

Serological Studies on an α -D-Galactosyl-Binding Lectin Isolated from *Bandeiraea simplicifolia* Seeds

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Abstract. The serological characteristics of a highly purified α -D-galactosyl-binding lectin, isolated from extracts of *Bandeiraea simplicifolia* seeds, are described. These studies show that the lectin preferentially agglutinates group B red cells and in addition has the capacity to distinguish between group A₁ and group A₂ erythrocytes.

Since the lectin is not absolutely specific for group B red cells, it is therefore unsuitable as an anti-B blood typing reagent. However, the reactions obtained against Tn and 'acquired-B' red cells suggest that it may be of value in the elucidation and classification of red cell polyagglutinable states.

Introduction

Anti-B hemagglutinating activity in extracts from *Bandeiraea simplicifolia* seeds was first reported by MÄKELÄ and MÄKELÄ [5]. Recently, HAYES and GOLDSTEIN [3] purified an α -D-galactosyl-binding lectin from these seeds, and suggested that this reagent may be a suitable alternative to human group A serum as a source of anti-B for blood typing purposes.

This report details the serological behavior of the lectin, and describes a method for enhancing its *in vitro* activity. The reactions obtained against normal and anomalous ABO blood group phenotypes are presented, and the significance of these results is discussed.

Materials and Methods

An α -D-galactosyl-binding lectin was isolated by affinity chromatography from extracts of *Bandeiraea simplicifolia* seeds as described previously [3].

Unless otherwise stated, serological studies were performed as follows: 2 vol of lectin (3 mg of protein per ml in phosphate-buffered saline at pH 7.2) were mixed with 1 vol of a 5% suspension of red cells (washed three times in isotonic saline at pH 7.2) and immediately centrifuged at 3,400 rpm for 15 sec. Tests were examined macroscopically for agglutination.

Details of other serological techniques have been described elsewhere [4].

Reactions were graded and scored according to the method of RACE and SANGER [7].

Initial studies with the lectin revealed that avid agglutination reactions against group B red cells could only be achieved following the addition of human serum or bovine albumin to the reaction medium. These reactions were extremely difficult to read owing to the adhesion of agglutinates to the surface of glass test tubes. In view of this and the dependence of the lectin's agglutinating activity on the presence of calcium ions, the lectin was diluted in a solution containing 5 parts of a 22% bovine albumin, 4 parts of phosphate-buffered saline at pH 7.2, and 1 part of a 0.1 M calcium chloride. To facilitate the reading of reactions, 0.02 ml of Tween 20 was added to each 10 ml of diluent.

Results

The reactions obtained with red cells from selected ABO phenotypes at various dilutions of *Bandeiraea simplicifolia* lectin are shown in table I. These results clearly demonstrate the lectin's affinity for group B red cells, and its capacity to distinguish between group A₁ and group A₂ blood samples. A considerable variation in the reactivity of different samples of the same phenotype was found, as shown in table II. In addition, group B cord blood samples were shown to give weaker reactions than those obtained with group B red cells from adults.

Table III shows the results of tests using a 1:100 dilution of the lectin. All ten group O samples tested were nonreactive, and one of 24 group A red cell samples was weakly agglutinated. Of 18 group B bloods studied, all were reactive; however, weak agglutination was observed with four samples. Increase in the incubation time to 5 min enhanced the reactions with group B red cells, but agglutination was also obtained with many of the group A red cell samples.

Absorption and elution studies showed that the activity of the lectin with group A red cells cannot be separated from that against group B cells.

Serological reactions were enhanced following the enzyme treatment of red cells as shown in table IV. Studies with enzyme-treated red cells from

Table I. Reactions of selected ABO phenotypes with *Bandeiraea simplicifolia* lectin

ABO phenotype	Reciprocals of lectin dilutions							Score
	10	20	40	80	160	320	640	
A ₁	3+	3+	2+	1+	0	0	0	23
A ₂	1+	0	0	0	0	0	0	5
B	4+	4+	4+	4+	3+	1+	0	63
O	0	0	0	0	0	0	0	0

Table II. Titration of *B. simplicifolia* lectin against randomly selected red cell samples from various ABO phenotypes

ABO phenotype	Number tested	Titer		Score	
		mean	range	mean	range
A ₁	12	100	40-160	30	22-37
A ₂	3	20	0-40	14	0-20
B	16	580	320-640	53	50-58
A ₁ B	9	320	160-640	46	41-56
A ₂ B	5	256	160-320	50	48-51
B (cord)	4	120	80-160	29	21-37

Table III. Reactions of different examples of groups A, B and O red cell samples with a 1:100 dilution of lectin

ABO phenotype	Total number tested	Number of samples giving reactions of				
		4+	3+	2+	1+	0
O	10					10 (10)
A	24	(1)	(2)	(2)	1 (5)	23 (14)
B	18	5 (18)	9	4		

Figures in parentheses represent reactions after 5 min incubation.

