

## OUTCOMES OF STATUS EPILEPTICUS

### Is neurogenesis reparative after status epilepticus?

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Neurogenesis persists in the hippocampal dentate gyrus of mammals, including human and nonhuman primates, throughout life (Altman and Das, 1965; Cameron et al., 1993; Kuhn et al., 1996; Eriksson et al., 1998; Gould et al., 1999; Kornack and Rakic, 2001). Neural stem-like cells reside in the subgranular zone (SGZ) at the border of the hilus and DGC layer (Seri et al., 2001; Filippov et al., 2003), where they generate neuroblasts that migrate into the layer and differentiate into granule neurons (Cameron et al., 1993; Kuhn et al., 1996). Adult-born dentate granule cells (DGCs) send axonal projections to appropriate targets (Stanfield and Trice, 1988; Markakis and Gage, 1999; van Praag et al., 2002) and acquire electrophysiological characteristics of mature DGCs (van Praag et al., 2002; Wadiche et al., 2005; Ge et al., 2006). Although the precise function is unknown, evidence implicates DGC neurogenesis in certain forms of hippocampus-dependent learning and memory (reviewed in Doetsch and Hen [2005]).

#### DGC NEUROGENESIS IS ALTERED AFTER SE OR KINDLING

Studies in adult rodent models of mesial temporal lobe epilepsy (mTLE) indicate that status epilepticus (SE) potentially stimulates DGC neurogenesis (Parent et al., 1997; Gray and Sundstrom, 1998; Parent et al., 1998). Kainate- or pilocarpine-induced SE increases dentate gyrus cell proliferation approximately 5- to 10-fold within 3 days (Parent et al., 1997; Gray and Sundstrom, 1998), and over 80% of the cells become DGCs. Electrical kindling of amygdala (Parent et al., 1998; Scott et al., 1998), hippocampus (Benzon et al., 1997), or perforant path (Nakagawa et al., 2000) also stimulates DGC neurogenesis. SE-induced neurogenesis may be short-lived, however, as chronically epileptic rats show decreased DGC production 5 months after SE (Hattiangady et al., 2004).

In addition to increased production, adult rodent DGCs generated after SE show abnormalities of structure and

location. Basal dendrites persist on adult-born DGCs much more frequently after SE (Scharfman et al., 2000; Dashtipour et al., 2001), and cells with basal dendrites appear both in the inner DGC layer and hilus after SE. Interestingly, new DGCs with abnormal basal dendrites continue to form many months after SE (Jessberger et al., 2007a), suggesting that the responsible cues persist well after the initial insult. Another abnormality involves dispersion of the DGC layer and abnormal locations of DGCs outside the layer. The dentate gyrus in human mTLE often shows excessive DGC dispersion and ectopic granule-like neurons in the hilus and molecular layer (Houser, 1990; Parent et al., 2006). DGC layer dispersion and hilar-ectopic DGCs appear in rodent mTLE models and persist chronically (Parent et al., 1997; Scharfman et al., 2000; Dashtipour et al., 2001; Jessberger et al., 2005; Parent et al., 2006). Ectopic cells in human and experimental mTLE are similar in terms of their morphology and expression of DGC-specific markers (Scharfman et al., 2000; Dashtipour et al., 2001; Parent et al., 2006).

#### MECHANISMS UNDERLYING SE-INDUCED NEUROGENESIS

The mechanisms by which SE stimulates neurogenesis are unknown. Huttman et al. (2003) used a neural stem cell reporter mouse to show that kainate-induced SE influences proliferation of the SGZ radial glia-like progenitors. SE may increase neurogenesis indirectly through the stimulation of astrocytosis, as astrocytes stimulate hippocampal neurogenesis via wnt signaling and perhaps other mechanisms (Song et al., 2002; Lie et al., 2005). SE also increases the expression of growth factor and other molecules with the potential to influence neurogenesis.

