

Localization of the Genes Encoding the Melanocortin-2 (Adrenocorticotrophic Hormone) and Melanocortin-3 Receptors to Chromosomes 18p11.2 and 20q13.2–q13.3 by Fluorescence *In Situ* Hybridization

Ira Gantz,* Takao Tashiro,† Christine Barcroft,‡ Yoshitaka Konda,† Yoshimasa Shimoto,† Hiroto Miwa,† Thomas Glover,‡§ Gerd Munzert,† and Tadataka Yamada†¶¹

Departments of *Surgery, †Internal Medicine, ‡Pediatrics, §Genetics, and ¶Physiology, University of Michigan Medical Center, Ann Arbor, Michigan

Received March 5, 1993; revised June 2, 1993

Adrenocorticotrophic hormone (ACTH) and α -, β -, and γ -melanocyte-stimulating hormone (MSH) are products of pro-opiomelanocortin post-translational processing (1). These compounds are collectively labeled as melanocortins (MC). Aside from their established effects on the regulation of the adrenal cortex (ACTH) and melanocytes (α -MSH), the melanocortins have been implicated in a broad array of physiological events. In the central nervous system, melanocortins have been shown to affect behavior, learning and memory, control of the cardiovascular system, thermoregulation, the release of other neurohumoral agents, and analgesia (2, 3). Peripherally, melanocortins have been identified to have immunomodulatory and neurotrophic properties and to be involved in the events surrounding parturition (4–6). Melanocortins mediate their effects through cell membrane receptors belonging to the superfamily of seven transmembrane G-protein-linked receptors. Using the technique of polymerase chain reaction with primers based on conserved areas of the seven transmembrane G-protein-linked receptor family, we recently isolated an "orphan" subfamily of this receptor group. Within the past year, two of these receptors were identified as specific for α -MSH (MC1) (7, 8) and ACTH (MC2) (7). We have recently described a third melanocortin receptor (MC3) that appears to recognize the core heptapeptide sequence of melanocortins with equal potency and efficacy and identified its presence in the brain, placenta, and gut (9). We have also characterized a fourth melanocortin (MC4) receptor primarily present in the brain (10). Unlike the α -MSH (MC1) and ACTH (MC2) receptors, the MC3 and MC4 receptors do not appear to be expressed in either the adrenal gland or melanocytes. In this paper we report the chromosomal localization of the ACTH (MC2) and the melanocortin-3 (MC3) receptors.

The technique of fluorescence labeled *in situ* hybridization (FISH) according to a protocol modified from Pinkel *et al.* (11) Lichter *et al.* (12), and Lemieux *et al.* (13) was used for the chromosomal localization of the MC2 and MC3 receptors. DNA of EMBL3 phage clones containing the genomic se-

quences of the MC2 and MC3 receptors was isolated and biotinylated using a Bionick kit (BRL, Gaithersburg, MD). Metaphase chromosomes from a normal female were prepared from peripheral blood lymphocytes following overnight synchronization with 5-bromodeoxyuridine and thymidine release. Cells were harvested and slides were prepared using standard cytogenetic techniques. Probe labeling, preannealing, hybridization, and washing were performed as described using 330 ng biotinylated DNA per slide (11, 12). Signal detection was achieved by incubations of 30 min at 37°C with fluorescein goat anti-biotin and fluorescein-labeled anti-goat IgG (Vector, Burlingame, CA) in 4× SSC/0.1% Tween/1% BSA. Each incubation was followed by washes in 4× SSC/0.1% Tween at 37°C. Slides were counterstained with propidium iodide (PI), rinsed in phosphate-buffered saline, and coverslipped with PPD11 anti-fade solution (13). Photographs were taken on Kodak ASA 400 Gold film using a Zeiss Axioskop epifluorescence microscope equipped with a Zeiss filter set allowing simultaneous visualization of FITC and PI. The results of hybridization are shown in Fig. 1.

