Met Phe Gin Trp Asp Ala Phe (578)
GAA GCA UUU GAA AGC AUA AUC Glu Ala Phe Glu Ser Ile Ile	CCC CAG AAG AUG GCU GGC CAG UAC AGU GGA UUU GCA AGA GCA GUG CUC AAA CAA AUG AGA GAC CAA GAG GUU AUG AAA ACU GAC CAG UUC (1868) Pro Gln Lys Met Ala Gly Gln Tyr Ser Gly Phe Ala Arg Ala Val Leu Lys Gln Met Arg Asp Gln Glu Val Met Lys Thr Asp Gln Phe (615)
AUA AAG UUG UUG CCU UUC UGU Ile Lys Leu Leu Pro Phe Cys	A (1913). UUC UCA CCA CCA AAA UUA AGG AGC AAU GGG GAG CCU UAU CAA UUC UUG AGG CUU AUG UUG AAG GGA GGA GGA GAA AAU UUC AUC AAA GUA (1979) Phe Ser Pro Pro Lys Leu Arg Ser Asn Gly Glu Pro Tyr Gln Phe Leu Arg Leu Met Leu Lys Gly Gly Gly Gly Asn Phe Tle Glu Val (652)
AGG AAA GGG UCC CCU CUA UUC Arg Lys Gly Ser Pro Leu Phe	.C (2003)
AUG GGG AAU GCA GUA UUG GCA Met Gly Asn Ala Val Leu Ala	GGC UUU CUC GUU AGU GGC AAG UAU GAU CCG GAU CUU GGA GAU UUC AAA ACU AUU GAA GAA CUU GAA AAG CUA AAA CCG GGG GAA AAA GCA (2201) Gly Phe Leu Val Ser Gly Lys Tyr Asp Pro Asp Leu Gly Asp Phe Lys Thr Ile Glu Glu Leu Glu Lys Pro Gly Glu Lys Ala (726)
AAC AUC UUA CUU UAU CAA GGA Asn Ile Leu Leu Tyr Gln Gly	ÁÁG CCC GUU ÁÁA GUÁ ÁGÚ ÁÁÁ ÁGÁ MÁÁ ÁGÁ MÁU ÁGÚ GCÚ MÚÁ MCC ÁÁÚ GÁU AUU MCÁ CÁÁ ÁGÁ ÁÚÚ ÁÁG ÁGÁ ÁGÁ ÁGÁ ÁGÁ ÁGÁ ÁGÁ Á
	UAAUAUAAAUUUAUCCAUUAAUUCAAUAAAUACAAUGGUGAAAAAUGGUGGUGUUUCUCAU (2396)

Fig. 2. PB2 gene. The can ucleotide sequence differences are presented above the wt sequence and the amino acid changes are presented below the wt sequence. Each site is followed or preceded by its position number in parentheses and, again, at the end of each line. The position (and unless otherwise noted) the sequence of each oligodeoxynucleotide primer is indicated by underlining of the wt sequence. A (—) in the sequence represents the deletion of the nucleotide in the corresponding sequence.

