Leukotrienes Increase Blood-Brain Barrier Permeability Following Intraparenchymal Injections in Rats

Keith L. Black, MD, and Julian T. Hoff, MD

To examine whether leukotrienes could increase blood–brain barrier permeability, rats were anesthetized and injected intravenously with Evans blue. Ten microliters of, vehicle, of leukotrienes B₄, C₄, or E₄, or of arachidonic acid was injected over 1 hour directly into the brain parenchyma. The percentage of the total surface area of Evans blue extravasation in a coronal section of brain centered on the injection site was then determined as an estimate of blood–brain barrier permeability. Leukotrienes B₄, C₄, and E₄, and arachidonic acid all increased blood–brain barrier permeability, but this effect was lost when the total dose was reduced to 20 ng. Increased blood–brain barrier permeability induced by arachidonic acid could be prevented by pretreatment with the lipoxygenase inhibitor BW755C, but not with indomethacin. Leukotrienes may play a role in the development of increased blood–brain barrier permeability after cerebral injury.


Arachidonic acid is metabolized by means of two pathways: cyclooxygenase, leading to the formation of prostaglandins, and lipoxygenase, leading to the production of leukotrienes. Prostaglandins have been implicated in the pathogenesis of cerebral ischemia. This is in part supported by studies showing that cerebral blood flow in ischemic brain is increased after treatment with cyclooxygenase inhibitors and prostacyclin [3]. Leukotrienes have been shown to increase vascular permeability in postcapillary venules, increase blood flow, and elicit erythema and wheal formation in skin [10]. Leukotriene concentrations are reportedly increased in cerebral ischemia [7], and we recently noted that the injection of leukotriene C₄ directly into brain parenchyma increased blood–brain barrier (BBB) permeability [2]. Consequently, leukotrienes may play a role in the pathogenesis of vasogenic cerebral edema [2, 3, 7].

This study was designed to determine (1) whether intraparenchymal injections of leukotriene (LT) B₄, LTC₄, or LTE₄, or arachidonic acid increase BBB permeability; and (2) whether the lipoxygenase inhibitor BW755C [3-amino-1-(3-trifluoromethylphenyl)-2-pyrazoline] [8, 9] prevents the increased BBB permeability induced by arachidonic acid.

Materials and Methods

Wistar rats weighing 250 to 350 gm were anesthetized with pentobarbital (40 mg per kilogram of body weight) injected intraperitoneally. PE-10 polyethylene catheters were inserted into the femoral artery and vein. The arterial catheter was connected to a blood pressure transducer and systemic arterial blood pressure was monitored continuously. Buffered Evans blue solution (0.5 ml) was then injected into the femoral vein. A burrhole was made over the left frontal cortex. A 10-μl syringe (J and W Scientific Inc) with a deactivated fused silica needle (125 μm in diameter) was mounted on the arm of a stereotaxic frame. With the aid of magnification, the tip of the needle was manipulated 3.5 mm deep into the brain parenchyma and then withdrawn 1 mm. The final depth into the parenchyma was therefore 2.5 mm, corresponding to the gray–white matter interface.

Ten microliters of either saline solution, water, 6.5% methanol in saline, LTB₄, LTC₄, LTE₄, or arachidonic acid was injected into the parenchyma slowly over 1 hour. Five animals injected with arachidonic acid were given BW755C (40 mg/kg) intraperitoneally 1 hour prior to arachidonic acid injection, and 5 animals were given indomethacin (6 mg/kg) intraperitoneally 1 hour prior to arachidonic acid injection. Rats were decapitated 1 hour after the intraparenchymal injection and their brains were removed and frozen. The frozen brains were sliced coronally through the plane of injection. The surface of the slice was photographed and, using polar planimetry, the area of Evans blue extravasation was estimated as the percentage of total surface area.

Leukotrienes were supplied by Dr R. Rokach of Merck-Frosst Inc, Canada. LTC₄ and LTE₄ were received in water at a concentration of 0.2 mg/ml. LTB₄ was received in 65% methanol in water at a concentration of 0.1 mg/ml. Leukotrienes were subsequently diluted in saline. They were di-
vided into small aliquots and stored in glass vials that were vacuum sealed and frozen at -70°C. The total dose of LTB₄ injected did not exceed 100 ng per brain since the methanol content would then have been greater than 6.5%. In preliminary studies we found that saline solutions with a methanol concentration greater than 6.5% produced some disruption in the BBB. Indomethacin and free arachidonic acid were obtained from Sigma. Free arachidonic acid was dissolved in 0.3 M sodium hydroxide and diluted in saline to a concentration of 1.0 mg/ml. BW755C was a gift of Wellcome Research Laboratories, Kent, England. BW755C was dissolved in saline to a final concentration of 20 mg/ml.

