Short communication

Effect of substance P and protein kinase inhibitors on β-amyloid peptide-induced proliferation of cultured brain cells

Vijendra K. Singh a,b,*, Jui-Fen Cheng a, Sy-Jye C. Leu a

Program in Molecular Biology, Department of Biology, Utah State University, Logan, UT 84322-6800, USA
b Department of Psychiatry, University of Michigan Medical Center, MSRB-I, A520B, Ann Arbor, MI 48109-0656, USA

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Abstract

The present study investigated the effect of substance P (SP) and protein kinase inhibitors (H7 and HA1004) on β-amyloid peptide-induced proliferation of neonatal rat brain cells in primary cultures. The β-amyloid peptide 1-28 (designated as βAP28), at nanomolar concentrations (10^-9 M), significantly (P < 0.05) increased the proliferation of brain cells (presumably non-neuronal) as measured by [3H]thymidine uptake into DNA (mitogenesis). The effect was dependent on time of culture, concentration of βAP28, and presence of fetal calf serum. The supplementation of SP into cell cultures at time zero reversed the proliferative response of βAP28. Moreover, the βAP28-induced proliferation was inhibited by protein kinase inhibitor H7, but not by HA1004. Since H7 is a selective protein kinase C (PKC) inhibitor and SP action involves PKC, we conclude that βAP28 induces normal brain cell proliferation through PKC pathway of cell signaling.

Keywords β-Amyloid protein; Brain cell proliferation; Growth factor; Signal transduction, Alzheimer's disease

Cerebral amyloidosis or accumulation of β-amyloid protein 1-40 (designated as βAP40) in brain occurs during normal aging. However, the amount of βAP40 increases several fold in Alzheimer's disease (AD), suggesting a pathological link of the protein with disease [22]. The βAP40, a small molecular weight (4 kDa) protein containing approximately 40 amino acids [4], is produced by normal proteolytic processing [14] of a larger precursor molecule (90 to 130 kDa) known as β-amyloid precursor protein (βAPP). Both βAP40 and βAP28 fragment thereof have been shown to induce neurotrophic and neurotoxic effects on primary cultures of hippocampal cells [20,21]. Moreover, βAP40 induces growth factors in glial cell cultures [1] and is localized extraneuronally in the skin fibroblasts [7], eukaryotic cultured cells [2,5] and biological fluids [13]. This finding suggested that βAP40 is produced during normal cell metabolism presumably having a physiological function throughout life. The present study describes the effect of βAP28 on the proliferation of primary brain cell cultures and its sensitivity to SP and protein kinase inhibitors.

The primary brain cell cultures were prepared according to our previous work [15]. Sprague–Dawley neonatal rats (18 to 24 h old) were sacrificed under ether anesthesia. By using pre-sterilized dissecting tools, the brain was removed quickly and collected in RPMI-1640 growth medium. Brain tissue was dissociated and passed through a nylon sieve under aseptic conditions of a Laminar flow-hood and the cell suspension was passed through a 21G needle. The cell suspension was centrifuged at 800 × g for 10 min. The cell pellet, after one washing, was resuspended in RPMI-1640 containing penicillin (100 U/ml)-streptomycin (100 µg/ml), 10% heat-inactivated fetal calf serum (FCS) and 1% glucose. The cells were seeded into tissue culture flasks (25 cm² size) at a count of 2.5 × 10⁵ cells in 5 ml per flask and allowed to grow for 4 to 5 days inside the humified chamber of a CO₂ (5%)-incubator at 37°C. The confluent cultures were subcultured by trypsin (0.2%) treatment for about 5 min and
the cells were washed three times with RPMI-1640 medium. They were either subcultured or used to set up the experiments.

The cell proliferation was measured by $[^{1}H]$thymidine uptake as described before [16]. The cell suspension from freshly trypsinized cultures was pipetted into microwells of a flat-bottomed 96-well microtiter plate (Corning). Triplicate microwells were set up in the absence or presence of $\beta$AP28 in a final volume of 200 $\mu$l which included 100 $\mu$l of cell suspension containing approximately 10,000 cells per well. After 5 days of incubation in CO$_2$-incubator, the cells were labelled for 4 h with 0.4 $\mu$Ci/well of [methyl-$^{1}H$]thymidine (spec. act. 2 Ci/mmol, purchased from NEN, Boston, MA) and harvested using a semi-automatic cell harvester (Titertek model, Flow Labs.) onto Whatman glass-fiber filter paper. The filter paper discs (10 mm in diameter) were transferred into counting vials, mixed with 2 ml of Ready Safe scintillant (Beckman) and counted for radioactivity using a liquid scintillation spectrometer (Packard Model Tri-Carb 1500) with approximately 58% counting efficiency. The data are presented as the counts per minute (CPM) of radioactivity or the per cent of growth medium control. The statistical significance ($P \leq 0.05$) was assessed by Student's $t$-test using Statview software for the Macintosh computer.

