

## EFFECT OF METHIONINE AND METHIONINE SULPHOXIMINE ON RAT BRAIN S-ADENOSYL METHIONINE LEVELS

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**Abstract**—Rat brain SAM levels were markedly increased after methionine administration, whereas the convulsant, L-methionine-dl-sulphoximine (MSO), produced a 35 per cent decrease in whole brain content of S-adenosyl-L-methionine (SAM). When methionine was given in combination with MSO, SAM levels were not decreased. Studies on the regional distribution of SAM revealed only a small variation between regions (from 24 nmol/g in midbrain to 49.5 nmol/g in striatum). SAM levels were reduced by about 50 per cent in the cerebellum, striatum, cortex and hippocampus 3 and 6 h after MSO. It is proposed that aberrant cerebral methylation processes may be involved in the genesis of the MSO seizure.

THE USE of the convulsant, methionine sulphoximine (MSO), to study seizure mechanisms is advantageous in that its rather long preconvulsive latency period (3–6 h) (REINER *et al.*, 1950) allows one to follow the course of specific cellular and metabolic alterations resulting from its administration. Previous studies (LODIN & KOLOUSEK, 1958; LAMAR & SELLINGER, 1965; SELLINGER *et al.*, 1970) have demonstrated that L-methionine administered concurrently with MSO in a 5:1 molar ratio markedly reduces the incidence and severity of seizures.

The importance of S-adenosyl methionine (SAM) as a methyl donor (LOMBARDINI & TALALAY, 1971), the structural similarities between MSO, methionine and SAM and the fact that both BALDESSARINI (1966) and SALVATORE *et al.* (1971) reported that methionine administration increased SAM levels in rat and rabbit brain, respectively, led us to investigate the effect of MSO and methionine on SAM levels in rat whole brain and brain regions.

### MATERIALS AND METHODS

#### Materials

L-Methionine-dl-sulphoximine (MSO) was obtained from Sigma Chemical Co. (St. Louis, Missouri); L-methionine, from ICN Nutritional Biochemicals (Cleveland, Ohio) and S-adenosyl-L-methionine, from P-L Biochemicals Inc. (Milwaukee, Wisconsin). Radioactive S-adenosyl-L-methionine [methyl- $^{14}\text{C}$ ] (SAM) (specific activity 52 mCi/mmol) and

[ $^{14}\text{C}$ ]toluene ( $4.26 \times 10^5$  d.p.m./ml) were purchased from New England Nuclear (Boston, MA). PCS (a tissue solubilizer-scintillant mixture) was obtained from Amersham Searle (Arlington Heights, IL). Silica gel TLC plates F-254 (with fluorescent indicator) were from EM Laboratories (Elmsford, NY). Dowex 50W-X8 resin was from Baker Chemical Co. (Phillipsburg, NJ) and was converted to the  $\text{Na}^+$  form prior to use (Shapiro & Ehninger, 1966).

#### Injection and sacrifice

Male Sprague-Dawley rats (100–200 g) were injected intraperitoneally with saline (10 ml/kg), methionine (4.7 mmol/kg), MSO (0.94 mmol/kg), or both MSO and methionine 1, 3 and 6 h prior to sacrifice. Sacrifice was always between the hours of 11 a.m. and 2 p.m. After sacrifice by decapitation, whole brains were rapidly removed, frozen in liquid nitrogen and stored at  $-65^\circ\text{C}$ . For regional analyses, brains were rapidly dissected on ice (GLOWINSKI & IVERSEN, 1966), regions pooled in groups of three and immediately frozen and stored as above. Samples remained stored no longer than 21 h before being analysed for SAM content.

