INHIBIN SECRETION DURING THE RAT ESTROUS CYCLE: RELATIONSHIPS TO FSH SECRETION AND FSH BETA SUBUNIT mRNA CONCENTRATIONS

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Summary

Serum inhibin and FSH and FSH beta subunit mRNA levels were measured at 3h intervals throughout the 4 day estrous cycle in female rats and hourly between 1000 and 2400 h of proestrus. On proestrus, serum inhibin concentrations fell during the late morning-early afternoon, then increased transiently during the late afternoon gonadotropin surges. Inhibin levels decreased during the late evening of proestrus, coincident with the FSH surge-related rise in FSH beta mRNA levels. Serum inhibin remained relatively stable during estrus and early metestrus, but rose during the late evening of metestrus and remained elevated until early diestrus. FSH beta mRNA levels were elevated on late estrus and early metestrus and declined during the evening of metestrus as serum inhibin levels increased. These data show that concentrations of serum inhibin change during the estrous cycle and that a general inverse relationship exists between serum inhibin and FSH levels and FSH beta mRNA concentrations in the pituitary. This suggests that inhibin may inhibit FSH beta gene expression and FSH secretion during the 4 day cycle in female rats.

The pattern of gonadotropin secretion during the 4 day rat estrous cycle has previously been characterized (1-4). On proestrus, a preovulatory surge of luteinizing hormone (LH) occurs during the late afternoon. Follicle stimulating hormone (FSH) also expresses a surge on proestrus, but this has a biphasic nature and evidence suggests that it is controlled by different mechanisms. Prior work has shown that the first phase of FSH release (as well as LH) is stimulated by an increase in pulsatile secretion of the hypothalamic peptide gonadotropin-releasing hormone (GnRH) (5,6). The secondary FSH surge appears to be regulated by ovarian inhibin (7,8). Inhibin is a glycoprotein composed of 2 subunits, alpha and beta (9). Inhibin has been shown to suppress FSH secretion (10-12) and FSH beta gene expression (12-14) by a direct action on the gonadotroph cell.

We recently characterized the changes in pituitary FSH beta mRNA during the rat estrous cycle to increase our understanding of the physiological relationship between FSH secretory activity and gene expression (15). Serum samples collected in that study were used to measure serum inhibin concentrations in order to establish the relationships between serum inhibin and FSH secretion and FSH beta gene expression during the rat estrous cycle.
Materials and Methods

Animal Protocol
Adult female Sprague-Dawley rats were maintained in controlled lighting (14h of light; lights on at 0600h), temperature (23°C) with food and water ad libitum. Vaginal smears were taken for 2 weeks and only rats that exhibited at least 2 consecutive 4-day cycles were used. Groups of rats (n = 5-11) were decapitated at 3h intervals throughout the cycle and hourly between 1000 and 2400h on proestrus. Trunk blood was collected and sera stored at -20°C. Anterior pituitaries were removed, snap frozen and stored at -70°C.

Radioimmunoassays (RIAs)
Serum inhibin levels were measured using an antibody generated against the synthetic rat inhibin alpha1-27 (Tyr27) peptide fragment as previously described (16). The radioligand and standard used in the RIA were the same synthetic peptide. Results were expressed as femtomoles/ml (as determined from the 32 KD inhibin molecule). Serum FSH was measured by RIA using NIADDK reagents and RP-1 as standard. Serum LH was measured in samples on proestrus in order to determine the onset and duration of the proestrus LH surge (using NIADDK reagents and RP-1 as standard).

mRNA Assessment
Pituitaries were homogenized under sterile conditions and cytoplasmic RNA extracted with a phenol-chloroform-isomyl alcohol mixture (100:100:1) and quantitated by absorbance at 260 nm. Nuclear pellets were sonicated and DNA measured by fluorometric assay. RNA samples were spotted on nitrocellulose filters (8 ug/spot) in duplicate and hybridized to saturating amounts of 32p labeled FSH beta cDNA probe (generously provided by Dr. Richard A. Maurer, Univ of Iowa) as described previously (17). FSH beta mRNA levels were expressed as femtomoles of cDNA bound/100 ug pituitary DNA as previously reported (15).

Statistical Analysis
Data was analyzed using one way analysis of variance with differences between timepoints determined by Duncan’s multiple range test.

Results
Figure 1C shows serum inhibin concentrations during the rat estrous cycle. Inhibin levels remained stable between 0500 h of diestrus through the late morning of proestrus, and decreased abruptly between 1100h and 1600h. This decline was followed by a transient rise (2 fold vs 1300h) in serum inhibin between 1700 and 1900h of proestrus. Serum inhibin declined during the late evening of proestrus and remained relatively stable throughout estrous until the afternoon of metestrus. During the evening of metestrus, inhibin concentrations rose reaching a peak (2 fold increase vs morning of metestrus) at 0200h of diestrus.

