

## Mouse Chromosome 3

Michael F. Seldin,<sup>1</sup> Jan-Bas Prins,<sup>2</sup> Nanda R. Rodrigues,<sup>2</sup> John A. Todd,<sup>2</sup> Miriam H. Meisler<sup>3,\*</sup>

<sup>1</sup>Duke University Medical Center, Departments of Medicine and Microbiology, Research Drive, Room 247, CARL Building, Box 3380, Durham, North Carolina 27710-3380, USA

<sup>2</sup>Nuffield Department of Surgery, John Radcliffe Hospital, Headington, Oxford OX3 9 DU, UK

<sup>3</sup>Department of Human Genetics, University of Michigan, Ann Arbor, Michigan 48109-0618, USA

Received: 10 May 1993

### Introduction

During the past year, more than 80 new loci have been assigned to Chromosome (Chr) 3, including 15 genes and a large number of anonymous DNA markers. This wealth of markers makes it increasingly difficult to represent the genetic maps of mouse chromosomes in traditional formats. In this report, map positions for Chr 3 loci are presented in tabular form. A traditional graphic representation of Chr 3 is available in the first and second committee reports (103, 103a).

This report provides an updated locus list and map, updated strain distribution patterns for recombinant inbred lines, recombination data from seven large multilocus crosses, a new map entirely based on PCR-based microsatellite loci that span Chr 3, and primer sequences for a large number of markers that can be detected by the PCR.

### Locus list and chromosome map

The main features of Chr 3 loci are presented in Table 1. Map positions were calculated by Seldin using the methods previously described (103, 103a). Entries that have been added or changed since the previous report are marked with an asterisk. The map positions are composites based on a large number of measurements. The 95% confidence intervals for the primary data are in most cases greater than 2 cM, and those for the composite data are greater than 5 cM. As discussed in the previous reports, there may be errors in the indicated gene orders for closely linked loci that have not been mapped in the same cross.

The map positions that are listed alphabetically in Table 1 are also listed in order of increasing distance from the centromere in Table 2, providing a tabular representation of the Chr 3 map.

Data on gene order obtained from seven multilocus crosses are summarized in Fig. 1A. Unambiguous gene order can be determined only for loci that were mapped within the same cross. The data from the seven crosses generate a consistent gene order. The quantitative data from these crosses are presented in Fig. 1B.

### PCR primers for Chr 3 loci

The development of PCR-based assays that detect genetic variation has greatly reduced the time and effort required for genotyping, as well as the amount of genomic DNA required per assay. PCR primers amplify products of different lengths as a result of variation in simple sequence repeat length. Published gene sequences have been used to derive primers that detect variation at known loci. In addition, a large number of primers that detect (CA)<sub>n</sub> repeat length variation have been developed from anonymous genomic clones by the Mouse Genome Center at the Massachusetts Institute of Technology (MIT) (37). Primer sequences for 72 Chr 3 loci of both types are presented in Table 3.

### Microsatellite map of Chr 3

During the past year, microsatellite markers have been widely adopted by many investigators as convenient and reliable genetic markers. The presence of variation among inbred strains, the reproducibility of as-

\* Chair of Committee for Mouse Chromosome 3

Table 1. Locus list for mouse Chr 3.

