

FACTORS INFLUENCING CLINICAL OXYGEN TOXICITY

John W. Bean

*Department of Physiology, University of Michigan
Ann Arbor, Mich.*

Introduction

Since the early work of Priestley¹ and Lavoisier,² it has been recognized, though questioned by many,³ that breathing increased concentrations of O₂ may have adverse as well as therapeutic effects on animal organisms. The exact mechanism of causation of these adverse reactions—which for want of a better name have been referred to as those of O₂ poisoning—is not known. But there are numerous factors which influence its occurrence and severity. If the clinical use of O₂ in increased concentrations, especially at high pressure, is to achieve the success which it certainly seems to promise, recognition of its potential dangers and limitations, as well as knowledge of the more important factors which contribute to O₂ toxicity, is imperative.

Fundamental Factors Influencing O₂ Toxicity

Fundamentally, the occurrence of this phenomenon is determined by: (1) an intensity factor, i.e., the O₂ concentration and (2) a duration factor or length of exposure, and both of these depend on (3) the inconstant and frequently unpredictable susceptibility of the subject. Manifestation of O₂ poisoning induced by increased concentrations of O₂ at normal atmospheric pressure (OAP) and at higher pressures (OHP), i.e., partial pressures in excess of pure O₂ at normal atmospheric pressure (hyperbaric O₂) have many features in common, but in OHP there are additional manifestations of CNS involvement not usually seen in OAP. Nevertheless, for brevity and convenience, here we shall draw no sharp line of demarcation.

Pulmonary Changes in O₂ Toxicity

The first toxic effects of O₂ to be observed were those noted by Priestley¹ and described by Lavoisier² as an “incendiary” action on the lungs of animals breathing O₂ at atmospheric pressure. These pulmonary effects which constitute a major manifestation of O₂ toxicity involve capillary and alveolar membrane changes which have been described in considerable detail by numerous investigators over the years.³⁻⁶ These changes may be so severe that pulmonary function fails and the animal may succumb from what in effect is an asphyxia or an “hyperoxic anoxia.”³ Fortunately, these changes can be prevented or their severity mitigated by appropriate adjustment of the fundamental factors of intensity and duration.

Inherent Differences in Susceptibility as Factors Influencing O₂ Toxicity

Evidence of pulmonary pathology in O₂ toxicity has been limited almost entirely to that in experimental animals, whose susceptibility to pulmonary damage by O₂ is much greater than that of man. Indeed, human lungs are so much more resistant to the toxic effect of O₂ that it has been said that human lung damage by O₂ is negligible. But man is by no means immune to pulmonary damage, either by OAP or OHP.^{3,7,8} Recent reports^{9,10} indicate that continuous therapeutic administration to man, for as short a time as two days, produces histopathology of capillary congestion and proliferation with capillary tufts projecting

into alveoli, diffuse fibrosis, edema, and thickening of alveolar membranes—changes not attributable to simple atelectasis.

In addition to species differences in susceptibility, there are also pronounced individual differences, not only in the pulmonary involvement,⁷ but also and especially in the CNS effects. Knowledge concerning human tolerance to O₂ toxicity is presently limited. Thus, there is urgent need of very careful observation and rigid controls in all therapeutic, clinical, and experimental procedures, and methods of O₂ administration must be such as to insure accurate determination of the concentration of O₂ actually inhaled if reliable thresholds are to be established. This has not always been the case.

The factor of susceptibility is complicated also by the fact that it differs in different tissues. This is well illustrated by another predominantly vascular manifestation of O₂ poisoning, viz., that of retrolental fibroplasia. The retina of the premature infant is particularly susceptible to injury by O₂ in concentrations very little above 35 to 40 per cent at atmospheric pressure.^{11,12} Infant lungs are much more resistant than the retina,¹¹ but they are not immune to O₂ damage.¹³ This relationship is reversed with increasing maturity: the retinal vasculature becomes more resistant than that of the lungs. However, the adult retina, at least in animals, is not entirely immune and may be permanently damaged by exposure to OHP.¹⁴ Permanent partial loss of visual function in adult chickens and rats has accompanied permanent motor paralysis induced by repeated exposure to O₂ at 65 psi.¹⁵ Exposure to OHP causes severe temporary visual disturbances in man,¹⁶ but apparently no permanent alterations have as yet been reported. Nevertheless, the adult human eye may prove to be quite vulnerable to damage by OHP—a fear which has been voiced also by others.¹⁷ Thus, age is another important factor in determining the reaction to O₂ poisoning.

Generally speaking, young individuals are less susceptible to O₂ toxicity, especially as manifested by O₂ convulsions, than are older ones. Infant rats have shown few or no convulsions in OHP until 18 or 19 days old. This is probably related to the immaturity of the adrenals.

