Workshop Report

In vivo percutaneous penetration/absorption

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This workshop, 'In Vivo Percutaneous Penetration/Absorption' was held in Washington, DC, on May 1-3, 1989. The first workshop in this series, 'In Vitro Percutaneous Penetration', took place in November 1986 (the report of this earlier meeting was published in *Pharmaceutical Research*, 4 (1987) 265-267). The objectives of the workshop were to review the relevant literature and to address in detail: (1) In vivo percutaneous penetration/absorption methodology; (2) The characteristics of dosage forms designed for application to the skin; (3) Critical factors controlling in vivo drug transport into and across the skin; (4) The use of models in the assessment and evaluation of in vivo percutaneous penetration/absorption; and (5) Bioavailability/bioequivalence considerations for topical drug products. Scientific knowledge and technology are rapidly evolving in the topical and transdermal drug products area. This report focuses on the methodologies available for the measurement of percutaneous penetration in vivo; each scientific approach is discussed briefly followed by advantages and disadvantages of the methodology.

**Introduction**

Drug products applied to the skin can be subdivided into two categories: (1) Dermatological formulations (creams, ointments, gels, lotions) intended for the treatment of local (i.e., application site) skin disorders. (2) Transdermal delivery systems (ointments and patches) intended for the treatment or prevention of systemic disease.

The skin is a barrier to the absorption of topically administered drugs (Schaefer et al., 1982; Barry, 1983; Bronaugh and Maibach, 1989). As a result, the rate of percutaneous transport is typically slow, and the extent of drug delivery, although concentrated in the skin beneath the application site, is usually low. One advantage of these features is that the incidence of systemic toxicity in topical therapy is far less than that encountered with systemic routes of drug delivery (e.g., oral). Still, topical therapy may fall short of delivering a clinically sufficient local concentration of drug. In such an instance, drug penetration/absorption/bioavailability can be increased through the use of chemical enhancers or through physical enhancing techniques such as iontophoresis and ultrasound. A general problem in ascertaining bioavailability/bioequivalence of topical drug therapy is that quantification of the drug in the body (skin and/or systemic circulation) is difficult because the absolute amounts of drug present are (generally) too small. In vivo drug penetration studies following topical application may clarify the poorly resolved bioavailability/bioequivalence issues.

The specific aims of in vivo skin penetration studies may be summarized as: (a) To verify and quantify the cutaneous bioavailability of a topically applied drug. (b) To verify and quantify the systemic bioavailability of a transdermally delivered drug. (c) To establish the bioequivalence of different topical formulations of the same drug. (d) To determine the incidence of and, if necessary, to quantitate local and systemic toxicological risk following the topical application of a specific drug.

Each of the in vivo percutaneous absorption/penetration approaches discussed at the workshop is now considered in turn. Typically, following an introductory outline of the method, the consensus advantages and drawbacks are listed. Finally, our conclusions are summarized, and future research directions are highlighted.

**Animal Models**

It is important to emphasize at the outset that the most relevant in vivo data on percutaneous absorption in man will be obtained from studies in humans themselves (Guy et al., 1987b). However, animal models are needed for the development of conceptual insights and to investigate
mechanisms. If the study objective is prediction of percutaneous absorption in man, then rate and extent of skin absorption in the animal should be: (1) quantitatively the same as in man; or (2) consistently related to the absorption in man by a constant ratio. Moreover, the animals chosen for the studies must respond to treatments (e.g., vehicle effects, especially the use of enhancers) in the same way and to roughly the same degree as in man (Bond and Barry, 1988; Choi et al., 1990). On the basis of the currently available data, the only animals in which permeation data are consistently qualitatively and quantitatively similar to human permeation data, are the pig (Reifenrath et al., 1984) (particularly the weanling pig), the Rhesus monkey (Wester and Maibach, 1989) and the hairless rat (Rougier et al., 1987). However, it should be noted that the extra body fat on the pig may alter drug distribution relative to man and thereby confound the results. In the case of the Rhesus monkey, skin applications should be limited to the non-hairy regions on the ventral surfaces of the animal. Regional variation in skin properties (thickness, composition, etc.) in animal and man should be considered. Absorption studies in the guinea pig are sometimes predictive of results in man but skin absorption rates in the rabbit, rat and mouse, appear to be substantially greater than that in man (Bartek et al., 1982; Reifenrath et al., 1984). Furthermore, some laboratory animals do not consistently respond to treatments (e.g., with an absorption enhancer) in the same fashion as man; rather, they seem to be considerably more responsive to such treatments (Bond and Barry, 1988).

Four approaches, which sample drug levels in: (a) the skin; (b) the venous blood draining the application site; (c) the systemic circulation; and (d) the excreta, can be identified. Principles of pharmacokinetics can be used when the drug concentrations are followed over time. For example, coupled with clearance parameters measured from intravenous administration, such data can be used to estimate the absolute extent of systemic bioavailability of a topically or transdermally applied drug. Using this methodology, drug application to both normal and pathological skin can be considered. For topical dermatological products, toxicokinetic and safety studies should, when possible, be performed in the same species.