New Symbol	Name	A	Map	T	Method	H. symbol	H. location	Notes	References
<i>Acrb-2</i>	acetylcholine receptor beta 2 neural		43.0	D	L			8	11
<i>Acts</i>	skeletal alpha actin		X	D	S,R	ACTA1	1p21-qter	8,12	33
<i>Adh-1</i>	alcohol dehydrogenase-1	1	68.1	B,D	L,R	ADH1	4q21-q23	4,5	13,15,20,22,54, 68,150,159
<i>Adh-1ps</i>	alcohol dehydrogenase-1 pseudogene		52.0	D	R				20
<i>Adh-1t</i>	alcohol dehydrogenase-temporal		68.1	B	L			13	6,71
<i>Adh-3</i>	alcohol dehydrogenase-3		68.1	B	R,L	ADH3	4q21-q23	12	43,67,68,69,72, 112,124
<i>Adh-3t</i>	alcohol dehydrogenase-3-temporal		68.1	B	L			12	66,69
<i>Adh-5</i>	alcohol dehydrogenase-5		X	B	S	ADH5	4q21-q25		53
<i>Alr-1</i>	aldehyde reductase-1		68.1	B	R,L			12	43
<i>Ampd-1</i>	AMP deaminase-1 (muscle form)		47.4	D	L	AMPD1	1p13	1	79
<i>Ampd-2</i>	AMP deaminase-2 (nonmuscle form)		50.2	D	L			1	79,111,118
<i>Amy-1</i>	amylase, salivary	1	49.6	B,D	L,R	AMY1	1p21	8,10,12	13,15,45,52, 59,86,121, 137,138,156
<i>Amy-2</i>	amylase, pancreatic	1	49.6	B,D	L,R	AMY2	1p21	1,2	13,14,109,121
* <i>Ank-2</i>	brain specific ankyrin-2		58.4	D	L			12	122
<i>Ap2</i>	adipocyte protein aP2		9.7	B,D	R,L			4	61,150
<i>Arnt</i>	aryl hydrocarbon receptor nuclear translator		X	B	S	ARNT	1pter-q12		17
<i>Atplal</i>	Na, K ATPase alpha-1		47.3	D	L	ATP1A1	1p13	1,13	77,79,81,109
* <i>Atpa-1</i>	see Atplal								
* <i>Bglap</i>	bone matrix Gla protein			D	S	BGLAP			76
<i>Bmn</i>	Beta-mannosidase activity (liver, kidney)		68.1	B	R				97
<i>BRS-3</i>	see Odc-rs3								
<i>Cacy</i>	calcyclin		42.9	D	L	CACY	1q21-q25	1,12	39,79,109,118
<i>Call1</i>	calpactin I light chain		42.9	D	R				131,151
<i>Calla</i>	see Mmc								
<i>Capl</i>	calcium binding protein, placental		42.9	D	R	CAPL	1q12-q22		39,151
<i>Car-1</i>	carbonic anhydrase-1		6.3	B	L	CA1	8q13-q22	13	44
<i>Car-2</i>	carbonic anhydrase-2	1	6.3	B,D	L,R	CA2	8q13-q22	1,7,8,13	15,23,28,44, 47,109, 117,121,156,161
<i>Car-3</i>	carbonic anhydrase-3		7.5	D	L	CA3	8q13-q22	13	9
* <i>Ccna</i>	Cyclin a		15.0	D	L			1,2	92,129,134
<i>Cd1</i>	cluster designation 1		43.9	D	L	CD1	1q22-23	1	23,110,111,118
<i>Cd2</i>	cluster designation 2		46.4	D	L	CD2	1p13	1	78,79,109,118
<i>Cd10</i>	see Mmc								
* <i>Cd53</i>	cluster designation 53		47.4	D	L			1	168
<i>cdm</i>	cadmium resistance		61.6	V	R				145,146
* <i>Cf-3</i>	coagulation factor 3		49.6	D	R	F3	1p22-p21		98
<i>Cnp-2</i>	cyclic nucleotide phosphodiesterase-2		38.8	D	R				10
<i>Cnx40</i>	connexin (see Gja-5)								
<i>coa</i>	cocoa		8.3	V	L			13	117,142
<i>Csfm</i>	colony stimulating factor, macrophage (alternative for op)		48.4	B,D,V	L	CSF1	1p13-21	2,8	18,54,86,93,169
* <i>D3Byu1</i>	DNA segment, Brig Young Univ. 1 (RAPD)		6.9	D	R				167
* <i>D3Byu2</i>	DNA segment, Brig Young Univ. 2 (RAPD)		6.9	D	R				167
* <i>D3Byu3</i>	DNA segment, Brig Young Univ. 3 (RAPD)		8.1	D	R				167
* <i>D3Byu4</i>	DNA segment, Brig Young Univ. 4 (RAPD)		45.3	D	R				167
* <i>D3Byu5</i>	DNA segment, Brig Young Univ. 5 (RAPD)		34.4	D	R				167
* <i>D3Byu6</i>	DNA segment, Brig Young Univ. 6 (RAPD)		42.3	D	R				167
* <i>D3Byu17</i>	DNA segment, Brig Young Univ. 17 (RAPD)		30.1	D	R				167
* <i>D3Do1</i>	P40-4 clone ? high affinity laminin		76.4	D	R				40
* <i>D3Hun1</i>	DNA segment, Chr 3, Hunter 1		56.1	D	L			1	73
* <i>D3J1</i>	DNA segment, Chr 3, Jackson Lab 1		33.1	D	R				114
* <i>D3J2</i>	DNA segment, Chr 3, Jackson Lab 2		65.6	D	R				114
* <i>D3J3</i>	DNA segment, Chr 3, Jackson Lab 3		54.7	D	R				114
* <i>D3Jfr1</i>	DNA segment, Chr 3, MJeffers-1		47.4	D	P				75
* <i>D3Jkn1</i>	DNA segment, Chr 3, Ian Jackson 1		69.0	D	R				99
* <i>D3Leh1</i>	DNA segment, Chr 3, Lehrach 1		17.6	D	L			2	31
* <i>D3Leh2</i>	DNA segment, Chr 3, Lehrach 2		10.2	D	L			2	31
* <i>D3Ler1</i>	DNA segment, Chr 3, Leroy 1		41.5	D	L			8	91
* <i>D3Ler2</i>	DNA segment, Chr 3, Leroy 2		41.5	D	L			8	91
* <i>D3Mit1</i>	DNA segment, Chr 3, MIT 1		7.0	D	L			3,4,5	37,150,159
* <i>D3Mit3</i>	DNA segment, Chr 3, MIT 3		20.0	D	L			3	37
* <i>D3Mit4</i>	DNA segment, Chr 3, MIT 4		20.0	D	L			3	37
* <i>D3Mit5</i>	DNA segment, Chr 3, MIT 5	1	24.0	D	L			3,4	37,150
* <i>D3Mit6</i>	DNA segment, Chr 3, MIT 6		22.0	D	L			3	37,150
* <i>D3Mit7</i>	DNA segment, Chr 3, MIT 7		30.0	D	L			3	37
* <i>D3Mit9</i>	DNA segment, Chr 3, MIT 9		38.6	D	L			3	37
* <i>D3Mit10</i>	DNA segment, Chr 3, MIT 10		48.6	D	L			4,5	37,159
* <i>D3Mit11</i>	DNA segment, Chr 3, MIT 11		48.6	D	L			3,4	37,150
* <i>D3Mit12</i>	DNA segment, Chr 3, MIT 12		50.3	D	L			3	37
* <i>D3Mit13</i>	DNA segment, Chr 3, MIT 1		57.7	D	L			3	84
* <i>D3Mit14</i>	DNA segment, Chr 3, MIT 14		61.0	D	L			3	37
* <i>D3Mit15</i>	DNA segment, Chr 3, MIT 15		63.1	D	L			3	37
* <i>D3Mit16</i>	DNA segment, Chr 3, MIT 16		63.1	D	L			3	37
* <i>D3Mit17</i>	DNA segment, Chr 3, MIT 17		68.7	D	L			3	37
* <i>D3Mit18</i>	DNA segment, Chr 3, MIT 18		73.0	D	L			1,3	37,134,161
<i>D3Mit19</i>	DNA segment, Chr 3, MIT 19	1	81.0	D	L			1,3,4	134,150,159
<i>D3Mit21</i>	see Il-2								
* <i>D3Mit22</i>	DNA segment, Chr 3, MIT 22	1	35.0	D	L			3,4	84,150
* <i>D3Mit23</i>	DNA segment, Chr 3, MIT 23		0.0	D	L			3	84
* <i>D3Mit24</i>	DNA segment, Chr 3, MIT 24		20.0	D	L			3	84

Continued on next page

Table 1. Continued.