B/AA/1/66 ca PB1 B/AA/1/66 wt PB1 B/AA/1/66 wt B/AA/1/66 ca		N-terminus Met Asr	ı Ile Asn Pro Tyr Phe Le	ÚC ƯƯC AUA GAU GUA CCC AUA CAG GCA GC eu Phe Ile Asp Val Pro Ile Gln Ala Ai	la Ile Ser Thr Thr Phe Pro (23)
	Pro Pro Tyr Ser His G	ly Thr Gly Thr Gly Tyr	·Thr Ile Asp Thr Val Il	UÙ AGA AGA CAÙ GAG UAC UCA AAC AAG GI le Arg Thr His Glu Tyr Ser Asn Lys Gl	ly Lys Gin Tyr Ile Ser Asp (59)
Val Thr Gly Cys	Ala Met Val Asp Pro T	hr Asn Gly Pro Leu Pro	o Glu Asp Asn Glu Pro Se	GÙ GCC UAU GCA CAA UUG GAU UGC GUU CI er Ala Tyr Ala Gln Leu Asp Cys Val Le	eu Glu Ala Leu Asp Arg Met (95)
GAU GAA GAA CAU Asp Glu Glu His	Pro Gly Leu Phe Gln A	la Ala Ser Gin Asn Ala	; AUG GAG GCA CUA AUG GL Met Glu Ala Leu Met Va	UC ACA ACU GUA GAC AAA UUA ACC CAG GO al Thr Thr Val Asp Lys Leu Thr Gln Gl	ly Arg Gln Thr Phe Asp Trp (131)
Thr Val Cys Arg	Asn Gln Pro Ala Ala T	hr Ala Leu Asn Thr Thi	r Ile Thr Ser Phe Arg Le	UG AAU GAU UUG AAU GGA GCC GAC AAG GC eu Asn Asp Leu Asn Gly Ala Asp Lys G	ly Gly Leu Val Pro Phe Cys (167)
CAA GAU AUC AUU Gln Asp Ile Ile	Asp Ser Leu Asp Lys P	ro Glu Met Thr Phe Pho	e Ser Val Lys Asn Ile Ly	AG AAA AAA UUG CCU GCU AAA AAC AGA A ys Lys Leu Pro Ala Lys Asn Arg L	ys Gly Phe Leu Ile Lys Arg (203)
Ile Pro Met Lys	Val Lys Asp Arg Ile T	hr Arg Val Glu Tyr Ilo	E Lys Arg Ala Leu Ser Le	UA AAC ACA AUG ACA AAA GAU GCU GAA A eu Asn Thr Met Thr Lys Asp Ala Glu A	rg GTy Lys Leu Lys Arg Arg (239)
GCA AUU GCC ACC Ala Ile Ala Thr	GCU GGG AUA CAA AUC A Ala Gly Ile Gln Ile A	GA GGG UUU GUA UUA GU rg Gly Phe Val Leu Va	A GUU GAA AAC UUG GCU A/ I Yal Glu Asn Leu Ala Ly	.A (801)	GGU UUG CCA GUA GGU GGA (846) er Gly Leu Pro Val Gly Gly (275)
Asn Glu Lys Lys	Ala Lys Leu Ser Asn A	la Val Ala Lys Met Lei	ı Ser Asn Cys Pro Pro Gl	GA GGG AUC AGC AUG ACA GUG ACG GGA G ly Gly Ile Ser Met Thr Val Thr Gly A	sp Asn Thr Lys Trp Asn Glu (311)
Cys Leu Asn Pro	Arg Ile Phe Leu Ala M	et Thr Glu Arg Ile Th	r Arg Asp Ser Pro Ile Ti	GG UUC CGG GAU UUU UGU AGU AUA GCA C rp Phe Arg Asp Phe Cys Ser Ile Ala P	ro Val Leu Phe Ser Asn Lys (347)
Ile Ala Arg Leu	Gly Lys Gly Phe Met I	le Thr Ser Lys Thr Ly:	s Arg Leu Lys Ala Gin Ii	UA CCU UGU CCC GAU CUG UUU AAC AUA C le Pro Cys Pro Asp Leu Phe Asn 11e P	ro Leu Glu Arg Tyr Asn Glu (383)
Glu Thr Arg Ala	Lys Leu Lys Lys Leu L	ys Pro Phe Phe Asn Gli	ı Glu Gly Thr Ala Ser Lo	UG UCG CCU GGG AUG AUG AUG GGA AUG U eu Ser Pro Gly Met Met Met Gly Met P	he Asn Met Leu Ser Thr Val (419)
UUG GGA GUA GCC Leu Gly Val Ala	Ala Leu Glv Ile Lvs A	AC AUU GGA AAC AGA GA sn Ile Gly Asn Arg Gl	ı Tvr Leu Tro Aso Glv Le	ĐỘ CÁA ƯỢU ƯỢU GẦU GÂU LƯƯ GẦU CỦG Ủ eu Gln Ser Ser Asp Asp Phe Ala Leu P	he Val Asn Ala Lvs Asp Glu (455)
Glu Thr Cys Met	Glu Gly ITe Asn Asp P	he Tyr Arg Thr Cys Ly	s Leu Leu Gly Ile Asn M	UG AGC AAA AAG AAA AGU UAC UGU AAU G let Ser Lys Lys Lys Ser Tyr Cys Asn G	lu Thr Gly Met Phe Glu Phe (491)
ACA AGC AUG UUC Thr Ser Met Phe	UAC AGA GAU GGA UUU G Tyr Arg Asp Gly Phe V	VA UCU AAU UUU GCA AU al Ser Asn Phe Ala Me	G GAA CUU CCU UCA UUU GI	.A (1557). GGG GUU GCU GGA GUA AAU GAA UCA GCA G GIY Val Ala GIY Val Asn Glu Ser Ala A	AU AUG GCA AUA GGA AUG ACA (1602) sp Met Ala Ile Gly Met Thr (527)
Ile Ile Lys Asn	Asn Met Ile Asn Asn G	GG AUG GGU CCA GCA AC ly Met Gly Pro Ala Thi	r Ala Gln Thr Ala Ile G'	ÀÀ UÙÀ ƯƯC AVÀ GCỦ GAU ỦAU ÁCA VÁC Á 11n Leu Phe Ile Ala Asp Tyr Arg Tyr T	hr Tyr Lys Cys His Arg Gly (563)
Asp Ser Lys Val	Glu Gly Lys Arg Met L	ys lie lie Lys Glu Le	A UGG GAA AAC ACU AAA G u Trp Glu Asn Thr Lys G	GA AGA GAU GGC CUG UUA GUA GCA GAU G ily Arg Asp Gly Leu Leu Yal Ala Asp G	ly Gly Pro Asn Ile Tyr Asn (599)
Leu Arg Asn Leu	His Ile Pro Glu Ile V	al Leu Lys Tyr Asn Le	u Met Asp Pro Glu Tyr Lj	AÀ GGG CGG UNA CUG CAU CCU CAA AAU C ys Gly Arg Leu Leu His Pro Gln Asn P	ro Phe Val Gly His Leu Ser (635)
lie Glu Giv Ile	Lvs Glu Ala Asp Ile I	hr Pro Ala His Glv Pr	o lie Lvs Lvs Met Asd i	AN GAN GCG GUA NCU GGA ACU CAN AGU N yr Asp Ala Val Ser Gly Thr His Ser T	rd Arg inr Lvs Arg Asn Arg (6/1)
Ser Ile Leu Asn	Thr Asp Gln Arg Asn M	et Ile Leu Glu Glu Gl	n Cys Tyr Ala Lys Cys C	GC AAC CUU UUU GAG GCU UGU UUU AAC A ys Asn Leu Phe Glu Ala Cys Phe Asn S	er Ala Ser Tyr Arg Lys Pro (707)
Val Gly Gln His	Ser Met Leu Glu Ala M	let Ala His Arg Leu Ar	g Met Asp Ala Arg Leu A	AU UAU GAA UCA GGA AGA AUG UCA AAG G Isp Tyr Glu Ser Gly Arg Met Ser Lys A	sp Asp Phe Glu Lys Ala Met (743)
GCU CAC CUU GGU Ala His Leu Gly	GAG AUU GGG CAC AUA U Glu Ile Gly His Ile (751) Tyr	AAGCUUCGAAGAUGUCUAUGG C-terminus (752) C-terminus (752)	GGUUAUUGGUCAUCAUUGAAUAC	AUGCGGUACACAAAUGAUUAAAAUGAAAAAAGGCUC	3' (2369) GUGUUUCUACU-3' (2369)

Fig. 3. PB1 gene. The ca and wt sequences are presented as in Fig. 2, with identical conventions for numbering and primer location and sequences.

