Urinary Concentrations of Di(2-ethylhexyl) Phthalate Metabolites and Serum Reproductive Hormones: Pooled Analysis of Fertile and Infertile Men

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ABSTRACT: Urinary concentrations of metabolites of the anti-androgenic xenobiotic di-(2-ethylhexyl) phthalate (DEHP) were previously shown to be weakly associated with serum levels of several hormones in 2 disparate US populations: partners of pregnant women participating in the Study for Future Families and partners in infertile couples from Massachusetts General Hospital infertility clinic. The observed associations between phthalate metabolites and reproductive hormones were robust and insensitive to the characteristics of the subpopulation or the laboratory in which the hormones were measured, despite the fact that these 2 populations span a range of fertility, urinary phthalate metabolite concentrations, and reproductive hormone levels. We therefore examined associations between urinary metabolites of DEHP and reproductive hormones—follicle-stimulating hormone, luteinizing hormone, testosterone (T), inhibin B, and estradiol (E2)—and sex hormone–binding globulin (SHBG) in the pooled population. The magnitude of the associations seen were similar to those reported for each population separately, but effect estimates were more precise because of the increased sample size and the greater range of phthalate metabolite concentrations and hormone levels. Urinary concentrations of 3 metabolites of DEHP [mono(2-ethylhexyl) phthalate (MEHP), mono(2-ethyl-5-hydroxyhexyl) phthalate (MEHHP), and mono(2-ethyl-5-oxohexyl) phthalate (MEOHP)] were inversely associated with the free androgen index (FAI = T/SHBG) and calculated free testosterone. Urinary concentrations of MEHHP and MEOHP were positively associated with SHBG, and MEHP was inversely associated with E2. No other phthalate metabolites were associated with serum hormones, consistent with results in each population. Our results in this diverse population suggest that DEHP exposure is robustly associated with some male sex steroid hormones.

Key words: Anti-androgens, DEHP metabolites, endocrine disruptor, male hormones.

J Androl 2012;33:488–498

This study is supported by The Danish Agency for Science, Technology, and Innovation, grant 271070678 to N.J.; a University of Iowa Center for Health Effects of Environmental Contamination cooperative project grant to A.E.S.; the General Clinical Research Center at Harbor–UCLA Medical Center (MO1 RR00425); and NIEHS grant ES009718 to R.H.

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Received for publication February 25, 2011; accepted for publication May 11, 2011.

DOI: 10.2164/jandrol.111.013557

Recent studies have reported secular shifts in male reproductive hormone levels (Andersson et al, 2007; Travison et al, 2007) which might be associated with decreases in semen quality (Carlsten et al, 1992; Swan et al, 1997). While exposure data are limited, it has been hypothesized that these changes could, at least in part, reflect the widespread use, and human exposure to, environmental endocrine-disrupting compounds (EDC) (Sharpe and Skakkebæk, 2008; Jørgensen et al, 2010).

Phthalates, man-made chemicals extensively used in industry and commerce, are among the most widely
studied EDCs, and several, including di(2-ethylhexyl) phthalate (DEHP) and di-n-butyl phthalate (DBP) have been shown to have anti-androgenic activity (Agency for Toxic Substances and Disease Registry, 2002; Centers for Disease Control and Prevention [CDC], 2011). A growing body of literature has shown relationships between several of these phthalates and adverse reproduction and development (National Research Council, 1999; Hauser and Calafat, 2005; Talness et al, 2009; Thompson et al, 2009). Laboratory studies have shown that DEHP and/or its metabolites are associated with the induction of testicular toxicity in neonatal, pubertal, and adult rodents (Parmar et al, 1986; Heindel et al, 1989; Srivastava et al, 1990; Li et al, 1998, 2000). However, adult animals are usually less sensitive than young pubertal animals or animals exposed in utero (Dostal et al, 1988; Higuchi et al, 2003). For example, several toxicological studies have demonstrated that DEHP, DBP, benzylbutyl phthalate, and di-isononyl phthalate disrupt reproductive tract development (eg, hypospadias, reduced fetal testosterone synthesis) in male rodents due to anti-androgenic action (Gray et al, 2000; Parks et al, 2000).