Although the factor of age is of considerable importance in the occurrence of O₂ poisoning in man, almost all of the current information on thresholds of O₂ toxicity in man³ has been obtained from relatively young healthy adult males—those chosen for naval and diving training programs. Such select subjects at rest remain relatively free of CNS symptoms in O₂ at 3 atm. abs. for exposures of two to three hours duration. But even in such select groups, there is considerable individual variation in susceptibility.¹⁶ Some investigators¹⁹ set the period of freedom from CNS reaction at two hours in O₂ at 2.8 atm. abs., but 80 per cent of their subjects suffered symptoms after two hours exposure to O₂ at 3.4 atm. abs. Others²⁰ have held that probably no absolutely safe threshold can be established for all subjects even in a select group, because of the wide individual variation in susceptibility. The state of physical rest of the individual, as well as his emotional state, are important factors which are not always readily controllable.

If it is difficult to set absolutely safe thresholds for select groups of young adult human subjects, one might expect it would be even more difficult, if not impossible, to set absolutely safe thresholds for human patients differing widely in age, physical and mental health, and emotional stability. However, a recent study of volunteers and patients indicates that subjects breathing pure O₂ at 3 atm. abs. without a mask are “unlikely” to have symptoms of O₂ poisoning for periods of at least one-half hour.²¹ Perhaps one may question whether in some cases the occurrence of some CNS symptoms may not be the price one has to pay for the clinical benefits sought in OHP.

It is disconcerting, however, that one cannot safely assume that because an individual has been free of symptoms during the period of maintained high pressure, he is "out of the woods," so to speak, and that he will remain so during decompression. Even under rigidly controlled decompression, animals which have been free of reactions during the maintenance of high pressure, not infrequently suffer very severe reactions during and after decompression and great difficulty is sometimes encountered in returning them safely to normal atmospheric pressure.³ Ruling out lung rupture, which may result from too rapid decompression where airways have been occluded, gas embolism, bends, or decompression sickness, leaves the cause of this "off effect" as it has been called, a matter of speculation. Apparently, as might be expected from these animal experiments, this "off effect" occurs also in human subjects.²¹

Metabolism as a Factor Influencing O₂ Toxicity

Low temperature has long been recognized as a deterrent to the occurrence of O₂ toxicity, whereas an elevation of temperature has the opposite effect.²²⁻²⁴ The reason for this is believed to be due largely to the attendant changes in metabolism. Thus, the use of hypothermia in clinical procedures in OHP should diminish the possibility of O₂ poisoning as well as reduce O₂ consumption.

Thyroid Effects

Since an elevation of metabolism by various means augments the toxic effects of O₂,²⁵ thyroid activity becomes another significant factor in O₂ toxicity.^{23,26,27} The administration of thyroid or thyroxin in experimental animals results in very pronounced enhancement of O₂ toxicity of both OAP and OHP, and thyroidectomy has a deterrent effect. One may reasonably assume that these factors would also be operative in human subjects selected for clinical exposure to OHP; thus, a routine BMR determination might be of help in anticipating a subject's possible tolerance to OHP.

Anesthesia

It has been known since the demonstration by Paul Bert²⁸ that anesthesia induced by a variety of agents³ protects against the occurrence of convulsive seizures of O₂ poisoning and is essential in many experimental and clinical procedures involving exposures to OHP. But it may be well to emphasize that OHP tends to counteract and lighten various anesthetics.³ The protective effect of anesthesia has been attributed, in part, to the attendant lowering of metabolism.²⁹ However, it is of special interest that anesthesia (sodium pentobarbital), paradoxical as it may seem, may also potentiate the toxic influence of OHP on the CNS and lead to permanent motor paralyzes in animals exposed to O₂ at 45 psi in as short a time as 15 min.³⁰ These paralyzes are apparently identical with those induced in our unanesthetized animals by repeated exposures to O₂ at 65 psi with which there was associated cerebral vascular pathology³¹ and permanent visual defect.³² However, some investigators, finding no very significant cerebral lesions following less severe exposures to OHP, believe the CNS effects of OHP are entirely functional in nature.³³ Happily, no permanent CNS effects have as yet been reported in men after exposures to OHP, either in the many experimental procedures^{34,35} or in the extensive clinical procedures initiated and carried out by Boerema and associates at Amsterdam or those of Illingsworth and Smith and associates at Glasgow.