B/AA/1/66 ca B/AA/1/66 wt B/AA/1/66 wt B/AA/1/66 ca	cRNA (+) 5 PA Protein	S'-AGCAGAAGCG	GUGCG <u>UUUGAUUI</u> N-te	rminus Met		r Phe	Ile Th	A AGA r Arg	Asn Ph	C CAG . e Gln	ACU ACA Thr Thr	He	lle Glr	Lys						
GAA UUU AGU G Glu Phe Ser G																				
AAA ACA UAU A Lys Thr Tyr T	Thr Ala Leu	Glu Gly Gln		iln Asn Lei	ı Arg Pr	o Gln	Tyr G1	u Val	Ile Gl	u Gly :	Met Pro	Arg .	Asn Ile	Ala	Trp M	et Val	Gln	Arg .	Ser	
UUA GCC CAA G Leu Ala Gln G		Ile Glu Thr		eu Ala Asp	Leu Ph	e Asp	Tyr Ly	s Thr	Lys Ar	g Phe	Ile Glu	٧a١	Gly Ile							
UAC UUU UGG A Tyr Phe Trp L	AAA AAG AAA .ys Lys Lys	Glu Lys Leu	GGG AAU AGC / Gly Asn Ser I	let Glu Lei	Met Il	e Phe	Ser Ty	r Asn	G1n As	p Tyr	Ser Leu	Ser	AAU GA/ Asn Gli	CAC	Ser L	JG GAU eu Asp	GAG G1 u	G1 u	G1y	(527) (166)
AAA GGG AGA G Lys Gly Arg V		Arg Leu Thr		lla Glu Lei	ı Ser Le	u Lys	Asn Le	u Trp	Gln Va	1 Leu	Ile Gly	Glu	Glu Asp	lle:						
AAA CUU GGA C Lys Leu Gly G	iln Thr Ile	Ser Lys Leu	AGG GAC AUA I Arg Asp Ile S	icu guu cc <i>i</i> Ser Val Pro	A1a G1	y Phe	Ser As	n Phe	Glu Gl	y Met	Arg Ser	Tyr	Ile Asp	Asn	Ile A	sp Pro	AAA	GGA G1y	GCA Ala	(743) (238)
AUA GAG AGA A ITe Glu Arg A	Isn Leu Ala	Arg Met Ser	CCC UUA GUA (Pro Leu Val	er Val Thi	· Pro Ly	s Lys	Leu Ly	s Trp	Glu As	p Leu	Arg Pro	He	Gly Pro	His	Ile T	∕r Ser	His	G1 u	Leu	(851) (274)
CCA GAA GUU C Pro Glu Yal F	CA UAU AAU Pro Tyr Asn	GCC UUU CUU Ala Phe Leu	CUA AUG UCU (Leu Met Ser /	AU GAG UUG Isp Glu Lei	GGG CU Gly Le	G GCU u Ala	AAU AU Asn Me	G ACU t Thr	GAA GG Glu Gl	G AAG y Lys	UCC AAG Ser Lys	AAA Lys	G (93 CCA AAG Pro Lys	S5) ACC Thr	UUA G Leu A	C AAA la Lys	GAA G1u	UGU Cys	CUA Leu	(959) (310)
GAA AAG UAC L Glu Lys Tyr S				le Leu Ile	Met Ly	s Ser	Glu Ly	s Ala	Asn G1	u Asn	Phe Leu	UGG Trp		UGG	AGG G	AC UGU				
AUA AGU AAU G Ile Ser Asn G																				
GAC GAA ACA A Asp Glu Thr N	UG VAC CAA let Tyr Gln	GAA GAG CCC Glu Glu Pro	AAA AUA CCU /	.C (1214). AU AAA UGU Isn Lys Cys	I AGA GU	G GĆŪ 1 Ala	GCU UG Ala Tr	G GUU p Val	CAA AC Gln Th	A GAG r Glu	AUG AAU Met Asn	CUA Leu	UUG AGO Leu Sei	ACU Thr	CUG A	A AGU nr Ser	AAA Lys	AGG Arg	GCC Ala	(1283) (418)
CUG GAU CUA C Leu Asp Leu F	ro Glu Ile	Gly Pro Asp	GUA GCA CCC (Val Ala Pro	al Glu His	: Val G1	y Ser	Glu Ar	g Arg	Lys Ty	r Phe	Val Asn	G1 u	Ile Ası	ı Tyr	Cys L	ys Ala	UCU Ser	ACC Thr	GUU Val	(1391) (454)
AUG AUG AAG L Met Met Lys T	JAU GUA CUU [yr Yal Leu	UUU CAC ACU Phe His Thr	UCA UUA UUA / Ser Leu Leu /	AU GAA AGO Isn Glu Sei	AAU GC Asn Al	C AGC a Ser	AUG GG Met G1	A AAA y Lys	UAU AA Tyr Ly	A GUA s Val	AUA CCA Ile Pro	AUA Ile	ACC AAC Thr Asi	AGA Arg	GUA G Val V	JA AAU al Asn	GAA Glu	AAA Lys	GGA G1y	(1499) (490)
GAA AGU UUU G Glu Ser Phe A	sp Ile Leu	UAU GGU CUG Tyr Gly Leu	Ala Val Lys (lly Gln Sei	· His Le															
AGA GUG GAC U Arg Val Asp S	ICA GGA AAG Ser Gly Lys	UGG CCA AAA Trp Pro Lys	Tyr Thr Val I	IUU AGA AUL	Gly Se	C UÚA r Leu	UUU GU Phe Va	1 Ser	Gly Ar	g Glu	Lys Ser	٧a١	Tyr Lei	UAU Tyr	UGC C Cys A	GA GUG rg Val	AAU Asn	GGU G1y	ACA Thr	(1715) (562)
U (1718) AAC AAG AUC O Asn Lys Ile G	iln Met Lys	Trp Gly Met	Glu Ala Arg /	irg Cys Let	ı Leu G1	n Ser	Met G1	n Gln	Met G1	u Ala	Ile Val	Asp	Gln Glu	Ser	Ser I	le GTr	GTy	UAU Tyr	GAC Asp	(1823) (598)
AUG ACC AAA G Met Thr Lys A	lla Cys Phe	Lys Gly Asp	AGA GUG AAU / Arg Val Asn S	ier Pro Lys	Thr Ph	e Ser	Ile G1	y Thr	G1n G1	u G1y	Lys Leu	٧al	Lys G1 ₃	Ser	Phe G	ly Lys	Ala	Leu	Arg	(1931) (634)
GUA AUA UUC A Val Ile Phe T	hr Lys Cys	Leu Met His	UAU GUA UUU (Tyr Yal Phe (ily Asn Ala	G1n Le	u G1u	Gly Ph	e Ser	A1a 61:	u Ser	Arg Arg	Leu	Leu Lei	Leu	Ile G	In Ala	Leu	Lys	Asp	(2039) (670)
AGA AAG GGC C Arg Lys Gly P	ro Trp Val	Phe Asp Leu	GAG GGA AUG E Glu Gly Met l	yr Ser Gly	· Ile Gl	u Glu	Cys Il	e Ser	Asn Asi	n Pro	Trp Val	Ile :	Gln Ser	Ala	Tyr Ti	p Phe	Asn	Glu	Trp	(2147) (706)
BUG GGC DUU G Leu Gly Phe G	AA AAA GAG Glu Lys Glu	GGG AGU AAA Gly Ser Lys	GUA UUA GAA I Val Leu Glu S	CA AUA GAU Ger Ile Asp	GAA AU Glu Il	A AUG e Met	GAU GA	A UGAA u C-te	AGAAGG erminus	GCAUAG (726)	CGCUCAA	G UUUGA	. (2235) Jacuaul	เบ้นดีบัน	ĊĂŮŮĀI	JGUAUC	ÜÄÄÄ	ĊĀŪĆŪ	ĊĀĀ	(2270)
AAAAAGAAUUGAG	AAUUAAAAUG	CACGUGUUUCUA	ců-3' (2308)																	

Fig. 4. PA gene. The ca and wt sequences are presented as in Fig. 2, with identical conventions for numbering and primer location and sequences.

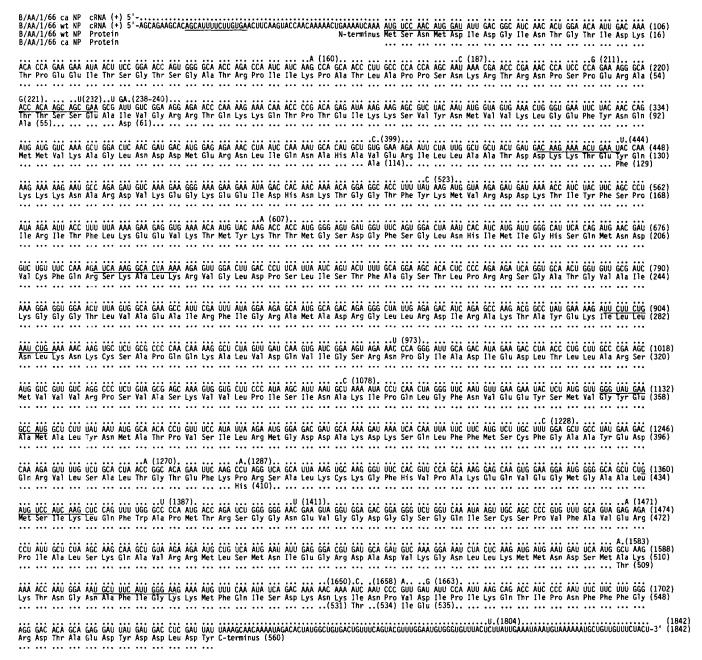


Fig. 5. NP gene. The ca and wt sequences are presented as in Fig. 2, with identical conventions for numbering and primer location, with one exception. The primer sequence starting at position 221 is actually ACC ACA AGC AGT GAA (see text).