Nevertheless, only a small number of human studies have investigated the relationship between male reproductive hormones and phthalate exposure. In those studies, relationships have been shown between human prenatal and perinatal exposure to some phthalate metabolites and alterations in reproductive hormones (sex hormone-binding globulin [SHBG], luteinizing hormone [LH], and free testosterone [FT]) (Main et al, 2006) and markers of male reproductive development (Swan et al, 2005; Swan, 2008). In a population of young men, Jönsson et al (2005) reported an inverse association between urinary monoethyl phthalate (MEP) concentrations and circulating LH, although no associations were found between other phthalate metabolites and reproductive hormones. Pan et al (2006) studied adult men occupationally exposed to some phthalates (DEHP and DBP) and reported that phthalate exposure was inversely associated with serum FT levels.

Meeker and collaborators (2009) investigated this issue and extended their previous work (Duty et al, 2005) by including a larger sample size and expanding the number of hormones and phthalate metabolites measured. In a male population attending a fertility clinic, the authors reported an association between increased urinary concentration of mono(2-ethylhexyl) phthalate (MEHP) with decreased testosterone (T), estradiol (E2), and free androgen index (FAI) levels, showing that exposure to DEHP might be associated with altered steroid hormones in these men. Recently, Mendiola et al (2011) investigated these associations in a population of fertile men. Both Meeker et al (2009) and Mendiola et al (2011) showed significant inverse association between FAI levels and urinary concentrations of several DEHP metabolites. In both studies, SHBG was positively associated with urinary concentrations of MEHP, but not with other DEHP metabolites. Neither study found notable associations between metabolites of any other phthalate and hormone under investigation. There were, however, some discrepancies between these studies. For instance, Duty et al (2005) reported a dose-response relationship between mono-benzyl phthalate (MBzP) and follicle-stimulating hormone (FSH) and between mono-n-butyl phthalate (MBP) and inhibin B, but no strong evidence of an association between MEHP and T. Meeker et al (2009) reported a significant relationship between MEHP and T, and mono(2-ethyl-5-hydroxyhexyl) phthalate (MEHHP) and mono(2-ethyl-5-oxohexyl) phthalate (MEOHP) and FAI (P < .05), but for FAI and MEHP, the adjusted P value was <.1. Mendiola et al (2011) reported a significant association between several DEHP metabolites and FAI, but no relationship between DEHP metabolites and T.

The aim of the current study was to use a pooled analysis of a large heterogeneous population of both fertile (Mendiola et al, 2011) and infertile men (Meeker et al, 2009) to more precisely examine the relationships of urinary phthalate metabolite concentrations with serum reproductive hormone levels. Although data from both populations were previously published, this new pooled analysis adds to our understanding of the human health effects of phthalates by allowing us to investigate systematically whether associations differ by populations on the basis of fertility status.

Materials and Methods

Study Populations

The present study includes men from 2 large ongoing studies of environmental influences on reproductive health. One of these, the Study for Future Families (SFF; n = 425), is a multicenter study of pregnant women and their male partners conducted at prenatal clinics affiliated with university hospitals in 5 US cities (Harbor-UCLA and Cedars-Sinai Medical Center in Los Angeles, California; University of Minnesota Medical Center in Minneapolis; University Physicians in Columbia, Missouri; Mt Sinai School of Medicine, New York City, and University of Iowa, Iowa City) between 1999 and 2005. In this study, couples were eligible only if the pregnancy was conceived without assisted reproduction (Swan et al, 2003). The second study included men who were male partners of infertile couples seeking evaluation at the Vincent Memorial Obstetrics and Gynecology Service, Andrology Laboratory and In Vitro Fertilization Unit, Massachusetts General Hospital (MGH) (n = 425) in Boston between January 2000 and May 2004.
Methods for serum hormone analyses at the 2 laboratories (MGH and SFF)

