

Active Site Studies on Muscle Carbonic Anhydrase III^a

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INTRODUCTION

Carbonic anhydrase occurs, in reptiles, birds and mammals, in at least three genetically distinct isozyme forms designated CA I, CA II, and CA III.¹ The CA III isozymes, which are abundantly present in red skeletal muscle, possess notably low CO₂ hydratase and esterase (*p*-nitrophenyl acetate) activities^{2,3} and are remarkably resistant to inhibition by some sulfonamides^{3,4} when compared with CA I and CA II. CA III isozymes from rabbit, pig and bovine muscle have also been shown to possess a novel low acid phosphatase activity.² These data tempt speculation that the CA III isozymes may have an as yet unknown physiological role other than catalysis of CO₂ hydration. The active site structure of the CA III isozymes shows striking differences from those of CA I and CA II.¹ Of the five putative active site residues unique to CA III and invariant in all CA III isozymes examined, two, at positions 67 and 91, are arginine.

It has been previously demonstrated that the treatment of bovine and gorilla CA III with butanedione results in the activation of the bicarbonate dehydration reaction, whereas the bovine and human CA I and CA II isozymes were unaffected by similar treatment.⁵ This report is an extension of those studies.

METHODS

Human CA I and CA II were purified from red cell hemolysates by affinity chromatography.⁶ Human CA III was purified from autopsied psoas major muscle by affinity chromatography and gel filtration⁷ followed by chromatography on DEAE-cellulose. Similar procedures were used in purifying CA III from bovine and chicken skeletal muscle. Bicarbonate dehydration activity was measured using a pH-stat assay system similar to that of Hansen and Magid⁸ with 1.0 M sulfuric acid as titrant. Assays were performed at pH 7.1 and 2° C in a volume of

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10 ml. The reaction mixture comprised 6.7 mM Na₂HPO₄, 6.7 mM NaH₂PO₄, 30 mM NaHCO₃, and 0.5 mM EDTA. Esterase activity towards *p*-nitrophenyl acetate (1 mM) was measured spectrophotometrically at 348 nm in 30 mM sodium diethylmalonate buffer, pH 7.2.⁹ Modification of the enzyme with freshly prepared butanedione was performed in the dark at 25° C in 50 mM borate buffer, pH 8.3. These conditions are known to selectively modify arginine residues.¹⁰ Tryptic digestion of CA III and BD-modified CA III was effected, after preliminary denaturation with 1 M HCl followed by 1 M NaOH, by incubation with 2% w/w trypsin in 0.05 M Tris/Cl buffer, pH 8.65, for 16 hours at 37° C. Tryptic peptides were separated by HPLC and then sequenced and identified by HPLC as described by Hewett-Emmett *et al.*⁷

RESULTS AND DISCUSSION

Modification with butanedione caused a pronounced activation of both the bicarbonate dehydration and esterase activities of human and bovine CA III, but only the esterase activity of chicken CA III (TABLE 1). No effect was observed when human CA I and CA II were similarly incubated with butanedione. Low butanedione concentrations gave rapid activation of the bicarbonate dehydration reaction of 70–100% (FIG. 1). Activation of the esterase activity followed a slower time course and required higher butanedione concentration (25 mM) but produced a greater degree of activation (TABLE 1).

Tryptic digestion and sequence analysis were performed on human CA III (56 μM) after modification with butanedione: (A) at 2.5 mM concentration for 10 min at 25° C and (B) at 25 mM concentration for 45 min at 25° C and on appropriate controls in the absence of butanedione. The results for (A) showed very little difference from its control apart from a slight increase in a peptide containing amino acids 90–113, suggesting partial modification of Arg-91. In contrast, (B) showed extensive modification compared to its control. Similar peptide analysis indicated modification of arginine residues at positions 39, 67, 80 or 89, and 91. Position 39 is not arginine in bovine CA III and position 89 is arginine in CA I and CA II as well as CA III. Arg-80, which may have been modified, is not at the active