references). Both methods were applied directly to vRNA isolated from purified virus. Thus, the determined sequences represent consensus sequences based on the population of vRNA segments. The chemical sequencing method identified the first 50–90 nucleotides of each gene and overlapped the first primer-determined sequence by at least 15 nucleotides. Therefore the sequences on the 5' side of the first primer positions were determined exclusively by the direct chemical sequencing method. Oligo-

deoxynucleotide primers were usually 15 nucleotides in length and the sequence from each preceding primer overlapped the next determined sequence by at least 15–20 nucleotides. Their positions in the sequences are shown by underlining of the wt cRNA sequence of each gene. Only the differences are presented for the ca RNA and predicted ca polypeptide sequences, and these are shown above and below the corresponding wt sequences, respectively. The results for each individual gene are presented below.

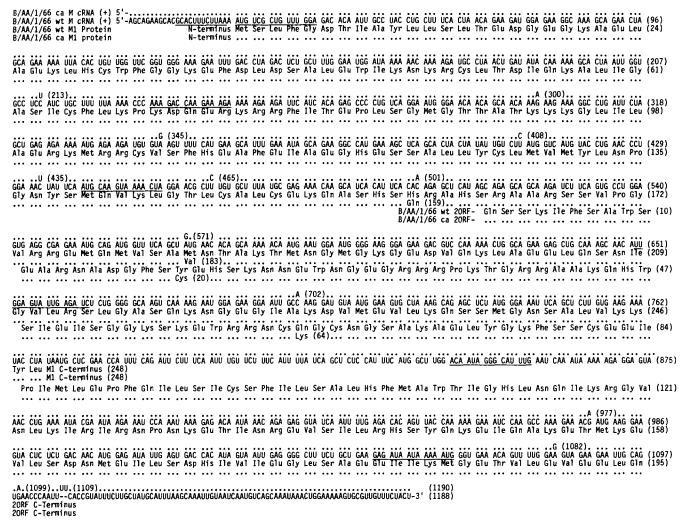


Fig. 6. M gene. The ca and wt sequences are presented as in Fig. 2, with the addition the second open reading frame (2 ORF) presented as two additional lines as indicated in the figure.

PB2 gene

Both wt and ca influenza B/AA/1/66 PB2 genes were 2396 nucleotides long. They encoded polypeptides of 770 amino acids. The first methionine codon in the open reading frame began at position 24 (in from the 5' end of the cRNA) and the first termination codon began at position 2334. A consecutive run of five adenosine residues began at position 2375 and was the probable polyadenylation site. The predicted wt and ca PB2 proteins had molecular weights of 88,035 and 88,072 Da, and both had a net charge of +24.5 at pH 7.0, with no large hydrophilic domains. The ca PB2 gene varied from the wt gene by 26 base substitutions. Of these changes, 24 occurred in the coding region. Only 4 of these changes were reflected in amino acid changes. Of these 4 changes, 1 was unique to the ca PB2 gene,

and 3 were indentical to sequences noted in other wt influenza B virus PB2 genes (B/Melbourne/43 and B/AA/1/86), see Fig. 8. A reassortant containing the PB2 gene of ca B/AA/1/66 virus, and all other genes from a wt virus, B/Houston/76, is non-ts, non-ca, and as virulent as the wt virus (Donabedian *et al.*, 1987). Hence, the PB2 gene of the ca donor virus apparently contributed to none of the three phenotypes of interest in an independent manner. It is possible that it may be involved in any of these phenotypes in combination with one or more other ca B/AA/1/66 genes, especially those involved in the replication complexes.

PB1 gene

The general characteristics of the influenza B PB1 gene have been previously described by Kemdirim and

colleagues (1986) for B/Lee/40 virus and by DeBorde and colleagues (1987) for B/AA/1/66 wt virus. The ca B/AA/1/66 PB1 gene varied from the wt PB1 gene by 21 base substitutions all within the coding region of the gene, which resulted in a total of three predicted amino acid changes. By comparison of the ca B/AA/1/66 gene sequence to that of B/Lee/40 (Kemdirim *et al.*, 1986) only one of these amino acid changes remained unique to the cold-adapted virus (Fig. 8). From the reassortment analysis of Donabedian and colleagues (1987), PB1, like PB2, apparently played no significant independent role in the establishment of the three phenotypes.

PA gene

The PA gene sequence has been reported for wt B/AA/1/66 virus (DeBorde *et al.*, 1987) and is also known for B/Singapore/79 (Akoto-Amanfu *et al.*, 1987). Our laboratory has also sequenced the PA gene of B/Houston/76 virus. The ca PA gene varied from the

wt gene by 17 base substitutions, one of which was in the cRNA 3'-noncoding region. The 16 changes in the coding region resulted in six amino acid changes between the wt and ca PA polypeptides. Comparison with the PA sequences of B/Houston/76 and B/Singapore/79, showed that only two of the six amino acid changes were unique to the ca B/AA/1/66 virus. As shown in the following article (Donabedian *et al.*, 1988), the codon of one of these unique positions (position 1320) underwent a change resulting in the reversion of both ts and att phenotypes.

NP gene

The NP gene has the largest number of base substitutions per nucleotide of any of the six gene pairs examined. Of the 25 changes in the RNA, 24 of them were in the coding region, accounting for nine amino acid changes from wt B/AA/1/66 NP protein to ca B/AA/1/66 NP protein. Comparison of these nine changes with the known NP gene sequences of

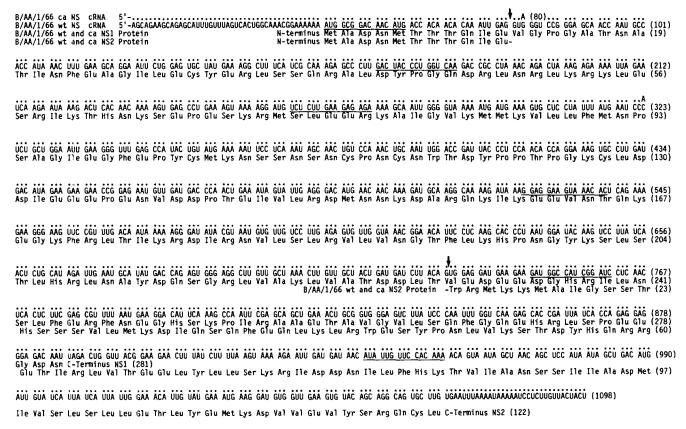


Fig. 7. NS gene. The two nucleotide substitutions in the cavirus NS gene sequence are shown above the wt gene sequence. Because neither base change resulted in an amino acid change, the ca and wt NS1 and NS2 proteins are shown together rather than separately as in the other sequence figures. The arrows represent the presumed splicing sites based on the sequence data given for B/Lee/40 virus (Briedis and Lamb, 1982).