*	<i>D3Mit25</i>	DNA segment, Chr 3, MIT 25	30.0	D	L			3	84
*	<i>D3Mit26</i>	DNA segment, Chr 3, MIT 26	38.6	D	L			3	84
*	<i>D3Mit28</i>	DNA segment, Chr 3, MIT 28	44.9	D	L			3	84
*	<i>D3Mit31</i>	DNA segment, Chr 3, MIT 31	73.0	D	L			3,4	84,150
*	<i>D3Mit32</i>	DNA segment, Chr 3, MIT 32	77.0	D	L			3,4	84,150
*	<i>D3Mit36</i>	DNA segment, Chr 3, MIT 36	53.4	D	L			3	84
*	<i>D3Mit38</i>	DNA segment, Chr 3, MIT 38	67.2	D	L			3	84
*	<i>D3Mit39</i>	DNA segment, Chr 3, MIT 39	53.4	D	L			3	84
*	<i>D3Mit40</i>	DNA segment, Chr 3, MIT 40	40.1	D	L			3	84
*	<i>D3Mit41</i>	DNA segment, Chr 3, MIT 41	46.1	D	L			3	84
*	<i>D3Mit42</i>	DNA segment, Chr 3, MIT 42	56.1	D	L			3	84
*	<i>D3Mit45</i>	DNA segment, Chr 3, MIT 45	75.3	D	L			3	84
*	<i>D3Mit46</i>	DNA segment, Chr 3, MIT 46	9.6	D	L			3	84
*	<i>D3Mit49</i>	DNA segment, Chr 3, MIT 49	40.1	D	L			3	84
*	<i>D3Mit51</i>	DNA segment, Chr 3, MIT 51	36.2	D	L			3	84
*	<i>D3Mit53</i>	DNA segment, Chr 3, MIT 53	35.0	D	L			3	84
	<i>D3Nds1</i>	DNA segment, Chr 3, Nottingham Dept. Surgery 1	27.0	D	L			4,5,9	150,159
*	<i>D3Nds2</i>	DNA segment, Chr 2, Nottingham Dept. Surgery 2	61.0	D	L,R			3	84
*	<i>D3Nds3</i>	DNA segment, Chr 2, Nottingham Dept. Surgery 3	63.0	D	R				30
	<i>D3Nds6</i>	see Il-2							
	<i>D3Nds8</i>	see Tshb							
	<i>D3Nds9</i>	see Adh-1							
*	<i>D3Pas1</i>	DNA segment, Chr 3, Pasteur Institute-1	20.9	D	R				135
*	<i>D3Pas2</i>	DNA segment, Chr 3, Pasteur 2	13.2	D	L			8	123
*	<i>D3Pas501</i>	DNA segment, Chr 3, Pasteur 501-temporary designation	8.2	D	L			8	26
*	<i>D3Pas502</i>	DNA segment, Chr 3, Pasteur 502-temporary designation-FP25	42.9	D	L			8	123
	<i>D3Sell</i>	DNA segment, Chr 3, Seldin 1	48.1	D	L			1	162
	<i>D3Sel2</i>	DNA segment, Chr 3, Seldin 2	30.1	D	L			1	162
	<i>D3Tu33</i>	DNA segment, Chr 3 Tubingen-33	57.9	D	R			3	153
	<i>D3Tu51</i>	DNA segment, Chr 3 Tubingen-51	42.9	D	L,R			1	118
*	<i>D3Uf1</i>	DNA segment, Chr3 Wakeland 1 (RAPD)	43.5	D	L			5	159
*	<i>D3Uf2</i>	DNA segment Chr 3 Wakeland 2 (RAPD)	52.3	D	L			5	159
	<i>de</i>	droopy ear	48.4	V	L			12,13	32,68,86,87
	<i>Egf</i>	epidermal growth factor	1 62.1	D	L,R	EGF	4q25	1,2,7	52,109,112,137,161,170
	<i>Emv-27</i>	endogenous ecotropic MuLV-27	49.6	D	L			13	148
	<i>Es-16</i>	esterase-16	10.1	B	L			12	154,156,158
	<i>Es-26</i>	esterase-26	33.3	B	L			12	117,154,156,158
	<i>Es-27</i>	esterase-27, serum cholinesterase	23.3	B	L			12	157,158
	<i>Evi-1</i>	ecotropic viral integration site-1	10.2	D	L,R	EV11	3q24-q28	1,2,6	21,23,27,28,31,54,62,92,109,112,141
	<i>Fabpi</i>	fatty acid binding protein intestinal	54.7	D	R	FABP2	4q28-q31		143
	<i>Fcgr1</i>	high affinity FC gamma receptor	42.9	D	L	FCCGR1	1q		1,12,118,120
	<i>Fgf2</i>	fibroblast growth factor basic	15.7	D	L,L	FGF2	4q25-27	1,2	23,31,62,92,102,141
	<i>Fgg</i>	gamma fibrinogen	42.3	D	L,R	FGG	4q28	2	13,31,59,92,93,94
	<i>Fim-3</i>	Friend MuLV integration site-3	10.2	D	L	FIM3	3q27	2,8	7,27,54,139
	<i>Fpsl-rs1</i>	farnesyl pyrophosphate synthetase - like 1	43.9	D	L	FPSL	1q24-q31	1	134
	<i>ft</i>	flaky tail	42.4	V	L				85,86
	<i>Gba</i>	beta glucocerebrosidase	1 42.6	B,D	L	GBA	1q21	1,12	109,118
	<i>Gbp-1</i>	guanine nucleotide-binding protein-1	64.3	D	L,R			12	124
*	<i>Gja-5</i>	gap junction protein - connexin 40	45.5	D	L			2	59,94
*	<i>Glur-2</i>	glutamate receptor 2	34.9	D	L	GLUR2	4q25-34	1	58
	<i>Glut-2</i>	glucose transporter 2	10.2	D	L	GLUT2	3q26	2,5	62
*	<i>Gnai-2</i>	guanine nucleotide binding protein, alpha inhibiting activity-2	48.4	D	L			2	166
	<i>Gnai-3</i>	guanine nucleotide binding protein, alpha inhibiting activity-3	48.4	D	L	GNAI3	1p13	2,4	166
*	<i>Gsl-4</i>	globoglycolipid expression-4	X	V	R				115
	<i>H-23</i>	histocompatibility-23	59.6	B	L,R			12	4,106
	<i>H-28</i>	histocompatibility-28	79.3	B	L,R			12	4,106
	<i>H-37</i>	histocompatibility-37	(X)	B	R				4
*	<i>H3f2</i>	histone 3, family 2	44.9	D	R	H3F2	1q21-21		132
*	<i>Hao-2</i>	hydroxyacid oxidase-2 (kidney)	40.0	B	L			8,12	54,64,65
*	<i>Hc3</i>	heterochromatin, Chr 3	0.0		L			6	21
	<i>Hist2</i>	histone gene (2)	X	D	S				56
	<i>Hnl</i>	hypothalamic norepinephrine level	59.6	V	L				46
	<i>Hsd3b</i>	3-beta-hydroxy steroid dehydrogenase	45.0	D	L	HSDB3	1p11-p13		5
	<i>Hsp86-ps2</i>	heat shock protein 86- pseudogene 2	19.3	D	S,L				107,108
*	<i>Iap1a1-7</i>	Intracisternal A-particle a1-7	42.3	D	R				95
*	<i>Iap1a2-9</i>	Intracisternal A-particle a2-9 (near Fgg)	X	D	R				94
*	<i>Iap1a2-14</i>	Intracisternal A-particle a2-14	74.6	D	R				94
*	<i>Iap1a3-13</i>	Intracisternal A-particle a3-13	28.0	D	R				95
	<i>Id4-3</i>	insulin dependent diabetes 3	X	V	L				149
	<i>If-1</i>	interferon inducibility locus	84.6	V	L,R			12	36,106
	<i>Il-2</i>	interleukin 2	1 15.0	D	S,L	IL2	4q26-q27	1,3,4,9	49,134,149,150,159,162,163
	<i>Il-7</i>	interleukin 7	2.0	B,D	L			2,6	21,62,137
*	<i>Kv1.2</i>	potassium channel gene	48.2	D	L			2	93

Continued on next page

Table 1. Continued.