CO₂ and Metabolic Products

The basic cause of oxygen convulsion—like that of pulmonary damage by O₂—is not known. It may be a variable and inconstant combination of several factors. But among these an increase in tissue CO₂ and the attendant changes in tissue pH, such as might occur with increased metabolism, are especially important.^{3,36} In this connection it is particularly significant that oxygen convulsions seldom occur either in animals or man in O₂ pressures much below 3 atm. abs. At this O₂ pressure and above there is sufficient extra O₂ dissolved in the plasma to supply the ordinary needs of the tissues. Thus, the oxyhemoglobin is not normally reduced, and as a result, the blood loses a large part of its buffer capacity.^{3,36,37} This interferes with the normal carriage of CO₂ by the blood and results in an increase in the tissue CO₂ and acidity.³⁸⁻⁴⁰ Although this elevation in tissue CO₂ has been shown to be less³⁷ than has been claimed by some authors,^{41,42} it assumes very considerable significance. For it has been shown by numerous investigators that low concentration of CO₂, which under normal conditions is innocuous (e.g., such as might be present in the patient's mask), has peculiarly toxic effects in the presence of O₂ in concentrations above 3 atm.^{3,34,36,38,39,43} This emphasizes a fact of great general importance in OHP, viz, that it is not safe to assume that the effect of a drug or biological agent when administered in OHP should be exactly the same as that induced when administered under normal conditions. This implies the need of pharmacological studies in OHP.

The influence of CO₂ and O₂ poisoning is complicated further, because, among other things, of its well-known dual action, i.e., of excitation and depression.^{44,45} The toxicity of O₂ at 3 atm. and above is augmented by CO₂ in its lower concentrations (about five per cent of one atm.),^{3,36,43,44,46} whereas in higher concentrations (e.g., about 16 per cent), it eliminates O₂ convulsions but not death.^{34,40,43} This seemingly "protective" influence of CO₂, as it is sometimes called, might better be designated as a CO₂ depression or CO₂ narcosis. Thus, in clinical application of OHP, any elevation in tissue CO₂, whether of endogenous or exogenous origin, should be given careful consideration. The administration of OHP or even OAP to an hypercapneic animal or man may complicate the situation by severe depression of respiratory control.

But where there is an anoxic component in O₂ poisoning or where surgical procedures involving anoxia are carried out under O₂ at high pressure, the presence of CO₂ (e.g., two per cent in exposures to O₂ at 3 atm. abs. may well be advantageous,^{47,48} because CO₂ counteracts the influence of anoxia.^{45,49} Both the augmentative and inhibitive effects of CO₂ in toxicity are, in part, at least attributable to the influence of CO₂ on the cerebral vasculature and blood flow.

The finding that Tris buffer [tris(hydroxymethyl) aminomethane] or THAM (an efficacious *in vivo* buffer for hypercapneic acidosis)⁵⁰ affords an appreciable degree of protection against the various aspects of O₂ poisoning by OHP^{51,52} (FIGURE 1), including that of lung damage⁵¹ (FIGURE 2), reemphasizes the importance of tissue CO₂ and acidity, which have long been stressed as important contributory factors in the causation of the toxic action of OHP.^{53,39}

Endocrines

The endocrine system provides a host of factors which influence O₂ toxicity, as demonstrated in experimental animals. The stress of O₂ toxicity induces hypertrophy of the hypophysis and the typical stress reaction in the adrenal cortex,⁵³

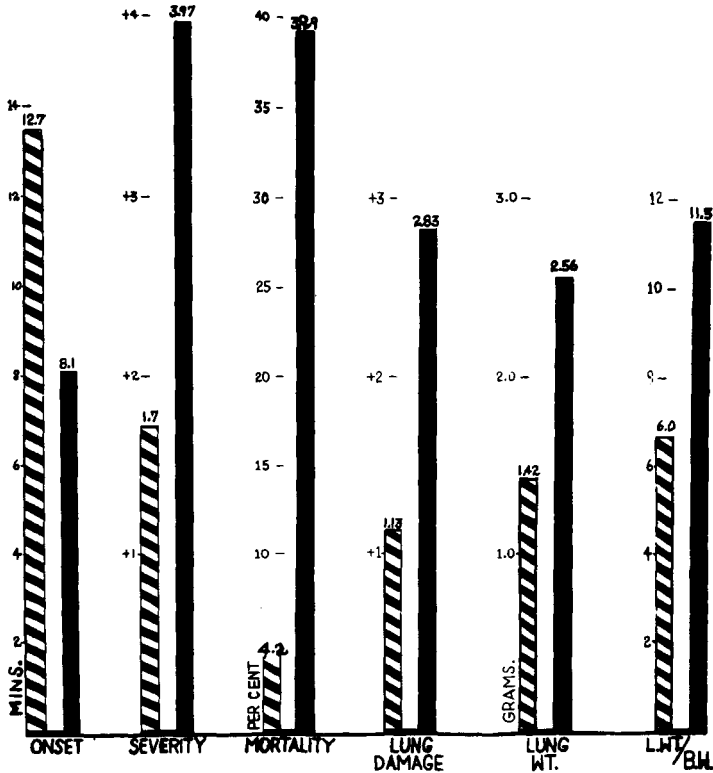


FIGURE 1. Protective action of Tris (THAM) against toxic action of OHP as determined by time of onset of seizures, severity, mortality, lung damage, lung weight, and lung weight/body weight ratio. The crosshatched column represents the Tris-treated rat, the solid column, the control. Both test and controls were exposed to OHP together.