#	PB2 Gen	a AA #	Pos.	PB1 Gen Lee wt	ca #	Pos.	PA G	a HN	Sing	#	Pos.	NP G Lee w	t ca S	ing #	Pos.	M G Lee w	t ca	Sing
1	1913 AGC AGC A			AUA AUA Ile Ile		1320		AUG GUG Met Val		1			CC GCC A hr Ala T			CAU C.		
2	2362 A A 3'-NONC	G A ODING 2		AAA (AGA	[AAA]	1518		CAU UAU lis Tyr		2			UG GCG G al Ala V			AUG A Met M		
3	1334 CAA CAA CA Gln Gln G		2272	Lys Arg	UAC 3			<u>icc</u> ucc		3			CU CAU C			UAU U Tyr T		
4	256 CAA CGA C Gln Arg G		345	Tyr His CAG CAA Gln Gln	CAG 5		AGC A	Ser Ser AAC AAC Asn Asn	AAC	4			CU ACU G la Thr A			บCบ บ Ser S		
5	828 AUC GUC A	UC AUC 5	399	AGA AGG Arg Arg	AGA 6	1514	AUA A	AUG AUG	AUG	5			CU GCC G LA ALA A		465	CUC C	UU CUC	cuc
6	50 UUA UUG U Leu Leu L	UA UUA 6	537	GAU GAC Asp Asp	GAU 7	1796	GAU (GAA GAA Glu Glu	GAA	6	1804		C U on-codin			ACA A Thr T		
7	626 ACU ACC A			AAA AAG Lys Lys		53		AAC AAC		7 8			UU GAU G le Asp A			GAG G Glu G		
8	1008 AGA AGG C			ACA ACG Thr Thr		668	GAC (Asn Asn GAU GAU	GAU	9			AU GAG G sp Glu G		1107	? UU ~	rtion - UU noncod	
9	1052 GAU GAC G Asp Asp A			AAU AAC Asn Asn		791	CCC (Asp Asp CCU CCU Pro Pro	CCU	10	1658		UU AUU G al Ile V		300	AAG A		
10	1088 AAG AAG A Lys Lys L			UAC UAU Tyr Tyr		800	UUA L	JUG JUG Leu Leu	UUG	11			AU AAC A sn Asn A	\AC	Ml	Lys L	ys Lys	s Lys
11	2003 UCC UCU U Ser Ser S			GGA GGG Gly Gly		935	CCA (CCG CCG Pro Pro	CCG	12		Ser S	GC AGU A er Ser S	Ser 10	M1	VAL V	AL VAL	_ VAL
12	572 AUG AUG A Met Met I	UA AUA 1e I1e		UAC UAU Tyr Tyr	Tyr 13	1044		CUG CUG Leu Leu				Phe P	UU UUC L he Phe P	he 11	435	Leu L	AC AAL	JAAU
13	1160 GGC AGU A	GC AGC		GGU GGC Gly Gly	Gly 14	1214		AAC AAC Asn Asn				Asn A	AC AAU A sn Asn A	lsn 12	702	Asn A	CC GC/	A GCA
14	341 CCC CCC C Pro Pro P			ACA UCA Thr Ser		1655		GGC GGC Gly Gly					AG GAA G lu Glu G		702	CAA C Gin G	AA AA	AAA
15	359 GGG GGG G GLY GLY G		822	GAG GAG Glu Glu		1718		AAU AAU Asn Asn		16	444	UAC (U	AC UUC L yr Phe F	JUC Phe 14	1099	UGA U	GA UA/	A UAA
16	476 CCC CCC C Pro Pro P			GGA GGA Gly Gly			3'-	G G -NONCOD	ING	17	1650		UC ACC A le Thr 1					
17	728 CAU CAU C His His H			AGU AGU Ser Ser					,	18			AG AAA G ys Lys A	\rg		NS Lee w	Gene	
18	899 GAC GAC G Asp Asp A	sp Asp		ACG ACG Thr Thr	Thr					19		Thr A	CG GCU G la Ala A	GCU == Ala I	 80	GUG G Val V	UG GU	A GUG
19	983 AAA AAA A Lys Lys L	ys Lys		AUU AUU Ile Ile	: Ile					20		Pro P	CA CCG C ro Pro F	Pro 2		ccc c		
20	1058 GAG GAG G	lu Glu		GCU GCU Ala Ala	Ala					21		Tyr T	AU UAC L yr Tyr 1	fyr -		Pro P		
	1184 UUG CVG C Leu Leu L	eu Leu 		CUG UUG Leu Leu						22		Lys L	AG AAA A ys Lys L	.ys				
	2081 AGA AGA A Arg Arg A	irg Arg										Asn A	AC AAU A sn Asn A CC ACA A	\sn				
23	2138 GAU GAU G ASP ASP A 2141 CCG CCG C	sp Asp										Thr T	hr Thr 1 CC GCU G	Thr .				
	Pro Pro P	ro Pro									1507		la Ala A					
26	Asp Asp A	isp Asp																
	3'-NONC	ODING																

Fig. 8. Unique and nonunique wt to ca changes. The sequence differences between the wt and ca B/AA/1/66 nonsurface genes are compared to sequences of other known influenza type B viruses. They are presented as part of the codon of which they are a member, rather than just a single nucleotide difference. The encoded amino acid is listed below the codon. Below each gene the sites are arranged as follows: changes unique to ca B/AA/1/66 followed by changes that match the sequence in one or more other influenza type B virus. These nonunique changes have been further grouped by matching to one or another known viral sequence. The columns labeled Pos. refer to the nucleotide position of the change as found in Figs. 2–7. Those nucleotide differences that result in amino acid differences have been enclosed in boxes. The viruses listed are as follows: Mel, B/Melbourne/43; Lee, B/Lee/40; HN, B/Houston/76; Sing, B/Singapore/222/79; AA, B/Ann Arbor/1/86; wt, wt B/AA/1/66; and ca, ca B/AA/1/66.

B/Lee/40 (Briedis and Tobin, 1984) and B/Singapore/72 (Londo *et al.*, 1983) left only four of these changes as unique to the ca NP gene (Fig. 8).

The primer sequence starting at position 221 is actually ACC ACA AGC AGT GAA. Thus its sequence varies from that of the wt sequence at position 232, and from that of the ca sequence at position 221. This primer was made based on B/Lee/40 sequences, but despite the one nucleotide difference for each B/AA/1/66 virus, it worked well. (All other primers which were made prior to actual B/AA/1/66 viral sequence knowledge and which were later shown to vary by even one nucleotide, would not function.)