Skin Sectioning

Definition. Cutting, stripping or otherwise separating skin into its constituent layers for independent assay of their drug content (Schaefer et al., 1978; Schaefer and Lamaud, 1987).

Advantages. The technique can establish distribution and disposition of a given compound in the tissue as a function of time and tissue depth following application under various conditions (different vehicles, diseased skin, etc.). The procedure is best performed in vivo but can, under well-defined circumstances, be conducted in vitro as well.

Disadvantages. Skin sectioning cannot normally be performed in vivo in humans unless limited to punch biopsies. If the experiments are performed in vitro, their relevance is greatest for short application times. The technique is generally limited to radiolabeled studies. The samples collected from the treated skin site in the experiments do not indicate the drug target nor the ultimate fate of the drug in the body. Assessments of metabolic processes and metabolites are especially difficult (Guzek et al., 1989).

Sampling of Excreta

Definition. Sampling of urine, feces, and expired air for drug content following a topical administration (Guy et al., 1987b).
Advantages. This is useful to assess total absorption. It is relatively non-invasive, and can be routinely performed in human subjects. Mass balance of the applied dose must be performed (Bucks et al., 1988).

Disadvantages. A control study involving parenteral injection of the drug is required. Availability of a radiolabeled drug or a sensitive chemical assay is necessary. Metabolism of the drug by the skin and/or systemic metabolism may confound precise bioavailability measurements. The technique has limitations for drugs which are naturally present in the body (e.g., certain steroids). The approach is difficult to apply to fat soluble drugs with long elimination half-lives.

Blood Sampling

Definition. Periodic sampling of blood as routinely performed in all standard pharmacokinetic analyses.

Advantages. For drugs delivered transdermally to elicit systemic pharmacological effect, assay of drug levels in the blood/plasma is essential. Drug concentrations of the order of those achieved by other administration routes are anticipated. A highly specific and sensitive assay may be necessary permitting high quality pharmacokinetic data to be obtained. Parent drug and metabolites can be identified and quantitated. With an intravenous control, absolute bioavailability can be determined.

Disadvantages. The method is generally unsuitable for assessment of locally active dermatologic preparations because relationships between systemic and locally effective levels are unclear. The blood concentrations are typically too low for bioavailability assessment. Interference by endogenous substances can also prove problematic.

Residual Analysis – ‘Difference’, ‘Disappearance’ Methods

Definition. Techniques in which the amount of drug absorbed into the skin are assessed as the difference between the amount applied and that recovered at a subsequent time.

Two approaches have been described: (a) After drug application for a fixed time, the residual formulation is washed from the skin surface, and the amount removed is analyzed (Yano et al., 1986). This is a single point determination (amount absorbed = (amount applied) - (amount remaining)). (b) The formulation is applied and drug content in the outer skin layers is then followed as a function of time by spectroscopic (e.g., infrared) or radioisotopic monitoring techniques (Guy et al., 1987a; Mak et al., 1990).

Approach (a): single point measurements of drug disappearance

Advantages. The procedure requires very small amounts of active formulation. Skin toxicology concerns, while present, are minimal due to the limited exposure (drug amount and skin area). The method is inexpensive and relatively rapid. It is suitable for clinical studies. Radioisotope use is reasonable, due to the very low levels required;
the radioisotopes need not have high specific activities; very little isotope is consumed.

Disadvantages. Only one assay per site per application is possible; full characterization of the drug uptake profile requires multiple site studies. Uniform recovery from different sites must be demonstrated. Drug removed inadvertently from the surface (on clothing, etc.) or by evaporation may be counted as absorbed. For poorly penetrating compounds, the method quantitates the small difference between two large numbers. The application technique is critical; the amount applied and uniformity of spreading must be validated and reproducible. The method does not lend itself to theoretical analysis (separation of non-stationary state from steady or quasi-steady state periods of diffusion is virtually impossible). Another potential problem is that the material used (soap solution, solvent, etc.) to remove the remaining preparation from the skin surface may influence drug penetration. Furthermore, the procedure does not actually measure the amount of drug at the target tissue in the skin. In other words, this type of measurement fails to reflect diffusive migration of drug into the critical zone, i.e., the region where the pharmacodynamic, pharmacological or biochemical event of relevance takes place.

Approach (b): continuous or periodic monitoring of drug uptake

The drug, radiolabeled (preferably) with \(^{14}\text{C}\), or containing a non-ambiguous spectrophotometric marker, is applied to the skin surface in the test formulation. Over time, the disappearance of radioactivity or of spectral signal is monitored using, for example, an appropriate Geiger-Muller tube or attenuated total reflectance, Fourier transform infrared spectroscopy (ATR-FTIR). Penetration kinetics are assessed from the decay of the respective signal.

Advantages. Pharmacologically insignificant drug doses can be used. Full characterization of the drug uptake profile from a single experiment is possible. The method is relatively sensitive. The methods are non-invasive; they are also precise and objective.