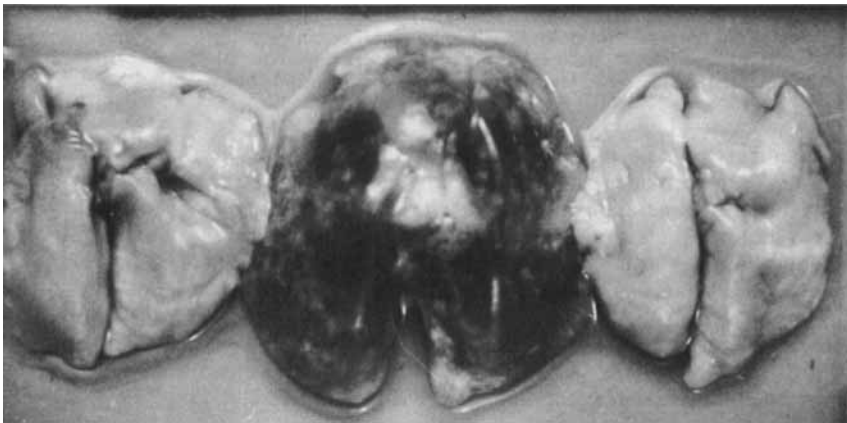


FIGURE 2. Protective action of Tris (THAM) buffer against pulmonary damage by OHP. (Left, TOHP) Lungs from test rat treated with Tris before exposure to OHP; (center, SOHP) lungs from control injected with saline before exposure, both rats exposed to OHP together; (right, T, Air) lungs from rat injected with Tris and kept in air at atmospheric pressure.

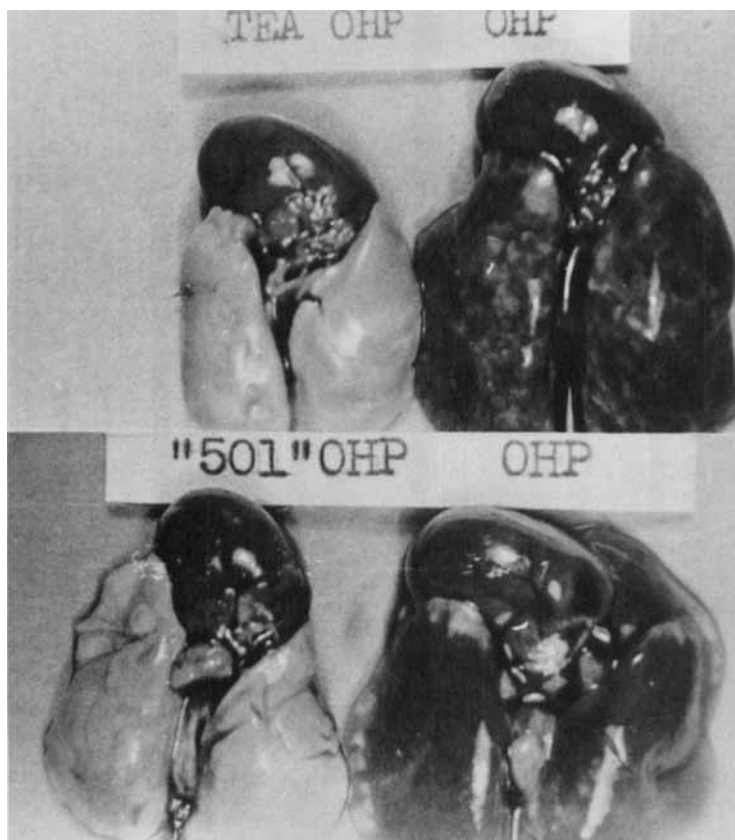


FIGURE 3. Protection against toxic action of OHP on lungs by sympathetic blocking agents: (a, top) by TEA; (b, bottom) by SKF501. In each case the lungs from the test animal are on the left. Both the test and control animals were exposed to OHP together. Essentially, similar effects are induced by Hexamethonium, Dibenamine, and Dibenzylene.

but, contrary to expectation, it was found that hypophysectomy⁵⁴ and adrenalectomy^{55,56} each provides protection against the toxic action of OHP and in accord with this, cortisone augments it.^{57,58} Thus, the hypophyseal-adrenocortical axis, which serves as a "hormone" defense mechanism against the usual types of stress, actually becomes a liability in the stress of OHP and its increased activity augments rather than diminishes the severity of O₂ toxicity.