M gene

There was a total of 12 nucleotide substitutions between wt and ca B/AA/1/66 M RNA. In addition, the ca B/AA/1/66 M gene had an insertion of two uridine nucleotides within positions 1107-1110. The insert position was ambiguous due to the presence of two uridine nucleotides at positions 1107-1108 in the wt B/AA/1/66 M gene. This insertion was in the 3'-noncoding region for M₁ protein and was also beyond the conserved second open reading frame (2 ORF) previously described for both B/Lee/40 (Briedis et al., 1982) and B/Singapore/222/79 (Hiebert et al., 1986) M genes. In fact, both of these viruses have a run of four uridine nucleotides at this position like the ca M gene. There were two amino acid changes in the M₁ protein region, and two predicted amino acid changes in the possible product of the 2 ORF. None of the M₁ changes are found in the other known influenza B M gene sequences, while one of the changes in the 2 ORF is also seen in B/Singapore/222/79 M gene (Hiebert et al., 1986). Thus, at this time only the two changes in the M₁ protein and possibly one in the 2 ORF appeared to be unique to the cold-adapted virus (Fig. 8).

NS gene

The NS gene had the lowest number of changes between wt and ca B/AA/1/66 viruses. Only two base substitutions were present, both exclusively in the NS₁ coding region, and neither of these resulted in amino acid changes in the NS₁ protein. One change, at position 80, occurred in the 5'-donor-splice region, but it was an acceptable change based on the consensus signal from Mount (1982). Unless the wt NS gene itself was inherently attenuating, cold-adapted, or temperature-sensitive, we would predict that this gene contributes little to the overall phenotypes of the ca B/AA/1/66 virus.

Unique changes of ca B/AA/1/66 virus

The total nucleotide and amino acid changes between the six nonsurface genes of wt and ca B/AA/1/ 66 viruses are given in Fig. 8 showing a comparison to the corresponding site in other known influenza B sequences. Except for those changes which occur in the noncoding regions of the genes, the sites are presented in their codon, with their encoded amino acid. Where possible the changes have been divided into groups: (1) those nucleotide changes that were unique to ca B/AA/1/66 virus (the top group for each gene), and (2) those changes that were also seen in one or more other wt viruses (the bottom two groups for each gene). Sequences for PB2, NP, M, and NS genes have been determined from viral isolates that occurred both earlier and later in time than the B/AA/1/66 viruses. Only one other sequence for the PB1 gene has been reported to which B/AA/1/66 viral PB1 sequences can be compared, and it was determined from an earlier virus isolate (B/Lee/40). Therefore, the group of changes, PB1 positions 14-21, may contain some unique ca nucleotide changes even though no amino acid change results. The PA gene has two other PA sequences to which it can be compared, but both genes are from later viral isolates. The nucleotide changes that result in amino acid changes are enclosed in boxes. Out of 15 unique nucleotide changes, 10 of these resulted in 11 unique amino acid changes. (One change in the M gene caused a unique amino acid change both in M₁ and in the 2 ORF.) The 89 remaining nonunique nucleotide changes resulted in only 15 additional amino acid changes.

Total changes

Table 1 presents a summary of the total numbers of nucleotide and amino acid changes that were selected in the cold-adaptation of wt B/AA/1/66 virus. The lack of changes occurring in the 5'-noncoding regions was probably not indicative of an active conservation of sequence, since overall, only one or two changes would have been predicted by the observed, average rate of change: i.e., 199 nucleotides/5'-noncoding region × 0.0094 changes/nucleotides = 1.88 changes/ 5'-noncoding region. More disturbing is the lack of change in the NS gene. While all other genes range around 0.1 change/nucleotide, that of the NS gene is fivefold less. The reasons for this difference are not known. Only 26 amino acid changes are predicted to occur from a total of 105 nucleotide changes. This indicates a high degree of functional constraint, since on a random basis, 105 changes should result in approximately two-thirds as many amino acid changes,

TABLE 1
SUMMARY OF B/AA/1/66 Wt TO CA SEQUENCE CHANGES

			cRNA		
	5′-	Coding region	-3'	Total	%
PB2	0/23	24/2310	2/63	26/2396	(1.11)
PB1	0/21	21/2256	0/92	21/2369	(0.89)
PA	0/29	16/2178	1/101	17/2308	(0.74)
NP	0/58	24/1680	1/104	25/1842	(1.36)
M	0/24	12/1073*	2/93	14/1190	(1.18)
NS	0/44	2/1021 ^b	0/33	2/1098	(0.18)
Total	0/199	99/10,518	6/486	105/11,203	(0.94)

		P	rotein	
	Total chan	ges/length	Unique cha	nges/length
PB2	4/770	(0.52%)	1/770	(0.13%)
PB1	3/752	(0.40%)	1/752	(0.13%)
PA	6/726	(0.83%)	2/726	(0.28%)
NP	9/560	(1.61%)	4/560	(0.71%)
M1	2/248	(0.81%)	2/248	(0.81%)
2 ORF°	2/195	(1.03%)	1/195	(0.51%)
NS1	0/281	(0%)	0/281	(0%)
NS2	0/122	(0%)	0/122	(0%)
Total	26/3654	(0.71%)	1 1/3654	(0.30%)

Dratain

i.e., 71. Of the 26 total amino acid differences that exist between the six pairs of wt and ca genes, only 11 are unique to ca B/AA/1/66 virus.

DISCUSSION

As seen in Fig. 1, the initial CK₁ isolate of wt B/AA/1/66 virus had not been cloned by plaguing prior to the cold-adaptation process. Therefore, it was most likely a pool of viruses whose individual genomic sequences were highly related, but to some extent different for each virus. This assumption is based on the data of Parvin and colleagues (1986) which give a mutation rate of 1.5×10^{-5} mutations/nucleotide/generation for influenza A virus. At this rate of mutation, they estimate that every virion in a virus pool will vary by an average of one mutation per genome after five generations. The further removed in time a virus line is from its last plaguing step, the more diverse its individual viruses. Thus, when a virus pool is sequenced directly using either dideoxynucleotide chain-termination or chemical methods and purified vRNA, it is a consensus sequence that is determined and not the absolute sequence of a particular virus.

It should be noted that mutations which occur during every generation of the virus become fixed in the dominant line (and hence appear in the consensus sequence) only if they are in the infecting virus selected by a plaquing step, or if they confer a very great growth advantage compared to the standard population. Mutations which confer growth advantages at suboptimal temperatures have a better chance of predominating in the population in the cold adaptation process, and hence a better chance of becoming selected during plaquing. In addition to these selected mutations, nonlethal mutations may also be selected at random by each plaquing step. Thus, in comparing the final two populations of ca and wt B/AA/1/66 viruses, the sequence changes that exist will primarily represent (1) sequence differences existing between the particular wt virus, that was the progenitor of the ca virus, and the wt virus consensus sequence, (2) cold-selected mutations (these differences may have already existed in the wt pool or have occurred spontaneously during the cold-adaptation process and then been selected, and (3) other spontaneous nonlethal mutations that occur randomly with each generation, and become fixed by the mechanism of plaquing as part of the final ca virus consensus sequence.