TABLE 1. Effect of 2,3-Butanedione (BD) on the Activities of Carbonic Anhydrase Isozymes

Isozyme	Bicarbonate Dehydration Activity	Ester Hydrolysis Activity (<i>p</i> -Nitrophenyl Acetate)
Human CA III	Activation ^a	Activation ^b
Bovine CA III	Activation	Activation
Chicken CA III	No effect	Activation ^c
Human CA II	No effect	No effect ^d
Human CA I	No effect	No effect

^a See FIGURE 1.

^b Incubation of Human CA III (56 μM) with BD (25 mM) for 24 hours produced >5-fold activation.

^c Incubation of chicken CA III (23 μM) with BD (25 mM) for 24 hours produced >10-fold activation.

^d Similar incubation with BD for 24 hours caused a slight decline in activity. Other conditions were as described in the text.

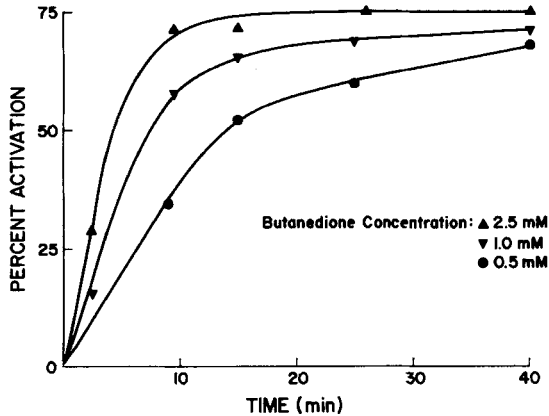


FIGURE 1. Activation of the bicarbonate dehydration activity of human CA III by incubation with 2,3-butanedione in 50 mM borate, pH 8.3 at 25° C. The enzyme concentration was 13.6 μ M.

site. The results suggest, therefore, that activation is associated with modification of arginyl residues at 67 and 91, which are unique to CA III. Limited sequencing studies on chicken CA III indicate the presence of either lysine or arginine at positions 67 and 91.¹ Absence of activation of the bicarbonate dehydration activity of this isozyme by butanedione modification suggests that arginine may not be present in both positions.

REFERENCES

1. TASHIAN, R. E., D. HEWETT-EMMETT & M. GOODMAN. 1983. *In* Isozymes: Current Topics in Biological and Medical Research. Vol. 7: 79-100. Alan R. Liss. New York.
2. KOESTER, M. K., L. M. PULLAN & E. A. NOLTMANN. 1981. *Arch. Biochem. Biophys.* **211**: 632-642.
3. SANYAL, G., E. R. SWENSON, N. I. PESSAH & T. H. MAREN. 1982. *Mol. Pharmacol.* **22**: 211-220.
4. MAREN, T. H. & G. SANYAL. 1983. *In* Annual Review of Pharmacology and Toxicology. Vol. 23: 439-459. Annual Reviews Inc., Palo Alto, CA.
5. TASHIAN, R. E., J. T. JOHANSEN & E. CHRISTIANSEN. 1980. *Fed. Proc.* **39**: 1303.
6. OSBORNE, W. R. A. & R. E. TASHIAN. 1975. *Anal. Biochem.* **65**: 297-303.
7. HEWETT-EMMETT, D., R. J. WELTY & R. E. TASHIAN. 1983. *Genetics* **105**: 409-420.
8. HANSEN, P. & E. MAGID. 1966. *Scand. J. Clin. Lab. Invest.* **18**: 21-32.
9. ARMSTRONG, J. MCD., D. V. MYERS, J. A. VERPOORTE & J. T. EDSALL. 1966. *J. Biol. Chem.* **241**: 5137-5149.
10. BORDERS, C. L. & J. F. RIORDAN. 1975. *Biochemistry* **14**: 4699-4704.