The formation of ectopic DGCs after SE likely involves developmental cues that persist in the adult. The migration guidance cue reelin, for example, is expressed in the adult human and rodent dentate gyrus and is implicated in DGC layer dispersion in human mTLE (Haas et al., 2002). Recent work shows that hilar-ectopic DGCs in experimental mTLE arise from aberrantly migrating DGC progenitors (Parent et al., 2006). This altered migration is associated with loss of reelin signaling that may disrupt the migratory behavior of DGC progenitors (Gong et al., 2007).

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Brain-derived neurotrophic factor (BDNF) is another candidate, as hippocampal BDNF infusion in adult rat increases DGC neurogenesis and induces ectopic DGCs (Scharfman et al., 2005). The combined action of multiple factors likely are involved, each potentially acting on different steps in the neurogenic cascade.

## CONSEQUENCES OF SE-INDUCED NEUROGENESIS

Recent evidence suggests that DGCs generated after SE integrate abnormally into existing networks (Scharfman et al., 2000; Dashtipour et al., 2001; Jessberger et al., 2007a). Intracellular recordings from hilar-ectopic DGCs in hippocampal slices from epileptic rats show that they are hyperexcitable and burst synchronously with CA3 pyramidal cells (Scharfman et al., 2000). This increased excitability may relate to excessive excitatory input onto basal dendrites of ectopic DGCs (Dashtipour et al., 2001). Moreover, granule cells that extend hilar basal dendrites stably integrate into the dentate circuitry (Jessberger et al., 2007a), leading to lasting changes in connectivity that may contribute to epileptogenesis. Further support for the epileptogenicity of SE-induced neurogenesis comes from the work of Jung et al. (2004). They inhibited DGC neurogenesis after pilocarpine treatment by antimetabolic agent infusion and found that rats developed fewer and shorter spontaneous recurrent seizures than controls. These data suggest that DGCs integrate abnormally after SE, are hyperexcitable, and may contribute to seizure generation or propagation (Fig. 1).

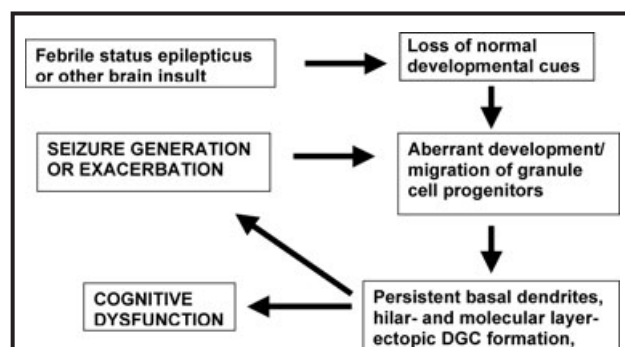
Other work suggests a more complex influence of adult-born DGCs on hippocampal excitability after SE. In contrast to chemoconvulsant SE models, the morphology of developing DGCs after electrically-induced SE appears unaltered, at least in the DGC layer (Jakubs et al., 2006). In this model, newborn DGCs receive increased inhibitory in-

put resulting in less excitability than newborn cells generated after exercise (Jakubs et al., 2006). Some DGCs generated after SE therefore could have compensatory antiepileptogenic effects.

As mentioned above, adult hippocampal neurogenesis likely underlies specific learning and memory functions, raising the possibility that defective neurogenesis may contribute to the progressive memory dysfunction seen in mTLE (Fig. 1; Helmstaedter, 2002). This idea is supported by recent findings of Jessberger et al.. They suppressed neurogenesis after kainic acid-induced SE with the histone deacetylases inhibitor and anticonvulsant valproic acid and found that decreased neurogenesis was associated with better performance in a hippocampus-dependent object recognition task than in SE controls (Jessberger et al., 2007b). These data may be reconciled with the decline in neurogenesis chronically after the experimental SE described above (Hattiangady et al., 2004), if one considers that memory function may be impaired both by aberrant neurogenesis or a reduction of normal DGC neurogenesis. However, further progress in understanding the functional consequences of SE-induced neurogenesis for epileptogenesis or cognitive impairment awaits more specific and selective experimental strategies to manipulate adult neurogenesis.

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**Figure 1.**

Aberrant neurogenesis hypothesis of mTLE pathophysiology.

*Epilepsia* © ILAE

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