Thus the cold-adaptation procedure and the plaquing procedures not only selected changes that resulted in the ca, ts, and att phenotypes, but also at the time of the first plaquing, a particular individual virus was selected from the mixed wt pool that would inherently contain a number of sequence differences from the consensus sequence of the wt virus pool, which probably have no bearing on these three phenotypes. It is conceivable that a change in the nucleic acid sequence might enable the expression of the caphenotype without a corresponding amino acid change, but it is more likely that the cold-adaptation procedure operates by selecting viruses whose proteins function better at the low temperature. Thus, those nucleotide changes that were cold-selected should most likely be reflected in an amino acid change. This condition certainly fits the set of 15 unique nucleotide changes, since they were responsible for 11 of the 26 total amino acid changes. The other 15 amino acid changes were encoded by a subset of the other 90 nonunique nucleotide changes. While it is impossible to go back now and determine which changes occurred at which stage in the cold adaptation process, it is very likely that a large part of the unique changes represented cold-selected changes, while most nonunique changes represented nonlethal random site differences that existed between the wt consensus

^{a,b} Coding region length includes both open reading frames.

^c 2 ORF, Second open reading frame, and length is given for second open reading frame.

sequence and an individual wt virus that was the progenitor of the ca virus.

The nonunique changes can be ordered into groups as mentioned earlier, in which the ca sequence remains the same as that of another wt virus. Sequences for the PB2, NP, M, and NS genes have been determined from isolates which bracket the B/AA/1/66 sequences in time. Thus in these genes it can be determined if the ca virus sequence change resembles that of an earlier or later virus. It should be mentioned that in those sites where the ca sequence is identical to an earlier virus, except in one instance (NP gene, position 1658) it is identical to a later virus as well, with the wt B/AA/1/66 sequence being unique. There are 22 sites in which the ca sequence is identical to an earlier virus, and 33 sites in which the ca sequence resembles that of a later virus for these four genes. Because B/AA/1/66 ca virus is not an evolutionary progenitor to the later viruses, these changes either existed between the sequence of the individual wt virus that was to be the progenitor of the ca virus and the wt consensus sequence prior to the cold-adaptation process, or there is some unknown mechanism which causes convergent changes at specific sites. This latter possibility is unlikely for two reasons: (1) Convergent changes might appear to happen if some sites have a higher than normal rate of mutation and simply switch back and forth between purine to purine, or pyrimidine to pyrimidine, manifesting either as one or the other depending upon the isolate sequenced. However, if this were the case, then out of the 22 sites in which the ca virus sequence is identical to the earlier virus sequence, a good number should show the wt B/AA/1/66 site identical to the later virus isolate's sequence. As mentioned above, this sequence configuration actually occurred only once in 22 times. (2) True convergent change implies that all the changes that appeared in the ca virus sequence occurred as mutations to the wt sequence. Based on the mutation rate determined previously for influenza A virus (Parvin et al., 1986), the 55 differences for these four internal protein genes could not have arisen during the limited number of passages that occurred during the cold-adaptation and plaquing process. From the original wt isolate there were seven chick kidney passages, nine egg passages, and seven plaquing steps to arrive at the virus pool responsible for the ca consensus sequence. Estimating five generations for each of these steps as in Parvin and colleagues (1986), we would calculate 23 steps \times 5 generations/steps = 115 generations. Using the mutation rate of 1.5×10^{-5} mutations/nucleotide/generation (Parvin et al., 1986), and a total of 6526 nucleotides for the PB2, NP, M, and NS genes, we can determine the total number of mutations expected to have arisen during the cold-adaptation procedure: 1.5×10^{-5} mutations/nucleotide/generation \times 6526 nucleotides \times 115 generations = 11.26 mutations expected to have occurred during the ca selection and plaquing procedure, not 55 changes. This mutation rate was determined for influenza A virus. Comparison of the rate of change for influenza A and B viruses had previously indicated that influenza A has a higher rate of change than influenza B virus, by as much as fivefold for known HA gene sequences (Palese and Young, 1983). Thus, it is even more unlikely that all 55 mutations occurred during the cold-adaptation process. Thus, while some of the nonunique nucleotide changes that exist between the wt and ca B/AA/1/66 viruses may be accounted for by mutations that occurred during the ca process, they would be predicted to account for only a small percentage of the number actually observed. The mechanism of convergent mutation cannot be entirely ruled out, however, since the number of viral generations in man prior to isolation are not known, and the convergent changes could have occurred then.

Of the six genes examined, the NS gene was quite restricted in the rate of change/nucleotide from the other five genes. If this restriction were due to a functional constraint inherent to the NS gene, then a similar restriction might be observed between wt NS genes as well. The NS gene of B/Lee/40 is 1096 nucleotides long. Both ca and wt B/AA/1/66 NS genes had an additional two nucleotides due to the insertion of two additional adenosine residues within a run of three existing adenosine residues at positions 39 to 41 of the B/Lee/40 sequence. This two-nucleotide insertion occurred in the 5'-noncoding region of mRNA and does not affect the NS1 and NS2 protein sequences. In addition to the two unmatched adenosine residues. there are 61 base mismatches between the B/AA/1/66 wt virus and B/Lee/40 virus. This amount of change is roughly the same as that found between the other known genes of B/Lee/40 and B/AA/1/66 viruses (De-Borde et al., 1987), indicating that the minimal nucleotide change occurring between wt and ca B/AA/1/66 NS gene pair is probably not due to an inherent functional restriction in the NS gene, but to some other reason. Whether it is specific to cold-adaptation or random chance is not known.

Genetic studies utilizing reassortants that were made between ca B/AA/1/66 and wt B/HN/76 have indicated that the PA gene is the primary determinant of the ts and att phenotypes (Donabedian *et al.*, 1987). A combination of other ca B/AA/1/66 genes (without the PA gene) could also contribute to attenuation. A

reassortant that contained the NP gene from ca B/AA/1/66 virus and all other genes from B/Texas/1/84 was shown to be ca and ts in PCK cells, and partially attenuated in ferrets (unpublished data, this laboratory). In the study of Donabedian and colleagues (1987) no single-gene NP reassortant was isolated, but reassortants containing the NP and other genes from B/AA/1/66 ca virus, without the PA gene, were shown to have a 2-log₁₀ reduction in growth at 39° compared to growth at 33°. One of these reassortants is also restricted for viral growth in ferret lungs, a probable marker for attenuation (Donabedian et al., 1987). Thus the NP gene may also carry mutations involved in the final level of ts and att phenotypes of ca B/AA/1/66 virus and its ca vaccine reassortants. From the studies of the reassortants made with the wt B/HN/76 virus. the ca marker appears to arise from a combination of the genes of ca B/AA/1/66, since no individual gene conferred this phenotype. However, with the wt B/ Texas/1/84 parent, as mentioned above, a single-gene NP reassortant did become cold-adapted in PCK cells. In addition to being tested in PCK instead of MDCK cells, one other possible reason for this inconsistency is that because the ca mutations enable growth, any wt gene which cannot function efficiently at 25° will abrogate the appearance of the ca phenotype, obscuring any number of ca enabling genes. In our studies, all genes except the NS gene, showed at least one unique amino acid change in its predicted protein sequence. Thus, every ca viral gene which has been implicated in the direct expression of one or more of the three phenotypes, by the above genetic studies, was shown to contain nucleotide and amino acid changes from those of the wt gene. Recently, a non-ts revertant of the PA gene was isolated and sequenced, and the sequence reversion has been shown to involve the codon position, 1320, PA gene (Fig. 8). The experiments detailing this revertant gene are presented in the companion paper (Donabedian et al., 1988).