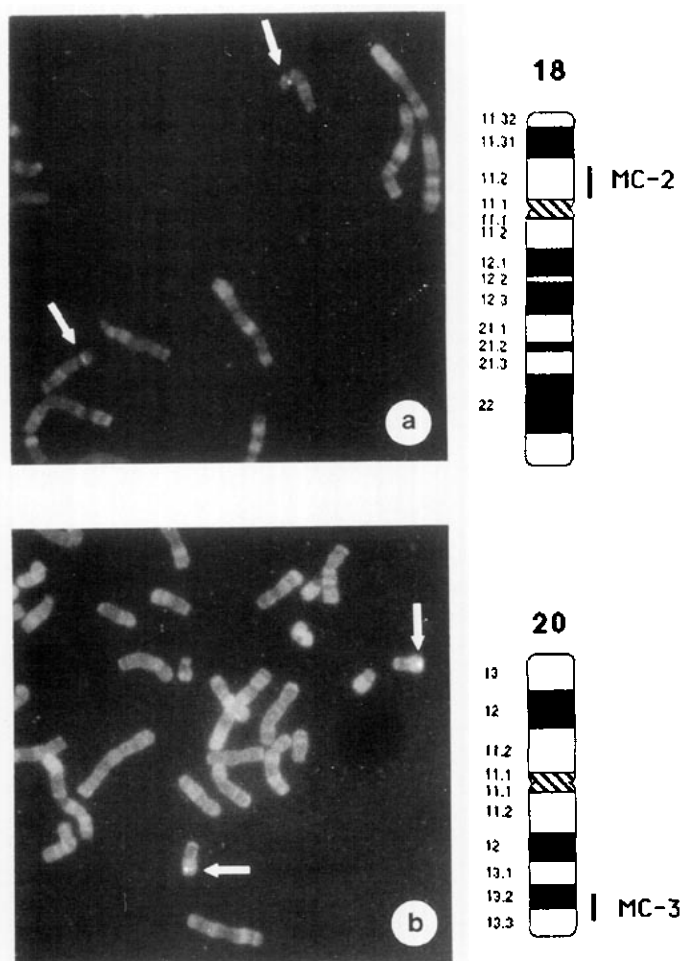


FIG. 1. Regional mapping of (a) the melanocortin-2 (ACTH) receptor gene to 18p11.2 and (b) the melanocortin-3 receptor gene to 20q13.2–q13.3 using the FISH technique. Hybridization signals were detected with FITC on propidium iodide-stained R-banded chromosomes.

¹ To whom correspondence should be addressed at 3101 Taubman Center, Ann Arbor, MI 48109-0386. Telephone: (313) 936-4770.

Using the FISH technique, we localized the ACTH and the melanocortin-3 receptors to chromosome loci 18p11.2 and 20q12.3-q13.2, respectively. We have reported previously the localization of the melanocortin-4 receptor to chromosome 18q21.3 (10). The scattered chromosomal distribution of the melanocortin receptors MC2, MC3, and MC4 appears to conform to a similar pattern of chromosomal localization seen in other human multisubtype seven transmembrane receptors (α_2 , β_1 , β_2 -adrenergic receptors, m1-5 muscarinic cholinergic receptors, D1, D2, and D4 dopamine receptors) (14). Only the human α -1A, α -1B adrenergic receptors share a common arm on the same chromosome. Apparent exceptions are the chemotactic receptors (complement 5A receptor, *N*-formyl peptide receptor, and two "orphan" receptors that are structural homologues of the *N*-formyl peptide receptor), which have all been mapped to chromosome 19 (15), and the receptors for the glycoproteins luteinizing hormone and follicle-stimulating hormone, which have both been localized to 2p21 (16). According to the Genome Database, the only gene previously localized at 18p11.2, the site of the melanocortin-2 receptor, encodes a putative receptor-like protein tyrosine phosphatase (17). No genes have been previously localized to 20q13.2-q13.3, the site of the melanocortin-3 receptor. The gene encoding the melanocortin-4 receptor shares 18q21 with the genes for *bcl-2* gene products (18) and plasminogen activator inhibitor, type II (19).

ACKNOWLEDGMENTS

This work was supported by NIH Grant R01DK34306 and funds from the University of Michigan Gastrointestinal Peptide Research Center (NIH Grant P30DK34933). Dr. Gantz is a recipient of a Veterans Administration Research Associate Award. Dr. Glover and Christine Barcroft are members of the University of Michigan Genome Center.