Neuropeptide $\beta$AP28 (cat. #PNPE269) was a synthetic product of > 99% purity (HPLC analysis) purchased from Bachem California, Torrance, CA. Synthetic substance P (code 7451) was purchased from Peninsula Labs., Inc., Belmont, CA. The protein kinase inhibitors H7 (code 120805) and HA1004 (code 120804) were purchased from Seikagaku America, St. Petersburg, FL. The fetal calf serum (FCS) and penicillin-streptomycin mixture were purchased from Hyclone Labs. (Logan, UT) and Gibco (Grand Island, NY), respectively. The $\beta$AP28 and all other reagents were dissolved aseptically in RPMI containing penicillin-streptomycin but no FCS and stored frozen at $-70^\circ$C. Whenever needed, they were thawed and used fresh.

As shown in various figures, $\beta$AP28 displayed a stimulatory effect on the proliferation of brain cells in primary cultures. The proliferative response was significantly higher in the presence of $\beta$AP28 as compared to growth medium control (Fig. 1); the $P$ values were 0.003, 0.05 and 0.006 for 31 nM, 153 nM and 765 nM concentrations, respectively. However, it required fetal calf serum (FCS) since in the absence of FCS there was virtually no cell proliferation. The $\beta$AP28-induced cell proliferation increased up to 5 days of incubation that was used in subsequent experiments. The stimulatory effect of $\beta$AP28 was observed both pre- and post-cell plating (Fig. 2) although the overall response was slightly lower if the cells had been pre-seeded overnight (16 to 18 h). The supplementation of substance P (SP) into cultures completely blocked the proliferative response of $\beta$AP28 (Fig. 3). In addition, the $\beta$AP28-induced stimulation of cell proliferation was inhibited by protein kinase inhibitor H7, but not by HA1004; the two inhibitors were tested at a concentration of 20 $\mu$M against the three concentrations of $\beta$AP28 (Fig. 4).

Based on their pathologic link with Alzheimer's disease [22], the amyloid proteins of neuritic plaques have been intensely studied during the last decade. However, the pathological role of $\beta$AP40 in AD continues to be a matter of debate and controversy especially in light of genetic associations of apolipoprotein E [12] and non-neuronal expression of $\beta$AP40 during
normal cell metabolism [2, 5, 7, 13]. While a causative role of \( \beta \)AP40 in AD continues to be established, this protein may have some biological function during normal cell metabolism. This line of thinking stems from several studies: (i) \( \beta \)AP40 and its fragment \( \beta \)AP28 exert trophic and toxic effects on hippocampal neurons [20, 21]; (ii) \( \beta \)AP40 expression occurs predominantly in astrocytic glial cells [2]; (iii) \( \beta \)AP40 occurs naturally in cells that are not brain-derived [5, 7, 13]; (iv) \( \beta \)AP40 induces glia-derived growth factors [1]; (v) \( \beta \)AP40 eliminates a functional potassium channel but not calcium channel in human fibroblasts [3]; (vi) \( \beta \)AP40 and \( \beta \)AP28 induce DNA synthesis in immunocytes of normal blood donors but not AD patients [18]; and (vii) \( \beta \)AP40 increases proliferation of neonatal rat brain cells as reported in our present study. Since neuronal proliferation ceases early in embryogenesis whereas glial cells continue to divide into postnatal periods of development, the \( \beta \)AP28-induced proliferative activity would be related to non-neuronal cells presumably astrocytes. Based on this consideration and the fact that astrocytes are a major source of \( \beta \)AP in brain [2], we suggest that this peptide acts as an autocrine hormone on glial cells and a paracrine hormone on neurons under physiological conditions in vivo.