An analysis of variance was used for statistical analysis.

Results
Except for negligible blue staining along the insertion tract of the needle, none of the rats injected with saline or water had evidence of Evans blue extravasation. The percentage of blue area in rats injected with 6.5% methanol was also negligible (0.68 ± 0.21% of the area of the left hemisphere in the plane of injection). Animals injected with LTB₄, LTC₄, or LTE₄ had significant extravasation of Evans blue in the area of injection (Fig 1).

Arachidonic acid injection also resulted in extravasation of Evans blue. This effect was, however, prevented by lipoxygenase inhibition with BW755C, but not by indomethacin (Figs 2, 3).

Blood pressure remained within physiological limits in all groups.

Discussion
Moskowitz and colleagues [7] reported increased levels of leukotrienes after cerebral ischemia and reperfusion in the gerbil brain and suggested that cortical
gray matter was the most likely source of leukotriene synthesis. Since leukotrienes are potent vasoconstrictors [11] and they increase vascular permeability [2, 10], some investigators suspect that leukotrienes are involved in the pathogenesis of brain damage following cerebral ischemia [2, 7, 11].

Our study demonstrates that LTB₄, LTC₄, or LTE₄ (>20 ng) injected directly into brain parenchyma significantly increases BBB permeability. This effect is lost when the total local dose is less than 20 ng. The amount of leukotriene required to produce vasogenic edema by injection into the brain is, therefore, several orders of magnitude higher than that present in the cortex after ischemia-reperfusion experiments. The discrepancy may be attributed to: (1) leukotrienes have a short half-life; the baseline level of leukotriene during the 1-hour injection may be lower than the actual dose injected; (2) a longer time interval of exposure may be required for smaller doses of leukotrienes to increase BBB permeability; and (3) measurement of leukotrienes in lesions that primarily cause the vasogenic form of edema may reveal leukotriene levels higher than those in ischemic edema. Studies to determine whether a lipoxygenase inhibitor will decrease vasogenic edema in these conditions are warranted.

Other investigators have shown that arachidonic acid produces cerebral edema. The induction of brain edema by arachidonic acid is dose dependent and the resulting edema becomes maximal between 24 and 48 hours. The edema is reduced by pretreatment with dexamethasone but not by indomethacin [5]. Interestingly, dexamethasone is reported to decrease leukotriene synthesis [4].

In the present study the lipoxygenase inhibitor BW755C prevented the vasogenic edema normally caused by arachidonic acid. BW755C has also been shown to inhibit cyclooxygenase [8]. The protective effect of BW755C is unlikely to be related to its effect on cyclooxygenase, since indomethacin, at a dose reported to inhibit cyclooxygenase [1], did not provide a similar protective effect. Our findings suggest that the production of vasogenic edema by arachidonic acid is dependent on products of the 5-lipoxygenase enzyme, i.e., leukotrienes, rather than the products of cyclooxygenase.

Caronna and co-workers [5] suggested that the ability of arachidonic acid to produce brain edema is not specific. They also reported increased edema after injections of other polyunsaturated fatty acids, but not saturated fatty acids. Horna and colleagues [6], however, recently reported that polyunsaturated fatty acids, but not saturated fatty acids, stimulate the formation of lipoxygenase products from arachidonic acid. Theoretically, therefore, other polyunsaturated fatty acids might increase edema by increasing the conversion of arachidonic acid to leukotrienes.

In our study, LTB₄ produced a greater increase in BBB permeability than the other leukotrienes. Whether this relates to the ability of LTB₄ to act as a chemotactic factor attracting leukocytes, which in turn may produce more leukotrienes, is speculative.

We believe that the development of vasogenic cerebral edema is multifactorial and depends on the stimuli inducing the edema process. This study demonstrates that leukotrienes do induce vasogenic edema and that the ability of arachidonic acid to induce edema can be prevented by pretreatment with a lipoxygenase inhibitor.

References