<table>
<thead>
<tr>
<th>Hormone</th>
<th>Method</th>
<th>Manufacturer/System</th>
<th>Sensitivity</th>
<th>CVs, %</th>
</tr>
</thead>
<tbody>
<tr>
<td>MGH assay details</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FSH</td>
<td>MEIA</td>
<td>Abbott AxSYM</td>
<td>1.1 IU/L</td>
<td>3–7</td>
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<tr>
<td>LH</td>
<td>MEIA</td>
<td>Abbott AxSYM</td>
<td>1.2 IU/L</td>
<td>4–7</td>
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<td>Testosterone</td>
<td>RIA</td>
<td>Coat-A-Count kit, Diagnostic Products Corp</td>
<td>0.14 nmol/L</td>
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<td>SHBG</td>
<td>Solid-phase two-site enzyme chemiluminescent immunometric assay</td>
<td>Immulite, Diagnostic Products Corp</td>
<td>1 nmol/L</td>
<td>2–5</td>
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<tr>
<td>Inhibin B</td>
<td>Double-antibody ELISA</td>
<td>Oxford Bioinnovation</td>
<td>50 pg/mL</td>
<td>8</td>
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<tr>
<td>Estradiol</td>
<td>MEIA</td>
<td>Abbott AxSYM</td>
<td>73 pmol/L</td>
<td>3–11</td>
</tr>
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<td>SFF assay details</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>FSH</td>
<td>TR-IFMA</td>
<td>DELFIA, Perkin Elmer</td>
<td>0.05 IU/L</td>
<td>1.3–2.1</td>
</tr>
<tr>
<td>LH</td>
<td>TR-IFMA</td>
<td>DELFIA, Perkin Elmer</td>
<td>0.05 IU/L</td>
<td>1.5–3.0</td>
</tr>
<tr>
<td>Testosterone</td>
<td>TR-FIA</td>
<td>DELFIA, Perkin Elmer</td>
<td>0.23 nmol/L</td>
<td>1.4–2</td>
</tr>
<tr>
<td>SHBG</td>
<td>TR-IFMA</td>
<td>DELFIA, Perkin Elmer</td>
<td>0.23 nmol/L</td>
<td>3–5</td>
</tr>
<tr>
<td>Inhibin B</td>
<td>Specific two-sided enzyme immunometric assay</td>
<td>Oxford Bioinnovation, in-house standard</td>
<td>20 pg/mL</td>
<td>15</td>
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<tr>
<td>Estradiol</td>
<td>RIA</td>
<td>Pantex</td>
<td>18 pmol/L</td>
<td>3–8</td>
</tr>
</tbody>
</table>

Abbreviations: CVs, coefficients of variation; ELISA, enzyme-linked immunosorbent assay; FSH, follicle-stimulating hormone; LH, luteinizing hormone; MEIA, microparticle enzyme immunoassay; MGH, Massachusetts General Hospital; RIA, radioimmunoassay; SFF, Study for Future Families; SHBG, sex hormone–binding globulin; TR-IFMA, time-resolved immunofluorometric assay; TR-FIA, time-resolved fluoroimmunoassay.

(Meeker et al, 2009). That infertility clinic population includes men with male factor infertility as well as men who are partners of women with female factor infertility. Methods for clinical examination, data collection, and semen analysis have been described previously for each study (Swan et al, 2003; Meeker et al, 2009). Briefly, in both studies, the men completed a questionnaire and gave urine, blood, and semen specimens. Information was collected on demographics, medical history, and lifestyle factors. Human subject approvals were obtained from institutional review boards at all participating institutions. The involvement of a CDC laboratory in SFF was limited and determined not to constitute engagement in human subjects research.