Similarly, the sympatho-adrenomedullary axis, which serves as another defense mechanism (nervous) against the usual types of stress, becomes something of a liability in the stress of OHP.⁵⁹ That this is so is shown by the finding that sympathetic blocking agents (TEA, SKF501,* Dibenamine®, Dibenzylene®, Hexamethonium) protect against the toxic action of OHP (FIGURES 3a and 3b) by: (1) diminishing the release of epinephrine which augments O₂ toxicity⁵⁹ and (2) by blocking sympathetic nerves to blood vessels. Thus, the vascular damage

* N-(9-fluorenyl)-N-ethyl-β-chlorethylamine.

by OHP appears to be mediated, in large part, by the sympathetics. The fact that such sympathetic blocking agents prevent, in large measure, the pulmonary damage by OHP suggests that this damage is largely of neurogenic origin.⁶⁰ The combined hypophyseal-adrenocortical and sympatho-adrenomedullary axes, which serve as defense in the usual types of stress but augment the stress of O₂ toxicity, are diagrammed in FIGURE 4. Chlorpromazine, an effective depressant of the higher centers of the sympathetics which has been found to afford a very pronounced measure of protection against the various aspects (FIGURE 5) of O₂ toxicity in experimental animals,⁶¹ is also a valuable adjunct in the clinical use of OHP.⁶²

Thus, neuroendocrine factors have a very significant influence in O₂ toxicity relating possibly to: (1) the cause of the wide, individual variation in the susceptibility and unpredictability of reaction to OHP; (2) the emotional and temperamental state of the subjects and patients selected for exposure to OHP; and (3) in the selection of medication, if any, to be used in the clinical application of such exposures. If then one could determine just what factors are responsible

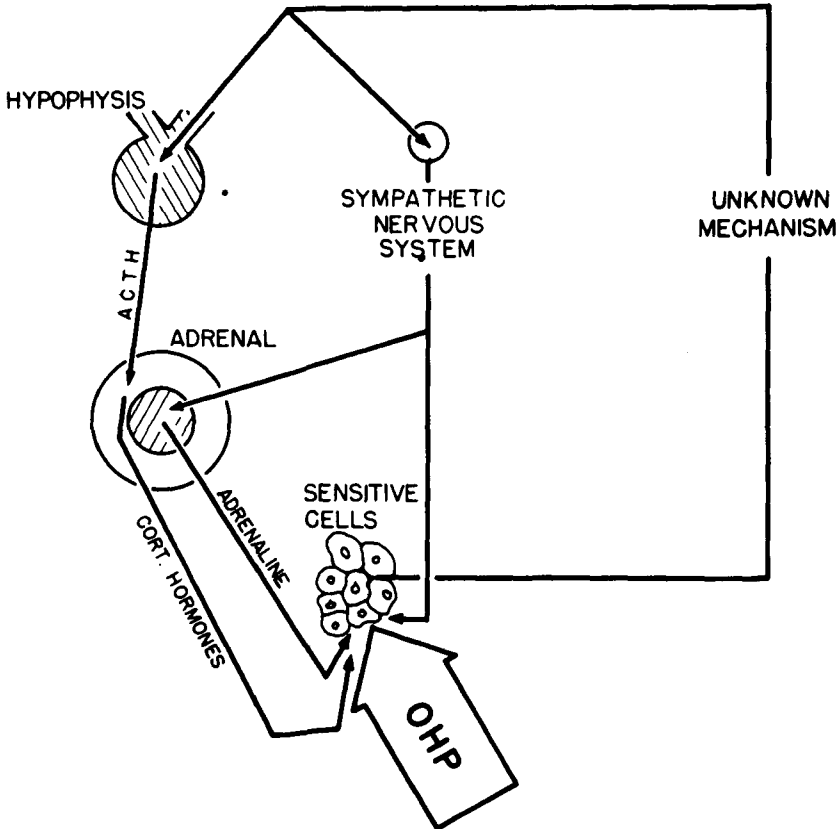


FIGURE 4. Diagrammatic representation of combined hypophyseal-adrenocortical and the sympatho-adrenomedullary "defense" mechanisms, each of which enhances the toxic reaction to the stress of OHP (diagram by P. C. Johnson).