#### Analytical

S-adenosyl methionine was measured by the radioisotope dilution technique of SALVATORE *et al.* (1971). Brains or regions were homogenized in 1.5 M-perchloric acid containing from 0.08 to 0.2  $\mu\text{mol}$  SAM adjusted to a specific activity of  $0.5 \times 10^5$ – $2 \times 10^5$  c.p.m./ $\mu\text{mol}$  by the addition of non-radioactive SAM purified according to SHAPIRO & EHNINGER (1966). After centrifugation at 9000 g for 15 min, the supernatants were adjusted to pH 6 by very slow dropwise addition of 1.0 N KOH in the cold with constant stirring. After pH adjustment, samples were centrifuged for 15 min at 9000 g and the supernatants added to Dowex 50 ( $\text{Na}^+$ ) columns ( $3 \times 0.4$  cm). The columns were thistle top

*Abbreviations used:* MSO, L-methionine-dl-sulphoximine; SAM, S-adenosyl-L-methionine.

funnel tubes (30 × 0.4 cm) to which were attached 2 cm lengths of tygon tubing and open jaw screw clamps. Eluting agents were 0.1 M NaCl followed by 6 N HCl and elution was continued until the absorbance of the acid eluate was less than 0.01 at 256 nm. One ml of acid eluate was combined with 10 ml PCS and counted in a Nuclear Chicago Unilux II liquid scintillation counter. Absorbances of the acid eluates were measured in a Beckman Acta II spectrophotometer at 256 nm and SAM concentration calculated using the molar extinction coefficient of 14,700 (SHAPIRO & EHNINGER, 1966). Two standards were carried through the experimental procedure each day. Acid eluates were intermittently checked for purity of SAM on silica gel F-254 TLC plates by isographic migration with authentic SAM. Solvent systems used were *n*-butanol-acetic acid-water (60:15:25), ethanol-acetic acid-water (64:1-35) (SALVATORE *et al.*, 1971) and 20% KCl-5% acetone-water (COHN *et al.*, 1972). Ninhydrin spray (0.25% w/v, in acetone) for amino acids, the u.v. quenching reaction for purine compounds (SALVATORE *et al.*, 1971), the chloroplatinate spray for sulfur compounds (WONG, 1971) and autoradiography were used for identification of SAM.

Reproducibility of the SAM assay was markedly decreased when tissue weights of less than 150 mg were assayed. Indeed, there was a significant correlation between tissue weight and standard error ( $r = 0.64$ ,  $P < 0.05$ ).

## RESULTS

Table 1 represents the regional distribution of SAM in rat brain. The range of variation in SAM levels between brain regions is rather small with the highest level in the striatum (49.5 nmol/g) and the lowest in the midbrain (24.0 nmol/g).

Administration of MSO resulted in a gradual decrease of whole brain SAM levels reaching a maximum at 6 h (-47%) (Fig. 1). Methionine, on the other hand, produced a marked increase in SAM that reached a

TABLE 1. REGIONAL DISTRIBUTION OF S-ADENOSYL-METHIONINE IN RAT BRAIN

Brain region	S-Adenosyl-methionine
Cerebellum	44.6 ± 4.16 (16)
Brainstem	29.6 ± 2.08 (17)
Striatum	49.5 ± 4.77 (17)
Hypothalamus	26.0 ± 4.41 (11)
Midbrain	24.0 ± 2.20 (17)
Hippocampus	32.9 ± 3.60 (17)
Cerebral cortex	30.5 ± 1.98 (17)
Whole brain	31.1 ± 2.15 (15)

Values are expressed as nmol/g brain and are means ± S.E.M.

Numbers in parentheses represent numbers of determinations.

Brain regions from three rats were pooled for each determination.

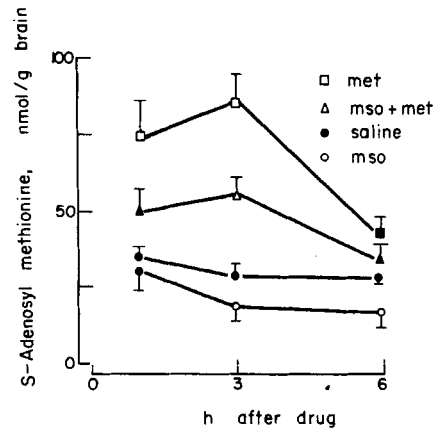


FIG. 1. Effect of methionine or methionine sulphoximine on rat brain S-adenosyl-methionine levels. L-Methionine (4.7 mmol/kg) or L-methionine sulphoximine (0.94 mmol/kg) were administered intraperitoneally. Each value represents the mean ± S.E.M. of 5-6 rats. Open symbols denote a significant difference from corresponding saline values at the 0.05 level using the Student *t*-test.