Serum Hormone Analysis

In both populations, venous blood samples were drawn, and the serum was separated and frozen at −80°C on the same day the urinary sample was collected. Samples were analyzed for hormones in 2 different laboratories: SFF samples at the Rigshospitalet Andrology Laboratory (Copenhagen, Denmark) and MGH samples at the REU Laboratory at MGH (Boston). Each methodology has been described previously elsewhere (Bang et al, 2005; Asklund et al, 2007; Meeker et al, 2009; Mendiola et al, 2011). The MGH lab is Clinical Laboratory Improvement Amendments–certified (Centers for Medicare and Medicaid Services, Department of Health and Human Services, Baltimore, Maryland), and the Rigshospitalet Andrology Laboratory participates in Bio-Rad Laboratories external quality immunoassay program (Bio-Rad Laboratories, Copenhagen, Denmark). Table 1 summarizes the serum hormone analysis methods that were employed at the 2 laboratories. FAI was calculated as total testosterone (×100/SHBG), and FT concentration was calculated using the equation of Vermeulen et al (1999).

Urinary Phthalate Metabolites Measures

In both populations, the concentrations of urinary phthalate metabolites were determined at the Division of Laboratory Sciences, National Center for Environmental Health (CDC, Atlanta, Georgia), which had no access to participant data. SFF samples were analyzed in 2006, and MGH samples were analyzed throughout a 3-year period (2003–2006). Urinary samples were frozen and stored at −80°C and then shipped to CDC on dry ice. Phthalate metabolites were measured in urine to avoid potential sample contamination from the parent diester and because the metabolites (not the parent diesters) are the active toxicants (Li et al, 1998). The analytical approach for the analysis of urinary phthalate metabolites in the MGH population of men has been previously described (Silva et al, 2007; Meeker et al, 2009). A modification of that approach was used in the SFF population and has been described and published elsewhere (Swan et al, 2005). Limits of detection (LOD) are in the low nanogram per milliliter range. Isotopically labeled internal standards were used along with conjugated internal standards to increase precision and accuracy of the measurements. The method is accurate (spiked recoveries are near 100%) and precise, with between-day relative standard deviations of <10%. Quality control samples and laboratory blanks were analyzed along with unknown samples to monitor performance of the method (Swan et al, 2005). Concentrations are reported in nanograms per milliliter (ng/mL). Although different metabolites were assessed in our separate studies, we report here only the 6 urinary phthalate metabolites that were measured in both populations: MEHP,
MEHHP, MEOHP, MEP, MBzP, and MBP (as the sum of MBP and mono-iso-butyl phthalate concentrations). We also calculated the percentage of these DEHP metabolites excreted as MEHP (MEHP%). To calculate MEHP%, we converted MEHP, MEHHP, and MEOHP concentrations to nanomoles per milliliter; divided MEHP concentrations by the sum of concentrations of MEOHP, MEHHP, and MEHP; and multiplied by 100 (Hauser et al, 2006).

### Statistical Analyses

Data from Meeker et al (2009) and Mendiola et al (2011) were pooled for statistical analysis. Serum hormones (except E₂) and urinary phthalate metabolite concentrations were log transformed to normalize their asymmetric distributions. In preliminary analyses, we used the Mann-Whitney U test and Pearson correlation coefficients to explore the relationship between each hormone concentration and each phthalate metabolite concentration. We then used multiple linear regression analysis to control for appropriate covariates, including age, age square, body mass index (BMI), smoking status (current smoker vs never smoked), ethnicity (African American vs others), time of sample collection (hours after 7:00 AM), and time of sample collection squared. Urinary dilution was measured differently in the 2 populations: SFF models were adjusted by urinary creatinine concentrations and MGH models by specific gravity (SG). Although these methods of adjusting for urinary concentration are different, the rank of urinary concentrations assigned by each method should be comparable (Box and Tidwell, 1962). Therefore, the measure of urinary dilution used in the combined analysis was the rank of creatinine or SG in the respective data sets. We also included a term for study center (SFF vs MGH) that reflects between-center differences, including those due to differing methods of hormone analysis and measurement for urinary dilution. Age, BMI, and time of collection were modeled as continuous variables and all others as dichotomous indicator variables. Most metabolite concentrations were above the LOD; those below the LOD were assigned the value LOD divided by the square root of 2, which has been recommended when the data are not highly skewed (ie, geometric standard deviation < 3; Hornung and Reed, 1990), as was the case in the present analysis. Two analysts (J.D.M. and J.M.) conducted all analyses independently using SAS version 9.1 (SAS Institute Inc, Cary, North Carolina) and SPSS version 18.0 (SPSS Inc, Chicago, Illinois).