REFERENCES

- Nakanishi, S., Inoue, A., Kita, T., Nakamura, M., Chang, A. C. Y., Cohen, S. N., and Numa, S. (1979). Sequence of cloned cDNA for bovine corticotropin- β -lipotropin precursor. *Nature* **278**: 423-427.
- Tatro, J. B. (1993). Melanotropin receptors of the brain. In "Methods in Neurosciences" (P. M. Conn, Ed.), Vol. 11, pp. 87-104, Academic Press, New York.
- Walker, J. M., Akil, H., and Watson, S. J. (1980). Evidence for homologous actions of pro-opiomelanocortin products. *Science* **210**: 1247-1249.
- Cannon, J. G., Tatro, J. B., Reichlin, S., and Dinarello, C. A. (1986). Inflammatory action of interleukin 1. *J. Immunol.* **137**: 2232-2236.
- Gispén, W. H. (1992). Therapeutic potential for melanocortins in peripheral nerve disease. *Trends Pharmacol. Sci.* **11**: 221-222.
- Clark, D., Thody, A. J., and Shuster, S. (1978). Immunoreactive α -MSH in human plasma in pregnancy. *Nature* **273**: 163-164.
- Mountjoy, K. G., Robbins, L. S., Mortrud, M. T., and Cone, R. D. (1992). The cloning of a family of genes that encode the melanocortin receptors. *Science* **257**: 1248-1251.
- Chhajlani, V., and Wikberg, J. E. S. (1992). Molecular cloning and expression of the human melanocyte stimulating hormone receptor cDNA. *FEBS Lett.* **309**: 417-420.
- Gantz, I., Konda, Y., Tashiro, T., Shimoto, Y., Hiroto, M., Munzert, G., Watson, S. J., Delvalle, J., and Yamada, T. (1993). Cloning of a novel melanocortin receptor. *J. Biol. Chem.* **268**: 8246-8250.
- Gantz, I., Miwa, H., Konda, Y., Shimoto, Y., Tashiro, T., Watson, S. J., DelValle, J., and Yamada, T. (1993). Molecular cloning, expression, and gene localization of a fourth melanocortin receptor. *J. Biol. Chem.*, in press.
- Pinkel, D., Straume, T., and Gray, J. W. (1986). Cytogenetic analysis using quantitative, high-sensitivity, fluorescence hybridization. *Proc. Natl. Acad. Sci. USA* **83**: 2934-2938.
- Lichter, P., Tang, C.-J. C., Call, K., Hermanson, G., Evans, G. A., Housman, D., and Ward, D. C. (1990). High-resolution mapping of human chromosome 11 by *in situ* hybridization with cosmid clones. *Science* **247**: 64-69.
- Lemieux, N., Dutrillaux, B., and Viegas-Pequignot, E. (1992). A simple method for simultaneous R- or G-banding and fluorescence *in situ* hybridization of small single-copy genes. *Cytogenet. Cell Genet.* **59**: 311-312.
- Human Gene Mapping 11 London Conference. (1991). *Cytogenet. Cell Genet.* **58**: 1-2200.
- Bao, L., Gerard, N. P., Eddy, R. L., Shows, T. B., and Gerard, C. (1992). Mapping of genes for the human C5a receptor (C5AR), human FMLP receptor (FPR), and two FMLP receptor homologue orphan receptors (FPRH1, FPRH2) to chromosome 19. (1992). *Genomics* **13**: 437-440.
- Rousseau-Merck, M. F., Atger, M., Loosfelt, H., and Berger, R. (1993). The chromosomal localization of the human follicle-stimulating hormone receptor gene (FSHR) on 2p21-p16 is similar to that of the luteinizing hormone receptor gene. *Genomics* **15**: 222-224.
- Gebbink, M. F. B. G., van Etten, I., Hateboer, G., Suijkerbuijk, R., Beijersbergen, R. L., van Kessel, A. G., and Moolenaar, W. H. (1991). Cloning, expression and chromosomal localization of a new putative receptor-like protein tyrosine phosphatase. *FEBS Lett.* **290**: 123-130.
- Tsujimoto, Y., and Croce, C. M. (1986). Analysis of the structure, transcripts, and protein products of *bcl-2*, the gene involved in human follicular lymphoma. *Proc. Natl. Acad. Sci. USA* **83**: 5214-5218.
- Samia, J. A., Alexander, S. J., Horton, K. W., Auron, P. E., Byers, M. G., Shows, T. B., and Webb, A. C. (1990). Chromosomal organization and localization of the human urokinase inhibitor gene: Perfect structural conservation with ovalbumin. *Genomics* **6**: 159-167.