We have presented a catalog of changes that existed between the wt and ca B/AA/1/66 viruses that should provide a valuable basis for understanding the mechanisms by which the ca donor virus confers the ca, ts, and att phenotypes to new ca vaccine reassortants. The changes that have occurred in the sequences of genes from phenotypic revertant viruses can be compared to those of the wt and ca B/AA/1/66 viruses in order to link specific mutations to specific phenotypes. This sequence information also provides a basis for the design of site mutagenesis experiments aimed at establishing directly the importance of some sequence sites to phenotypes not amenable to sin-

gle-gene or revertant analysis (like the ca phenotype). Finally, understanding which of the six nonsurface genes of ca B/AA/1/66 are essential for the stability and attenuation of the ca reassortant live virus vaccine, and which genes are not, can lead to the laboratory creation of more effective ca reassortant donor viruses.

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REFERENCES

- AKOTO-AMANFU, E., SIVASUBRAMANIAN, N., and NAYAK, D. P. (1987).
 Primary structure of the polymerase acidic (PA) gene of an influenza B virus (B/Sing/222/79). Virology 159, 147–153.
- BRIEDIS, D. J., and LAMB, R. A. (1982). Influenza B virus genome: Sequences and structural organization of RNA segment 8 and the mRNAs coding for the NS₁ and NS₂ proteins. *J. Virol.* 42, 186–193.
- BRIEDIS, D. J., LAMB, R. A., and CHOPPIN, P. W. (1982). Sequence of RNA segment 7 of the influenza B virus genome: Partial amino acid homology between the membrane proteins (M₁) of influenza A and B viruses and conservation of a second open reading frame. *Virology* **116**, 581–588.
- BRIEDIS, D. J., and TOBIN, M. (1984). Influenza B virus genome: Complete sequence of the influenza B/Lee/40 virus genome RNA segment 5 encoding the nucleoprotein and comparison with the B/Singapore/222/79 nucleoprotein. *Virology* 133, 448–455.
- COX, N. J., KITAME, F., KLIMOV, A., KOENNECKE, I., and KENDAL, A. P. (1986). Comparative studies of wild-type and cold-mutant (temperature-sensitive) influenza virus: Detection of mutations in all genes of the A/Ann Arbor/6/60(H2N2) mutant vaccine donor strain. *Microbiol. Pathol.* 1, 387–397.
- DeBorde, D. C., Naeve, C. W., Herlocher, M. L., and Maassab, H. F. (1986). Resolution of a common RNA sequencing ambiguity by terminal deoxynucleotidyl transferase. *Anal. Biochem.* **157**, 275–282.
- DeBorde, D. C., Naeve, C. W., Herlocher, M. L., and Maassab, H. F. (1987). Nucleotide sequences of the PA and PB1 genes of B/Ann Arbor/1/66 virus: Comparison with genes of B/Lee/40 and type A influenza viruses. *Virus Res.* **8**, 33–41.
- Donabedian, A. M., DeBorde, D. C., Cook, S., Smitka, C. W., and Maassab, H. F. (1988). A mutation in the PA protein of cold-adapted (ca) B/Ann Arbor/1/66 influenza virus associated with reversion of the ts phenotype and attenuated virulence. *Virology* **163**, 444–451.
- DONABEDIAN, A. M., DEBORDE, D. C., COOK, S., SMITKA, C. W., and MAASSAB, H. F. (1988). A mutation in the PA protein of cold-adapted (ca) B/Ann Arbor/1/66 influenza virus associated with reversion of the ts phenotype and attenuated virulence. *Virology* 163, 444–451.
- HIEBERT, S. W., WILLIAMS, M. A., and LAMB, R. A. (1986). Nucleotide sequence of RNA segment 7 of influenza B/Singapore/222/79:

- Maintenance of a second large open reading frame. *Virology* **155**, 747–751.
- KEITEL, W. A., CATE, T. R., and COUCH, R. B. (1986). Evaluation of a cold-recombinant influenza B vaccine. *In* "Options for the Control of Influenza" (A. P. Kendal and P. A. Patriarca, Eds.), pp. 287–291. A. R. Liss, New York.
- Kemdirim, S., Palefsky, J., and Briedis, D. J. (1986). Influenza B virus PB1 protein: Nucleotide sequence of the genome RNA segment predicts a high degree of structural homology with the corresponding influenza A virus polymerase protein. *Virology* 152, 126–135.
- LONDO, D. R., DAVIS, A. R., and NAYAK, D. P. (1983). Complete nucleotide sequence of the nucleoprotein gene of influenza B virus. *J. Virol.* 47, 642–648.
- MAASSAB, H. F., DEBORDE, D. C., DONABEDIAN, A. M., and SMITKA, C. W. (1985). Development of cold-adapted "master" strains for type B influenza virus vaccines. *In* "Vaccines 85" (R. A. Lerner, R. M. Chanock, and F. Brown, Eds.), pp. 327–332. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY.
- MAASSAB, H. F., DEBORDE, D. C., DONABEDIAN, A. M., and SMITKA, C. W. (1986). Prospects for influenza type B live attenuated vaccines. *In* "Cptions for the Control of Influenza" (A. P. Kendal and P. A. Patriarca, Eds.), pp. 271–286. A. R. Liss, New York.

- MARTIN, R. (1987). Overcoming DNA sequencing artifacts: Stops and compressions. Focus 9, 8–10.
- MOUNT, S. M. (1982). A catalog of splice junction sequences. *Nucleic Acids Res.* **10**, 459–472.
- PALESE P., and Young, J. F. (1983). Molecular epidemiology of influenza virus. In "Genetics of Influenza Viruses" (P. Palese and D. W. Kingsbury, Eds.), pp. 321–336. Springer-Verlag, New York.
- Parvin J. D., Moscona, A., Pan, W. T., Leider, J. M. and Palese, P. (1986). Measurement of the mutation rates of animal viruses: Influenza A virus and poliovirus type 1. *J. Virol.* **59**, 377–383.
- PEACOCK, A. C., and DINGMAN, C. W. (1968). Molecular weight estimation and separation of ribonucleic acid by electrophoresis in agarose–acrylamide composite gels. *Biochemistry* 7, 668–674.
- PEATTIE, D. A. (1979). Direct chemical method for sequencing RNA. *Proc. Natl. Acad. Sci. USA* **76**, 1760–1764.
- QUEEN, C., and KORN, L. J. (1984). A comprehensive sequence analysis program for the IBM personal computer. *Nucleic Acids Res.* **12**, 581–599.
- REEVE, P., PIBERMANN, M., BACHMAYER, H., LIEHL, E., MORITZ, A., GANZINGER, U., HOFMANN, H., and KUNZ, C. (1982). Studies in man with a cold-recombinant live influenza B virus vaccine. *J. Med. Virol.* 9, 1–9.