value about 200 per cent that of saline controls at 3 h, but was not significantly increased (+29 per cent) at 6 h. Animals receiving a combination of MSO and methionine showed no significant differences in SAM levels at 1 and 6 h but at 3 h SAM levels were significantly elevated (Fig. 1).

One hour after MSO, SAM was decreased by about

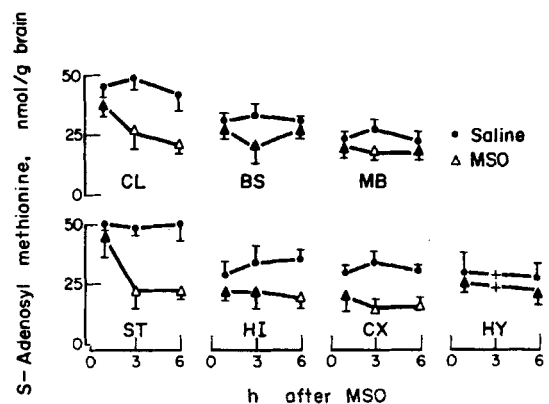


FIG. 2. Effect of methionine sulphoximine on S-adenosyl methionine levels in rat brain regions. CL—cerebellum, BS—brainstem, MB—midbrain, ST—striatum, HI—hippocampus, CX—cortex, HY—hypothalamus. Each value represents the mean of 5-6 determinations ± S.E.M. Regions from the brains of three rats were pooled for each determination. Open symbols denote a significant difference from corresponding saline value at the 0.05 level using the Student *t*-test. (+) Values not available.

25 per cent in the hippocampus and cortex and by 14 per cent in the cerebellum, whereas other regions showed no significant changes in SAM content (Fig. 2). Three and six hours after MSO, SAM was about 50 per cent of control values in the cerebellum, striatum, hippocampus and cortex. At these time periods SAM levels in the brainstem, midbrain and hypothalamus were slightly decreased, the only significant reduction being in the midbrain at 3 h (Fig. 2).

#### DISCUSSION

There were no striking differences in the regional distribution of SAM in rat brain (Table 1). These data are generally in agreement with previous findings (BALDESSARINI & KOPIN, 1966), the only exception being the high cerebellar SAM values in the present study. As yet, this result remains unexplained but TLC chromatography of the cerebellar eluates revealed no detectable spots other than SAM (see Methods).

Methionine has been shown to elevate rat (BALDESSARINI, 1966) and rabbit brain SAM levels (SALVATORE *et al.*, 1971). Such a result has been demonstrated in this study (Fig. 1). Further, the decrease in SAM levels after MSO is prevented by the concomitant administration of methionine. The reason for this is thought to be that methionine competitively decreases the amount of MSO entering the brain (GHITTONI *et al.*, 1970), a similar competitive transport relationship recently having been noted in unicellular algae (MEINS & ABRAMS, 1972). Moreover, the ability of methionine to antagonize MSO seizures is markedly decreased if methionine is given 60 min or more after MSO (SELLINGER *et al.*, 1968). Thus the concomitant administration of methionine and MSO probably does not allow sufficient MSO to enter the CNS either to produce seizures or to reduce SAM levels.

The marked increase in SAM levels after methionine (Fig. 1) is probably the result of increased precursor availability, especially since the conversion of methionine to SAM in rat brain occurs very rapidly *in vivo* (BALDESSARINI & KOPIN, 1966).