*	<i>Kvj.3</i>	potassium channel gene	48.2	D	L			2	93
*	<i>Kvj.3rs3</i>	potassium channel gene 1.3 related sequence	48.4	D	L			2	93
	<i>Lef-1</i>	lymphoid enhancer-binding factor 1	X	D	R,S	LEF1	4q23-q25		105
*	<i>M6pr-ps</i>	cation-dependent mannose 6-phosphate receptor pseudo gene	45.5	D	L			2	94
	<i>ma</i>	matted	40.4	V	L			10,12	85,86,87,106
	<i>Mme</i>	membrane metallo-endo peptidase (neutral endopeptidase)	30.1	D	L	MME	3q21-27	1,7	23,161
	<i>Mmv-2</i>	MCF endogenous virus-2	X	D	S				63
	<i>Mmv-12</i>	MCF endogenous virus-12	X	D	S				63
	<i>Mov-10</i>	Moloney leukemia virus-10	X	D	S				74,113
	<i>Mpmv-9</i>	modified polytropic murine leukemia virus-9	88.2	D	L,R			11	51
	<i>Mpmv-20</i>	modified polytropic murine leukemia virus-20	9.6	D	R				51
	<i>my</i>	blebs	30.4	V	L			10	19,35,45
	<i>Ngfb</i>	nerve growth factor beta	47.4	D	L	NGFB	1p13	1,2	18,42,52,59,79,94, 109,118,166,171
	<i>Nras</i>	Nras oncogene	47.4	D	L	NRAS	1p13	1,2	18,93,118,128
*	<i>Nscl-2</i>	see Tau-1							
	<i>Oat-rs2</i>	ornithine aminotransferase related sequence 2	52.4	D	L				125
	<i>Odc-rs3</i>	ornithine decarboxylase-3	X	D	R				126
	<i>op</i>	osteopetrosis (Csfm mutation see Csfm)							
	<i>Otf-3rs3</i>	octamer transcription factor -3 related sequence 3	0.0	D	L			2,7	137,161
	<i>Otf-3rs4</i>	octamer transcription factor-3 related sequence 4		62.1	D	L			1,2,7,137,161
*	<i>Otf-3rs9</i>	octamer transcription factor-3 related sequence 9		16.4	D	L			7,161
	<i>Oua-1</i>	ouabain resistance-1	X	V	S				83
	<i>Pgk-1ps3</i>	phosphoglycerate kinase-1 pseudogene 3	8.7	D	S,R				1
	<i>Pk-1</i>	pyruvate kinase (may be the same as Pklr)	33.6	B	L			8	54,139
	<i>Pklr</i>	pyruvate kinase liver, red blood cells (see Pk-1)		42.6	D	L	PKLR	1q21	1,118
*	<i>Pmp-1</i>	peroxisomal membrane protein (70k)	54.6	D	L	PMP1	1p21-22	2	52
	<i>Pmv-26</i>	polytropic murine virus-26	71.8	D	R				50
	<i>Pmv-28</i>	polytropic murine virus-28	42.9	D	R				50
	<i>Pmv-38</i>	polytropic murine virus-38	43.1	D	R				50
	<i>Pmv-39</i>	polytropic murine virus-39	53.8	D	R				50
	<i>Rap1a</i>	member of RAS oncogene family	47.4	D	L	RAP1A	1p12-p13	1	41
	<i>rcm</i>	rostral cerebellar malformation	65.4	V	L			12	89,90
*	<i>Rn7s-3</i>	7s RNA related sequence -3	63.0	D	R				147
	<i>Rnu1b-1</i>	U1b1 small nuclear RNA	42.7	D	R	RNU1			96
	<i>Rnu1b-3</i>	U1b3 small nuclear RNA	42.9	D	R				13,96
	<i>soc</i>	soft coat	43.4	V	L			10	45,140
	<i>spa</i>	spastic	37.4	V	L			10	85,86
	<i>suc-1</i>	see Suc-1r							
	<i>Suc-1r</i>	sucrase-isomaltase, regulatory	33.3	B	L				12
	<i>Suc-1s</i>	sucrase-isomaltase, structural	33.3	D	R	SI	3q25-26		12
	<i>sut</i>	subtle gray	12.2	V	L			12	88
	<i>Tau-1</i>	basic domain helix-loop-helix (bHLH)	50.6	D	L			12	55
*	<i>Thbs3</i>	thrombospondin 3	43.9	D	L				1
*	<i>Tkr</i>	tyrosine kinase receptor (Ngf is ligand)	43.9	D	L				1
	<i>Tmevd-2</i>	TMEV induced demyelinating disease susceptibility	8.4	V	R				104
*	<i>Tpi-2</i>	triosephosphate isomerase related sequence-2	36.4	D	L			2	138
	<i>Tshb</i>	thyrotropin stimulating hormone beta subunit	1	47.4	D	L	TSHB	1p13	1
	<i>Va</i>	varitint-waddler	71.6	V	L			10,12	13,42,79,82,109, 116,118,150,159 86,87,106
	<i>Xmmv-22</i>	xenotropic-MCF leukemia virus - 22	42.3	D	R				13
	<i>Xmmv-47</i>	xenotropic-MCF leukemia virus - 47	31.0	D	R				164
	<i>Xmmv-65</i>	xenotropic-MCF leukemia virus - 65	42.3	D	L,R			12	164
*	<i>Yb1d</i>	YB-1 DNA binding protein related sequence d	14.6	D	L			2	141

The Chr 3 map positions are an estimate of distances, in cM, from the centromere. The position of the centromere is determined by heterochromatin mapping data (21, 101). Since recombination frequencies may vary depending on the specific cross, composite map positions may distort gene order when loci have not been mapped in an individual backcross. In deriving the composite map, RI strain data was used to determine gene position only as a supplement to backcross data. For a fuller discussion of the generation of map positions, see text and (133). In addition to the references cited for each locus, data used to derive map positions are described in the following "Notes:" (1) Duke University cross: complete haplotypes in 114 and incomplete haplotypes in 338 interspecific [(C3H/HeJ-*gld* × *M. spretus*)F<sub>1</sub> × C3H/HeJ-*gld*] backcross mice (134); (1a) same cross as above, but fewer than 40 meiotic events examined; (2) Frederick cross: complete haplotypes in 83-198 interspecific [(C57BL/6J × *M. spretus*)F<sub>1</sub> × C57BL/6J] backcross mice; (2a) same cross as 2, but fewer than 50 meiotic events examined; (3) Rockefeller University cross and MIT microsatellite mapping panel: complete haplotype data