Serum FSH concentrations are shown in Figure 1A. On proestrus, serum FSH began to increase at 1800h and peaked at 2000h, coincident with the decline in serum inhibin levels. Serum FSH fell on late proestrus - early estrus before the secondary rise at 0500h of estrus. Serum FSH remained stable during the later portion of estrous and through metestrus and diestrus. The onset and duration of the proestrus LH surge is shown as a horizontal line above the serum FSH data in Figure 1A. Serum LH began to rise (>200 ng/ml) between 1600 and 1700h and remained elevated (> 2100 ng/ml) through 2100h. The peak of the LH surge (2440±1273 ng/ml) was seen at 1800h. FSH beta mRNA concentrations (Fig 1B) rose during the FSH surge beginning at 2000h (as serum inhibin levels were declining) and reached a peak at 0200h of estrus. A second increase in FSH beta mRNA began at 2300h of estrus and continued through the late afternoon of metestrus. FSH beta mRNA declined as serum inhibin levels were increasing during the evening of metestrus.
Fig. 1

Serum FSH (A), FSH beta mRNA (B) and serum inhibin (C) concentrations during the rat estrous cycle. Mean ± SEM for each group (5-11 rats/group) are shown. The horizontal line between the arrows above serum FSH values (1A) show the timing and duration of the proestrus LH surge. Panel A: *P < 0.05 vs 0200-1200h of proestrus. B * P < 0.05 vs all timepoints with mean values less than or equal to 0.1 fmol cDNA bound/100 ug pit DNA. Panel C: a P < 0.05 vs 0200-0800h of metestrus, b P < 0.05 vs 0500-2000h of diestrus; c P < 0.05 vs 0200-1000h of proestrus; d P < 0.05 vs 1100-1600h of proestrus. The FSH beta mRNA concentrations have been previously reported (Endocrinology 123:2149, 1988) and are shown here for comparison with the permission of the Editor.
Discussion

The present study describes the first characterization of the relationships between serum inhibin, FSH secretion and FSH beta gene expression throughout the rat estrous cycle. The data agree with recent findings on more limited studies of proestrus (18,19) which showed that serum inhibin increases during the evening of proestrus before falling on late proestrus-early estrus. The timecourse of this decline in serum inhibin has been shown to parallel observed decreases in ovarian inhibin alpha and beta subunit mRNAs (20). Data from other studies (21,22) suggest that the decrease in serum inhibin during late proestrus may be regulated by the late afternoon gonadotropin surges (either LH, FSH or both) and could be responsible for the secondary FSH surge seen during the early morning of estrus. It is of interest that this decrease in serum inhibin levels on late proestrus was coincident with the initial FSH surge-related rise in FSH beta mRNA concentrations and preceded the FSH beta mRNA peak (0200h of estrus) by 6h. Serum inhibin also showed a transient decrease during the late morning-early afternoon of proestrus. The physiological significance of this decrease is uncertain, but could relate to previously observed changes in serum estradiol (E2) concentrations on proestrus. Serum E2 levels have been shown to decline on the afternoon of proestrus (1,3,18). Other studies (23,24) have shown that E2 can stimulate inhibin secretion as well as inhibin subunit gene expression, suggesting a possible paracrine regulatory mechanism. It is possible that a fall in ovarian E2 concentrations might reduce inhibin synthesis and secretion resulting in the transient decrease in serum inhibin during the early afternoon of proestrus.

Beginning on late estrus through the evening of metestrus, FSH beta subunit mRNA levels were elevated (approx 2 fold). This was not coupled to a measurable increase in FSH secretion similar to previously described increases for pituitary alpha and LH beta subunit mRNAs on diestrus (25). The late metestrus decrease in FSH beta mRNA levels was coincident with a rise in serum inhibin levels. This increase in serum inhibin during metestrus was also observed in a recent study by Hasegawa and colleagues (18). In vitro studies have shown (12,14) that inhibin can rapidly and selectively suppress FSH beta gene expression in gonadotroph cells, suggesting that the metestrus rise in inhibin secretory activity could be responsible for the decrease in FSH beta mRNA levels.

During the early morning of diestrus, serum inhibin concentrations decreased and remained stable for a period of 29h. Approximately one half of the developing follicles recruited as a result of the proestrus FSH surge become atretic during metestrus (26). Since the levels of inhibin in blood appear to be positively correlated to the number of available follicles (27) the decline in serum inhibin on early diestrus might reflect the loss of viable follicles.

Recent studies provide support for a physiological role of the rise in serum inhibin during late metestrus-early diestrus (19). Removal of endogenous inhibin activity by administration of inhibin antisera on metestrus and diestrus resulted in a rise in serum FSH, a 2 fold increase in ovulation rate and a 2 fold increase in the number of conceptuses in animals that were mated. These results suggest that the rise in inhibin between metestrus and diestrus may provide a protective effect against overstimulation of the ovary by FSH.

In summary, alterations in serum inhibin concentrations occur during the rat estrous cycle. The data provide support for an inverse relationship between serum inhibin and FSH secretory activity and FSH beta gene expression. However, further studies are required to delineate the exact physiological interactions between inhibin and FSH synthesis and secretion, and await the availability of recombinant inhibin for in vivo studies.

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