Since SAM levels were depressed prior to MSO seizures, the possibility of a causal relationship between levels of SAM and the MSO seizure may be entertained. At present, however, this appears untenable since pargyline (BALDESSARINI, 1966) and DOPA (WURTMAN *et al.*, 1970) both decrease SAM but neither is known to produce convulsions. It is possible, however, that localized pools of SAM exist in cellular compartments (JUDES & JACOB, 1972), and that DOPA and pargyline alter levels of SAM in a pool other than that affected by MSO.

The mechanism(s) whereby MSO lowers brain SAM levels may involve a decreased synthesis of SAM by the methionine activating enzyme (ATP: L-methionine adenosyl transferase; EC 2.4.2.13), an increased utilization of SAM by various cerebral methyl transferases or a combination of both of these processes. LOMBARDINI *et al.* (1970) have reported that MSO is neither a substrate nor an inhibitor of the rat liver methionine activating enzyme *in vitro*, and we have reported no inhibitory effects of MSO on this enzyme in rat cerebral cortex and cerebellum (SCHATZ *et al.*, 1973).

DE ROBERTIS *et al.* (1967) have provided preliminary evidence for the alternate mechanism, that is, one operating to increase utilization of SAM by cerebral methyl transferases, for they noted a marked increase in rat brain catechol-*O*-methyltransferase activity 6 h after MSO administration.

The striking effect of MSO on cerebral SAM levels most probably reflects marked alterations of methylation processes in brain. Perhaps aberrant methylation of brain amines, or of certain key macromolecules, is at least in part responsible for the ability of MSO to elicit seizures.

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#### REFERENCES

- BALDESSARINI R. J. (1966) *Biochem. Pharmac.* **15**, 741–748.  
 BALDESSARINI R. J. & KOPIN I. J. (1966) *J. Neurochem.* **13**, 769–777.  
 COHN C. K., VESELL E. S. & AXELROD J. (1972) *Biochem. Pharmac.* **21**, 803–809.  
 DE ROBERTIS E., SELLINGER O. Z., RODRIGUEZ DE LORES ARNAIZ G., ALBERICI M. & ZIEHER L. M. (1967) *J. Neurochem.* **14**, 81–89.  
 GHITTONI N. E., OHLSSON W. G. & SELLINGER O. Z. (1970) *J. Neurochem.* **17**, 1057–1068.  
 GLOWINSKI J. & IVERSEN L. L. (1966) *J. Neurochem.* **13**, 655–669.  
 LAMAR C. Jr. & SELLINGER O. Z. (1965) *Biochem. Pharmac.* **14**, 489–506.  
 LODIN Z. & KOLOUSEK J. (1958) *Physiol. Bohemoslov.* **7**, 292–298.  
 LOMBARDINI J. B., COULTER A. W. & TALALAY P. (1970) *Molec. Pharmac.* **6**, 481–499.  
 LOMBARDINI J. B. & TALALAY P. (1971) *Advances in Enzyme Regulation* **9**, 349–384.  
 MEINS F. Jr. & ABRAMS M. L. (1972) *Biochim. biophys. Acta* **266**, 307–311.  
 REINER L., MISANI F. & WEISS P. (1950) *Archs. Biochem. Biophys.* **25**, 447–454.  
 SALVATORE F., UTILI R., ZAPPALÀ V. & SHAPIRO S. K. (1971) *Analyt. Biochem.* **41**, 16–28.

- SCHATZ R., DIEZ-ALTARES M. C. & SELLINGER O. Z. (1973) *Trans. Am. Soc. Neurochem.* **4**, 74.
- SELLINGER O. Z., AZCURRA J. M. & OHLSSON W. G. (1968) *J. Pharmac. exp. Ther.* **164**, 212-222.
- SHAPIRO S. K. & EHNINGER D. J. (1966) *Analyt. Biochem.* **15**, 323-333.
- WONG F. F. (1971) *J. Chromatogr.* **59**, 448-457.
- WURTMAN R. J., ROSE C. M., MATTHYSSE S., STEPHENSON J. & BALDESSARINI R. J. (1970) *Science, N.Y.* **169**, 395-397.