in 40-48 (C57BL/6J-*ob* × CAST/Ei)F<sub>2</sub> intersubspecific intercross mice; (4) European Collaborative Interspecific Backcross [(C57BL/6 × SPR)F<sub>1</sub> × SPR] produced by the UK's Human Genome Mapping Project, with the support of the Medical Research Council (15); (5) data from the cross [(NOD/*Uf* × C57BL/6J)F<sub>1</sub> × NOD/*Uf*] from the laboratory of E.K. Wakeland. The RAPD polymorphism *D3Uf1* was detected with the primer GTGCCTAACC, and *D3Uf2* with the primer TGCTCACTGA; (6) RPMI cross: complete haplotypes in 130-140 [(C57BL/6 × *M. spretus*)F<sub>1</sub> × *M. spretus*]; (7) Duke University cross #2: complete haplotype data in 100-182 [(MRL/MpJ-*lpr* × CAST/Ei)F<sub>1</sub> × MRL/MpJ-*lpr*] intersubspecific backcross mice (161); (8) Pasteur cross: incomplete haplotypes in 38-74 interspecific backcross mice (J.-L. Guénet, unpublished data); (9) same as 4 with complete haplotype data in 92-299 mice; (10) included in nine overlapping three- or four-point crosses that derive from analysis of 125-500 meiotic events in each of multiple individual crosses; (11) haplotype data in 75 interspecific backcross mice (W.N. Frankel, unpublished data); (12) three-point mapping data; (13) two-point mapping data.

**Table 2.** Tabular map of Chr 3. Loci described in Table 1 are arranged in order of increasing distance from the centromere. The 95% confidence intervals for these composite data are greater than 5 cM. Unambiguous gene order can only be determined for loci which were mapped within the same cross (Fig. 1B).

Map	Locus				
0.0	D3Mit23	28.0	<i>Iap1a3-13</i>	42.9	<i>Cap1</i>
0.0	<i>Hc3</i>	30.0	D3Mit25	42.9	D3Pas502
0.0	<i>Otf-3rs3</i>	30.0	D3Mit7	42.9	D3Tu51
2.0	<i>Il-7</i>	30.1	D3Byu17	42.9	<i>Fcgr1</i>
6.3	<i>Car-1</i>	30.1	D3Sel2	42.9	<i>Pmv-28</i>
6.3	<i>Car-2</i>	30.1	<i>Mme</i>	42.9	<i>Rnu1b-3</i>
6.9	D3Byu1	30.4	<i>my</i>	43.0	<i>Acrb-2</i>
6.9	D3Byu2	31.0	<i>Xmmv-47</i>	43.1	<i>Pmv-38</i>
7.0	D3Mit1	33.1	D3J1	43.4	<i>soc</i>
7.5	<i>Car-3</i>	33.3	<i>Es-26</i>	43.5	D3Uf1
8.1	D3Byu3	33.3	<i>Suc-1r</i>	43.9	<i>Cdl</i>
8.2	D3Pas501	33.3	<i>Suc-1s</i>	43.9	<i>Fpsl-rs1</i>
8.3	<i>coa</i>	33.6	<i>Pk-1</i>	43.9	<i>Thbs3</i>
8.4	<i>Tmevd-2</i>	34.4	D3Byu5	43.9	<i>Tkr</i>
8.7	<i>Pgk-1ps3</i>	34.9	<i>Ghur-2</i>	44.9	D3Mit28
9.6	D3Mit46	35.0	D3Mit22	44.9	<i>H3f2</i>
9.6	<i>Mpmv-20</i>	35.0	D3Mit53	45.0	<i>Hsdb3b</i>
9.7	<i>Ap2</i>	36.2	D3Mit51	45.3	D3Byu4
10.1	<i>Es-16</i>	36.4	<i>Tpi-2</i>	45.5	<i>Gja-5</i>
10.2	D3Leh2	37.4	<i>spa</i>	45.5	<i>M6pr-ps</i>
10.2	<i>Evi-1</i>	38.6	D3Mit26	46.1	D3Mit41
10.2	<i>Fim-3</i>	38.6	D3Mit9	46.4	<i>Cd2</i>
10.2	<i>Glut-2</i>	38.8	<i>Cnp-2</i>	47.3	<i>Atplal</i>
12.2	<i>sut</i>	40.0	<i>Hao-2</i>	47.4	<i>Ampd-1</i>
13.2	D3Pas2	40.1	D3Mit40	47.4	<i>Cd53</i>
14.6	<i>Yb1d</i>	40.1	D3Mit49	47.4	D3Jfr1
15.0	<i>Ccna</i>	40.4	<i>ma</i>	47.4	<i>Ngfb</i>
15.0	<i>Il-2</i>	41.5	D3Ler1	47.4	<i>Nras</i>
15.7	<i>Fgf2</i>	41.5	D3Ler2	47.4	<i>Rap1a</i>
16.4	<i>Otf-3rs9</i>	42.3	D3Byu6	47.4	<i>Tshb</i>
17.6	D3Leh1	42.3	<i>Fgg</i>	48.1	D3Sell
19.3	<i>Hsp86-ps2</i>	42.3	<i>lap1a1-7</i>	48.2	<i>Kvl.2</i>
20.0	D3Mit24	42.3	<i>Xmmv-22</i>	48.2	<i>Kvl.3</i>
20.0	D3Mit3	42.3	<i>Xmmv-65</i>	48.4	<i>Csfm</i>
20.0	D3Mit4	42.4	<i>ft</i>	48.4	<i>de</i>
20.9	D3Pas1	42.6	<i>Gba</i>	48.4	<i>Gnai-2</i>
22.0	D3Mit6	42.6	<i>Pklr</i>	48.4	<i>Gnai-3</i>
23.3	<i>Es-27</i>	42.7	<i>Rnu1b-1</i>	48.4	<i>Kvl.3rs3</i>
24.0	D3Mit5	42.9	<i>Cacy</i>	48.6	D3Mit10
27.0	D3Nds1	42.9	<i>Call1</i>	48.6	D3Mit11
49.6	<i>Amy-1</i>	68.1	<i>Ahr-1</i>		
49.6	<i>Amy-2</i>	68.1	<i>Bmn</i>		
49.6	<i>Cf-3</i>	68.7	D3Mit17		
49.6	<i>Emv-27</i>	69.0	D3Jkn1		
50.2	<i>Ampd-2</i>	71.6	<i>Va</i>		
50.3	D3Mit12	71.8	<i>Pmv-26</i>		
50.6	<i>Nscl-2</i>	73.0	D3Mit18		
52.0	<i>Adh-1ps</i>	73.0	D3Mit31		
52.3	D3Uf2	74.6	<i>lap1a2-14</i>		
52.4	<i>Oat-rs2</i>	75.3	D3Mit45		
53.4	D3Mit36	76.4	D3Dol		
53.4	D3Mit39	77.0	D3Mit32		
53.8	<i>Pmv-39</i>	79.3	<i>H-28</i>		
54.6	<i>Pmp-1</i>	81.0	D3Mit19		
54.7	D3J3	84.6	<i>If-1</i>		
54.7	<i>Fabpi</i>	88.2	<i>Mpmv-9</i>		
56.1	D3Hun1	X	<i>Ida-3</i>		
56.1	D3Mit42	(X)	<i>H-37</i>		
57.7	D3Mit13	X	<i>Act5</i>		
57.9	D3Tu33	X	<i>Adh-5</i>		
58.4	<i>Ank-2</i>	X	<i>Arnt</i>		
59.6	<i>H-23</i>	X	<i>Bglap</i>		
59.6	<i>Hnl</i>	X	<i>Gsl-4</i>		
61.0	D3Mit14	X	<i>Hist2</i>		
61.0	D3Nds2	X	<i>lap1a2-9</i>		
61.6	<i>cdm</i>	X	<i>Lef-1</i>		
62.1	<i>Egf</i>	X	<i>Mmv-12</i>		
62.1	<i>Otf-3rs4</i>	X	<i>Mmv-2</i>		
63.0	D3Nds3	X	<i>Mov-10</i>		
63.0	<i>Rn7s-3</i>	X	<i>Odc-rs3</i>		
63.1	D3Mit15	X	<i>Oua-1</i>		
63.1	D3Mit16				
64.3	<i>Gbp-1</i>				
65.4	<i>rcm</i>				
65.6	D3J2				
67.2	D3Mit38				
68.1	<i>Adh-1</i>				
68.1	<i>Adh-1t</i>				
68.1	<i>Adh-3</i>				
68.1	<i>Adh-3t</i>				