Table IV. The effect of enzyme (ficin) treatment of red cells in enhancing the agglutinating activity of the lectin

ABO type	Untreated red cells		Ficin-treated cells	
	titer	score	titer	score
O	0	0	80	18
A ₁	80	26	5,000	99
A ₂	20	13	320	48
B	640	53	20,000	117

Table V. Reactions obtained with red cells from unusual blood group phenotypes following treatment with ficin

Red cell phenotype	Untreated red cells		Ficin-treated red cells	
	titer	score	titer	score
Oh (Bombay)	0		40	23
B ^{hm}	0		80	26
O, Sd(a+)	0		40	25
O, Sd(a++), strong reactor	0		80	33
O, Sd(a-)	0		40	25
O, p (Tj ^{a-})	0		40	23
O, P ₁	0		80	26
O, P ₂	0		80	33
O, Tn-poly-agglutinable	2,560	81	80	33
O, T-activated	40	23		
A ₁ (B) ('acquired B')	80	35		

Table VI. Reactions of 'acquired-B' [A₁(B)] red cells with *B. simplicifolia* lectin and human anti-B reagents

Reagent	Cell type	Titer	Score
<i>B. simplicifolia</i> lectin	A ₁ (B)	80	27
	normal group B	320	63
Human anti-B	A ₁ (B)	640	69
	normal group B	640	71

individuals of unusual ABO phenotype, selected phenotypes within the P and Sd^a blood group systems, and cases of red cell polyagglutination are presented in table V. In marked contrast to the reactions of untreated cells, the lectin was found to react with enzyme-treated group O red cells. The strong reactions observed with group O Tn-polyagglutinable red cells were decreased following treatment of the cells with ficin. The lectin weakly agglutinated group A₁ (B) ('acquired B') red cells. When tested with anti-B from group A individuals, these cells were found to react comparably to normal group B red cells as shown in table VI.

The hemagglutinating activity of the lectin against either group A₁, group B, ficin-treated group O or group O Tn red cells was readily abolished by saliva samples from secretors of B substance, and to a lesser degree by hydatid cyst fluids and group A secretor saliva samples. Group O secretor and nonsecretor salivas, and one group B nonsecretor saliva sample, were noninhibitory.

Discussion

A number of lectins and protectins with blood group A specificity have been described, and the use of these reagents is well established. The search for a suitable alternative to human group A serum as a source of anti-B has, however, yet to be fulfilled. A method for the large scale production of anti-B from the fungus *Fomes fomentarius* has been described by PARDOE *et al.* [6], but this reagent is somewhat unstable. The findings by MÄKELÄ and MÄKELÄ [5] of anti-B activity in saline extracts of *Bandeiraea simplicifolia* seeds led to the isolation and purification from these seeds of an α -D-galactosyl-binding lectin by HAYES and GOLDSTEIN [3]. These workers suggested that this lectin may prove suitable as an alternative to human sources for an anti-B blood typing reagent.

The results of the studies presented in this publication demonstrate that the α -D-galactosyl-binding lectin from the seeds of *Bandeiraea simplicifolia* has limitations with regard to its routine use as an anti-B typing reagent. The results of titration experiments show such a wide variation in the reactivity of different examples of the same ABO type that at no one dilution will reactions specific for group B cells be obtained, although at a 1:100 dilution level the lectin preferentially agglutinated group B red cells. Furthermore, the activity for group A cells cannot be separated by absorption and elution techniques from the activity against group B red cells. This latter

finding confirms the biochemical analysis of the lectin [3] which indicates that the purified protein is homogeneous.

The inhibitory effect of hydatid cyst fluids on the lectin's hemagglutinating activity is not surprising in view of the studies of WATKINS and MORGAN [8], which show that α -D-galactosyl residues are the immunodominant sugars involved with P blood group specificity. However, we were unable to demonstrate any such specificity by hemagglutination tests using untreated and protease-treated P₁, P₂ and p red cells. Presumably this is because the P blood group determinants on erythrocyte membranes are not readily accessible to the lectin's antigen-combining sites.

The strong reactivity of Tn-polyagglutinable red cells with the lectin is most likely due to the cryptic A-like receptors on these red cells, as shown by DAHR *et al.* [2], and the weak binding of *N*-acetyl-D-galactosamine to the lectin in addition to its affinity for D-galactose [3]. Since we were unable to demonstrate any increase in agglutination reactions with group O Sd^a + + (Cad 2) red cells, it is suggested that *Bandeiraea simplicifolia* lectin may provide a more readily available source of reagent than the *Salvia* lectins described by BIRD and WINGHAM [1] for the elucidation and classification of red cell polyagglutinable states. Some initial studies performed on crude saline extracts of *Bandeiraea simplicifolia* seeds suggest that they may be used for this purpose. Similarly, the results obtained when 'acquired B' red cells are tested with the purified lectin, in contrast to when human anti-B reagents are used, indicate the potential value of the lectin in the serological diagnosis of the acquired-B phenomenon.

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