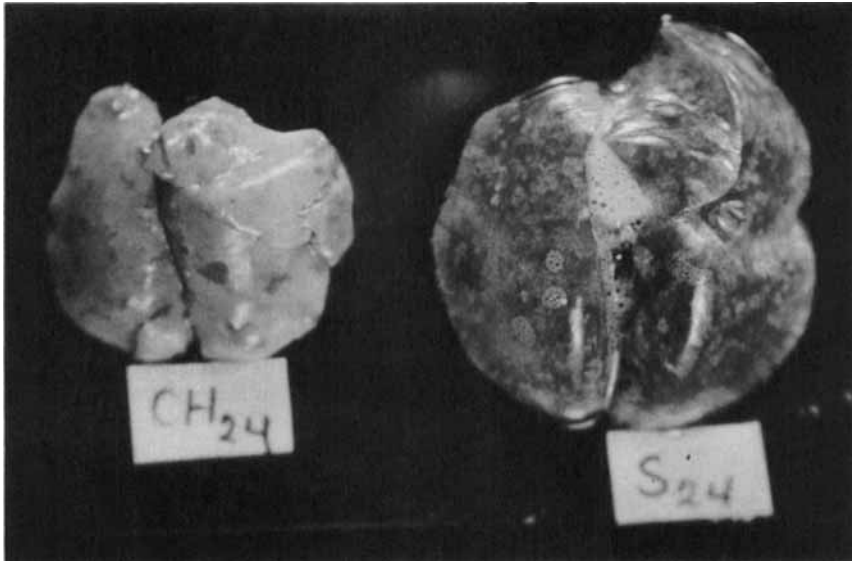


FIGURE 5. Protection against toxic action of OHP on lungs by chlorpromazine (Thorazine®). Lungs from test animal (CH₂₄) on the left; lungs from the control (saline, S₂₄), on the right; both animals exposed to OHP together.

for the wide and inconstant individual variation in susceptibility to O₂ poisoning, it would help immeasurably in an understanding of the fundamental mechanisms of its causation.

Factors of Changing Cerebral Blood Flow and O₂ Tension

Still other major factors influencing clinical O₂ toxicity are changes in brain blood flow and cerebral O₂ tension. But currently there are no entirely satisfactory means of determining absolute values of O₂ tension in cerebral tissue. There is evidence³⁵ that in unanesthetized men exposed to O₂ at 3.5 atm. the mean pO₂ of mixed venous brain blood and inferentially that of cerebral tissue, is not much, if any, above normal levels. This has been attributed to cerebral vasoconstriction³⁵ which occurs in breathing O₂.^{37,63,64} But the finding of minute bubbles in blood after its withdrawal to atmospheric pressure from the superior sagittal sinus of anesthetized dogs exposed to O₂ at pressures of 3.5 to 6 atm. abs., together with polarographic evidence, suggests that in such exposures, the pO₂ of brain tissue itself, as well as that of circulating sagittal sinus blood, rises well above normal levels, despite the cerebral vasoconstriction.⁶⁵⁻⁶⁷

The enhancement of the toxic action of OHP by CO₂ in respired O₂ may well be due, in large part, to its cerebral vasodilatory effect.^{64,68} Such addition of CO₂ causes a pronounced elevation in pO₂, not only of mixed venous brain blood,^{37,65} but also in the cerebral tissue itself, as indicated by polarographic recordings from both anesthetized and unanesthetized animals.⁶⁵⁻⁶⁷

Simultaneous recordings from several regions of the brain of either anesthetized or unanesthetized animals exposed to OHP have shown well-pronounced elevations in brain tissue O₂. But in unanesthetized animals, superimposed upon

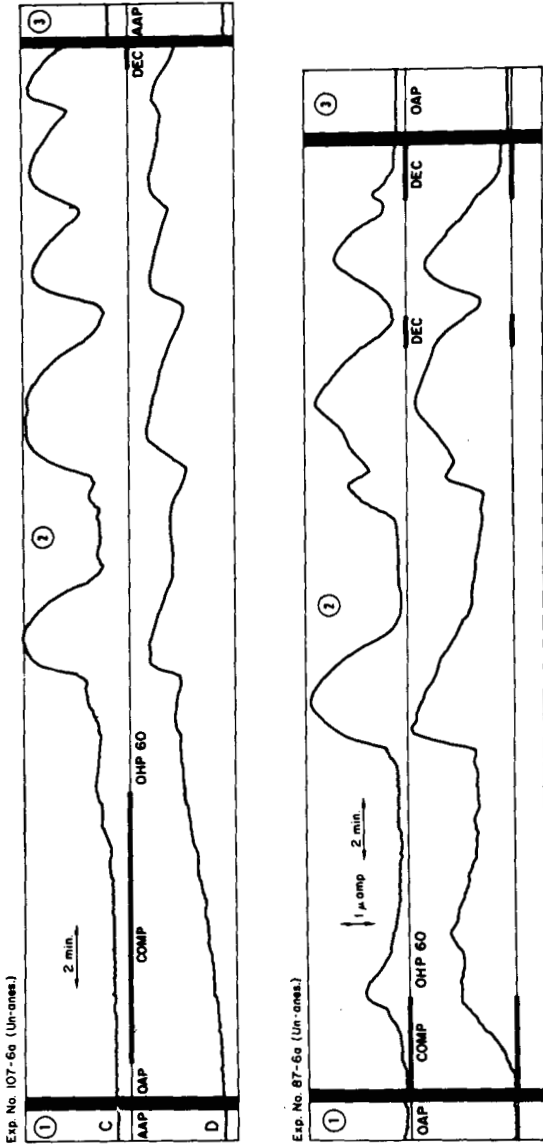


FIGURE 6. Simultaneous polarographic recordings of changes in O_2 availability in two regions of the brain, cortex (C) and hypothalamus (D), in an unanesthetized rat with chronically implanted matched electrodes. AAP indicates breathing air at atmospheric pressure; OAP, O_2 at atmospheric pressure; COMP indicates compression to OHP of 60 psi; Dec, decompression. Each pair of tracings was recorded from the same animal at an interval of several days. Decompression in each experiment was begun after signs of convulsive onset in order to avoid excessively severe seizures and to allow for recovery.