Our finding of a proliferation-enhancing effect by \( \beta \)AP28 is similar to a neurotrophic effect [20, 21] but whether the two involve same or completely different mechanisms is not presently known. One notable difference, however, is that the proliferative response can be induced by 100- to 3000-fold lower concentrations of \( \beta \)AP28 than the concentrations required for neurotrophic effect; we used a concentration range of 30 to 765 nM as compared to 3 to 30 \( \mu \)M in one study [20] and 100 \( \mu \)M in another [21]. The latter study [21] showed that the trophic effect of \( \beta \)AP40 was maximal by approximately 0.1 nM, but \( \beta \)AP28 was much less potent than \( \beta \)AP40 and required a 100 \( \mu \)M concentration to yield only 29% of the trophic response. Furthermore, the \( \beta \)AP28-induced proliferative response required serum since there was no proliferation in the absence of fetal bovine serum (FCS). The reason for this requirement is presently not known but we suggest that \( \beta \)AP28 may interact with some serum protein(s) ensuring a proliferative response. Also, the morphological differentiation was not a prerequisite since the stimulation of cell proliferation took place irrespective of the time of addition (at time zero or after overnight seeding) of \( \beta \)AP28 into cultures.

SP has been shown to prevent neurotrophic and neurotoxic effects of \( \beta \)AP40 on hippocampal neurons [21] We also found that SP completely reverses \( \beta \)AP28 action on cell proliferation. In contrast, SP did not modify \( ^{125} \)I-\( \beta \)AP40 binding to brain homogenates [9] or truncated \( \beta \)AP22–35-induced neurotoxicity [19]. This discrepancy may be related to assay differences in different studies: (i) \( ^{125} \)I-\( \beta \)AP40 binding was assayed with pathological tissue (AD brain) [9] but not with normal tissue and the homogenate contained bovine serum albumin and protease inhibitors (bacitracin, chymostatin, leupeptin and dimethyl sulfoxide) that may alter binding characteristics; and (ii) a truncated \( \beta \)AP22–25 form [19], instead of \( \beta \)AP40 or \( \beta \)AP28, was added to cultures after 5 days of cell differentiation but not at time zero. Since the \( \beta \)AP28-induced response was counteracted by SP, we suggest that this peptide interacts with a molecule structurally similar to SP receptor or functionally coupled to it but the nature of this putative molecule remains unknown.

Indirectly through the use of protein kinase inhibitors, an approach recently used by us for second messenger studies of another neuropeptide [17], we found that the proliferative response of \( \beta \)AP28 was preferentially inhibited by H7, a selective inhibitor of PKC [6], but not by HA1004 which is a PKA inhibitor and intracellular Ca\(^{2+}\) antagonist without any in-

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**Fig. 3** Effect of substance P on the action of \( \beta \)AP28. The cell concentration was approximately 10,000 cells per microwell. All other conditions of incubation were the same as those given under Fig. 1. Whenever present, the substance P (+ SP) was at a concentration of 20 \( \mu \)M. The data are given as the percent of control (100%) that did not contain \( \beta \)AP28.

**Fig. 4** Effect of protein kinase inhibitors on \( \beta \)AP28-stimulated cell proliferation. Three different concentrations of \( \beta \)AP28 were set up with approximately 10,000 cells per microwell. Depending on design, the H7 or HA1004 was added at a concentration of 20 \( \mu \)M and the response was measured as described under Fig. 1.
hibitory effect on PKC [6]. This finding implies that the mechanism of proliferation-enhancing action of βAP28 involves a PKC-activated pathway of signal transduction; a mechanism strengthened by a recent study [8] which showed that in the presence of glutamate βAP40 activates tachykinin receptors and phosphatidylinositol (PI) turnover. PKC is a key enzyme in mediating the action of neurotrophic factors to maintain neuronal survival and βAP28 certainly behaves like a growth factor [1,20,21, present study]. In addition, SP, which prevents βAP28-induced proliferation, also involves PKC-activated phosphoinositide turnover [10], indicating that the two peptides might compete with each other for interaction to PKC system. Based on PKC involvement in βAP28 action [present study] and PKC (βII isoform) reduction in AD brains [11], we postulate that βAP28 controls cellular (PKC activity) and molecular (DNA synthesis) functions of normal brain cells but aberration of its function at a purported site of (βII isoform) reduction in AD brains [11], we postulate other for interaction to PKC system. Based on PKC involvement in βAP28 action [present study] and PKC (βII isoform) reduction in AD brains [11], we postulate that βAP28 controls cellular (PKC activity) and molecular (DNA synthesis) functions of normal brain cells but aberration of its function at a purported site of action may form neuritic plaques in the CNS Thus, our study probes into the mechanism of cellular action of this peptide which may help in understanding the pathological link between β-amylloid protein and Alzheimer’s disease.

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