says, and the availability of the MIT primers from Research Genetics (Birmingham, Ala.) make these markers extremely useful. Additional microsatellite markers have been developed by J. A. Todd and co-workers at the Nuffield Department of Surgery, Oxford, U.K. (Nds). Table 4 presents recombination data for 21 microsatellite markers spanning Chr 3, obtained from the European Interspecific Backcross [(C57BL/6J × Spretus) F<sub>1</sub> × Spretus].

### Recombinant inbred lines

Like multilocus backcrosses, RI lines provide a cumulative mapping resource. New loci can be mapped by typing the existing RI lines and comparing strain distribution patterns with the corresponding data for previously typed markers. Strain distribution patterns for Chr 3 loci that have been typed on RI lines are presented in Fig. 2.

### Anchor loci

In the last report we recommended use of six anchor loci to facilitate integration of new genetic data with the current map. It is too early to judge the effectiveness of this recommendation, that is, the degree to which investigators will type these loci in new crosses. To fill in the gaps in the previous anchor map, we recommend two additional microsatellite markers. Either D3Mit5 or D3Mit27 will divide the 28-cM interval between *Il-2* and *Gba* approximately in half. D3Mit19 provides a marker for the distal end of Chr 3; it is located approximately 20 cM distal to *Adh-1*. A subset of these well-mapped anchor markers can be selected to divide Chr 3 into convenient intervals: *centromere*–6–*Car-2*–9–*Il-2*–9–D3Mit5, D3Mit27–11–D3Mit22–8–*Gba*–5–*Tshb*–2–*Amy-1*–12–*Egf*–6–*Adh-1*–13–D3Mit19.

### Conserved linkage relationships

Mapping of genes has identified syntenic relationships between mouse Chr 3 and four human chromosomes: 1, 3, 4, and 8. The position of these mouse Chr 3 genes and their relationships to the chromosomal positions of the human homologs are shown in Table 5. These data suggest that rearrangements of chromosomal segments during mouse evolution have resulted in three separate homology groups with human Chr 4 and two separate homology groups with human Chr 3. In addition, the conserved linkage relationship with human Chr 1 spans the centromere of this chromosome. Long-range restriction site analysis has also indicated very strong conservation of some of these relation-

A		CROSS									CROSS						
LOCI		1	2	3	4	5	6	7	LOCI		1	2	3	4	5	6	7
Hc3									Gja-5								
D3Mr23									M6pr-ps								
Otf-3rs3									D3Mit41								
Il-7									Cd2								
Car-2									Atp1a1								
D3Mr1									Ampd-1								
D3Mr46									Cd53								
Ap2									Ngfb								
Evi-1									Nras								
Fim-3									Rap1a								
Glur-2									Tshb								
Yb1d									D3Sel1								
Ccna									Kv1.2								
Il-2									Kv1.3								
Fgf2									Csfm								
Otf-3rs9									Gnai-2								
D3Mr24									Gnai-3								
D3Mr3									D3Mr10								
D3Mr4									D3Mit11								
D3Mr6									Amy-2								
D3Mr5									Ampd-2								
D3Nds1									D3Mit12								
D3Mr25									D3U12								
D3Mr7									D3Mit36								
Mme									D3Mit39								
Glur-2									Pmp-1								
D3Mr22									D3Mr42								
D3Mr53									D3Mit13								
D3Mr51									D3Tu33								
Tpi-2									D3Mr14								
D3Mr26									D3Nds2								
D3Mr9									Egf								
D3Mr40									Otf-3rs4								
D3Mr49									D3Nds3								
Fgg									D3Mit15								
Gba									D3Mit16								
Thbs3									D3Mit38								
Pkir									Adh-1								
Fpsl-rs1									D3Mit17								
Cd1									D3Mit18								
Trk									D3Mit31								
Cacy									D3Mit45								
D3Tu51									D3Mit32								
Fcgr1									D3Mr19								
D3U1																	
D3Mr28																	

**Fig. 1. (A)** Gene order in seven multilocus crosses. Loci are listed from proximal to distal on the chromosome. The **filled boxes** represent the loci that were typed in each cross. The **shaded boxes** represent loci that did not recombine with the locus listed directly above. The observed recombination frequencies from each of these crosses are presented in Fig. 1B. See Table 1 and Fig. 1B for additional information about the crosses.

**B**

**CROSS 1: (C3H/HeJ-*gld* x *M. spretus*)F<sub>1</sub> x C3H/HeJ-*gld* (n = 117 to 338)**  
*Car-2* - 3.5 - *Evi-1* - 2.6 - *Il-2* - 0 - *Cyca* - 0.9 - *Fgf2* - 13.5 - *Mme* - 4.4 - *Glur-2* - 7.10 - *Gba* - 0.9 - *Thbs3* - 0 - *Pkir* - 0 - *Fspl-rs1* - 0 - *Cd1* - 0 - *Trk* - 0.6 - *D3Tu51* - 0 - *Cacy* - 0.9 - *Fcgr1* - 2.6 - *Cd2* - 0.9 - *Atp1a1* - 0.3 - *Nras* - 0 - *Cd53* - 0 - *Ngfb* - 0 - *Tshb* - 0 - *Ampd-1* - 1.8 - *Amy-2* - 0.6 - *Ampd-2* - 13.4 - *Egf* - 11.7 - *D3Mit18* - 9.3 - *D3Mit19*