this general elevation in brain pO_2 , fluctuations appear, which usually increase in magnitude and may culminate in precipitous cyclic elevations frequently associated with overt convulsive seizures. These fluctuations commonly differ in different parts of the brain at any one time, and have been tentatively ascribed to regional changes in brain blood flow and metabolism^{65,66} (FIGURE 6). It may be inferred that there are attendant equally transient and asynchronous regional changes in brain pCO_2 , pO_2 , and pH which are masked in mixed venous brain blood. Thus, it would appear that in the unanesthetized animal and probably in man exposed to OHP, there are wide regional asynchronous swings in brain blood flow associated with wide regional swings in pO_2 , pCO_2 , and pH , all of which are important factors influencing the reaction of the animal as a whole in O_2 poisoning.

References

1. PRIESTLEY, J. 1906. Alembic Club Reprints. No. 7. Univ. Chicago Press. Chicago, Ill.
2. LAVOISIER, A. L. 1783. Soc. Roy. Med. (Paris) 5: 569.
3. BEAN, J. W. 1945. Physiol. Rev. 25: 1.
4. SMITH, L. 1899. J. Physiol. 24: 19.
5. CEDERGREN, B., L. GYLLENSTEN & J. WERSALL. 1959. Acta Paediat. 48: 477.
6. VAN DEN BRENK, H. A. S. & D. JAMIESON. 1962. Australian J. Exptl. Biol. 40: 37.
7. BECKER-FREYSENG, H. G. & H. CLAMANN. 1939. Klin. Wochschr. 18: 1382.
8. LEE, W. L., P. B. CALDWELL & H. S. SCHILDKRAUT. 1963. Federation Proc. 22: 305.
9. PRATT, P. C. 1958. Am. J. Pathol. 34: 104.
10. GABLE, W. D. & F. M. TOWNSEND. 1962. Aerospace Med. 33: 1343.
11. PATZ, A. 1957. Pediatrics 19: 509.
12. ASHTON, N., G. WARD & G. SERPELL. 1954. Brit. J. Ophthalmol. 38: 394.
13. GYLLENSTEN, L. J. & B. E. HELLSTRÖM. 1954. Acta Paediat. Suppl. 100. 43: 131.
14. NOELL, W. K. & W. H. BAKER. 1957. Am. J. Physiol. 87: 584.
15. BEAN, J. W. & E. C. SIEGFRIED. 1945. Federation Proc. 4: 6.
16. BEHNKE, A. R., H. S. FORBES & E. P. MOTLEY. 1936. Am. J. Physiol. 144: 436.
17. BROWN, I. 1963. An. Surg. 158: 360.
18. BEAN, J. W. & R. BAUER. 1951. Unpublished data.
19. YARBROUGH, D. D., W. WELHAM, E. S. BRITTON & A. R. BEHNKE. 1947. Project X-337 Sub No. 62). Rept. No. 1. Experimental Diving Unit. Naval Gun Factory. Washington, D. C.
20. DONALD, K. W. 1947. Brit. Med. J. 1: 667.
21. FOSTER, C. A. & I. CHURCHILL-DAVIDSON. 1963. J. Appl. Physiol. 18: 492.
22. CAMPBELL, J. A. 1937. J. Physiol. 89: 17.
23. GROSSMAN, M. S. & K. W. PENROD. 1949. Am. J. Physiol. 156: 182.
24. WILLIAMS, C. M. & H. K. BEECHER. 1944. Am. J. Physiol. 140: 556.
25. CAMPBELL, J. A. 1937. J. Physiol. 90: 91.
26. BEAN, J. W. & R. BAUER. 1952. Proc. Soc. Exptl. Biol. Med. 81: 693.
27. SMITH, C. W., J. W. BEAN & R. BAUER. 1960. Am. J. Physiol. 199: 883.
28. BERT, P. 1878. La Pression Barometrique. G. Masson. Paris, France.
29. CAMPBELL, J. A. 1939. Libro Homenagem. : 89. A. Ozario de Almeida. Rio de Janeiro, Brazil.
30. JAMIESON, D. & H. A. S. VAN DEN BRENK. 1962. Nature 194: 777.
31. BEAN, J. W. 1945. Proc. Soc. Exptl. Biol. Med. 58: 21.
32. BEAN, J. W. 1945. Federation Proc. 4: 6.
33. EDSTRÖM, J. E. & H. ROCKERT. 1962. Acta Physiol. Scand. 55: 255.
34. SHAW, L. A., A. C. MESSER & A. R. BEHNKE. 1934. Am. J. Physiol. 108: 652.
35. LAMBERTSEN, C. J., J. H. EWING, R. H. KOUGH, R. GOULD & M. W. STROUD. 1955. J. Appl. Physiol. 8: 255.
36. GESELL, R. 1923. Am. J. Physiol. 66: 5.
37. LAMBERTSEN, C. J., R. H. KOUGH, D. Y. COOPER, G. L. EMMEL, H. H. LOESCHKE & C. F. SCHMIDT. 1953. J. Appl. Physiol. 5: 471.
38. BEAN, J. W. 1929. Proc. Soc. Exptl. Biol. Med. 26: 832.
39. BEAN, J. W. 1931. J. Physiol. 72: 27.
40. BEHNKE, A. R., L. A. SHAW, C. W. SHILLING, R. M. THOMSON & A. C. MESSER. 1934. Am. J. Physiol. 107: 13.
41. TAYLOR, H. J. 1949. J. Physiol. 109: 272.