### Results

Serendipitously, 425 men in each population provided urine and blood. E₂ and inhibin B serum levels were available for 830 and 849 males, respectively, and 783 had complete information on all covariates and were included in the final multivariate analyses. MEHHP and MEOHP urinary concentrations were measured in 646 men, as these metabolites were not incorporated into the MGH study until after the study had already begun. Basic demographic data are presented in Table 2, including information about reproductive parameters in the separate and joint populations; Figure 1a through g present the frequency distribution of the reproductive hormones measured in the 2 populations. Summary statistics for the serum concentrations of men’s reproductive hormones are presented in Table 3. All hormone levels differed significantly between the 2 populations. Both FSH and LH were about 3-fold higher in MGH men compared with SFF men, and inhibin B levels were lower in MGH men.

Urinary concentrations (ng/mL) of DEHP metabolites (before urine dilution adjustment) are shown in Table 4, together with the LOD and percentage of samples above the LOD. Urinary concentrations of DEHP metabolites were notably higher in MGH men than in SFF men, whereas MEP, MBP, and MBzP were higher in SFF men. MEHP% was similar in the 2 populations.

### Table 2. Characteristics of the SFF and MGH study populations

<table>
<thead>
<tr>
<th></th>
<th>SFF (n = 425)</th>
<th>MGH (n = 425)</th>
<th>Total (n = 850)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>32.2 (6.2)</td>
<td>36 (5.3)</td>
<td>34.3 (6.1)</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>28.2 (5.4)</td>
<td>28 (4.5)</td>
<td>28.1 (4.9)</td>
</tr>
<tr>
<td>Current smoker</td>
<td>21</td>
<td>9</td>
<td>15</td>
</tr>
<tr>
<td>White, non-Hispanic</td>
<td>72.3</td>
<td>85</td>
<td>79</td>
</tr>
<tr>
<td>Sperm concentration &lt; 20 × 10⁶/mL</td>
<td>7.8</td>
<td>15.3</td>
<td>12</td>
</tr>
<tr>
<td>Sperm motility (A + B) &lt;50%</td>
<td>37.4</td>
<td>45.9</td>
<td>42</td>
</tr>
<tr>
<td>Made a partner pregnant</td>
<td>100</td>
<td>41.6</td>
<td>71</td>
</tr>
<tr>
<td>Had trouble fathering a child</td>
<td>4.3</td>
<td>100</td>
<td>52</td>
</tr>
</tbody>
</table>

Abbreviations: BMI, body mass index; MGH, Massachusetts General Hospital; SFF, Study for Future Families.

⁴ In SFF, all men were partners of pregnant women. In MGH, this is the percentage of men who self-reported that they had “ever made a partner pregnant.”

⁵ In MGH, all men were part of a couple seeking evaluation or treatment for infertility. In SFF, this is the percentage of men who responded positively to the question: “Have you ever seen a doctor because you thought you might be having trouble fathering a child?”
Figure 1. Distribution (density) of the reproductive hormone profiles for Study for Future Families (dashed line) and Massachusetts General Hospital (continuous line). All data have been truncated to fall between the 5th and 95th percentiles. (a) Follicle-stimulating hormone (FSH). (b) Inhibin B (INHIBINB). (c) Luteinizing hormone (LH). (d) Serum estradiol (E2). (e) Free androgen index (FAI). (f) Free testosterone (FT). (g) Testosterone (T).
Table 5 shows correlation coefficients for reproductive hormones and unadjusted urinary DEHP metabolite concentrations from initial bivariate analyses. We observed no associations between any hormone levels and any urinary metabolites of phthalates other than DEHP (data available on request). Therefore, here we report only the associations involving the 3 measured metabolites of DEHP (MEHP, MEHHP, and MEOHP).