**CROSS 2: (C57BL/6J x *M. spretus*)F<sub>1</sub> x C57BL/6J (n = 83 to 198)**  
*Otf-3rs3* - 2.0 - *Il-7* - 7.3 - *Evi-1* - 0 - *Fim-3* - 0 - *Glut2* - 7.6 - *Ybld* - 0 - *Ccna* - 0 - *Fgf2* - 21.8 - *Tpi-2* - 6.0 - *Fgg* - 6.8 - *Gja-5* - 0 - *M6pr-ps* - 5.2 - *Ngfb* - 0 - *Nras* - 2.2 - *Kv1.2* - 0 - *Kv1.3* - 0.6 - *Csfm* - 0 - *Kv1.3-rs3* - 0 - *Gnai-2* - 0 - *Gnai-3* - 1.5 - *Amy-2* - 4.9 - *Pmp-1* - 5.5 - *Egf* - 0 - *Otf-3rs4*

**CROSS 3: (C57BL/6J-*ob* x CAST/Ei)F<sub>2</sub> (n = 40 to 48)**  
*D3Mit23* - 6.8 - *D3Mit1* - 1.4 - *D3Mit46* - 2.4 - *Il-2* - 2.2 - *D3Mit4* - 0 - *D3Mit3* - 0 - *D3Mit24* - 1.1 - *D3Mit5* - 0 - *D3Mit6* - 4.7 - *D3Mit7* - 0 - *D3Mit25* - 3.4 - *D3Mit22* - 0 - *D3Mit53* - 1.1 - *D3Mit51* - 2.3 - *D3Mit26* - 0 - *D3Mit9* - 1.4 - *D3Mit40* - 0 - *D3Mit49* - 4.6 - *D3Mit28* - 1.1 - *D3Mit41* - 2.3 - *D3Mit10* - 0 - *D3Mit11* - 1.1 - *D3Mit12* - 1.8 - *D3Mit39* - 0.5 - *D3Mit36* - 1.8 - *D3Mit42* - 1.1 - *D3Mit13* - 2.2 - *D3Nds2* - 0 - *D2Mit14* - 1.1 - *D3Mit16* - 0 - *D3Mit15* - 3.5 - *D3Mit38* - 1.2 - *D3Mit17* - 4.3 - *D3Mit31* - 0 - *D3Mit18* - 3.1 - *D3Mit45* - 2.4 - *D3Mit32* - 7.3 - *D3Mit19*

**CROSS 4: C57BL/6J x SPR)F<sub>1</sub> x SPR: (n = 117 to 338)**  
*D3Mit1* - 7.6 - *Ap2* - 10.8 - *Il-2* - 7.6 - *D3Mit6* - 2.0 - *D3Mit5* - 3.0 - *D3Nds1* - 8.6 - *D3Mit22* - 8.6 - *Tshb* - 1.0 - *D3Mit11* - 1.9 - *Adh-1* - 10.8 - *D3Mit31* - 4.3 - *D3Mit32* - 3.0 - *D3Mit19*

**CROSS 5: (NOD/Uf x C57BL/6J)F<sub>1</sub> x NOD/Uf (n = 117 to 338)**  
*D3Mit1* - 19.0 - *Glut-2* - 4.0 - *Il-2* - 6.1 - *D3Nds1* - 18.8 - *D3U1* - 7.3 - *Tshb* - 4.3 - *D3Mit10* - 3.2 - *D3Uf2* - 14.1 - *Adh-1* - 20.5 - *D3Mit19*

**CROSS 6: (C57BL/6J x *M. spretus*)F<sub>1</sub> x *M. spretus* (n = 130 to 140)**  
*Hc3* - 0.7 - *Il-7* - 6.9 - *Evi-1*

**CROSS 7: (MRL/MpJ-*lpr* x CAST/Ei)F<sub>1</sub> x MRL/MpJ-*lpr*: (n = 117 to 338)**  
*Otf-3rs3* - 6.3 - *Car-2* - 10.6 - *Otf-3rs9* - 14.4 - *Mme* - 24.3 - *Otf-3rs4* - 0 - *Egf* - 26.8 - *D3Mit18*

**Fig. 1. (B)** Recombination frequencies in multilocus crosses. Crosses 1–7 are described in Table 1 and Fig. 1A. The approximate lengths, in cM, of the intervals observed in each cross are presented here. We included only crosses with at least six markers analyzed in at least 80 meiotic events, with the exception of cross 6, which provides data for anchoring the composite maps with respect to the centromere. (Note: Cross 4, n = 92)







Table 3. Continued.

Locus	Sequence	Primer forward (5'-3')	Primer reverse (5'-3')	Product size	Conditions	Size variation
D3Mit60	F107			168-198	1mM/55	CAST>>LP=AKR=BALB=DBA>C3H=A>NOD=NON=B6=OB; SPE:-
D3Mit51	J8	GGCACTGATAGCAGGCCCTAG	TCTCTTCGTGTAATTCCTTCGG	230-258	1mM/55	AKR=BALB=DBA>NOD=NON>>LP>>C3H=B6=A=OB>>CAST; SPE:-
D3Mit52	A1081	AGCCAGGATATGGAAATATGCC	TGACCAGATTGCATGCATTT	196-204	1mM/55	A=SPE>OB>NOD=NON=AKR=DBA=C3H>LP>BALB=B6; CAST:-
D3Mit54	B572	TTGGTCCACAGCAACTAGG	CAGGGAATGTATGTCAATGAGG	122-148	1mM/55	NOD=BALB=C3H=B6=A=OB>CAST>NON>>LP=AKR=DBA>>SPE
D3Mit55	B536	CTGGGACCCACAGTAGTACCA	TCAGGACTGCCAAGTGGGC	116-144	1mM/55	C3H=A>>LP=NOD=B6=AKR=BALB=DBA=CAST>>NON>>SPE; OB:-
D3Mit56	B713	TCTAGCTATGTGATGAGTGTGTGG	CAGGATTTCCAAAAACATCCA	138-148	1mM/55	A>LP=NOD=NON=AKR=DBA=C3H=B6=OB>BALB>>CAST; SPE:-
D3Mit57	B493	TCCAGTTACTTTGGTGAAGTCCA	ATATGTTACATGTTTCATGGTGTG	148-176	1mM/55	CAST>>SPE>>LP=B6=NON=AKR=BALB=C3H=A=OB>>NOD; DBA:-
D3Mit58	B527	ACATCAGAGAGTCAATTCATCA	GCTTTCAGTCACAGCTCTGC	140-152	1mM/55	CAST>>NOD=NON=BALB=DBA=C3H=B6=A=OB>>LP=AKR; SPE:-
D3Mit59	B543	GTTCGATGCCCAAGGAATGAT	CTACTGCATCCTGGCACAGA	204-212	1mM/55	OB>CAST>>LP=AKR=B6=NON=BALB=DBA=C3H=A=SPE>>NOD

Recommended magnesium concentrations and annealing temperatures are indicated for most loci. \*D3Nds23: *Hsp86-ps2*, unpublished sequence was kindly provided by S. Moore, Food and Drug Administration, Division of Metabolism and Endocrine Drug Products, Rockville, Md.; the STS and polymorphism are from N. Rodrigues. \*\*D3Nds24: *CD10*, clone obtained from M. Shipp, Dana-Farber Cancer Institute, Boston, Mass.; the STS and polymorphism are from N. Rodrigues. \*\*\*D3Nds25: *Gba*, (ref. 119). \*\*\*\*D3Nds26: *Gpi-1rs*, unpublished STS from J. Jones, MRC Radiobiology Unit, UK.