Disadvantages. (i) Radiometric methods: the use of radioisotopes on human subjects is necessary. The application of the method for bioequivalence measurements has not been demonstrated. Again, one is not measuring drug levels at the target site. (ii) Spectrophotometric methods: spectrophotometric interference is a major problem. Optimal application of the technique requires that the drug has one or more unique spectral features that distinguish it from the spectral characteristics of the skin. However, providing the drug with selected carbon–deuterium substitutions (for regular C-H bonds) could generalize the approach. The equipment is specialized and costly. As with radioisotope monitoring, one is not measuring drug levels at the target site.

In Vivo Measurement of Drug Concentrations in Stratum Corneum: Prediction of Penetration

This is a technique in which the permeability of drug is projected from the amount recovered in the stratum corneum by adhesive tape-stripping at a fixed time following drug application [20]. The method depends upon a correlation between short-time uptake by the stratum corneum and total percutaneous absorption.

Advantages. The method requires pharmacologically insignificant drug doses; the experiment is straightforward and inexpensive; radiolabeled drugs are not essential if the compound can be efficiently extracted from the tape-strips for conventional analysis; comparisons between formulations are easily performed, and can be well-controlled.

Disadvantages. Quantification of drugs in the stratum corneum tape-strips has generally been limited to radioisotope counting. Other approaches have yet to be optimized and validated. The correlation between the amount of the drug in the stratum corneum and total drug absorption has only been established for some drugs and formulations. Since different body sites of skin have different drug penetration properties, the site of application has to be specified for predicting drug absorption like for any other method. The method does not sample the epidermis or
the dermis (i.e., the normal ‘targets’ of topical drug products). The cleaning and preparation of the skin for stripping is a critical determinant of drug recovery.

**Mathematical Models in the Assessment of In Vivo Percutaneous Penetration/Absorption**

Mathematical (mechanistic) models of percutaneous penetration have been utilized to simulate drug delivery from the formulation, drug movement through skin into the cutaneous circulation, and the subsequent systemic distribution, metabolism and excretion (Berner, 1987; Berner and Cooper, 1987; Hadgraft, 1990; McDougal et al., 1990; Osborne, 1990). Using appropriate parameters, this approach may lead to experimentation with novel systems. Mathematical models of percutaneous absorption have been developed to extrapolate in vitro measurements to in vivo data, to test mechanistic hypotheses, and to interpret data.

**Bioavailability / Bioequivalence Considerations**

For topical dermatological products, a measurable pharmacodynamic response and/or quantification of the amount of drug absorbed/penetrated at the target site can be regarded as a measure of drug availability (bioavailability) from the dosage form (Guy et al., 1986; Caron et al., 1990). On the other hand, for transdermal drug products, measurement of systemic drug concentrations and/or pharmacodynamic response provide the necessary evidence of drug bioavailability.

Cutaneous bioavailability can be assessed by performing, as a function of time: (1) pharmacokinetic measurements of the drug concentrations in the skin or systemic circulation; or (2) pharmacodynamic measurements of the pharmacological response to the drug in the skin or elsewhere.

The combination of the two methods allows one to investigate problems of principal importance and subsequently to choose one of the two approaches for more routine purposes.

**Pharmacodynamic Responses for Assessing Bioavailability and Bioequivalence of Topical Dermatological Products**

Pharmacodynamic responses in skin function can serve as proof, and as measures, of local absorption. To date, vascular responses have been used in this way. The pharmacodynamic assays are worth pursuing (in particular, the skin blanching assay for glucocorticoids), but require development and generation of a rigid worldwide protocol before they can be used for bioavailability/bioequivalence determinations with confidence (Barry, 1983, 1989).

**Advantages.** There is a long history with these assays (in particular, the vasoconstriction or skin blanching assay) in dermatological research with glucocorticoids and there are good clinical efficacy correlations. The methods are internally consistent, and for clinical studies, they are inexpensive and facile. The assays are qualitatively reliable and have proven useful for evaluating vehicle effects on drug absorption.

**Disadvantages.** Only a limited number of compounds (e.g., topical corticosteroids) evoke a strong local pharmacological response and lend themselves to the pharmacodynamic method. Pharmacodynamic responses are difficult to validate. The methods require a rigid protocol. The pharmacodynamic measurements are subjective, and there is a need, therefore, to develop objective methods to quantify these pharmacodynamic responses (Queille-Roussel et al., 1991). For most topical drug products, the dose–response curves are yet to be defined. The ‘area under the curve’ (AUC) (response versus time) type of measurements and calculations are a necessary part of the evaluation.

**Bioavailability / Bioequivalence of Transdermal Delivery Systems**

Pharmacokinetic and pharmacodynamic studies are required as appropriate for innovator and generic drug products which are designed to elicit a systemic pharmacological effect. Special attention should be paid to the occurrence and sever-
ity of local (i.e., application site) skin reactions. Appropriate irritation and sensitization studies must be performed. Although these requirements are expensive and time-consuming, they are essential. Since transdermal drug products are administered for systemic effects, it is desirable to measure drug/metabolite concentrations in blood/plasma and/or urine.

References


Wester, R.C. and Maibach, H.I., In vivo animal models for...