Table 6 shows the results of the multivariate analysis for reproductive hormones and urinary DEHP metabolite concentrations in both populations separately and combined. After adjustment for covariates, many of the relationships (as described by the $b$ coefficients) were consistent with previously published results (Meeker et al, 2009; Mendiola et al, 2011), although the effect estimate for E$_2$ strengthened in the pooled analysis. Overall, an increase in statistical power due to increased sample size resulted in increased precision in the effect estimates compared with the individual studies. There were no significant associations between T and any urinary DEHP metabolites. FAI and FT were both inversely associated with the urinary concentrations of all 3 urinary DEHP metabolites measured in the study (MEHP, MEHHP, and MEOHP). Serum gonadotropin levels (FSH and LH) were not associated with DEHP metabolite concentrations in the separate or combined populations. There was a significant inverse association between E$_2$ levels and urinary MEHP concentrations, but not with the other DEHP metabolites. The T/E$_2$ ratio was positively associated with urinary MEHP metabolite concentrations. SHBG levels were positively related to urinary MEHHP and MEOHP concentrations, but not MEHP concentration. Figure 2 shows the percent change in men’s reproductive hormones expected with an interquartile increase in urinary DEHP metabolite concentrations for a 34-year-old nonsmoker with a BMI of 28 kg/m$^2$. For this typical subject, an increase in
urinary concentrations of MEHP and the oxidative metabolites (MEHHP and MEOHP) from the 25th to the 75th percentile would be predicted to decrease steroid hormone levels, the amounts ranging around 3.5% and 7%, for T and E₂, respectively.

Discussion

This is the first study to examine the associations between urinary concentrations of phthalate metabolites and reproductive hormone serum levels in a large cohort including both fertile men and male partners of infertile couples. Our results suggest that exposure to DEHP at environmental concentrations is associated with statistically significant declines in free testosterone (both FAI and FT) and serum estradiol (E₂). The other phthalate monoester metabolites we examined (MEP, MBP, and MBzP) were not associated with any reproductive hormones. These associations are not substantially different from those reported in separate analyses, which in turn do not differ appreciably between the 2 populations (Meeker et al, 2009; Mendola et al, 2011). However, each of the individual studies provides information only about a limited subset of the total population. When the 2 populations are combined, the effect estimates are more precise and more generalizable to men of reproductive age.

In this combined population of fertile and subfertile men, we saw no significant associations with total T levels and any phthalate metabolites. However, both FT and FAI were both inversely associated with urinary DEHP metabolite concentrations. This may be accounted for by a positive association between serum SHBG levels and urinary MEHP concentrations in the SFF cohort and with MEHHP and MEOHP in the combined analysis. Significant positive associations were seen between SHBG and MEHHP and MEOHP in the combined analysis. However, associations between SHBG and MEHP differed in these 2 cohorts, with a significant positive association in SFF men but a nonsignificant negative association in the MGH cohort.
This resulted in a nonsignificant positive association between SHBG and MEHP in the combined analyses. It should be noted that the serum SHBG concentration in all the subjects are within the physiological range of adult men. Thus, the small increases in serum SHBG levels associated with greater DEHP may result in a small reduction of FT without affecting the total serum T levels.

We did not see an association between DEHP metabolite concentrations and LH in this combined population of fertile and infertile men. In this mixed population the small changes in FT and FAI associated with DEHP may not be sufficient to elicit the negative feedback that would be expected to produce a positive association between LH and DEHP metabolites.