Table 5. Relationships between mouse Chr 3 genes and homologous human genes.

Map	Locus	Name	Human	Human position
6.3	<i>Car-1</i>	carbonic anhydrase-1	CA1	8q13-q22
6.3	<i>Car-2</i>	carbonic anhydrase-2	CA2	8q13-q22
7.5	<i>Car-3</i>	carbonic anhydrase-3	CA3	8q13-q22
10.2	<i>Evi-1</i>	ecotropic viral integration site-1	Evi1	3q24-q28
10.2	<i>Fim-3</i>	Friend MuLV integration site-3	FIM3	3q27
10.2	<i>Glur-2</i>	glucose transporter 2	GLUT2	3q26
15.0	<i>Il-2</i>	interleukin 2	IL2	4q26-q27
15.7	<i>Fgf2</i>	fibroblast growth factor basic	FGF2	4q25-27
30.1	<i>Mme</i>	membrane metallo-endo peptidase	MME	3q21-27
33.3	<i>Suc-1s</i>	sucrase-isomaltase, structural	SI	3q25-26
34.9	<i>Glur-2</i>	glutamate receptor 2	GLUR2	4q25-34
42.3	<i>Fgg</i>	gamma fibrinogen	FGG	4q28
42.6	<i>Pklr</i>	pyruvate kinase liver, red blood cells	PKLR	1q21
42.6	<i>Gba</i>	beta glucocerebrosidase	GBA	1q21
42.9	<i>Cacy</i>	calyculin	CACY	1q21-q25
42.9	<i>Capl</i>	calcium binding protein, placental	CAPL	1q12-q22
42.9	<i>Fcgr1</i>	high affinity FC gamma receptor	FCGR1	1q
43.9	<i>Cdl</i>	cluster designation 1	CD1	1q22-23
43.9	<i>Fpsl-rs1</i>	farnesyl pyrophosphate synthetase-like 1	FPSL	1q24-q31
44.9	<i>H3f2</i>	histone 3, family 2	H3F2	1q12-21
45.0	<i>Hsd3b</i>	3-beta-hydroxy steroid dehydrogenase	HSD3B	1p11-13
46.4	<i>Cd2</i>	cluster designation 2	CD2	1p13
47.3	<i>Atplal</i>	Na, K ATPase alpha-1	ATPIA1	1p13
47.4	<i>Ampd-1</i>	AMP deaminase-1 (muscle form)	AMPD1	1p13
47.4	<i>Ngfb</i>	nerve growth factor beta	NGFB	1p13
47.4	<i>Nras</i>	Nras oncogene	NRAS	1p13
47.4	<i>Rap1a</i>	member of RAS oncogene family	RAP1A	1p12-p13
47.4	<i>Tshb</i>	thyrotropin stimulating hormone beta subunit	TSHB	1p13
48.4	<i>Csfm</i>	colony stimulating factor, macrophage	CSF1	1p13-21
48.4	<i>Gnai-3</i>	guanine nucleotide binding protein, alpha inhibiting activity-3	GNAI3	1p13
49.6	<i>Amy-1</i>	amylase, salivary	AMY1	1p21
49.6	<i>Amy-2</i>	amylase, pancreatic	AMY2	1p21
49.6	<i>Cf-3</i>	coagulation factor 3	F3	1p22-p21
54.6	<i>Pmp-1</i>	peroxisomal membrane protein (70k)	PMP1	1p21-22
54.7	<i>Fabpi</i>	fatty acid binding protein intestinal	FABP2	4q28-31
62.1	<i>Egf</i>	epidermal growth factor	EGF	4q25
68.1	<i>Adh-1</i>	alcohol dehydrogenase-1	ADH1	4q21-q23
68.1	<i>Adh-3</i>	alcohol dehydrogenase-3	ADH3	4q21-q23
X	<i>Act5</i>	skeletal alpha actin	ACTA1	1p21-qter
X	<i>Adh-5</i>	alcohol dehydrogenase-5	ADH5	4q21-q25
X	<i>Arnt</i>	aryl hydrocarbon receptor nuclear translocator	ARNT	1pter-q12
X	<i>Lef-1</i>	lymphoid enhancer-binding factor 1	LEF1	4q23-q25

Table 4. Microsatellite map of Chr 3.

Loci	RF	Distance
<i>D3Mit1-D3Nds15</i>	7/92	7.65 cM
<i>D3Nds15-D3Nds27</i>	10/92	11 cM
<i>D3Nds27-D3Mit6</i>	7/92	7.65 cM
<i>D3Mit6-D3Mit5</i>	2/92	2.1 cM
<i>D3Mit5-D3Mit7</i>	3/92	3.2 cM
<i>D3Mit7-D3Nds1</i>	0/92	0
<i>D3Nds1-D3Nds24</i>	3/92	3.2 cM
<i>D3Nds24-D3Mit22</i>	5/92	5.4 cM
<i>D3Mit22-D3Mit51</i>	3/92	3.2 cM
<i>D3Mit51-D3Mit9</i>	1/85	1.1 cM
<i>D3Mit9-D3Nds25</i>	0/85	0
<i>D3Nds25-D3Nds22</i>	0/92	0
<i>D3Nds22-D3Nds8</i>	4/92	4.3 cM
<i>D3Nds8-D3Mit11</i>	1/91	1 cM
<i>D3Mit11-D3Mit42</i>	7/91	7.7 cM
<i>D3Mit42-D3Nds9</i>	12/92	13.3 cM
<i>D3Nds9-D3Mit38</i>	3/92	3.2 cM
<i>D3Mit38-D3Mit31</i>	7/90	7.8 cM
<i>D3Mit31-D3Mit32</i>	4/90	4.4 cM
<i>D3Mit32-D3Mit19</i>	3/92	3.2 cM

Data from the laboratory of J.A. Todd from the European Collaborative Interspecific Backcross [(C57BL/6 × SPR)F<sub>1</sub> × SPR] produced by the UK's Human Genome Mapping Project, with the support of the Medical Research Council. Distances were calculated with the Kosambi mapping function. RF, recombinant fraction (recombinants/total analyzed).