42. CAMPBELL, J. A. 1930. *J. Physiol.* **68**: 7.
43. MARSHALL, J. R. & C. J. LAMBERTSEN. 1955. *J. Appl. Physiol.* **183**: 662.
44. SMITH, C. W. & J. W. BEAN. 1955. *Am. J. Physiol.* **183**: 662.
45. GELLHORN, E. 1956. *Foundations of Neurology and Psychiatry*. : 43. U. Minn. Press. Minneapolis, Minn.
46. CHAPIN, J. 1955. *Proc. Soc. Exptl. Biol. Med.* **90**: 663.
47. MEIJNE, N. G., D. M. E. VERMUELEN-CRANCH, M. E. SUYTER, S. J. P. ELOFF, L. SCHRIPSEMA, L. DEEN, G. SCHOEMAKER & I. BOEREMA. 1962. *J. Thoracic Surg.* **44**: 749.
48. RICHARDS, V., D. PINTO & P. COOMBS. 1963. *Ann. Surg.* **158**: 349.
49. GIBBS, F. A., E. L. GIBBS, W. G. LENNOX & L. F. NIMS. 1943. *J. Aviation Med.* **14**: 250.
50. NAHAS, G. G., E. C. JORDON & J. C. LIGOU. 1959. *Am. J. Physiol.* **197**: 1308.
51. BEAN, J. W. 1961. *Am. J. Physiol.* **201**: 737.
52. SANGER, C., G. G. NAHAS, A. R. GOLDBERG & G. M. D'ALESSIS. 1961. *Ann. N. Y. Acad. Sci.* **92**: 710.
53. BEAN, J. W. 1951. *Federation Proc.* **10**: 11.
54. BEAN, J. W. 1952. *Am. J. Physiol.* **170**: 508.
55. GERSCHMAN, R. & W. O. FENN. 1954. *Am. J. Physiol.* **176**: 6.
56. BEAN, J. W., P. JOHNSON & C. W. SMITH. 1953. *XIX Intern. Physiol. Congr. Abst.* : 945.
57. BEAN, J. W. & C. W. SMITH. 1952. *Am. J. Physiol.* **172**: 169.
58. TAYLOR, D. W. 1953. *XIX Intern. Physiol. Congr. Abst.* : 821.
59. BEAN, J. W. & P. C. JOHNSON. 1955. *Am. J. Physiol.* **180**: 438.
60. JOHNSON, P. C. & J. W. BEAN. 1957. *Am. J. Physiol.* **188**: 593.
61. BEAN, J. W. 1956. *Am. J. Physiol.* **187**: 389.
62. CHURCHILL-DAVIDSON, I., C. SANGER & R. H. THOMLINSON. 1957. *Brit. J. Radiol.* **30**: 405.
63. GIBBS, F. A., E. A. GIBBS & W. G. LENNOX. 1935. *Am. J. Physiol.* **111**: 557.
64. KETY, S. S. & C. F. SCHMIDT. 1948. *J. Clin. Invest.* **27**: 484.
65. BEAN, J. W. 1961. *Am. J. Physiol.* **201**: 1192.
66. BEAN, J. W. & L. SULLIVAN. 1962. *Physiologist* **5**: 104.
67. JAMIESON, D. & H. A. S. VAN DEN BRENK. 1963. *J. Appl. Physiol.* **18**: 869.
68. LENNOX, W. G. & E. L. GIBBS. 1932. *J. Clin. Invest.* **11**: 1155.