Although all men had serum steroid hormones within the laboratory reference ranges, our findings suggest a somewhat anti-androgenic effect of DEHP. This is consistent with data showing that phthalates may inhibit expression of genes involved in steroidogenesis (cholesterol transport and the biosynthesis of testosterone) in rat fetal testis after in-utero exposure to large doses of DEHP (Borch et al., 2006).

E$_2$ plays a role in male germ cell survival in vitro (Pentikainen et al., 2000). In our study, urinary MEHP concentrations were inversely associated with serum E$_2$ levels and positively associated with T/E$_2$ ratio. In vitro and animal studies have shown that aromatase activity and E$_2$ production can be lowered by DEHP, MEHP, or both (Davis et al., 1994; Lovekamp and Davis, 2001; Andrade et al., 2006; Noda et al., 2007). Our results suggest that, as in rodent models, DEHP may be associated with a reduced aromatase activity.

We compared unadjusted urinary concentrations of DEHP metabolites in our subjects to those from male participants in the 2007-2008 National Health and Nutrition Examination Survey (NHANES; CDC, 2011). Median MEHP concentration was almost twice as high in our combined population (4.4 ng/mL compared with 2.3 ng/mL), whereas the other DEHP metabolites were
similar (eg, medians 20.9 and 23.2 ng/mL for MEHHP in NHANES and our population).

Our data were limited by the use of a single urine and blood sample to assess DEHP exposure and hormone function, respectively. However, several studies have reported that although phthalate metabolite concentrations are variable within an individual over time, the average concentration over the course of days, weeks, or months can be satisfactorily predicted by a single sample (Hoppin et al, 2002; Hauser et al, 2004; Teitelbaum et al, 2008). Similarly, a single sample can be used to classify reproductive hormone levels in men (Björnnerem et al, 2006).

It is generally accepted that hormone levels obtained in different laboratories, with different methods, or both are likely to differ. The variations among laboratories are more marked for steroid hormone levels at low levels (eg, T and E₂ levels in men) than for gonadotropins (Taieb et al, 2003; Wang et al, 2004; Sikaris et al, 2005; Rosner et al, 2007; Pitteloud et al, 2008). We included a center effect in our multivariate models to reflect between-laboratory differences. Adding this covariate did not alter associations between urinary DEHP metabolites and androgens (T, FT, and FAI). However, it did slightly increase effect estimates for E₂ and SHBG. One limitation of all previously published studies on phthalate metabolites and reproductive parameters is that their study populations (fertile men or men in infertility clinics) are not representative of the general population. Our combined analysis includes a wider range of men, although still not a representative sample of adult men.

Our results in this population, including both fertile and infertile men, suggest that DEHP exposure is associated with some changes in circulating levels of male sex steroid hormones, consistent with the known anti-androgenic effect of this chemical.
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Bang AK, Carlsen E, Holm M, Petersen JH, Skakkebaek NE, Asklund C, Jørgensen N, Skakkebaek NE, Jensen TK. Increased urinary concentrations of the phthalate metabolites.


Figure 2. Percent change in men’s reproductive hormones expected with an increase from the 25th to the 75th percentile in di-(2-ethylhexyl) phthalate metabolite concentrations for a standard subject (34 years old, nonsmoker with body mass index of 28 kg/m²). Error bars indicate 95% confidence intervals; MEHP, mono(2-ethylhexyl) phthalate; MEHHP, mono(2-ethyl-5-hydroxyhexyl)phthalate; MEOHP, mono(2-ethyl-5-oxohexyl) phthalate; SHBG, sex hormone-binding globulin; T, testosterone; FAI, free androgen index; FT, free testosterone; E₂, estradiol.

Acknowledgments

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the CDC. The authors gratefully acknowledge Manori J. Silva, Ella Samandar, James Preau, and Jack Reidy (CDC, Atlanta, Georgia) for measuring the urinary concentrations of the phthalate metabolites.

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